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Carbonyl reductase 1 catalyzes 20β-reduction of glucocorticoids, modulating receptor activation and metabolic complications of obesity

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39 Supplementary Materials:

40 SM1 Quantification of urinary glucocorticoid metabolites

Glucocorticoids were extracted from equine urine (20mL) by solid phase extraction on Bond
Elut Nexus mixed mode Large Reservoir Capacity, 60 mg columns (Agilent Technologies,
Santa Clara, CA, USA). Glucocorticoids were extracted from human urine (10mL) by solid
phase extraction on Sep-Pak columns (Waters, Milford, MA, USA).

Steroid conjugates were hydrolysed using β -glucuronidase followed by re-extraction. The 45 steroids obtained were derivatized to form methoxime-trimethylsilyl (MO-TMS) derivatives. 46 Steroidal derivatives were separated by gas chromatography using the TRACE GC Ultra Gas 47 Chromatograph (Thermo Fisher Scientific). Analysis was performed on a TSQ Quantum Triple 48 Quadrupole GC-tandem mass spectrometer (Thermo Fisher Scientific) using a 35HT 49 50 Phenonemex column (30m, 0.25mm, 0.25µm, Agilent Technologies) as previously described (45, 46). Epi-cortisol and epi-tetrahydrocortisol were used as internal standards (Steraloids, 51 Newport, RI, USA). The steroids analyzed were cortisol (F), cortisone (E), 5β-52 tetrahydrocortisol (5β-THF), 5β-tetrahydrocortisone (5β-THE), 5α-tetrahydrocortisol (5α-53 THF) (45, 46) with the inclusion of the following transitions (collision energy) α -cortol 54 $(535\rightarrow355, 20V)$ and β -cortol $(535\rightarrow455, 10V)$, α - and β -cortolone $(449\rightarrow269, 10V)$, 6β -55 hydroxycortisol (693 \rightarrow 513, 10V), 20 α - dihydrocortisol (578 \rightarrow 488, 10V) and 20 β -56 dihydrocortisol ($681 \rightarrow 578, 10V$). 57

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59 Steroid quantities in equine urine were expressed as a ratio to creatinine, which was measured
60 using a colorimetric method based on the modified Jaffe's reaction (IL650 analyser,
61 Instrumentation Laboratories, Barcelona, Spain).

62 SM2 Quantification of glucocorticoids in plasma

Plasma samples (1 mL Equine, 200 μ L Human) enriched with internal standard (D4-F, D4-E and D8-B; 250 ng of each) were extracted by liquid-liquid extraction. Chloroform (10 volumes) was added to each sample, mixed and the organic layer was dried under nitrogen (60 °C). The extracts were re-suspended in mobile phase (60 μ L, water: methanol 70:30 v/v) for quantification of steroids by LC-MS/MS. The injection volume was 30 μ L.

68 SM3 Quantification of glucocorticoids in adipose tissue

Adipose samples (100mg) were homogenized in ethyl acetate (1 mL) and enriched with internal 69 standard (D4-F, D8-E, D8-B; 250ng of each). The homogenate was slowly dripped onto chilled 70 ethanol: glacial acetic acid: water (95:3:2 v/v, 10 mL) and frozen at -80 °C overnight. Samples 71 were thawed (4 °C) prior to sonication (8 x 15 second bursts) and centrifugation (3000 x g, 30 72 mins, 4 °C). The supernatant was dried under nitrogen (60 °C), re-suspended in methanol (10 73 74 mL) and frozen at -80 °C overnight. Samples were thawed (RT) and hexane (10 mL) added and mixed. The hexane layer was removed, the remaining methanol dried down under nitrogen 75 (60 °C) and re-suspended in water (400 µL) and ethyl acetate (4 mL). The organic layer was 76 removed, dried under nitrogen (60 °C) and re-suspended in 30 % methanol (5 mL). C18 Bond 77 Elut columns (Agilent Technologies, Santa Clara, CA, USA) were conditioned (methanol 5 78 79 mL) and equilibrated (water 5 mL), samples were loaded and steroids eluted with methanol (2 mL). Eluates were dried down under nitrogen (60 °C) and re-suspended in 60 µL mobile phase 80 (water: methanol, 70:30 v/v) for quantification of steroids by LC-MS/MS. Injection volume 81 was 30 µL. 82

83 SM4 Quantification of mRNA by RT-qPCR

Total RNA was extracted from adipose and liver using the RNAeasy Mini Kit (Qiagen Inc,
Valencia, CA, USA). The tissue was mechanically disrupted in either QIAzol (Qiagen) for

adipose tissue or RLT buffer (Qiagen) for liver tissue. Total RNA was extracted from cells in
QIAzol lysis reagent using an RNeasy Mini Kit according to the manufacturer's instructions.

