

**Table S1** MICs of 39 fosfomycin-resistance *E. coli* isolates

Strain	MIC (mg/liter)							
	FOF <sup>a</sup>	FFC	GEN	AMK	CIP	TET	CTX	CST
SY301B	>512	128	128	2	16	64	>256	0.5
SY301F	>512	128	128	>256	8	128	256	0.5
SY303P	>512	256	64	>256	8	128	64	0.25
SY301L	>512	256	>256	>256	32	128	64	0.5
SY303M	>512	128	256	>256	16	128	128	0.5
SY305M	>512	128	128	>256	16	256	128	0.5
SY286M	>512	>512	128	>256	0.125	128	128	0.5
SY287M	>512	256	128	>256	0.25	128	256	0.5
SY284B	>512	256	64	>256	0.125	128	128	0.5
CH293B	>512	512	256	>256	128	64	>256	0.5
CH292B	>512	256	256	>256	128	128	>256	0.5
CH291M	>512	128	>256	>256	16	256	>256	0.5
CH281F	>512	128	128	256	32	128	>256	1
CH282M	>512	128	64	256	64	128	128	0.5
CH285F	>512	>512	128	>256	64	>256	128	0.5
DH286F	>512	256	64	>256	16	>256	128	0.5
DH286M	>512	256	64	>256	32	128	256	1
JL12G	>512	256	>256	>256	32	64	256	1
JL15P	>512	128	>256	>256	0.5	128	256	0.5
JL17P	>512	4	128	>256	0.125	128	32	0.5
JT14G	>512	128	256	256	0.125	32	128	0.5
JT14P	>512	256	256	128	0.25	32	128	1
HL12L	>512	256	256	4	0.125	16	128	0.5
HL36B	>512	128	128	4	0.125	128	256	0.5
HL40C	>512	128	128	2	0.125	128	>256	0.5
HB37B	>512	>512	256	256	64	256	64	0.5
HB35D	>512	512	256	256	0.25	256	128	0.5
HB38B	>512	4	64	128	8	16	128	0.5
HB38L	>512	256	128	>256	32	64	128	0.5
HB18F	>512	256	128	>256	64	64	64	1
HB13B	>512	128	>256	>256	64	256	>256	0.5
HB13M	>512	256	>256	>256	32	32	128	0.5
HB312G	>512	256	256	>256	16	64	128	1
SH312M	>512	512	64	256	16	64	>256	0.5
SH312N	>512	512	64	256	8	128	>256	0.5
SH21F	>512	128	32	2	8	256	>256	0.5
SH21G	>512	128	128	2	32	256	256	0.5
SH21M	>512	128	128	8	16	256	256	0.5
SH33L	>512	128	64	1	32	32	128	0.25

<sup>a</sup> FOF, fosfomycin; FFC, florfenicol; GEN, gentamicin; AMK, amikacin; CIP, ciprofloxacin; TET, tetracycline; CTX, cefotaxime; CST, colistin;

**Table S2** Characteristics of the *fosA3*-positive *E. coli* isolates.

Isolate <sup>a</sup>	Source	Farm	Isolation date	PFGE pattern	MLST	Phylogroup	Resistance profile <sup>b</sup>
SY301B	pig feces	Fa1	06/2015	1	ST761	B2	<u>FFC, CTX, TET, GEN, CIP</u>
SY301F	pig liver	Fa2	06/2015	2	ST3315	B2	<u>FFC, CTX, TET, GEN, AMK, CIP</u>
SY303P	pig liver	Fa3	06/2015	3	ST5714	B2	<u>FFC, CTX, TET, GEN, AMK, CIP</u>
SY301L	pig kidney	Fa4	06/2015	4	ST5442	B2	<u>FFC, CTX, TET, GEN, AMK, CIP</u>
SY303M	pig liver	Fa5	06/2015	5	ST3944	B2	<u>FFC, CTX, TET, GEN, AMK, CIP</u>
SY305M	pig feces	Fa5	06/2015	5	ST3944	B2	<u>FFC, CTX, TET, GEN, AMK, CIP</u>
SY286M	cow milk	Fa6	06/2015	6	ST93	B2	<u>FFC, CTX, TET, GEN, AMK</u>
SY287M	cow milk	Fa6	06/2015	6	ST98	B2	<u>FFC, CTX, TET, GEN, AMK</u>
SY284B	cow milk	Fa7	06/2015	7	ST4683	B1	<u>FFC, CTX, TET, GEN, AMK</u>
CH293B	chicken faeces	Fa8	08/2015	8	ST10	B2	<u>FFC, CTX, TET, GEN, AMK, CIP</u>
CH292B	chicken faeces	Fa8	08/2015	8	ST10	B2	<u>FFC, CTX, TET, GEN, AMK, CIP</u>
CH291M	chicken faeces	Fa9	08/2015	9	ST410	B2	<u>FFC, CTX, TET, GEN, AMK, CIP</u>
CH281F	chicken faeces	Fa10	08/2015	10	ST5889	A	<u>FFC, CTX, TET, GEN, AMK, CIP</u>
CH282M	chicken faeces	Fa11	08/2015	11	ST1437	A	<u>FFC, CTX, TET, GEN, AMK, CIP</u>
CH285F	chicken faeces	Fa12	08/2015	12	ST448	B2	<u>FFC, CTX, TET, GEN, AMK, CIP</u>
DH286F	chicken faeces	Fa13	08/2015	13	ST2518	B2	<u>FFC, CTX, TET, GEN, AMK, CIP</u>
DH286M	chicken faeces	Fa14	08/2015	14	ST617	B2	<u>FFC, CTX, TET, GEN, AMK, CIP</u>
JL12G	cow milk	Fa15	08/2015	15	ST559	B2	<u>FFC, CTX, TET, GEN, AMK, CIP</u>
JL15P	cow milk	Fa16	08/2015	16	ST209	B2	<u>FFC, CTX, TET, GEN, AMK</u>
<u>JL17P</u>	cow milk	Fa16	08/2015	17	ST359	A	<u>CTX, TET, GEN, AMK</u>
JT14G	cow milk	Fa17	08/2015	18	ST5689	B1	<u>FFC, CTX, TET, GEN, AMK</u>
JT14P	cow milk	Fa18	08/2015	19	ST5714	B1	<u>FFC, CTX, TET, GEN, AMK</u>
<u>HL12L</u>	cow milk	Fa19	11/2015	20	ST195	D	<u>FFC, CTX, TET, GEN</u>
HL36B	cow milk	Fa20	11/2015	21a	ST4680	B1	<u>FFC, CTX, TET, GEN</u>
HL40C	farmer sputum	Fa20	11/2015	21b	ST4680	B1	<u>FFC, CTX, TET, GEN</u>
HB37B	chicken faeces	Fa21	02/2016	22	ST1725	B1	<u>FFC, CTX, TET, GEN, AMK, CIP</u>
HB35D	chicken faeces	Fa21	02/2016	23	ST48	B2	<u>FFC, CTX, TET, GEN, AMK</u>
HB38B	cow milk	Fa22	02/2016	24	ST2055	B1	<u>CTX, TET, GEN, AMK, CIP</u>
HB38L	cow milk	Fa23	02/2016	25	ST5693	B1	<u>FFC, CTX, TET, GEN, AMK, CIP</u>
HB18F	cow milk	Fa24	02/2016	26	ST1081	B1	<u>FFC, CTX, TET, GEN, AMK, CIP</u>
HB13B	chicken faeces	Fa25	02/2016	27	ST167	B2	<u>FFC, CTX, TET, GEN, AMK, CIP</u>
HB13M	cow milk	Fa26	02/2016	28	ST4463	B2	<u>FFC, CTX, TET, GEN, AMK, CIP</u>
HB312G	cow milk	Fa27	02/2016	29	ST685	B2	<u>FFC, CTX, TET, GEN, AMK, CIP</u>
SH312M	pig faeces	Fa28	04/2016	30	ST1488	B2	<u>FFC, CTX, TET, GEN, AMK, CIP</u>
SH312N	pig faeces	Fa29	04/2016	30	ST1313	B2	<u>FFC, CTX, TET, GEN, AMK, CIP</u>
SH21F	pig faeces	Fa30	04/2016	31	ST209	B2	<u>FFC, CTX, TET, GEN, CIP</u>
<u>SH21G</u>	pig faeces	Fa31	04/2016	32	ST354	D	<u>FFC, CTX, TET, GEN, CIP</u>
SH21M	pig faeces	Fa32	04/2016	33	ST648	D	<u>FFC, CTX, TET, GEN, CIP</u>
SH33L	cow milk	Fa33	04/2016	29	ST3743	B1	<u>FFC, CTX, TET, GEN, CIP</u>

