

Supplementary Information

Shape-based separation of microalga *Euglena gracilis* using inertial microfluidics

Ming Li^{1,2}, Hector Enrique Muñoz¹, Keisuke Goda^{2,3,4*}, and Dino Di Carlo^{1,5,6*}

¹Department of Bioengineering, University of California, Los Angeles, USA

²Department of Electrical Engineering, University of California, Los Angeles, USA

³Department of Chemistry, University of Tokyo, Japan

⁴Japan Science and Technology Agency, Japan

⁵California NanoSystems Institute, University of California, Los Angeles, USA

⁶Jonsson Comprehensive Cancer Centre, University of California, Los Angeles, USA

* dicarlo@seas.ucla.edu, goda@chem.s.u-tokyo.ac.jp

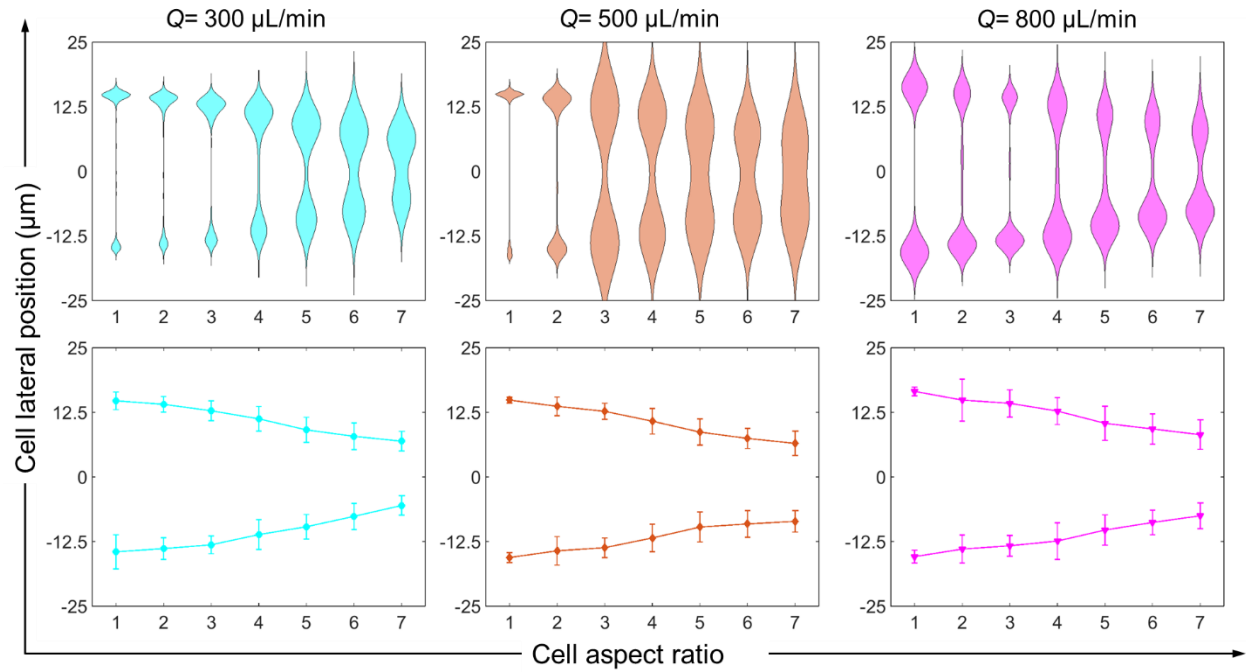


Figure S1 Inertial focusing distributions of *E. gracilis* cells with various aspect ratios ranging from 1 to 7 in a microchannel (width=50 μm , height=90 μm) at flow rates of 300, 500, and 800 $\mu\text{L}/\text{min}$.

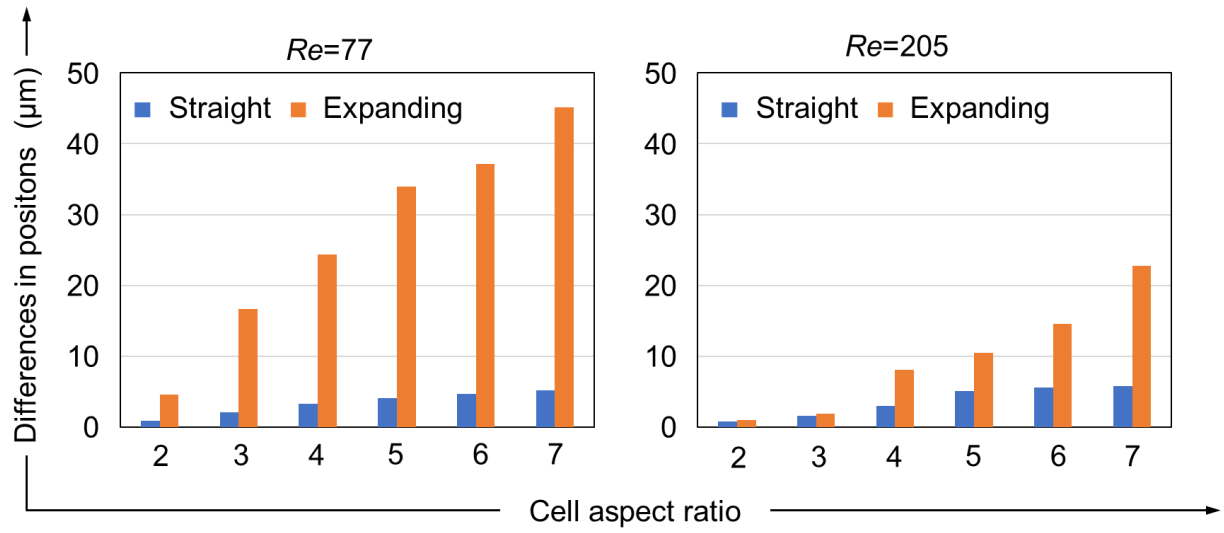


Figure S2 Comparison of differences in averaged lateral positions between *E. gracilis* cells with AR=1 and others (i.e. AR=2-7) in the straight region and the gradually expanding region for $Re=77$ (left) and $Re=205$ (right).

