Supporting Information for

Site- and Stereo-selective *trans*-Hydroboration of 1,3-Enynes Catalyzed by 1,4-Azaborine-Based Phosphine-Pd Complex

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Index	page
General	S2
Synthesis of compound S1	S3
General procedure A for the synthesis of amides	S3-S5
General procedure B for the synthesis enamines	S5-S6
General procedure C for the synthesis of 1,4-azaborines	S6-S7
General procedure D for the synthesis of Senphos ligands	S7-S9
Gram-scale synthesis of ligand L3	S10
General procedure E for the synthesis of enynes	S10-S11
Synthesis of enyne 4n	S11-S12
General procedure F for the synthesis of enynes	S12-S16
General procedure G for the hydroboration of terminal enynes	S17-S23
General procedure H for the hydroboration of internal enynes	S23-S29
Gram-scale hydroboration of internal enyne 61	S30
Suzuki-Miyaura reaction of 71	S30-S31
Diels-Alder reaction 71	S31-S32
Homologation of 71	S32-S33
Preparation of complex 12 and its use in trans-hydroboration	S33
Synthesis of CC-L3 and its performance in hydroboration reactions	S34-S37
Crystallographic data for 3a, 5a, 7a, 9, 12	S38-S42
References	S43
NMR spectra for all new compounds	S44-S143

General

All oxygen- and moisture-sensitive manipulations were carried out under an inert atmosphere using either standard Schlenk techniques or a glove box.

THF, Et₂O, CH₂Cl₂, toluene, benzene, and pentane were purified by passing through a neutral alumina column under argon. All other chemicals and solvents were purchased and used as received. Enynes **4a**¹ [CAS: 935-01-3], **4e**² [CAS: 16124-56-4], **4g**² [CAS: 1089304-22-2], **4l**³ [CAS: 3752-22-5], **4m**⁴ [CAS: 2807-10-5], **4q**⁵ [CAS: 73395-75-2], **6d**⁶ [CAS: 56392-49-5], **6g**⁷ [CAS: 1655-05-6], **6l**⁸ [CAS: 54147-31-2], and carbamate **10**⁹ [CAS: 145167-88-0] were synthesized according to the literature procedures and the characterization data are consistent with those reported in the literature.

¹¹B NMR spectra were recorded on a Varian Unity/Inova 500 spectrometer at ambient temperature. ¹¹B NMR chemical shifts are externally referenced to BF₃•Et₂O (δ 0). ¹H NMR spectra were recorded on a Varian Unity/Inova 500 spectrometer. ¹³C NMR spectra were recorded on a Varian Unity/Inova 500 or Unity/Inova 600 spectrometer. ¹⁹F NMR spectra were recorded on a Varian Unity/Inova 500 spectrometer. ³¹P NMR spectra were recorded on a Varian Unity/Inova 500 spectrometer. IR spectra were recorded on a Bruker FTIP Alpha (ATR mode) spectrometer. High-resolution mass spectroscopy data were obtained at the Mass Spectroscopy Facilities at Chemistry Department of Boston College.

Synthesis of S1

The preparation of S1 was adapted from literature procedures. 10 A 50-mL flask was charged with the aryl triflate (1.775 g, 5.000 mmol), which was prepared according to literature procedures. 11 Diphenylphosphine (931 mg, 5.00 mmol), Pd(PPh₃)₄ (251 mg, 0.220 mmol), toluene (13.0 mL), and Et₃N (0.77 mL, 5.5 mmol) were added to the reaction flask. The resulting mixture was then allowed to stir at 110 °C for 16 hr. At the conclusion of the reaction, the mixture was allowed to cool to the room temperature. H₂O (50 mL) was added, and the mixture was extracted with CH₂Cl₂ (3 x 30 mL). The combined organic layer was dried over anhydrous Na₂SO₄. After removal of the solvent, the crude residue was purified by column chromatography on silica gel with hexanes/EtOAc (20: 1) as the eluent to afford S1 as white solid (1.37 g, 70% yield) ¹H NMR (500 MHz, CDCl₃) δ 8.36 (d, J = 8.5 Hz, 1H), 7.79 (d, J = 7.5 Hz, 1H), 7.67 (d, J = 8.0 Hz, 1H), 7.60 (t, J = 8.0 Hz, 1H), 7.54 (t, J = 8.0 Hz, 1H), 7.30-7.37 (m, 10H), 6.86 (dd, J = 8.0, 2.0 Hz, 1H); ¹³C NMR (150 MHz, CDCl₃) δ 136.9, 136.8, 136.3, 136.2, 134.4, 134.0, 133.9, 132.5, 131.3, 131.1, 129.9, 128.9, 128.7, 128.6, 128.1, 127.7, 127.6, 127.3, 127.2 (complexity due to the P-C coupling); ³¹P NMR (202 MHz, CDCl₃) δ -3.2; IR (ATR) 3051, 1583, 1543, 1478, 1433, 1214, 1180, 1154, 998, 848, 769 cm⁻¹; HRMS (DART) calcd for $C_{22}H_{17}^{79}BrP$ ($[M+H]^+$) 391.02512, found 391.02594.

General procedure A for the synthesis of amides

To a 500-mL flask charged with 2-bromoaniline (17.2 g, 100 mmol), pyridine (7.9 mL, 100 mmol), and CH₂Cl₂ (250 mL) was added acyl chloride (100 mmol) in dropwise fashion at 0 °C. The resulting mixture was then allowed to stir at the same temperature for 0.5 h. Water (100 mL) was then added to quench the reaction. The biphasic mixture was then extracted with CH₂Cl₂ (100 mL x 3). The combined organic phase was dried over Na₂SO₄, and volatiles were removed under reduced pressure. The residue was then dissolved in THF (250 mL), and the mixture was cooled to 0 °C. NaH (6.0 g, 60% in

mineral oil, 150 mmol) was then added at 0 °C in 3 portions. The resulting mixture was then allowed to stir at 0 °C for 0.5 h. Methyl iodide (9.30 mL, 150 mmol) was then added in dropwise fashion at 0 °C. The reaction mixture was then allowed to stir at room temperature for 2 h. At the conclusion of the reaction, water (100 mL) was added in a dropwise fashion to quench the reaction at 0 °C. Then the mixture was extracted with CH₂Cl₂ (100 mL x 3), and the combined organic phase was dried over Na₂SO₄. The resulting crude residue was purified by distillation under attenuated pressure to afford amide 1 as an yellow oil.