<sup>a</sup>The first two capital letters in each strain name represent different cities. SY, Shenyang; CH, Changchun; DL, Dehui; JL, Jilin; JT, Jiutai; HL, Hailaer; HB, Harbin; SH, Suihua; Shenyang belongs to Liaoning province; Changchun, Dehui, Jilin and Jiutai belong to Jilin Province; Hailaer belongs to Inner Mongolia autonomous region; Harbin and Suihua belong to Heilongjiang province. Isolates from healthy animals are underlined.

<sup>b</sup>AMK, amikacin; CTX, cefotaxime; CIP, ciprofloxacin; FFC, florfenicol; GEN, gentamicin; TET, tetracycline; the MIC value of florfenicol ( $\geq 32\text{mg/L}$ ) was interpreted according to European Committee on Antimicrobial Susceptibility Testing (EUCAST). Transferable resistance phenotypes are underlined.

**Table S3** Primers were used to investigate the four epidemic plasmids and the genetic environment of the *fosA3* gene in other plasmids

Plasmid		Primer <sup>a</sup>	Nucleotide sequence (5' to 3')	Product size (bp)	Annealing temperature (°C)
<b>pECF12</b>	PCR 1	repA_fw	GTGCAGGATGGTGTGACTGA	4179	52
		tnpA_rv	AGAAAGTCGCCATGCCGTTG		
	PCR 2	tnpA_fw	CTGCCAGCGTGAAGGAATAA	2518	51
		fosA3_rv	GACTATACCCACTACGCCTT		
	PCR 3	fosA3_fw	TCCAGCTTATGGCCGTAGG	3790	49
		tetA_rv	TGATCACCGGCCCTGTAGCC		
	PCR 4	tetA_fw	CAAGCAGGATGTAGCCTGTG	3450	53
		strA_rv	GTGTCCCGCAATGCCGTCAA		
	PCR 5	strA_fw	GCCCAGTTCTTCGGCGTT	4947	54
		ΔTn3_rv	CATCGACCGGCCGCTACTACG		
	PCR 6	ΔTn3_fw	TCCGCCATCTTGGTCAGGCC	4014	53
		IS26_rv	GGCACTGTTGCAAATAGTCG		
	PCR 7	IS26_fw	TTCTTCACGTTGTTGAGGAT	3889	50
		stbA_rv	CGAACAGTATTCTTTGATC		
	PCR 8	stbA_fw	ATCTATAATTAATAAAGAAT	4371	50
		ycgC_rv	TCCAGCATCACCAGCCGGAAC		
	PCR 9	ycgC_fw	ACGATGAGCTGGAAACCGTC	4660	55
		psiB_rv	CAGTGACGCCGCCAGTGCCA		
	PCR 10	psiB_fw	TGGCAGCGAGTTGGCGGGCT	4707	55
		ydcA_rv	TCTCCTGCTGGCGTGGCCCG		
	PCR 11	ydcA_fw	CCATGTGTGATGCGGTGGAG	4211	52

		traJ_rv	TTGCATGACCAACACCCAGC		
PCR 12		traJ_fw	AGAGAGTTATATACGTCTGC	3489	50
		TraB_rv	AATGTACGGCAGTGATGGTC		
PCR 13		TraB_fw	CGTCGCGAGCTGGATGTCCT	2610	55
		traV_rv	CGCTATCCATAATGCAGCCG		
PCR 14		traV_fw	TCACTGCCGCGCCTGGCTGA	4814	55
		traU_rv	CAGGCTGAAGCCCAGGTTCA		
PCR 15		traU_fw	GCAGTATCAAAGTCAGTCAG	4309	50
		traN_rv	TTTATCCCGGTATGCCAGC		
PCR 16		traN_fw	CTGCCAGGATCACCGGAGAC	3933	53
		trbF_rv	CTCCCGGCGCATACGGCGTT		
PCR 17		trbF_fw	ACGGAATATTGAGTATAGTT	3914	50
		traG_rv	GGCTGCGGGTCATATTGTC		
PCR 18		traG_fw	CGCATAATATCAGTAATGAG	3711	50
		traD_rv	GAGACAACAAAGAAGGTAAT		
PCR 19		traD_fw	ATATCATGCTTACTGCCTG	4093	52
		tral_rv	CACCATTCCGGGCGGTACA		
PCR 20		tral_fw	GTGCGCTGATGCCATGAAG	4404	55
		traX_rv	TCACAGCGCCAGAACGCCCA		
<b>pECB11</b>	PCR 21	repA_fw	GTGACTGATCTCAACAA	3950	49
		tnp_rv	CTTGCCACGGACATATC		
	PCR 22	tir_fw	TTTGCCGATAAGTCCATA	3828	53
		ISCR2_rv	AAGCGGGTTAGGTTGA		
	PCR 23	ISCR2_fw	CTATCTGGCCTCGGATGC	4707	51
		tetA_rv	GCGCTGTCGGATCGTT		
	PCR 24	tetA_fw	AGGCAAGCAGGATGTAGC	3438	50

