

Supplementary Figure 1. Overview of the effect of siRNA on the elemental content of HeLa cells. A total number of genes showing significant increase (red) and decrease (blue) for each element is shown. Changes with standard scores (z-scores, derived from 3 replicates) above 5 were considered significant.

Supplementary Figure 2. An algorithm for processing the ionomics data. The ICP-MS data from the ionomics screen in HeLa cells was processed to remove bias. For each gene knockdown and element, we calculated standard scores (z-scores) and fold change. These data were further used for functional analysis.

Supplementary Figure 3. Efficiency of siRNA knockdown analyzed by real-time PCR. KCNA1, RPL14 and SBP2 were identified in the screen and validated using ⁷⁵Se labeling in HeLa cells (derived from 3 replicates). Total RNA from HeLa cells transfected with the corresponding siRNAs was used for realtime PCR analyses. scrRNA transfection was used as control. This figure shows gene expression changes normalized to control. One fold of standard variances for each bar were showed above.

Supplementary Figure 4. Characterization of genes that affect copper levels in HeLa cells. Following gene-specific knockdowns, HeLa cells were fixed and analyzed by confocal microscopy with antibodies against ATP7A (green) and the trans-Golgi network marker TGN46 (red). Scale bar: 25 µm. (b) 100x magnification images. ANKRD9 knockdown results in fragmented TGN and fragmented ATP7A staining pattern. Scale bar: 10 µm. (c) HeLa cells were transiently transfected with siRNAs, and cell lysates were analyzed by Western blotting. A representative blot is shown. A full blot is in Supplementary Figure 7.

Supplementary Figure 5. Assay development for high-throughput ionomics screening of human cells. (a) Standard score (derived from 3 replicates) for elemental ICP-MS profile of indicated human cell lines. Cells were grown to full confluence on 6-well plates in 10% serum-supplemented DMEM. Cells were washed 3 times in TE (10 mM Tris, 1mM EDTA, pH 7.4), followed by digestions with 1.5% nitric acid and 1.5 % hydrogen peroxide (50 ppb gallium was also added as an internal standard). Trace elements were analyzed by ICP-MS. The graph represents the number of standard deviations from the mean (zscore). (b) Optimization of medium removal. HeLa cells were washed 3 times with HEPES-bicarbonate, HEPES-citrate, or TRIS-EDTA (TE), all at pH 7.4, digested and analyzed by ICP-MS.

Supplementary Figure 6. Effect of target down-regulation on ATP7A protein levels and electrophoretic mobility. Following gene knockdowns, HeLa cell lysates were analyzed by Western blotting. The blot was cut at 130 kDa; the upper part was immunostained for ATP7A and the lower part was immunostained for Na/K-ATPase. This figure shows the entire blot, and Fig. 4b its reduced version.

Supplementary Figure 7. Characterization of genes that affect copper levels in HeLa cells. HeLa cells were transiently transfected with siRNAs, and cell lysates were analyzed by Western blotting. This is a full blot that accompanies Supplementary Figure 4c.

Supplementary Table 1. Biological processes associated with changes in iron levels. The table was generated using Gene Ontology analysis (Genesifter, Geospiza) for the set of genes down-regulation of which increased or decreased iron levels in cells with a z-score > 3. Increased iron levels are linked to more specific biologic processes, whereas iron depletion is linked to more global changes in cell.

Supplementary Table 2. Distribution of genes, whose knockdown increases Cu or Fe levels, according to their standard scores (z-scores, derived from 3 replicates).

Supplementary Table 3. Comparison of biological processes influencing copper and iron ionomes.

To generate this table, gene ontology analysis was performed for all genes down-regulation of which was associated with increased copper or iron levels (z-score above 4, derived from 3 replicates). Biological processes common for copper and iron ionomes are indicated in italic. The majority of processes were unique for each ionome.

Supplementary Table 4. Transport/trafficking proteins down-regulation of which produces copper elevation with the standard score (z-score, derived from 3 replicates) > 4. Blue color indicates protein with known/predicted roles in intracellular protein or vesicle trafficking. The Table was generated using GeneSifter.

Supplementary Table 5. Biological processes associated with changes in copper levels. The table was generated using Gene Ontology analysis (Genesifter) for the set of genes downregulation of which changes copper levels in cells with a standard score (z-score, derived from 3 replicates) above 3.

Supplementary Table 6. Comparison of the ICP-MS data for *Saccharomyces cerevisiae* **gene deletions producing significant changes of Fe or Cu levels and their orthologs in HeLa cells**. Yeast genes producing statistically significant changes of copper or iron levels upon knockdown were selected from the database available at<http://www.ionomicshub.org/> and the fold change was calculated using normalized ICP-MS value available at the website. Human orthologs of these yeast genes were identified through Ensembl database; and the ICP-MS data for these orthologs was taken from the dataset generated in the current study. The Z-score values for yeast data are defined in Yu and colleagues 21 ; the Z-score values for HeLa screen are as defined in the text. All the statistical results were derived from 3 replicates.

* outlier ICP-MS values were not included in the calculations; NR: non-reliable data due to wide range of values. Orthologs that result in similar effects on Cu or Fe ionomes in yeast and human cells are indicated in red.