1a [168335-51-1]: 19.8 g, 82% yield.

¹H NMR (500 MHz, CDCl₃) δ 7.68 (dd, J = 8.5, 1.5 Hz, 1H), 7.38 (td, J = 8.0, 1.5 Hz, 1H), 7.22-7.28 (m, 2H), 3.19 (s, 3H), 1.97 (q, J = 7.5 Hz, 2H), 1.06 (t, J = 7.5 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 173.6, 142.8, 133.8, 129.8, 129.6, 128.9, 123.5, 35.7, 27.3, 9.3; IR (ATR) 3058, 2978, 2937, 1665, 1584, 1476, 1420,

1378, 1321, 1283, 1250, 1132, 1046, 1028, 807, 766 cm⁻¹; HRMS (DART) calcd for $C_{10}H_{13}^{79}$ BrNO ([M+H]⁺) 242.01805, found 242.01860.

1b: 23.7 g, 93% yield.

¹H NMR (500 MHz, CDCl₃) δ 7.69 (dd, J = 7.5, 1.0 Hz, 1H), 7.38 (td, J = 8.0, 1.0 Hz, 1H), 7.22-7.27 (m, 2H), 3.19 (s, 3H), 1.93 (t, J = 7.0 Hz, 2H), 1.57-1.64 (m, 2H), 0.83 (t, J = 7.5 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 172.8, 142.9, 133.8, 129.9, 129.6, 128.9, 123.5, 35.8, 35.6, 18.5, 13.8; IR (ATR)

2961, 2929, 2873, 1659, 1584, 1476, 1436, 1417, 1382, 1340, 1309, 1290, 1250, 1223, 1131, 1109, 1049, 894, 764 cm⁻¹; HRMS (DART) calcd for $C_{11}H_{15}^{79}BrNO$ ([M+H]⁺) 256.03370, found 242.03468.

1c: 25.3 g, 94% yield.

¹H NMR (500 MHz, CDCl₃) δ 7.69 (dd, J = 7.5, 1.0 Hz, 1H), 7.38 (td, J = 8.0, 1.0 Hz, 1H), 7.22-7.26 (m, 2H), 3.19 (s, 3H), 2.13-2.18 (m, 1H), 1. 84 (d, J = 7.0 Hz, 2H), 0.86 (d, J = 6.5 Hz, 3H), 0.83 (d, J = 6.5 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 172.3, 142.9, 1c 133.8, 130.0, 129.6, 128.9, 123.5, 42.8, 35.7, 25.4, 22.6, 22.5; IR (ATR) 2956, 2929, 2870, 1662, 1584, 1476, 1419, 1376, 1337, 1305, 1262, 1140, 1116, 1053, 1031, 765 cm⁻¹; HRMS (DART) calcd for $C_{12}H_{17}^{79}BrNO$ ([M+H]⁺) 270.04935, found 242.05054.

General procedures B for the synthesis of enamines

To a 250-mL flask charged with 1 (20.0 mmol), PMHS (5.2 g, M_w 1700-3200, ~80.0 mmol hydride), and toluene (30 mL) was added a toluene (2.0 mL) solution of (PPh₃)₂(CO)IrCl (7.8 mg, 0.01 mmol). Gelation was observed immediately and the reaction mixture was allowed to sit at room temperature for 0.5 h. Diethyl ether (100 mL) was then added, and the mixture was passed through a pad of celite. The filter cake was then washed with ether 3 times (100 mL each time). The combined organic phase was then concentrated. The residue was then purified by distillation under attenuated pressure to afford 2 as light yellow oil.

2a [1527467-69-1]: 2.85 g, 63% yield. The characterization data are consistent with those reported in the literature. 12

2b: 3.41 g, 71% yield.

¹H NMR (500 MHz, CD₂Cl₂) δ 7.69 (dd, J = 7.5, 1.0 Hz, 1H), 7.27 (td, J = 7.5, 1.5 Hz, 1H), 7.11 (d, J = 7.5 Hz, 1H), 6.98 (td, J = 8.0, 1.5 Hz, 1H), 6.18 (d, J = 14.0 Hz, 1H), 4.56-4.62 (m, 1H), 2.96 (s, 3H), 2.04-2.10 (m, 2H), 1.02 (t, J = 7.5 Hz, 3H); ¹³C NMR (125 MHz, CD₂Cl₂) δ 149.0, 136.1, 134.3, 128.8, 126.9, 126.2, 120.4, 105.2, 38.8, 24.1, 16.2; IR (ATR) 3055, 2956, 2927, 2847, 2805, 1653, 1584, 1512, 1480, 1464, 1439, 1420, 1309, 1109, 969, 786 cm⁻¹; HRMS (DART) calcd for C₁₁H₁₅⁷⁹BrN ([M+H]⁺) 240.03879, found 240.03281. 2c: 4.38 g, 86% yield.

¹H NMR (500 MHz, CD₂Cl₂) δ 7.58 (dd, J = 8.0, 1.0 Hz, 1H), 7.29 Me (td, J = 8.0, 1.5 Hz, 1H), 7.11 (dd, J = 7.5 Hz, 1.5 Hz, 1H), 6.99 (td, J = 7.0, 1.5 Hz, 1H), 6.18 (dd, J = 13.0 Hz, 1.0 Hz, 1H), 4.56 Br (dd, J = 13.0, 7.0 Hz, 1H), 2.96 (s, 3H), 2.34-2.36 (m, 1H), 1.05 (dd, J = 6.0, 1.0 Hz, 6H); ¹³C NMR (125 MHz, CD₂Cl₃) δ 149.1, 134.7, 134.4, 128.8, 126.9, 126.1, 120.3, 111.4, 38.8, 29.9, 24.5; IR (ATR) 2953, 2866, 2806, 1651, 1584, 1513, 1482, 1464, 1420, 1397, 1357, 1111, 986, 765 cm⁻¹; HRMS (DART) calcd for C₁₂H₁₇⁷⁹BrN ([M+H]⁺) 254.05444, found 254.05436.

General procedure C for the synthesis of C3-Substituted 1,4-azaborines

To a 100-mL flask charged with 2 (15.0 or 20.0 mmol) was added *n*BuLi (6 or 8 mL, 15 or 20 mmol) at -78 °C. The resulting mixture was allowed to stir at -78 °C for 15 min. Diisopropylaminoboron dichloride (2.73 or 3.64 g, 15.0 or 20.0 mmol) was then added at -78 °C. The reaction mixture was allowed to stir at -78 °C for 1 h and then room temperature for another 1 h. At the conclusion of the reaction, solvents were removed under reduced pressure, and the resulting crude residue was purified by vacuum distillation under attenuated pressure to afford 3 as light yellow oil which slowly solidified. We were not able to obtain their HRMS data due to their extremely air and moisture sensitivity.