	strA_rv	GTCATCCGACTTCT		
PCR 25	strA_fw	TTCTCTCGCGTAGCA	4997	53
	ΔTn21_rv	GCGCCGACCGAGTTCAT		
PCR 26	ΔTn21_fw	GCGCTGCTGCTCGAAACT	4381	50
	res/mod_rv	TAACAGCGCGAAAGA		
PCR 27	res/mod_fw	TTGCCGGTATGTTCTC	3523	48
	res/mod_rv	CCATAAAGGCGAGGTAAA		
PCR 28	res/mod_fw	TACCGCTATCCTGTATT	4587	49
	exc1_rv	ATGGCACTGGTATCAATC		
PCR 29	exc1_fw	TGCCGGTAGCTTGGACTC	3529	49
	fosA3_rv	ATAGCGGAGCCTATCTCT		
PCR 30	fosA3_fw	TCACTGCGTTATCTTC	3353	50
	bla <sub>CTX-M-55</sub> _rv	GAAACGTTCCGTCTCGAC		
PCR 31	bla <sub>CTX-M-55</sub> _fw	GCCGACGCTAACATACATCG	2037	50
	IS26_rv	GGCACTGTTGCAAATAGTCG		
PCR 32	IS26_fw	CAGATACGCCAGCGGCCAT	4980	55
	met_rv	GTTGAGCCGCTGCCTGCAA		
PCR 33	met_fw	GGCTGCAACCAGCCTGTAAT	3220	55
	ycjA_rv	TGCCTGCCAGCGCCATCACT		
PCR 34	ycjA_fw	AGACGTGCCGGACAGTGAAT	5010	53
	ssb_rv	CGGTGACGTAACGGGTAATA		
PCR 35	ssb_fw	CATGGCAGTTCGTGGCATT	3939	55
	psiA_rv	TCCAGGTCATGCTCTGCCAC		
PCR 36	psiA_fw	TGGCTGTATGCCGTGATGT	4621	49
	traM_rv	CTCTGATATCGTCAACCATA		
PCR 37	traM_fw	TCAGGAACTGCTTCAATGCT	4526	50

		traB_rv	CGCTCCATTCCCTGACCGAT		
PCR 38		traB_fw	GGTGACGTACCAGTCTGTGC	4232	53
		traC_rv	CGCTGAACCACACC GG CCTC		
PCR 39		traC_fw	CACAGTGAAGCCAACCTGAA	4820	53
		trbC_rv	GGATTGCCCTGACCTGCTG		
PCR 40		trbC_fw	CTGACAGCAGGACTGTTTAT	4043	50
		traF_rv	ATTAATGACCTGCGCCAGCT		
PCR 41		traF_fw	AGGAGCCGGATATTATGCAG	4544	50
		traG_rv	ATGCCACCAGAATGGTCTG		
PCR 42		traG_fw	ACAGGCTGAATCTGATACA	3905	49
		hp_rv	TAATTCTTATGCCTGAGAG		
PCR 43		hp_fw	TGTTGGTGAGAATAATCAAT	4027	50
		tral_rv	GTTCTGCCAGCACGTCCGT		
PCR 44		tral_fw	TCACAGGCTATCCGAGAGGC	3004	55
		tral_rv	CGCGCCGGTCCTCATTCAAGG		
PCR 45		tral_fw	GAAC TGCGGGAGGC CGGTATA	3250	50
		finO_rv	TTATTTCTCATCAAGCACGG		
<b>pECXH3</b>	PCR 46	IS903_fw	TCAGGCAATACGCACGCTTT	4178	53
		IS26_rv	CACGATT TACCGCTGGGTTTC		
PCR 47		IS26_fw	AGCAAGCGCGCGACCATAGG	3860	53
		orf2_rv	CCAGCGCCCTGTTAACTAT		
PCR 48		orf2_fw	GCCATGTACCAGCACATCCC	4583	54
		hp_rv	CCCAACGCTGCCGAGATGC		
PCR 49		hp_fw	GTAGGCGCCGCCCTATACCT	3173	50
		hp_rv	TCAGTCAGATCCATCCGACT		
<b>pECM13</b>	PCR 50	repZ_fw	GTGACCGGCGAGCAGAGCCT	4699	52

	IS26_rv	TCATGGATTAAATCCCAGC		
PCR 51	IS26_fw	CACTGTCGATTTTATCTCT	4358	49
	IS26_rv	CCTACGTGAAGGTCAATGGC		
PCR 52	IS26_fw	ATCCGTTTCAGTTGCCATG	5621	55
	ISCfr1_rv	CGTTCGCAGCGAGAGACGAG		
PCR 53	ISCfr1_fw	CTGGTCGTCGGCTACTGCTA	5585	52
	ISCR1_rv	TAGACGAAGTCTTCCCCAAG		
PCR 54	ISCR1_fw	TGAGCGAGCGTATTAGCTC	4705	53
	Intl1_rv	TGATCCGCATGCCGTTCCA		
PCR 55	Intl1_fw	ATGTCCACTGGGTTCGTGCC	4797	49
	smc_rv	AGTTAACGCTGAACAATACA		
PCR 56	smc_fw	AGGCCAGATCCATCACTCGT	3718	49
	tir_rv	CTCAAGCTTCTCGATTATGG		
PCR 57	tir_fw	CAGTACCGGAAGCAGTTATT	4250	50
	res_rv	TTATGGAATTCTTTGAGCA		
PCR 58	res_fw	CACATGCTGTATGCCGGTAT	4195	50
	umuC_rv	GCAGTTGAGCCCTACTCAAT		
PCR 59	umuC_fw	GCCAGCTGCAGTGCTGTGTT	4541	55
	ychA_rv	TTCGTCTTCGGTCAGCCACT		
PCR 60	ychA_fw	TATCCGGGCTGGATCAGCAT	4241	55
	parB_rv	TGTTCGGCAGGGTGATATC		
PCR 61	parB_fw	TGAGCGTGCCGCTGGCCTCA	2958	55
	psiA_rv	CGCCAGAGTGGTCTCCTCCTG		
PCR 62	psiA_fw	TGTGGCTGGCTGCCGTGGAT	4552	55
	tnp_rv	GCCTGCTTAATCTGCTCCGG		
PCR 63	tnp_fw	ACTCCCGCAGGTATGGTGG	4282	50

