

## **Hyaluronic acid based extracellular matrix regulates surface expression of GluN2B containing NMDA receptors**

Barbara Schweitzer<sup>1</sup>, Jeet Singh<sup>1</sup>, Anna Fejtova<sup>2,3</sup>, Laurent Groc<sup>4</sup>, Martin Heine<sup>5\*</sup> & Renato Frischknecht<sup>1,6,7\*</sup>

<sup>1</sup> Leibniz Institute for Neurobiology, Dept. for Neurochemistry and Molecular Biology, Brenneckestr. 6, Magdeburg 39118 Germany

<sup>2</sup> RG Presynaptic Plasticity, Leibniz Institute for Neurobiology, Magdeburg, Germany

<sup>3</sup> Molecular Psychiatry, Department of Psychiatry and Psychotherapy, University Hospital, Friedrich-Alexander-University Erlangen-Nuremberg, Erlangen, Germany

<sup>4</sup> Centre National de la Recherche Scientifique, Interdisciplinary Institute for Neuroscience, UMR 5297, 33000 Bordeaux, France.

<sup>5</sup> RG Molecular Physiology, Leibniz Institute for Neurobiology; Brenneckestr. 6, Magdeburg 39118 Germany

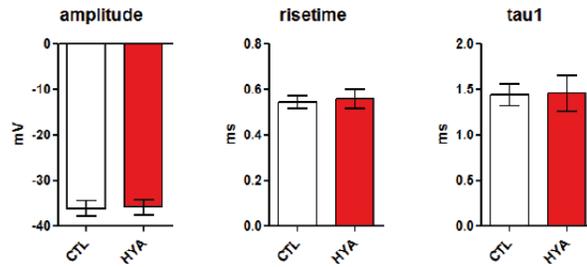
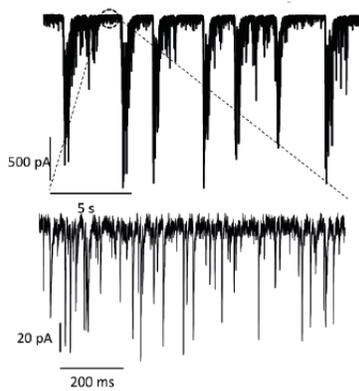
<sup>6</sup> Center for Behavioral Brain Sciences, D-39118 Magdeburg, Germany

<sup>7</sup> Department of Biology, Animal Physiology, University of Erlangen-Nuremberg, Staudtstrasse 5, 91058 Erlangen, Germany

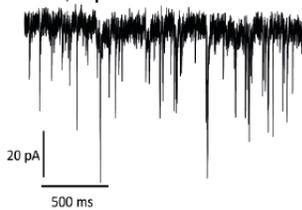
\* Correspondence to: [renato.frischknecht@fau.de](mailto:renato.frischknecht@fau.de) or [martin.heine@lin-magdeburg.de](mailto:martin.heine@lin-magdeburg.de)

## Supplementary Material

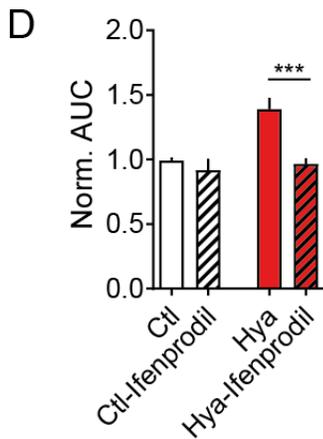
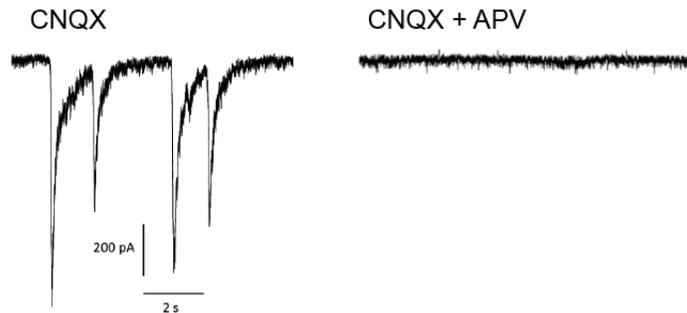
**A** Extracellular: + 10  $\mu$ M Bicuculline, 10 mM APV, 0.5 MgCl<sub>2</sub>, 2mM CaCl<sub>2</sub>