Quantitative real-time polymerase chain reaction was performed using a Light-cycler 480 88 89 (Roche Applied Science, Indianapolis, IN, USA). Primers were designed using sequences from the National Centre of Biotechnological Information and the Roche Universal Probe Library 90 (see Table S7-9) for details of primers for genes of interest and housekeeping genes). Samples 91 were analysed in triplicate and amplification curves plotted (y axis fluorescence, x axis cycle 92 number). Triplicates were deemed acceptable if the standard deviation of the crossing point 93 94 was < 0.5 cycles. A standard curve (y axis crossing point, x axis log concentration) for each gene was generated by serial dilution of cDNA pooled from different samples and fitted with 95 a straight line and deemed acceptable if reaction efficiency was between 1.7 and 2.1. 96

Supplementary Tables and Figures

	Lean (n=14)	Obese (n=14)
Age (years)	15.6 ± 5.6	13.8 ± 7.8
Sex	4 Females	8 Female
	10 Castrated males	5 Castrated males
Breeds	11 Thoroughbred	2 Thoroughbred
	2 Native pony	3 Cob
	1 Percheron	7 Native ponies
		1 Arab pony
		1 Clydesdale
Body condition score	2.3 ± 0.3	$3.8 \pm 0.7*$
(/5)		

Table S 1 Characteristics of equine study subjects

Data are expressed as mean \pm SEM. Student's t-test or Mann-Whitney U test: *p<0.05

Table S 2 Characteristics of obese human participants with type 2 diabetes providing 24 hour urine samples

n	19
Age (years)	58.9 ± 1.5
Body Mass Index (kg/m ²)	32.60 ± 1.2
Concurrent medications	4 no medication 15 metformin

All participants were male and diagnosed with diabetes. Participants provided 24 hour urine samples. The additional lean and obese participants providing urine were recruited as part of a different study (Upreti et al 2014). Data are expressed as mean ± SEM.

	Lean (n=10)	Obese (n=10)
Age (years)	50.5 ± 10.4	50.0 ± 11.8
Body Mass Index (kg/m ²)	23.8 ± 1.2	32.9 ± 2.7
Concurrent mediciations	 6 No medication 1 perindopril, nifedipine, 1 citalopram, simvastatin, clopidogrel, ranitidine 1 clomipramine 1 ranitidine 	 8 No medication 1 tamsulosin, lansoprazole 1 pantoprazole

Table S 3 Characteristics of lean and obese study participants providing plasma samples

Plasma was collected (between 8 and 9am) from lean and obese but otherwise healthy men. Participants with diabetes or prior corticosteroid treatment were excluded prior to recruitment. Data are expressed as mean \pm SEM.

Table S 4 Characteristics of l	lean and	l obese	men	providing	adipose	biopsy	samples	during
surgery.								

	Lean (n=8)	Obese (n=8)
Age (years)	55.7 ± 12.7	52.8 ± 13.7
Body Mass Index (kg/m ²)	23.0 ± 1.7	38.8 ± 6.7
Surgery	 Cholecystectomy Laparoscopic cholecystectomy Removal of gastric band Hernia repair Open cholecystectomy 3 Abdominal hernia repairs 	 Gastric bypass Laparoscopic sleeve gastrectomy Laparoscopic cholecystectomy Cholecystectomy 2 Laparoscopic fundoplications Laparoscopic cholecystectomy Laparoscopic removal of gastric band and gastric bypass

All the participants were male. Samples were obtained at the time of surgery. Data are

expressed as mean \pm SEM.

Table S 5 Estimates of the phenotypic associations with CBR1 expression in the liver based on Mendelian Randomisation

Outcome	Beta	Se	P value	Ν	Sample
BMI	-0.01	0.01	0.25	339224	(55)
Body Fat	-0.01	0.01	0.06	100716	(56)
2hr glucose adjusted for BMI	<0.01	0.03	0.89	15234	(57)
Fasting glucose	0.01	< 0.01	0.02	58074	(58)
Fasting insulin	< 0.01	< 0.01	0.45	51750	(58)
HOMA-B	< 0.01	< 0.01	0.59	46186	(59)
HOMA-IR	0.01	0.01	0.27	46186	(59)
HbA1c	0.01	<0.01	0.01	46368	(60)

These analyses used rs1005696 as the instrument for CBR1 expression in liver and associations were analysed using MR-base. See supplementary references for further details of studies used.