	nikB_rv	TGGTACTGGCGTGTAGTCAC		
PCR 64	nikB_fw	TATGACAGTGTACGCCATAC	4500	49
	trbC_rv	TGATATCGATATCCGTGACA		
PCR 65	trbC_fw	GTGGCTGTTGCTGTCCTC	3320	52
	trbA_rv	CTGTACTGGCAGATGGTGAT		
PCR 66	trbA_fw	AGCCATTGCTGTGCTTCGTT	4218	49
	traY_rv	GCAGGATTCTCTTATCTAT		
PCR 67	traY_fw	TCCTCCTAGGAAATTAATTAA	4883	49
	traU_rv	AGGGATAACTCAACGTTCC		
PCR 68	traU_fw	CGAACTGAACCTAACATTATT	3612	50
	traT_rv	ACACAATCAGTGACACACGA		
PCR 69	traT_fw	GTACTGGTGAATTTCAGCTTT	4250	49
	traN_rv	TCTCTGACAGTTAACATCTGTC		
PCR 70	traN_fw	CCACTGGTTACGTTAGGAT	3655	52
	SogL/SogS_rv	GATGACAAACAAAGCACCGG		
PCR 71	SogL/SogS_fw	AGGTATCGGTGGAGTCGTCG	4747	55
	tral_rv	CCATCATTGCCCTGAACGC		
PCR 72	tral_fw	TTGGCTTCAAGCGTCAGGTC	3925	50
	traE_rv	TACATCATGCTGTCGCATGA		
PCR 73	traE_fw	TCAATATTACCCCGCAATAT	4645	50
	pilV_rv	TTACAGGCATATGTGGTTCG		
PCR 74	pilV_fw	ACGTTATTCAAGGTTATTGA	4180	49
	pilQ_rv	CATTACATACAAACTCAGCT		
PCR 75	pilQ_fw	CCTGCCGGTGAGGCCTTAC	4766	50
	pilN_rv	CACCACCGGATTCAATATGA		
PCR 76	pilN_fw	ATAATGGATTGCTGGCGTA	2920	52

PCR 77	pilL_rv	AGAGTCAGCGTGCCAGAGAC		
	pilL_fw	GGCTGCATTCTCTGTCTGA	3368	50
	traC_rv	ATGACCACGACAGAACATCC		

<sup>a</sup>PCR1 to PCR20 were designed per the complete sequence of pECF12 and PCR2 to PCR6 were used to investigate the type I structure of *fosA3* environment; PCR21 to PCR45 were designed per the complete sequence of pECB11 and PCR22 to PCR31 were used to investigate the type II structure of *fosA3* environment; PCR46 to PCR49 were designed per the sequence of pECXH3, and which were used to investigate the type III structure of *fosA3* environment; PCR50 to PCR77 were designed per the complete sequence of pECM13 and PCR50 to PCR55 were used to investigate the type IV structure of *fosA3* environment.

Table S4. Annotation of ORFs in plasmid pECF12

Gene name	location (start-end)	size (aa <sup>a</sup> )	aa identity (%)	Alignment region of the match	homolog description
<i>repA1</i>	1-870	290	100	1-290/290	plasmid replication protein RepA [ <i>Escherichia coli</i> 7A8]
<i>tir</i>	1809-2462	217	100	1-217/217	transfer inhibition protein [ <i>Escherichia coli</i> ]
<i>peml</i>	2555-2812	85	100	1-85/85	stable plasmid inheritance [ <i>Escherichia coli</i> ]
<i>pemk</i>	2736-3146	136	100	1-136/136	stable inheritance protein PemK [ <i>Escherichia coli</i> UMNK88]
$\Delta$ <i>tnpA</i>	4409-3846	177	100	1-177:14-190/177 <sup>b</sup>	Tn3 transposase, tnpA [ <i>Escherichia coli</i> ]
<i>fosA3</i>	6617-6201	138	100	1-138/138	Fosfomycin resistance protein FosA3 [ <i>Escherichia coli</i> ]
IS26	6991-7707	238	100	1-238/238	Tnp1 superfamily, IS26 transposase [ <i>Escherichia coli</i> ]
<i>res</i>	8339-9076	245	100	1-245/245	Ser-recombinase, resolvase [ <i>Escherichia coli</i> 75]
<i>tetA</i>	10937-9663	424	100	1-424/424	tetracycline efflux protein TetA [ <i>Escherichia coli</i> UMNK88]
<i>tetR</i>	10941-11618	225	100	1-225/225	tetracycline resistance repressor protein TetR [ <i>Escherichia coli</i> ]
<i>strB</i>	13034-12198	278	100	1-278/278	streptomycin phosphotransferase, StrB [ <i>Escherichia coli</i> W26]
<i>strA</i>	13525-13034	163	100	1-163/163	streptomycin phosphotransferase, StrA [ <i>Escherichia coli</i> ]
$\Delta$ <i>sul2</i>	14713-13898	271	100	1-271:13-283/271	dihydropteroate synthase type-2 [ <i>Escherichia coli</i> UMEA 3215-1]
IS5075	16405-15401	334	100	1-334/334	IS5075 transposase [ <i>Escherichia coli</i> SMS-3-5]
$\Delta$ Tn3	18190-16484	568	100	1-568/568	Tn3 family transposase [ <i>Escherichia coli</i> SMS-3-5]
IS26	19018-18302	238	100	1-238/238	transposase of IS26 [ <i>Escherichia coli</i> ]
<i>bla</i> <sub>CTX-M-55</sub>	20392-19517	291	100	1-291/291	Beta-lactamase CTX-M-55 [ <i>Escherichia coli</i> ]
IS26	21530-20814	238	100	1-238/238	Tnp1 superfamily IS26, transposase of IS26 [ <i>Escherichia coli</i> ]
<i>stbA</i>	25179-24217	320	100	1-320/320	stable plasmid inheritance protein A, stbA [ <i>Escherichia coli</i> ]
<i>ycgA</i>	25696-26379	227	100	1-227/227	DNA methylase family protein, YcgA [ <i>Escherichia coli</i> ]
<i>ycgB</i>	26615-27049	144	100	1-144/144	transcription-repair coupling factor, YcgB [ <i>Escherichia coli</i> ]
<i>yciA</i>	27734-28321	195	100	1-195/195	YciA [ <i>Escherichia coli</i> ]
<i>ycgC</i>	28766-29188	140	100	1-140/140	y cgC [ <i>Escherichia coli</i> DEC8C]
<i>ssb</i>	30177-30743	188	100	1-188/188	single-strand binding protein, Ssb [ <i>Escherichia coli</i> ]
<i>parB</i>	31095-32516	473	100	1-473/473	parB, ParB-like partition protein [ <i>Escherichia coli</i> ]
<i>psiB</i>	33187-33621	144	100	1-144/144	plasmid SOS inhibition protein B [ <i>Escherichia coli</i> ]
<i>psiA</i>	33618-34337	239	100	1-239/239	plasmid SOS inhibition protein PsiA [ <i>Escherichia coli</i> F18+]