**B** ± 0.5  $\mu$ M TTX

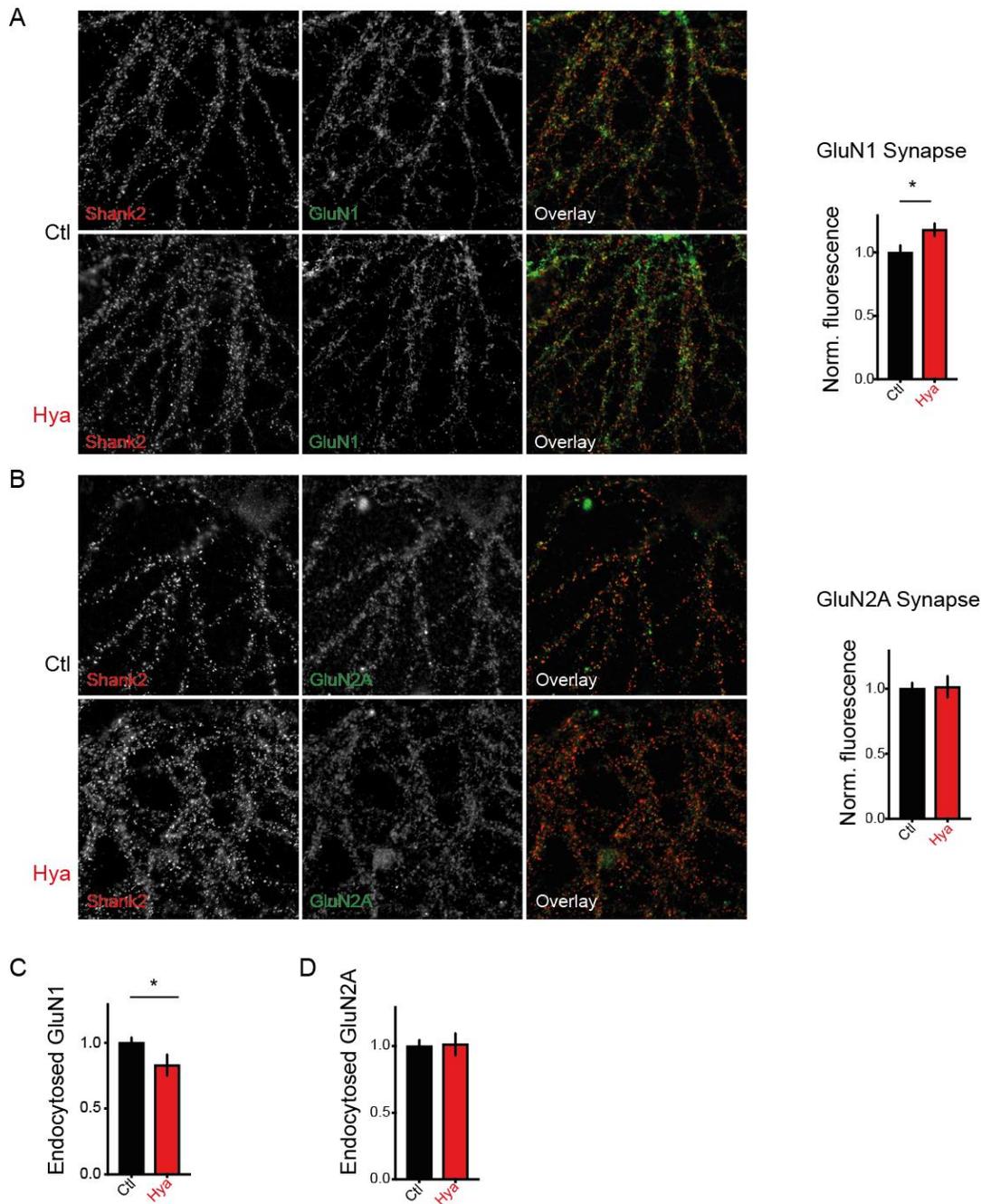


**C** Extracellular: + 10  $\mu$ M Bicuculline, 10 mM CNQX, 15  $\mu$ M Glycin, 0 mM MgCl<sub>2</sub>, 1 mM CaCl<sub>2</sub>  
Intracellular: 10 mM QX314



**Supplementary Figure S1: Isolation of NMDA receptor mediated currents** **A)** sEPSCs under control conditions, driven by AMPARs and NMDA receptors. Large burst-like events alternated with periods with single events (see magnification) which were analyzed. There was no change in amplitude, rise time or decay time (tau1) in these AMPAR dominated events. **B)** mEPSCs under control conditions. **C)** sEPSCs in presence of CNQX, driven by NMDA receptors. Single peaks with large amplitudes were detected and analyzed, which were abolished by APV. **D)** Ifenprodil had no significant effect on charge transfer before hyaluronidase treatment (Ctl: 1.0 ± 0.02; Ctl-Ifenprodil: 0.92 ± 0.09; Hya: 1.39 ± 0.09; Hya-Ifenprodil: 0.96 ± 0.05, P = 0.0006, Unpaired Student's t-test).

## Supplementary Material



**Supplementary Figure S2: Regulation of surface expression of GluN1 and GluN2A subunits.** **A)** Dissociated hippocampal neurons were co-stained for surface expressed GluN1 subunit of the NMDA receptor and the excitatory synapse protein shank2. Quantification of fluorescence intensity at shank2 positive synapses revealed a significant increase of GluN1 subunit after 12 h hyaluronidase treatment (Ctl:  $1.00 \pm 0.06$ ,  $n = 28$ ; Hya:  $1.18 \pm 0.09$ ,  $n = 27$ ,  $P = 0.024$ , Unpaired Student's t-test). **B)** Quantification of synaptic GluN2A subunit co-stained with shank2 revealed no difference between Hya treated and control cells (Ctl:  $1.00 \pm 0.05$ ,  $n = 10$ ; Hya:  $1.01 \pm 0.08$ ,  $n = 10$ ). **C+D)** Quantification of endocytosed GluN1 and GluN2A subunits revealed decreased endocytosis for GluN1 but no change for GluN2A after hyaluronidase treatment (GluN1 Ctl:  $1.00 \pm 0.04$ ,  $n = 13$ ; Hya:  $0.82 \pm 0.08$ ,  $n = 6$ ,  $P = 0.04$ ; GluN2A Ctl:  $1.00 \pm 0.08$ ,  $n = 10$ ; Hya:  $1.09 \pm 0.10$ ,  $n = 8$ ; Unpaired Student's t-test).