3a: 20.0 mmol scale, 3.0 g, 79% yield.

¹H NMR (500 MHz, CD₂Cl₂) δ 8.34 (d, J = 7.5 Hz, 1H), 7.75 (s, 1H), 7.69 (td, J = 9.0, 1.5 Hz, 1H), 7.53 (d, J = 9.0 Hz, 1H), 7.34 (t, J = 7.5 Hz, 1H), 3.88 (s, 3H), 2.21 (s, 3H); ¹³C NMR (125 MHz, CD₂Cl₂) δ 148.6, 144.2, 133.7, 131.9, 127.6 (br), 121.9, 121.1 (br), 115.6, 42.5, 17.6; ¹¹B NMR (160 MHz, CD₂Cl₂) δ 44.6.

3b: 15.0 mmol scale, 2.14 g, 69% yield.

¹H NMR (500 MHz, CD₂Cl₂) δ 8.36 (dd, J = 8.0, 1.5 Hz, 1H), 7.68-7.73 (m, 2H), 7.55 (d, J = 8.0, Hz, 1H), 7.34 (t, J = 8.0 Hz, 1H), 3.89 (s, 3H), 2.62 (q, J = 7.5 Hz, 2H), 1.20 (t, J = 7.5 Hz, 3H); ¹³C NMR (125 MHz, CD₂Cl₂) δ 148.2, 144.2, 133.8, 131.9, 128.2 (br), 121.9, 115.6, 42.5, 25.8, 16.8 (a B-aryl carbon signal not observed); ¹¹B NMR (160 MHz, CD₂Cl₂) δ 44.2.

3c: 15.0 mmol scale, 3.09 g, 94% yield.

¹H NMR (500 MHz, CD₂Cl₂) δ 8.37 (dd, J = 7.5, 1.5 Hz, 1H), 7.68-7.73 (m, 2H), 7.54 (d, J = 8.5 Hz, 1H), 7.33 (td, J = 7.0, 1.0 Hz, 1H), 3.91 (s, 3H), 3.17-3.21 (m, 1H), 1.26 (d, J = 7.5 Hz, 6H); ¹³C NMR (125 MHz, CD₂Cl₂) δ 146.7, 144.0, 133.9, 132.0, 121.9, 115.6, 42.8, 30.5, 24.2 (two B-aryl carbon signals not observed); ¹¹B NMR (160 MHz, CD₂Cl₂) δ 44.1.

General procedure D for the synthesis of Senphos ligands

To a 20-mL vial charged with o-bromoaryldiarylphosphine (1.0 mmol) and THF (5.0 mL) was added nBuLi (0.40 mL, 2.5 M in hexanes, 1.0 mmol) at -78 °C. The resulting mixture was allowed to stir at -78 °C for 1-2 h. 1,4-Azaborine 3 (1.0 mmol) in THF (2.0 mL) was then added. The resulting mixture was allowed to stir at -78 °C for 2 h and then at room temperature for 2 h. At the conclusion of the reaction, volatiles were removed under reduced pressure. The resulting crude residue was purified by column chromatography on silica gel using pentane/Et₂O as the eluent to afford ligand L as a white powder.

L2: 240 mg, 58% yield.

¹H NMR (500 MHz, CD₂Cl₂) δ 7.76 (s, 1H), 7.62-7.64 (m, 1H), 7.58-7.60 (m, 2H), 7.43-7.45 (m, 1H), 7.25-7.39 (m, 11H), 7.16-7.20 (m, 2H), 7.08 (td, J = 7.5, 1.5 Hz, 1H), 3.98 (s, 3H), 1.99 (s, 3H); ¹³C NMR (150 MHz, CD₂Cl₂) δ 147.00, 143.48, 139.44, 139.41, 139.38,

139.30, 139.13, 139.05, 137.02, 133.95, 133.92, 133.83, 133.80, 131.89, 131.79, 130.94, 128.79, 128.74, 128.66, 128.62, 128.60, 128.53, 128.49, 127.22, 120.99, 114.98, 42.43, 19.44, 19.42 (complexity due to P-C coupling; B-aryl carbon signal not observed); 11 B NMR (160 MHz, CD₂Cl₂) δ 47.2; 31 P NMR (202 MHz, CD₂Cl₂) δ –10.3; IR (ATR) 3051, 2923, 1604, 1585, 1540, 1491, 1477, 1454, 1432, 1269, 1171, 1101, 1046, 1026, 943, 892, 763 cm⁻¹; HRMS (DART) calcd for C₂₈H₂₆BNP ([M+H]⁺) 418.13959, found 418.18939.

L3: 288 mg, 67% yield.

¹H NMR (500 MHz, CD₂Cl₂) δ 7.71 (s, 1H), 7.52-7.57 (m, 2H), 7.48 (d, J = 7.5 Hz, 1H), 7.39 (t, J = 7.0 Hz, 1H), 7.19-7.29 (m, 11H), 7.10-7.14 (m, 2H), 6.98 (td, J = 7.5, 1.0 Hz, 1H), 3.94 (s, 3H), 2.38-2.41 (m, 1H), 2.29-2.32 (m, 1H), 0.94 (t, J = 8.0 Hz, 3H); ¹³C NMR (150 MHz, CD₂Cl₂) δ 146.68, 143.36, 139.43, 139.35, 139.32, 139.28, 139.17, 139.09, 137.13, 133.98, 133.96,

Me N B Et PPh₂

133.90, 133.86, 133.78, 132.28, 132.17, 130.95, 128.77, 128.73, 128.64, 128.60, 128.53, 128.33, 127.15, 120.89, 114.94, 42.56, 26.98, 26.97, 17.01 (complexity due to P-C coupling; B-aryl carbon signal not observed); 11 B NMR (160 MHz, CD₂Cl₂) δ 48.5; 31 P NMR (202 MHz, CD₂Cl₂) δ –10.6; IR (ATR) 3051, 2955, 2924, 2864, 1604, 1584, 1539, 1490, 1432, 1405, 1374, 1172, 1067, 763, 763 cm⁻¹; HRMS (DART) calcd for C₂₉H₂₈BNP ([M+H]⁺) 432.20524, found 432.20696.