<i>ydbA</i>	36172-37533	453	100	1-453/453	YdbA [ <i>Escherichia coli</i> ]
<i>ydcA</i>	37580-38143	187	100	1-187/187	YdcA, DNA-methyltransferase-like protein [ <i>Escherichia coli</i> ]
<i>yubP</i>	38887-39708	273	100	1-273/273	YubP [ <i>Escherichia coli</i> ]
<i>ygfA</i>	40005-40652	215	100	1-215/215	lytic transglycosylase YgfA [ <i>Escherichia coli</i> ]
<i>traM</i>	40929-41312	127	100	1-127/127	type IV conjugative transfer protein TraM [ <i>Escherichia coli</i> ]
<i>traJ</i>	41443-42189	248	98	1-248/248	conjugal transfer protein TraJ [ <i>Escherichia coli</i> LAU-EC10]
<i>traA</i>	42544-42909	121	99	1-121/121	conjugal transfer protein TraA [ <i>Escherichia coli</i> O26:H11]
<i>traE</i>	43257-43823	188	100	1-188/188	type IV conjugative transfer system protein TraE [ <i>Escherichia coli</i> ]
<i>traK</i>	43810-44538	242	100	1-242/242	type IV conjugative transfer secretin TraK [ <i>Escherichia coli</i> ]
<i>traB</i>	44538-45965	475	100	1-475/475	type IV conjugative transfer system protein TraB [ <i>Escherichia coli</i> ]
<i>traP</i>	45955-46545	196	100	1-196/196	type IV conjugative transfer system protein TraP [ <i>Escherichia coli</i> ]
<i>traV</i>	46989-47504	171	100	1-171/171	type IV conjugative transfer system TraV [ <i>Escherichia coli</i> ]
<i>traW</i>	51027-51659	210	100	1-210/210	type IV conjugative transfer system TraW [ <i>Escherichia coli</i> ]
<i>traU</i>	51656-52648	330	100	1-330/330	conjugal transfer pilus assembly protein TraU [ <i>Escherichia coli</i> ]
<i>trbC</i>	53594-54232	212	100	1-212/212	type IV conjugative transfer system TrbC [ <i>Escherichia coli</i> ]
<i>traN</i>	55045-56895	616	100	1-616/616	conjugative transfer protein TraN [ <i>Escherichia coli</i> ]
<i>traF</i>	57142-57915	257	100	1-257/257	conjugative transfer pilus assembly protein TraF [ <i>Escherichia coli</i> ]
<i>trbB</i>	58299-58844	181	100	1-181/181	Putative conjugative transfer protein TrbB [ <i>Escherichia coli</i> ]
<i>trbF</i>	59102-59494	130	100	1-130/130	probable conjugal transfer protein TrbF [ <i>Escherichia coli</i> ]
<i>traH</i>	59481-60854	457	100	1-457/457	type IV conjugative transfer system pilus TraH [ <i>Escherichia coli</i> ]
<i>traG</i>	60851-63667	938	100	1-938/938	type IV conjugative transfer system TraG [ <i>Escherichia coli</i> ]
<i>traT</i>	64243-64977	244	100	1-244/244	type IV conjugative transfer system TraT [ <i>Escherichia coli</i> ]
<i>traD</i>	65968-68175	735	100	1-735/735	conjugal transfer protein TraD [ <i>Escherichia coli</i> ]
<i>tral</i>	68175-73445	1756	99	1-1756/1756	type IV conjugative transfer relaxase protein [ <i>Escherichia coli</i> ]
<i>traX</i>	73465-74211	248	100	248/248	type IV conjugative transfer pilin acetylase [ <i>Escherichia coli</i> ]

<sup>a</sup>aa, amino acids.

<sup>b</sup>Query alignment region:match alignment region/total aa of the match.

Table S5. Annotation of ORFs in plasmid pECB11

Gene designation	location (start-end)	size (aa <sup>a</sup> )	aa identity (%)	Alignment region of the match	homologue description
<i>repA1</i>	1-858	285	100	1-285/285	plasmid replication initiator RepA [ <i>Escherichia coli</i> ]
<i>repA4</i>	1221-1607	128	100	1-128/128	replication initiation protein RepA4 [ <i>Escherichia coli</i> ]
<i>tir</i>	1797-2450	217	100	1-217/217	CAAX amino terminal protease Tir protein [ <i>Escherichia coli</i> ]
<i>tnp</i>	4367-3834	177	100	1-177/177	Tn3 transposase [ <i>Escherichia coli</i> ]
<i>IS26</i>	4445-5161	238	100	1-238/238	transposase tnpAIS26 [ <i>Escherichia coli</i> ]
<i>ISCR2</i>	5374-6297	307	100	1-307/307	putative transposase of ISCR2 [ <i>Escherichia coli</i> EC1865]
<i>virD2</i>	6328-7212	294	100	1-294/294	type IV secretory pathway protein VirD2 [ <i>Escherichia coli</i> ]
<i>floR</i>	7429-8643	404	100	1-404/404	florfenicol-chloramphenicol exporter FloR [ <i>Escherichia coli</i> ]
<i>lysR</i>	8671-8976	101	100	1-101/101	putative transcriptional regulator protein LysR [ <i>Escherichia coli</i> ]
<i>tetA</i>	10517-9243	424	100	1-424/424	tetracycline repressor protein TetA class A [ <i>Escherichia coli</i> ]
<i>tetR</i>	10521-11198	225	100	1-225/225	tetracycline repressor protein TetR [ <i>Escherichia coli</i> ]
<i>strB</i>	12614-11778	278	100	1-278/278	aminoglycoside resistance protein StrB [ <i>Escherichia coli</i> ]
<i>strA</i>	13105-12614	163	100	1-163/163	streptomycin phosphotransferase, StrA [ <i>Escherichia coli</i> ]
$\Delta$ sul2	14293-13478	271	100	1-271:13-283/271 <sup>b</sup>	dihydropteroate synthase type-2 [ <i>Escherichia coli</i> UMEA ]
<i>IS5075</i>	15985-14981	334	100	1-334/334	Transposase, similar to IS5075 [ <i>Escherichia coli</i> ]
$\Delta$ tnp	17770-16064	568	100	1-568:421-988/568	transposase for transposon Tn21 [ <i>Escherichia coli</i> BIDMC 19A]
<i>mobA</i>	18113-18850	245	100	1-245/245	relaxation protein MobA [ <i>Escherichia coli</i> ]
<i>rel</i>	18863-19447	194	100	1-194/194	relaxation protein [ <i>Escherichia coli</i> ]
<i>res/mod</i>	22126-19490	878	100	1-878/878	type III restriction and modification system [ <i>Escherichia coli</i> ]
<i>res/mod</i>	24179-22131	682	100	1-682/682	type III restriction/modification enzyme [ <i>Escherichia coli</i> ]
<i>exc1</i>	27775-27326	149	100	1-149/149	Putative entry exclusion protein 1 (Exc1) [ <i>Escherichia coli</i> ]
<i>IS26</i>	28074-28790	238	100	1-238/238	transposase tnpAIS26 [ <i>Escherichia coli</i> ]
<i>orf2</i>	29290-30078	262	100	1-262/262	CadC-like protein, transcriptional regulator [ <i>Escherichia coli</i> ]
<i>fosA3</i>	31017-30601	138	100	1-138/138	fosfomycin resistance protein FosA3 (plasmid) [ <i>Escherichia coli</i> ]
<i>IS26</i>	31391-32107	238	100	1-238/238	transposase tnpAIS26 [ <i>Escherichia coli</i> ]
<i>bla<sub>TEM-1</sub></i>	32235-32900	221	100	1-221/221	beta-lactamase protein TEM-1b [ <i>Escherichia coli</i> UMNF18]