Table S 6 Human primer sequences for PCR

Gene Symbol, full name	Forward Primer (3'→ 5')	Reverse Primer (5'→ 3')
RNA18s (ribosomal RNA 18s)	CTTCCACAGGAGGCCTACA C	CGCAAAATATGCTGGAACT TT
DUSP1 (dual specificity phosphatase 1)	TTCAAGAGGCCATTGACTT	CCTGGCAGTGGACAAACA C
<i>GILZ</i> (glucocorticoid- induced leucine zipper)	CCGTTAAGCTGGACAACAG TG	ATGGCCTGTTCGATCTTGT T
<i>FKBP51</i> (FK506-binding protein 51)	GGATATACGCCAACATGTT CAA	CCATTGCTTTATTGGCCTCT
<i>IGFBP1</i> (insulin-like growth factor binding protein 1)	GCCTTGGCTAAACTCTCTA CGA	CCATGTCACCAACATCAAA AA
<i>IL-1β</i> (Interleukin 1β)	TGTAATGAAAGACGGCACA CC	TCTTCTTTGGGTATTGCTTG G
<i>CBR1</i> (Carbonyl Reductase 1)	TCCCTCTAATAAAACCCCA AGG	GGTCTCACTGCGGAACTTC T

Table S 7 Equine primer sequences for PCR

Gene Symbol, full name	Forward Primer (3'→ 5')	Reverse Primer (5'→ 3')
RNA18s (ribosomal RNA 18s)	TGACCCAAGGCTAGTAGCT GA	TTCAACACATCACCCACCA T
<i>SDHA</i> (Succinate	CTACGGAGACCTTAAGCAT CTGA	GGGTCTCCACCAGGTCAGT A

dehydrogenase complex)		
<i>Equine CBR1</i> (Carbonyl Reductase 1)	ACCCAGCCATGTCTTACAC C	CAGGATAGTGAAGCCGAT GC
Reductase 1)		

Table S 8 Murine primer sequences for PCR

Gene Symbol, full	Forward Primer (3'→ 5')	Reverse Primer $(5' \rightarrow 3')$	
name			
RNA18s (ribosomal	CTCAACACGGGAAACCT	CGCTCCACCAACTAAGA	
RNA 18s)	CAC	ACG	
Tbp (TATA-binding	GGGAGAATCATGGACCA	GATGGGAATTCCAGGAG	
protein)	GAA	TCA	
Per1 (Period 1)	GCTTCGTGGACTTGACAC CT	TGCTTTAGATCGGCAGT GGT	
<i>Pepck</i> (phosphoenolpyruvate carboxykinase)	GAGGCACAGGTCCTTTTC AG	GTTCCTGGGCCTTTGTG AC	
Adipq (Adiponectin)	GGTGAGAAGGGTGAGAA AGGA	TTTCACCGATGTCTCCCT TAG	
Lpl (Lipoprotein lipase)	CTCGCTCTCAGATGCCCT AC	GGTTGTGTGTTGCCAT T	
Sgk1 (serum glucocorticoid kinase 1)	TTTCCAAAGGGGGGATGC T	TGTTGGCATGATTACAT TGTTCT	
ENaC1 (Epithelial sodium channel 1)	AGCACAGAGAACACCCC TGT	TGGCTCTTCCTACCCTCT CTC	
Cbr1	AGGTGACAATGAAAACG	GGACACATTCACCACTC	
(Carbonyl Reductase 1)	AACITI	IGC	





Urinary glucocorticoids of lean and obese participants were extracted, derivitized and quantified by GC-MS/MS. Urinary excretion of 20β -dihydrcortisol was higher in the obese group.

Figure S 2 Equine liver cytosol metabolises cortisol to 20β -dihydrocortisol and this production is inhibited by quercetin



[A] In the presence of NADPH equine liver cytosol produced 20β -DHF as the predominant metabolite. (THF = tetrahydrocortisol, THE = tetrahydrocortisone). [B] CBR1 inhibitor quercetin prevented production of 20β -DHF by equine liver cytosol. Data are mean \pm SEM.



Figure S 3 20β-Dihydrocorticosterone is present in murine tissues

20 β -Dihydrocortisone (the murine equivalent of 20 β -dihydrocortisol) was measured by LC-MS/MS in murine [A] plasma and [B] liver. Data are mean \pm SEM.



Figure S 4 20β-Dihydrocorticosterone induces murine glucocorticoid receptor activation

HEK293 cells transfected with murine GR-MMTV-Luc were incubated with increasing concentrations of 20 β -dihydrocorticosterone (20 β -DHB) the murine equivalent of 20 β -dihydrocortisol. Data are mean \pm SEM, *P<0.05 compared to vehicle.

Supplementary References

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