L4: 360 mg, 81% yield. ¹H NMR (500 MHz, CD₂Cl₂) δ 7.73 (s, 1H), 7.52-7.57 (m, 2H), 7.46 (d, J = 7.0 Hz, 1H), 7.19-7.39 (m, 12H), 7.09-7.12 (m, 2H), 6.97 (t, J = 7.0 Hz, 1H), 3.96 (s, 3H), 2.72-2.77 (m, 1H), 1.03 (d, J = 7.0 Hz, 3H), 0.96 (d, J = 6.5 Hz, 3H); ¹³C NMR (150 MHz, CD₂Cl₂) δ 145.30, 143.22, 139.39, 139.36, 139.31, 139.28, 139.22, 137.26, 134.18, 134.00, 133.92, 133.87, 133.79, 132.38, 132.28, 130.99, 128.81, 128.77, 128.68, 128.64, 128.54, 128

132.38, 132.28, 130.99, 128.81, 128.77, 128.68, 128.64, 128.54, 128.34, 127.14, 120.95, 114.98, 42.81, 31.02, 26.29, 23.93, (complexity due to P-C coupling; B-aryl carbon signal not observed); 11 B NMR (160 MHz, CD₂Cl₂) δ 46.7; 31 P NMR (202 MHz, CD₂Cl₂)

 δ –11.2; IR (ATR) 3048, 1604, 1578, 1536, 1493, 1478, 1465, 1453, 1370, 1265, 1214, 1191, 1090, 1026, 989, 814, 764 cm⁻¹; HRMS (DART) calcd for C₃₀H₃₀BNP ([M+H]⁺) 446.22292, found 446.22089.

L5: 320 mg, 68% yield.

¹H NMR (500 MHz, CD₂Cl₂) δ 7.85 (d, J = 8.5 Hz, 1H), 7.78 (d, J Me = 8.0 Hz, 1H), 7.76 (s, 1H), 7.58-7.60 (m, 2H), 7.36-7.46 (m, 4H), 7.15-7.28 (m, 11H), 6.93-6.97 (m, 1H), 4.00 (s, 3H), 1.84 (s, 3H); Me ¹¹B NMR (160 MHz, CD₂Cl₂) δ 47.9; ³¹P NMR (202 MHz, CD₂Cl₂) δ -12.2; ¹³C NMR (150 MHz, CD₂Cl₂) δ 146.66, 143.29, 139.50, 139.40, 139.31, 139.21, 136.87, 136.09, 135.96, 133.77, **L5** 133.73, 133.65, 133.61, 131.11, 130.43, 130.35, 128.83, 128.79, 128.70, 128.67, 128.52, 128.50, 127.53, 126.85, 125.70, 121.17, 115.21, 42.49, 19.41 (complexity due to P-C coupling; B-aryl carbon signal not observed); IR (ATR) 3048, 2923, 1604, 1585, 1540, 1490, 1456, 1432, 1401, 1371, 1280, 1212, 1169, 1103, 1047, 1025, 935, 895, 764 cm⁻¹; HRMS (DART) calcd for C₃₂H₂₈BNP ([M+H]⁺) 468.20524, found 468.20468.

L6: 360 mg, 75% yield.

¹H NMR (500 MHz, CD₂Cl₂) δ 7.85 (d, J = 8.5 Hz, 1H), 7.79 (d, J Me = 6.5 Hz, 1H), 7.77 (s, 1H), 7.53-7.61 (m, 2H), 7.39-7.45 (m, 3H), 7.33 (d, J = 8.0 Hz, 1H), 7.27-7.30 (m, 6H), 7.17-7.23 (m, 4H), 7.14 (dd, J = 7.5, 1.0 Hz, 1H); 6.90 (td, J = 6.0, 1.5 Hz, 1H), 4.02 (s, 3H), 2.28-2.34 (m, 1H), 2.20-2.24 (m, 1H), 0.85 (t, J = 7.5 Hz, 3H); ¹³C NMR (150 MHz, CD₂Cl₂) δ 146.23, 143.21, 139.53, **L6** 139.45, 139.32, 139.24, 137.08, 136.49, 136.36, 135.97, 133.85, 133.75, 133.73, 133.63, 131.12, 130.84, 130.55, 128.81, 128.77, 128.68, 128.64, 128.58, 128.50, 128.48, 127.50, 126.82, 125.50, 121.07, 115.14, 42.65, 27.04, 16.28 (complexity due to P-C coupling; Baryl carbon signal not observed); ¹¹B NMR (160 MHz, CD₂Cl₂) δ 48.0; ³¹P NMR (202 MHz, CD₂Cl₂) δ -10.1; IR (ATR) 3048, 2954, 2924, 2865, 1604, 1581, 1539, 1490, 1431, 1406, 1374, 1169, 1064, 864, 763 cm⁻¹; HRMS (DART) calcd for C₃₃H₃₀BNP ([M+H]⁺) 482.22089, found 482.22084.

Gram-scale synthesis of ligand L3

To a 100-mL flask charged with *o*-bromophenyldiphenylphosphine (1.71g, 5.00 mmol) and THF (50 mL) was added *n*BuLi (2.0 mL, 2.5 M in hexanes, 5.0 mmol) at -78 °C. The resulting mixture was allowed to stir at -78 °C for 2 h. 1,4-Azaborine **3b** (1.027 g, 5.000 mmol) in THF (10.0 mL) was then added. The resulting mixture was allowed to stir at -78 °C for 2 h and then room temperature for 2 h. After removal of the solvent, the residue was passed through a pad of silica gel using CH₂Cl₂ as the eluent. After removal of the solvent, the resulting crude residue was purified by column chromatography on silica gel using pentane/Et₂O (20 : 1) then Et₂O/DCM (1 : 1) as the eluent to afford ligand **L3** as a white powder (1.502 g, 71%). The characterization data are consistent with the ones obtained from the small-scale synthesis.