<i>orf477</i>	33159-33635	158	100	1-158/158	ORF477 [ <i>Escherichia coli</i> ]
<i>bla</i> <sub>CTX-M-55</sub>	34557-33682	291	100	1-291/291	extended-spectrum beta-lactamase CTX-M-55 [ <i>Escherichia coli</i> ]
<i>IS26</i>	34979-35695	238	100	1-238/238	transposase <i>tnpAIS26</i> [ <i>Escherichia coli</i> ]
<i>stbB</i>	38382-38029	117	100	1-117/117	stable plasmid inheritance protein B [ <i>Escherichia coli</i> ]
<i>stbA</i>	39344-38382	320	100	1-320/320	stable plasmid inheritance protein A (plasmid) [ <i>Escherichia coli</i> ]
<i>met</i>	39861-40544	227	100	1-227/227	DNA methylase family protein [ <i>Escherichia coli</i> ]
<i>ycgB</i>	40780-41214	144	100	1-144/144	transcription-repair coupling factor, YcgB [ <i>Escherichia coli</i> ]
<i>ychA</i>	41214-42041	275	100	1-275/275	<i>YchA</i> (plasmid) [ <i>Escherichia coli</i> ]
<i>yciA</i>	41914-42486	190	100	1-190/190	<i>YciA</i> [ <i>Escherichia coli</i> ]
<i>kicA</i>	42459-42884	141	100	1-141/141	anti-restriction protein <i>kicA</i> [ <i>Escherichia coli</i> ]
<i>ycjA</i>	42931-43353	140	100	1-140/140	<i>YcjA</i> (plasmid) [ <i>Escherichia coli</i> ]
<i>ydbA</i>	44861-46222	453	100	1-453/453	<i>YdbA</i> (plasmid) [ <i>Escherichia coli</i> ]
<i>ydcA</i>	46269-46832	187	100	1-187/187	<i>YdcA</i> [ <i>Escherichia coli</i> ]
<i>ssb</i>	47648-48214	188	100	1-188/188	single-strand binding protein, Ssb [ <i>Escherichia coli</i> ]
$\Delta$ <i>yefA</i>	50128-50589	153	100	1-153:39-191/153	<i>YefA</i> (plasmid) [ <i>Escherichia coli</i> ]
<i>psiB</i>	50658-51092	144	100	1-144/144	plasmid SOS inhibition protein B [ <i>Escherichia coli</i> ]
<i>psiA</i>	51089-51808	239	100	1-239/239	plasmid SOS inhibition protein A [ <i>Escherichia coli</i> ]
<i>yubP</i>	53568-54389	273	100	1-172/273	<i>yubP</i> protein [ <i>Escherichia coli</i> FVEC1302]
<i>ygfA</i>	55333-54686	215	100	1-215/215	lytic transglycosylase <i>YgfA</i> [ <i>Escherichia coli</i> ]
<i>traM</i>	55610-55993	127	100	1-127/127	conjugal transfer protein <i>TraM</i> [ <i>Escherichia coli</i> ]
<i>traJ</i>	56124-56870	248	100	1-248/248	conjugal transfer protein <i>TraJ</i> [ <i>Escherichia coli</i> ]
<i>traA</i>	57225-57590	121	100	1-121/121	conjugal transfer protein <i>TraA</i> [ <i>Escherichia coli</i> ]
<i>traL</i>	57605-57916	103	100	1-103/103	conjugal transfer pilus assembly protein <i>TraL</i> [ <i>Escherichia coli</i> ]
<i>traE</i>	57938-58504	188	100	1-188/188	conjugal transfer protein <i>TraE</i> [ <i>Escherichia coli</i> ]
<i>traK</i>	58491-59219	242	100	1-242/242	conjugal transfer protein <i>TraK</i> [ <i>Escherichia coli</i> ]
<i>traB</i>	59219-60646	475	100	1-475/475	conjugal transfer protein <i>TraB</i> [ <i>Escherichia coli</i> ]
<i>traP</i>	60636-61226	196	100	1-196/196	conjugal transfer protein <i>TraP</i> [ <i>Escherichia coli</i> ]
$\Delta$ <i>traV</i>	61778-62185	135	100	1-135:37-171/135	conjugal transfer protein <i>TraV</i> [ <i>Escherichia coli</i> ]
<i>traC</i>	62803-65328	841	100	1-841/841	conjugal transfer protein <i>TraC</i> [ <i>Escherichia coli</i> ]