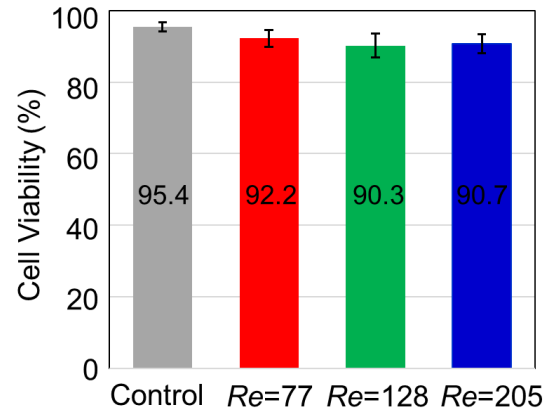


Figure S3 The viability of *E. gracilis* cells was not significantly affected by inertial effects. The standard deviations are obtained from three measurement.

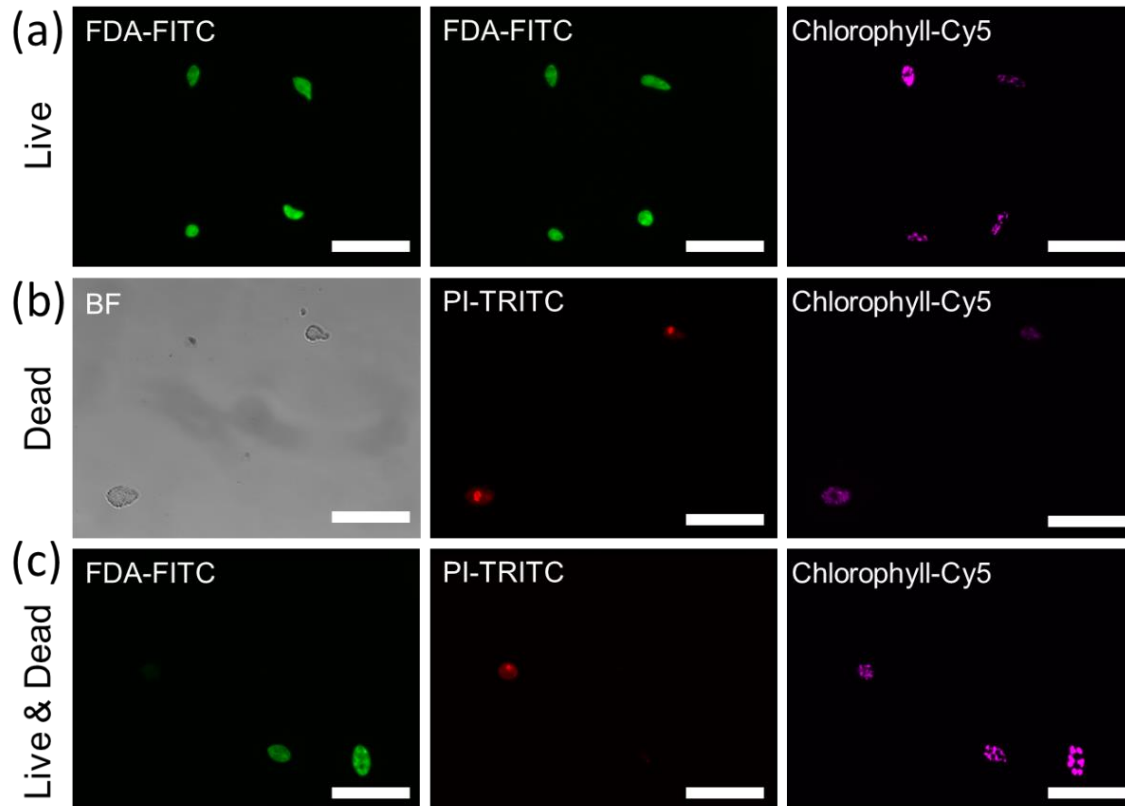


Figure S4 Viability assays of *E. gracilis* cells using fluorescence-based live-dead assays with FDA and PI. (a) Microscopic images of dead cells (from left to right: FDA signal in FITC channel, FDA signal in FITC channel, and chlorophyll auto-fluorescence signal in Cy5 channel). (b) Microscopic images of live cells (from left to right: bright field, PI signal in TRITC channel, and chlorophyll auto-fluorescence signal in Cy5 channel). (c) Microscopic images of live/dead cell mixtures (from left to right: FDA signal in FITC channel, PI signal in TRITC channel, and chlorophyll auto-fluorescence signal in Cy5 channel). Scale bar=50 μ m.

Supplementary Movie Captions

Movie S1

Shape-dependent lateral inertial focusing equilibrium positions for *E. gracilis* cells at an expansion region. High-speed microscopic video at a flow rate of 300 $\mu\text{L}/\text{min}$ ($\text{Re} = 77$).

Movie S2

Shape based separation of *E. gracilis* cells at outlets 1 and 2. High-speed microscopic video at a flow rate of 300 $\mu\text{L}/\text{min}$ ($\text{Re} = 77$).

Movie S3

Shape based separation of *E. gracilis* cells at outlet 3. High-speed microscopic video at a flow rate of 300 $\mu\text{L}/\text{min}$ ($\text{Re} = 77$).