General procedure E for the synthesis of enynes

OPO(OEt)₂

Ar

$$then KDA, THF$$
 $-78 °C$
 Ar
 Ar
 Ar
 Ar

To a 250-mL flask charged with diisopropylamine (1.40 mL, 10.0 mmol) and THF (40 mL) was added *n*BuLi (4.0 mL, 2.5 M in hexanes, 10 mmol) at -78 °C. The resulting mixture was allowed to stir at -78 °C for 15 min. Enone (10 mmol) was then added at -78 °C in one portion. After stirring the mixture at -78 °C for 30 min, the diethylchlorophosphate was added, and the mixture was allowed to stir at room temperature for 1 h. Then, the reaction mixture was cooled to -78 °C, and freshly prepared KDA (potassium diisopropyl amide)¹³ (50 mL, 0.5 M in THF, 25.0 mmol) was added to the mixture. The mixture was allowed to stir at -78 °C for 5-30 min. At the conclusion of the reaction, the reaction was then quenched by 1.0 M aq. HCl. The organic phase was separated, and the aqueous layer was extracted with ether (3 × 50 mL). The combined organic layer was then dried over anhydrous Na₂SO₄. After removal of the solvents, the resulting crude residue was purified by column chromatography on silica gel with hexanes as the eluent to afford desired enyne.

Compounds $\mathbf{4b}^2$ [CAS: 23517-04-6], $\mathbf{4d}^2$ [CAS: 23517-05-7], $\mathbf{4f}^{14}$ [CAS: 1499245-30-5], $\mathbf{4h}^2$ [CAS: 141735-20-8], $\mathbf{4i}^{13}$ [CAS: 61172-01-8], $\mathbf{4j}^{13}$ [CAS: 72450-98-7], and $\mathbf{4k}^{13}$ [CAS: 134987-93-2] were prepared according to this general procedure E, and the characterization data are consistent with those reported in the literature.

4c [CAS: 23517-06-8], 56%. ¹H NMR (500 MHz, CDCl₃) δ Me 7.18-7.23 (m, 3H), 7.12 (d, J = 7.5 Hz, 1H), 7.02 (d, J = 16.0 Hz, 1H), 6.11 (dd, J = 16.0, 2.0 Hz, 1H), 3.04 (d, J = 2.0 Hz, 4c 1H), 2.35 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 143.3, 138.3, 135.8, 129.7, 128.6, 127.0, 123.5, 106.7, 83.0, 79.0, 21.3; IR (ATR) 3290, 3029, 2920, 2863, 2097, 1614, 1600, 1488, 1454, 1271, 954, 776 cm⁻¹; HRMS (DART) calcd for $C_{11}H_{11}$ ([M+H]⁺) 143.08608, found 143.08589.

Synthesis of enyne 4n [CAS: 2807-14-9]

Synthesis of enyne **4n** was adapted from literature procedures¹⁵. To a 50-mL flask charged with trans-2-cyclohexylvinyl iodide¹⁶ (2.36 g, 10.0 mmol), CuI (20.4 mg, 0.1 mmol), Pd(PPh₃)₂Cl₂ (15.7 mg, 0.022 mmol), and Et₂NH (20.0 mL) was added trimethylsilylacetylene (1.18 g, 12.0 mmol) slowly at 0 °C. The resulting mixture was allowed to stir at room temperature for 16 h. At the conclusion of the reaction, the reaction was quenched with H_2O (20 mL) followed by addition of hexanes (30 mL). The organic layer was separated, and the aqueous layer was extracted with hexanes (3 × 20 mL). The combined organic layer was then concentrated and the residue was dissolved in MeOH (30 mL). To the resulting methanolic solution was added KF (4.6 g, 79 mmol) and KOH (200 mg, 3.60 mmol) in one portion. The resulting mixture was allowed to stir at room temperature for 0.5 h. The reaction mixture was then diluted by H_2O (100 mL) followed by extraction with hexanes (3 × 30 mL). The combined organic layer was then dried over anhydrous Na_2SO_4 . After removal of the solvent, the residue was purified by

column chromatography on silica gel with hexanes as the eluent to afford **4n** as a colorless liquid (2.14 g, 80%).

¹H NMR (500 MHz, CDCl₃) δ 6.24 (dd, J = 16.0, 7.0 Hz, 1H), 5.44 (dt, J = 16.0, 1.5 Hz, 1H), 2.80 (d, J = 2.0 Hz, 1H), 2.03-2.09 (m, 1H), 1.66-1.78 (m, 6H), 1.09-1.29 (m, 4H); ¹³C NMR (125 MHz, CDCl₃) δ 152.2, 106.1, 82.8, 75.7, 41.2, 32.1, 26.0, 25.8; IR (ATR) 3311, 2923, 2851, 2012, 1448, 957, 842 cm⁻¹; HRMS (DART) calcd for C₁₀H₁₅ ([M+H]⁺) 135.11738, found 135.11741.

General procedure F for the synthesis of enynes

R1
$$X$$

$$X = Br, I$$

$$R^{2}$$

$$R^{2}$$

$$THF/benzene (1/1)$$

$$RT, 16 h$$

$$R^{2}$$

$$R^{1}$$

To a 100-mL flask charged with *trans*-alkenyl bromide or iodide 12,17,18 (5.0 mmol), Pd(PPh₃)₄ (6.0 mg, 0.050 mmol), and benzene (12 mL) was added alkynylmagnesium bromide (12.0 mL, 0.5 M in THF, 6.0 mmol) slowly at 0 °C. The resulting mixture was allowed to stir at room temperature for 16 h. The reaction was then quenched by 1.0 M aq. HCl (20 mL), and the organic layer was separated. The aqueous layer was then extracted with diethyl ether (3 × 30 mL). The combined organic layer was then dried over Na₂SO₄. After removal of the solvent, the resulting crude residue was purified by column chromatography on silica gel with hexanes as the eluent to afford desired enynes.

Compound **6e** was prepared according to this general procedure F, and the characterization data are consistent with those reported in the literature.¹⁹

4o [CAS: 408305-79-3]: 65%. ¹H NMR (500 MHz, CDCl₃) δ 6.24 (dt, J = 16.0, 7.0 Hz, 1H), 5.50-5.54 (m, 1H), 3.66 (t, J = 6.5 TBSO Hz, 2H), 2.79 (d, J = 2.0 Hz, 1H), 2.30-2.35 (m, 2H), 0.89 (s, **4o** 9H), 0.05 (s, 6H); ¹³C NMR (125 MHz, CDCl₃) δ 143.2, 110.3, 82.4, 75.9, 62.0, 36.6, 25.9, 18.3, -5.3; IR (ATR) 3314,2954, 2929, 2895, 2857, 2739, 2106, 1631, 1471, 1361, 1094, 957, 933, 774 cm⁻¹; HRMS (DART) calcd for $C_{12}H_{23}OSi$ ([M+H]⁺) 211.15182, found 211.15118.