<i>trbI</i>	65325-65711	128	100	1-128/128	conjugal transfer protein TrbI (plasmid) [ <i>Escherichia coli</i> ]
<i>traW</i>	65708-66340	210	100	1-210/210	conjugal transfer pilus assembly protein TraW [ <i>Escherichia coli</i> ]
<i>traU</i>	66337-67329	330	100	1-330/330	F pilin assembly, TraU [ <i>Escherichia coli</i> ]
<i>ygeA</i>	67356-67664	102	100	1-102/102	ygeA protein [ <i>Escherichia coli</i> DEC8D]
<i>trbC</i>	68275-68913	212	100	1-212/212	conjugal transfer protein TrbC (plasmid) [ <i>Escherichia coli</i> ]
<i>HNH</i>	69307-69729	140	100	1-140/140	HNH endonuclease family protein [ <i>Escherichia coli</i> DEC1B]
<i>traN</i>	69726-71576	616	100	1-616/616	conjugal transfer protein TraN [ <i>Escherichia coli</i> ]
<i>traF</i>	71823-72596	257	100	1-257/257	conjugative transfer pilus assembly protein TraF [ <i>Escherichia coli</i> ]
<i>trbB</i>	72980-73525	181	100	1-181/181	conjugal transfer protein TrbB [ <i>Escherichia coli</i> ]
<i>trbF</i>	73783-74175	130	100	1-130/130	putative conjugal transfer protein TrbF [ <i>Escherichia coli</i> ]
<i>traH</i>	74162-75535	457	100	1-457/457	conjugal transfer pilus assembly protein TraH [ <i>Escherichia coli</i> ]
<i>traG</i>	75601-78348	915	100	1-915/915	conjugal transfer protein TraG, partial [ <i>Escherichia coli</i> ]
<i>traS</i>	78381-78902	173	100	1-173/173	conjugal transfer entry exclusion protein TraS [ <i>Escherichia coli</i> ]
<i>traT</i>	78924-79658	244	100	1-244/244	conjugative transfer protein TraT [ <i>Escherichia coli</i> ]
<i>hp</i>	79861-80598	245	100	1-245/245	transmembrane protein (plasmid) [ <i>Escherichia coli</i> ]
$\Delta$ <i>traD</i>	80649-82157	502	100	1-500/500	conjugal transfer protein TraD [ <i>Escherichia coli</i> ]
<i>tral</i>	82886-88156	1756	100	1-1756/1756	conjugal transfer protein TraI [ <i>Escherichia coli</i> ]
<i>traX</i>	88176-88922	248	100	1-248/248	conjugal transfer protein, TraX [ <i>Escherichia coli</i> ]
<i>finO</i>	88977-89537	186	100	1-186/186	conjugal transfer fertility inhibition protein FinO [ <i>Escherichia coli</i> ]

<sup>a</sup>aa, amino acids.

<sup>b</sup>Query alignment region:match alignment region/total aa of the match.

Table S6. Annotation of ORFs in plasmid pECXH3

Gene designation	location (start-end)	size (aa <sup>a</sup> )	aa identity (%)	Alignment region of the match	homologue description
IS903	1309-386	307	99	1-307/307	IS903 transposase (plasmid) [ <i>Escherichia coli</i> ]
<i>bla</i> <sub>CTX-M-65</sub>	2264-1389	291	100	1-291/291	CTX-M-65 beta-lactamase [ <i>Escherichia coli</i> ]
<i>fipA</i>	3395-2769	208	100	1-208/208	<i>fipA</i> [Plasmid pKM101]
IS26	4709-3993	238	100	1-238/238	IS26 transposase, (plasmid) [ <i>Escherichia coli</i> ]
IS26	6112-5396	238	100	1-238/238	IS26 transposase, (plasmid) [ <i>Escherichia coli</i> ]
<i>fosA3</i>	6486-6902	138	100	1-138/138	fosfomycin resistance protein FosA3 [ <i>Escherichia coli</i> ]
<i>hp</i>	7114-7443	109	100	1-109/109	hypothetical protein A6V31_17905 [ <i>Proteus mirabilis</i> ]
<i>orf1</i>	7425-6907	172	100	1-172/172	Orf1 (plasmid) [ <i>Escherichia coli</i> ]
<i>orf2</i>	8213-7425	262	100	1-262/262	Orf2 (plasmid) [ <i>Escherichia coli</i> ]
IS26	9429-8713	238	100	1-238/238	IS26 transposase, (plasmid) [ <i>Escherichia coli</i> ]
<i>tetR</i>	10909-10250	219	99	1-219/219	tetracycline repressor protein TetR [ <i>Escherichia coli</i> SE11]
<i>tet(A)</i>	11002-12192	396	100	1-396/396	Tetracycline efflux protein TetA [ <i>Proteus mirabilis</i> ]
<i>hp</i>	12096-12434	112	100	1-112/112	conserved hypothetical protein (plasmid) [ <i>Escherichia coli</i> ]
<i>orf</i>	13184-12717	155	100	1-155/155	orf [ <i>Escherichia coli</i> ]
<i>merR</i>	13782-13375	135	100	1-135/135	MerR family transcriptional regulator [ <i>Proteus mirabilis</i> ]
<i>hp</i>	13878-14774	298	100	1-298/298	sodium: proton antiporter [Gammaproteobacteria]
<i>hp</i>	14778-15290	170	100	1-170/170	lipoprotein signal peptidase [ <i>Escherichia coli</i> ]

<sup>a</sup>aa, amino acids.

Table S7. Annotation of ORFs in plasmid pECM13

Gene designation	location (start-end)	size (aa <sup>a</sup> )	aa identity (%)	Alignment region of the match	homologue description
<i>repZ</i>	1-765	255	100	1-255/255	IncFII RepA protein [ <i>Escherichia coli</i> 2.3916]
<i>rep</i>	900-307	197	99	1-197/197	replication initiation protein (plasmid) [ <i>Escherichia coli</i> ]
<i>IS26</i>	4111-4827	238	100	1-238/238	transposase of IS26, IS6 family [ <i>Escherichia coli</i> ]
<i>bla</i> <sub>CTX-M-14</sub>	5130-6005	291	100	1-291/291	extended-spectrum beta-lactamase CTX-M-14 [ <i>Escherichia coli</i> ]
<i>fosA3</i>	6617-7033	139	100	1-139/139	Fosfomycin resistance protein FosA3 [ <i>Escherichia coli</i> ]
<i>IS26</i>	9029-8313	238	100	1-238/238	transposase of IS26, IS6 family [ <i>Escherichia coli</i> ]
<i>aac(3)-IId</i>	9250-10110	286	100	1-286/286	aminoglycoside 3-N-acetyltransferase [ <i>Escherichia coli</i> ]
<i>tmrB</i>	10123-10665	180	100	1-180/180	Tunicamycin resistance protein [ <i>Escherichia coli</i> ]
<i>ISCfr1</i>	12892-14262	456	100	1-456/456	ISCfr1, TnpA [ <i>Escherichia coli</i> ]
<i>bla</i> <sub>TEM-1</sub>	15083-15943	286	100	1-286/286	TEM-1 beta-lactamase [ <i>Escherichia coli</i> ]
<i>rmtB</i>	16113-16868	251	100	1-251/251	16S rRNA methylase RmtB [ <i>Escherichia coli</i> ]
<i>ISCR1</i>	19002-17461	513	100	1-513/513	ISCR1 transposase [ <i>Escherichia coli</i> ]
<i>sul1</i>	20333-19407	279	100	1-279/279	dihydropteroate synthase, sul1 [ <i>Escherichia coli</i> ]
<i>qacEdelta1</i>	20587-20240	115	100	1-115/115	quaternary ammonium compounds [ <i>Escherichia coli</i> ]
<i>aadA2</i>	21542-20751	263	100	1-263:10-272/263 <sup>b</sup>	aminoglycoside nucleotidyltransferase [ <i>Klebsiella pneumoniae</i> ]
<i>intI1</i>	22592-23605	337	100	1-337/337	IntI1 integrase [ <i>Escherichia coli</i> ]
<i>IS26</i>	24598-23882	238	100	1-238/238	transposase of IS26, IS6 family [ <i>Escherichia coli</i> ]
<i>chem</i>	24577-25965	462	99	16-462/447	chemotaxis protein [ <i>Enterobacter hormaechei</i> subsp. <i>oharae</i> ]
<i>smc</i>	28744-27188	518	100	1-518/518	recF/RecN/SMC N terminal domain protein [ <i>Escherichia coli</i> ]
<i>tir</i>	30185-31288	367	100	1-367/367	nucleotide-binding containing TIR-like [ <i>Escherichia coli</i> ]
<i>res</i>	33953-34732	259	100	1-259/259	resolvase [ <i>Escherichia coli</i> KTE191]
<i>yefA</i>	34879-35556	225	100	1-225/225	ParB-like nuclease YefA [ <i>Escherichia coli</i> ]
<i>stbA</i>	36310-37344	344	100	1-344/344	plasmid partitioning and stability StbA [ <i>Escherichia coli</i> ]
<i>umuC</i>	39029-37755	424	100	1-424/424	UV protection protein UmuC [ <i>Escherichia coli</i> ]
<i>yubD</i>	41199-41882	227	100	1-227/227	putative DNA methylase YubD [ <i>Escherichia coli</i> ACN001]
<i>ychA</i>	42552-43379	275	100	1-275/275	YchA [ <i>Escherichia coli</i> ]
<i>parB</i>	46437-48485	682	100	1-682	ParB-like partition protein [ <i>Escherichia coli</i> KTE191]

<i>psiA</i>	48971-49690	239	100	1-239/239	plasmid SOS inhibition protein A, PsiA [ <i>Escherichia coli</i> ]
<i>ardA</i>	50512-51246	244	100	1-244/244	antirestriction protein ArdA [ <i>Escherichia coli</i> ]
<i>tnp</i>	52832-53794	320	100	1-320/320	transposase [ <i>Escherichia coli</i> ]
<i>ydiA</i>	54901-55749	282	100	1-282/282	YdiA protein [ <i>Escherichia coli</i> ACN001]
<i>nikB</i>	56831-59449	872	100	1-872/872	relaxase NikB [ <i>Escherichia coli</i> ]
<i>trbC</i>	62754-60463	763	100	1-763/763	conjugal transfer protein TrbC [ <i>Escherichia coli</i> EC4402]
<i>trbB</i>	63817-62747	356	100	1-356/356	TrbB protein [ <i>Escherichia coli</i> ACN001]
<i>trbA</i>	65044-63836	402	100	1-402	conjugal protein TrbA [ <i>Escherichia coli</i> ACN001]
<i>traY</i>	70091-67854	745	100	1-745/745	Incl1 conjugal transfer protein TraY [ <i>Escherichia coli</i> ]
<i>traW</i>	71934-70732	400	100	1-400/400	conjugal transfer protein, TraW [ <i>Escherichia coli</i> KO11FL]
<i>traU</i>	75559-72515	1014	100	1-1014/1014	conjugal protein TraU [ <i>Escherichia coli</i> W26]
<i>traT</i>	76449-75649	266	100	1-266/266	conjugal transfer protein TraT [ <i>Escherichia coli</i> ]
<i>traP</i>	78371-77667	234	100	1-234/234	conjugal transfer protein TraP [ <i>Escherichia coli</i> ]
<i>traO</i>	79660-78371	429	100	1-429/429	conjugal transfer protein TraO [ <i>Salmonella enterica</i> subsp]
<i>traN</i>	80646-79663	327	100	1-327/327	conjugal transfer protein TraN (plasmid) [ <i>Escherichia coli</i> ]
<i>traM</i>	81349-80657	230	100	1-230/230	conjugal transfer protein TraM [ <i>Escherichia coli</i> ]
SogL/SogS	85478-81711	1255	100	1-1255/1255	SogL/SogS DNA primase [ <i>Escherichia coli</i> ]
<i>traK</i>	86479-85568	303	100	1-303/303	conjugal transfer protein TraK (plasmid) [ <i>Escherichia coli</i> ]
<i>traJ</i>	87569-86421	382	100	1-382/382	conjugative transfer protein (plasmid) [ <i>Escherichia coli</i> ]
<i>tral</i>	88384-87566	272	100	1-272/272	plasmid conjugative transfer protein Tral [ <i>Escherichia coli</i> ]
<i>traF</i>	91080-89878	400	100	1-400/400	conjugal transfer protein TraF [ <i>Escherichia coli</i> ]
<i>traE</i>	91989-91165	274	100	1-274/274	plasmid conjugative transfer protein TraE [ <i>Escherichia coli</i> ]
<i>rcl</i>	93294-92140	384	99	1-384:12-395/384	Shufflon-specific DNA recombinase, Rcl [ <i>Escherichia coli</i> ]
<i>pilV</i>	96348-95239	369	100	1-369/369	conjugal transfer protein pilV [ <i>Escherichia coli</i> ]
<i>pilU</i>	97019-96336	227	100	1-227/227	conjugal transfer preilin peptidase PilU [ <i>Escherichia coli</i> ]
<i>pilR</i>	99276-98179	365	100	1-365/365	conjugal transfer protein PilR [ <i>Escherichia coli</i> ]
<i>pilQ</i>	99289-100842	517	100	1-517/517	conjugal transfer protein PilQ [ <i>Escherichia coli</i> ]
<i>pilO</i>	103923-102628	431	100	1-431/431	conjugative transfer protein PilO [ <i>Escherichia coli</i> ETEC]
<i>pilN</i>	105598-103916	560	100	1-560/560	Incl1 conjugal transfer protein PilN [ <i>Escherichia coli</i> ]

<i>pilL</i>	107116-106049	355	100	1-355/355	conjugal transfer protein PilL [ <i>Escherichia coli</i> ]
<i>traC</i>	110070-109387	227	100	1-227/227	conjugational TraC protein [ <i>Escherichia coli</i> ETEC 1392/75]

<sup>a</sup>aa, amino acids.

<sup>b</sup>Query alignment region:match alignment region/total aa of the match.