4p, [CAS: 129077-84-5], 62%. ¹H NMR (500 MHz, CDCl₃) δ 6.25 (dt, J = 16.0, 7.5 Hz, 1H), 5.45 (dd, J = 16.5, 2.0 Hz, 1H), 3.60 (t, J TBSO **4p** = 7.0 Hz, 2H), 2.77 (d, J = 2.0 Hz, 1H), 2.08-2.13 (m, 2H), 1.49 **4p** 1.54 (m, 2H), 1.37-1.40 (m, 2H), 2.27 (br, 10H), 0.90 (s, 9H), 0.05 (s, 6H); ¹³C NMR (125 MHz, CDCl₃) δ 147.0, 108.4, 82.6, 75.5, 63.3, 33.0, 32.9, 29.5, 29.4, 29.3, 29.1, 28.3, 26.0, 25.8, 18.4, -5.2; IR (ATR) 3314, 2926, 2854, 1462, 1360, 1095, 1005, 956, 833, 774 cm⁻¹; HRMS (DART) calcd for $C_{19}H_{37}OSi$ ([M+H]⁺) 309.26137, found 309.26197.

6a, [CAS: 2807-15-0], 87%. ¹H NMR (500 MHz, CDCl₃) δ 6.01 (dd, J = 16.0, 7.0 Hz, 1H), 5.38 (m, 1H), 1.98-2.00 (m, 1H), 1.92 (d, J = 2.0 Hz, 3H), 1.62-1.73 (m, 5H), 1.05-1.33 (m, 5H); ¹³C **6a** NMR (125 MHz, CDCl₃) δ 148.9, 107.4, 84.2, 78.5, 41.1, 32.4, 26.0, 25.8, 4.2; IR (ATR) 3017, 2922, 2850, 2222, 1447, 955 cm⁻¹; HRMS (DART) calcd for C₁₁H₁₇ ([M+H]⁺) 149.13303, found 149.13343.

6b, [CAS: 66717-35-9], 89%. ¹H NMR (500 MHz, CDCl₃) δ 6.04 (dt, J = 16.5, 7.0 Hz, 1H), 5.40-5.44 (m, 1H), 2.05 (t, J = 7.0 n Hex Hz, 2H), 1.92 (d, J = 2.0 Hz, 3H), 1.24-1.38 (m, 8H), 0.88 (t, J = **6b** 7.0 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 143.5, 109.7, 83.9, 78.4, 32.9, 31.6, 28.8, 28.7, 22.6, 14.1, 4.2; IR (ATR) 3020, 2956, 2925, 2855, 1458, 953 cm⁻¹; HRMS (DART) calcd for C₁₁H₁₇ ([M+H]⁺) 151.14856, found 151.14868.

6h, 97%. ¹H NMR (500 MHz, CDCl₃) δ 7.25 (d, J = 6.0 Hz, 2H), 7.12 (d, J = 8.5 Hz, 2H), 6.84 (d, J = 16.5 Hz, 1H), 6.08 (dq, J = 16.0, 2.0 Hz, 1H), 2.34 (s, 3H), 2.01 (d, J H₃C **6h** = 2.5 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 140.0, 138.2, 133.8, 129.3, 125.9, 107.7, 87.8, 79.0, 21.2, 4.6; IR (FT-ATR) 3025, 2914, 2850, 2217, 1609, 1513, 1440, 1376, 955, 798, cm⁻¹; HRMS (DART) calcd. for C₁₂H₁₃ ([M+H]⁺) 157.10173, found 157.10240.

6i 93%. ¹H NMR (500 MHz, CDCl₃) δ 7.31 (dd, J = 8.0, 5.0 Hz, 2H), 7.00 (t, J = 8.5 Hz, 2H), 6.82 (d, J = 16.5 Hz, 1H), 6.04 (dq, J = 16.5, 1.5 Hz, 1H), 2.09 (d, J = 2.0 Hz, 3H); ¹³C _F **6i** NMR (125 MHz, CDCl₃) δ 162.7 (d, J = 246.8 Hz), 138.8, 132.7 (d, J = 2.9 Hz), 127.6 (d, J = 8.5 Hz), 115.8 (d, J = 21.9 Hz), 108.6 (d, J = 2.9 Hz), 88.3, 78.7, 4.4; ¹⁹F NMR (470 MHz, CDCl₃) δ -113.2; IR (FT-ATR) 3037, 2915, 2850, 2219, 1599, 1506, 1227, 1156, 952, 868, 851 cm⁻¹; HRMS (DART) calcd. for C₁₁H₁₀F ([M+H]⁺) 161.07665, found 161.07606.

6j, 75%. ¹H NMR (500 MHz, CDCl₃) δ 7.28 (s, 4H), 6.82 (d, J = 16.0 Hz, 1H), 6.10 (dq, J = 16.5, 2.0 Hz, 1H), 2.01 (d, J = 2.5 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 138.7, 135.0, 133.9, 128.8, 127.2, 109.5, 88.9, 78.7, 4.5; IR (FT-

ATR) 3032, 2914, 2848, 2215, 1615, 1592, 1490, 1089, 1011, 954, 806 cm⁻¹; HRMS (DART) calcd. for $C_{11}H_{10}^{35}Cl([M+H]^+)$ 177.04710, found 177.04736.

6k, 60%. ¹H NMR (500 MHz, CDCl₃) δ 7.01 (d, J = 8.0 Hz, 2H), 6.84 (d, J = 9.0 Hz, 2H), 6.82 (d, J = 17.0 Hz, 1H), 5.98 (dq, J = 16.5, 2.5 Hz, 1H), 3.81 (s, 3H), 2.01 (d, MeO **6k** J = 2.0 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 159.7, 139.6, 129.3, 127.3, 114.0, 106.4, 87.3, 79.1, 59.2, 4.5; IR (FT-ATR) 3030, 3001, 2955, 2934, 2912, 2835, 2217, 1604, 1509, 1440, 1275, 1173, 1030, 952, 847 cm⁻¹; HRMS (DART) calcd. for C₁₂H₁₃O ([M+H]⁺) 173.09664, found 173.09594.

6m, 73%. ¹H NMR (500 MHz, CDCl₃) δ 7.26-7.36 (m, 5H), 6.90 (d, J = 16.0 Hz, 1H), 6.18 (dt, J = 16.0, 2.5 Hz, 1H), 2.39-2.44 (m, 2H), 1.21-1.25 (m, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 140.0, **6m** 136.6, 128.6, 128.2, 126.0, 108.8, 94.2, 79.1, 13.9, 13.3; IR (FT-ATR) 3059, 3027, 2975, 2936, 2842, 2211, 1594, 1491, 1448, 1375, 1316, 952, 746, 690 cm⁻¹; HRMS (DART) calcd. for C₁₄H₁₇ ([M+H]⁺) 157.10173, found 157.10136.

6n, 80%. ¹H NMR (500 MHz, CDCl₃) δ 7.36 (d, J = 7.0 Hz, 2H), 7.31 (t, J = 7.0 Hz, 2H), 7.25 (t, J = 7.0 Hz, 1H), 6.87 (d, J = 16.5 Hz, 1H), 6.15 (dt, J = 16.0, 7.5 Hz, 1H), 2.38 (td, J = 7.0, **6n** 2.5 Hz, 2H, 1.53-1.57 (m, 2H), 1.43-1.48 (m, 2H), 0.94 (t, J = 7.0 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 139.9, 136.6, 128.6, 128.2, 126.0, 108.9, 93.0, 79.7, 30.8, 22.0, 19.3, 13.6; IR (FT-ATR) 3060, 3027, 2956, 2871, 2212, 1614, 1595, 1464, 1027, 745, 689 cm⁻¹; HRMS (DART) calcd. for C₁₄H₁₇ ([M+H]⁺) 185.13033, found 185.13310.

60, 44%. ¹H NMR (500 MHz, CDCl₃) δ 7.36 (d, J = 7.5 Hz, 2H), 7.31 (d, J = 7.5 Hz, 2H), 7.25 (t, J = 7.5 Hz, 1H), 6.87 (d, J = 16.0 Hz, 1H), 6.15 (dt, J = 16.5, 2.5 Hz, 1H), 2.36 (td, J = 60 7.5, 2.5 Hz, 2H), 1.53-1.58 (m, 2H), 1.40-1.44 (m, 2H), 1.30 (br, 8H), 0.89 (t, J = 7.5 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 139.9, 136.6, 128.6, 128.2, 126.0, 108.9, 93.1, 79.7,

31.8, 29.2, 29.1, 29.0, 28.8, 22.7, 19.6, 14.1; IR (FT-ATR) 3060, 3027, 2953, 2924, 2854, 2210, 1596, 1490, 1465, 1429, 950, 745, 689 cm $^{-1}$; HRMS (DART) calcd. for $C_{18}H_{25}$ ([M+H] $^{+}$) 241.19563, found 241.19513.

6p, 43%. ¹H NMR (500 MHz, CDCl₃) δ 7.36 (d, J = 7.5 Hz, 2H), 7.31 (t, J = 7.5 Hz, 2H), 7.25 (t, J = 7.5 Hz, 1H), 6.88 (d, J = 16.0 Hz, 1H), 6.18 (dt, J = 17.0, 2.5 Hz, 1H), 2.27 (dd, **6p** J = 6.0, 1.0 Hz, 2H), 1.84-1.90 (m, 1H), 1.02 (d, J = 7.0 Hz, 6H); ¹³C NMR (125 MHz, CDCl₃) δ 139.9, 136.6, 128.6, 128.2, 126.0, 108.9, 91.9, 80.6, 28.8, 28.2, 22.0; IR (FT-ATR) 3060, 3027, 2957, 2828, 2211, 1615, 1463, 1343, 1027, 745, 689 cm⁻¹; HRMS

6q, 86%. ¹H NMR (500 MHz, CDCl₃) δ 7.36 (d, J = 7.5 Hz, 2H), 7.31 (t, J = 7.5 Hz, 2H), 7.26 (t, J = 6.5 Hz, 1H), 6.88 (d, J = 16.0 Hz, 1H), 6.13 (dt, J = 16.5, 2.5 Hz, 1H), 3.78 (t, J = 7.5 Hz, 2H), 2.59 (td, J = 7.0, 2.0

for $C_{18}H_{27}OSi([M+H]^+)$ 287.18312, found 287.18437.

(DART) calcd. for $C_{14}H_{17}$ ([M+H]⁺) 185.13303, found 185.13370.

Hz, 1H), 3.78 (t, J = 7.5 Hz, 2H), 2.59 (td, J = 7.0, 2.0 Hz, 2H), 0.90 (s, 9H), 0.10 (s, 6H); ¹³C NMR (125 MHz, CDCl3) δ 140.4, 136.5, 128.6, 128.3, 126.1, 108.6, 89.1, 80.8, 62.0, 25.9, 24.0, 18.4, -5.2; IR (FT-ATR) 3028, 2953, 2882, 2855, 2219, 1491, 1253, 1098, 951, 876,775, 745, 689 cm⁻¹; HRMS (DART) calcd.

OTBS

General procedure G for catalytic hydroboration of terminal 1,3-enynes

To a 5-mL vial charged with 1,3-enyne **4** (0.25 mmol), the catalyst (Pd₂dba₃/L4) solution (0.05 M in CH₂Cl₂, 0.20 mL, 0.01 mmol), and CH₂Cl₂ (0.80 mL) was added HBCat (39 μ L, 0.375 mmol). The resulting mixture was allowed to stir at room temperature until completion as monitored by TLC. At the conclusion of the reaction, the crude ¹H NMR was taken to determine ratio of the *trans/cis* hydroboration adducts (using ³ $J_{H,H}$ of CH next to boron as the diagnostic tool to assign the *trans/cis* hydroboration adducts). Then, pinacol (354 mg, 3.0 mmol) in CH₂Cl₂ (3.0 mL) was introduced, and the resulting mixture was allowed to stir at room temperature for 1 hr. After removal of the solvent, the crude residue was purified by column chromatography on silica gel with (Hex/EtOAc = 100: 1) as the eluent to afford dienyl boronates **5**. The ratio of *trans/cis* hydroboration adducts for the Bpin products may slightly differ from the ratio originally observed for the Bcat intermediates (this latter ratio is shown in Table 2). The spectra (including the integration of both diastereomers in the inset) for the Bpin products are provided in the NMR collection.

5a: 30 min, EE/EZ > 98: 2, 85% yield. Crystals of **5a** suitable for single crystal X-ray diffraction analysis were grown from slow evaporation of a pentane solution at -30 °C. 1 H NMR (500 MHz, CDCl₃) δ 7.66 (dd, J = 15.5, 11.0 Hz, 1H), 7.45 (d, J = 7.5 Hz, 2H), 7.34 (t, J = 7.5 Hz, 2H), 7.25 (t, J = 7.5 Hz, 1H), 7.02 (t J = 12.0 Hz, 1H), 6.65 (d, J = 15.5 Hz, 1H), 5.47 (d, J = 13.0 Hz, 1H), 1.33 (s, 12H); 13 C NMR (125 MHz, CDCl₃) δ 150.3, 137.2, 136.3, 129.4, 128.6, 127.9, 126.9, 119.5 (br), 83.1, 24.9; 11 B NMR (160 MHz, CDCl₃) δ 29.6; IR (ATR) 2978, 2930, 1624, 1587, 1571, 1451, 1423, 1379, 1330, 1279, 1258, 1215, 1113, 1007, 964, 782 cm⁻¹; HRMS (DART) calcd for C₁₆H₂₂BO₂ ([M+H]⁺) 257.17128, found 257.17055.

5b: 30 min, EZ/EE > 98 : 2, 86% yield. ¹H NMR (500 MHz, CDCl₃) δ 7.62 (ddd, J = 16.0, 11.0, 1.0 Hz, 1H), 7.34 (d, J = 8.0 Hz, 2H), 7.15 (d, J = 8.0 Hz, 2H), 7.01 (t, J = 12.0 **5b** Hz, 1H), 6.63 (d, J = 11.0 Hz, 1H), 5.42 (d, J = 13.0 Hz, 1H), 2.35 (s, 3H), 1.32 (s, 12H); ¹³C NMR (125 MHz, CDCl₃) δ 150.6, 137.8, 136.4, 134.4, 129.3, 128.5, 126.8, 119.3 (br), 83.1, 24.9, 21.3; ¹¹B NMR (160 MHz, CDCl₃) δ 29.3; IR (ATR) 2978, 2926, 1624, 1586, 1297, 1279, 1113, 1106, 965, 878, 846 cm⁻¹; HRMS (DART) calcd for C₁₇H₂₄BO₂ ([M+H]⁺) 271.18693, found 271.18643.

5c: 30 min, EZ/EE = 98: 2, 82% yield. ¹H NMR (500 MHz, CDCl₃) δ 7.65 (dd, J = 16.0, 11.5 Hz, 1H), 7.22-7.29 (m, 3H), ⁸Cpin) 7.07 (d, J = 7.5 Hz, 1H), 7.02 (t, J = 13.0 Hz, 1H), 6.63 (d, J = 13.0 Me **5c** 11.0 Hz, 1H), 5.46 (d, J = 13.0 Hz, 1H), 2.36 (s, 3H), 1.33 (s, 12H); ¹³C NMR (125 MHz, CDCl₃) δ 150.4, 138.4, 137.1, 136.5, 129.2, 128.7, 128.4, 127.8, 123.8, 119.4 (br), 83.1, 24.9; ¹¹B NMR (160 MHz, CDCl₃) δ 29.3; IR (ATR) 2980, 2931, 1620, 1597, 1587, 1421, 1402, 1372, 1316, 1279, 1138, 975, 868, 845 cm⁻¹; HRMS (DART) calcd for $C_{17}H_{24}BO_{2}$ ([M+H]⁺) 271.18693, found 271.18597.

5d: 30 min, EZ/EE = 98: 2, 80% yield. ¹H NMR (500 MHz, CDCl₃) δ 7.58 (dd, J = 15.5, 11.0 Hz, 1H), 7.54 (d, J = 7.5 Hz, Me B(pin) 1H), 7.15-7.21 (m, 3H), 7.08 (d, J = 11.5 Hz, 1H), 6.86 (d, J = 5d 15.5 Hz, 1H), 5.47 (d, J = 13.0 Hz, 1H), 2.38 (s, 3H), 1.31 (s, 12H); ¹³C NMR (125 MHz, CDCl₃) δ 150.7, 136.0, 135.9, 134.0, 130.6, 130.4, 127.7, 126.1, 125.8, 119.7 (br), 83.1, 24.9; ¹¹B NMR (160 MHz, CDCl₃) δ 29.3; IR (ATR) 2979, 2931, 1624, 1594, 1579, 1507, 1431, 1406, 1390, 1371, 1279, 1231, 1113, 1006, 965, 875, 860 cm⁻¹; HRMS (DART) calcd for C₁₇H₂₄BO₂ ([M+H]⁺) 271.18693, found 271.18753.

5e: 30 min, EZ/EE > 98: 2, 76% yield. ¹H NMR (500 MHz, CDCl₃) δ 7.54 (ddd, J = 16.0, 11.5, 1.0 Hz, 1H), 7.40 (dd, J = 8.5, 3.0 Hz, 2H), 7.00 (t, J = 12.5 Hz, 1H), **Se**

6.89 (dd, J = 8.5, 3.0 Hz, 2H), 6.60 (d, J = 15.5 Hz, 1H), 5.41 (d, J = 13.0 Hz, 1H), 3.82 (s, 3H), 1.32 (s, 12H); ¹³C NMR (125 MHz, CDCl3) δ 159.5, 150.7, 136.0, 130.0, 128.1, 127.5, 118.4 (br), 114.1, 83.0, 55.3, 24.9; ¹¹B NMR (160 MHz, CDCl₃) δ 29.4; IR (FT-ATR) 2994, 2976, 2935, 2837, 1624, 1605, 1584, 1570, 1510, 1463, 1414, 1389, 1371, 1329, 1329, 1297, 1245, 1174, 1141, 1111, 1030, 1107, 964, 877, 846 cm⁻¹; HRMS (DART) calcd. for C₁₇H₂₄BO₃ ([M+H]⁺) 287.18185, found 287.28239.

5f: 30 min, EZ/EE > 98: 2, 88% yield. ¹H NMR (500 MHz, CDCl₃) δ 7.57 (dd, J = 15.5, 11.0 Hz, 1H), 7.39-7.42 (m, 2H), 6.97-7.05 (m, 3H), 6.62 (d, J = 15.5 Hz, 1H), 5.46 (d, J = 13.0 Hz, 1H), 1.33 (s, 12H); ¹³C NMR (125 MHz, CDCl₃) δ 162.5 (d, J = 246.6 Hz), 150.1, 135.0, 133.4 (d, J = 3.7 Hz), 129.1 (d, J = 1.0 Hz), 128.3 (d, J = 11.0 Hz), 119.7 (br), 115.6 (d, J = 21.7 Hz), 83.1, 24.9; ¹¹B NMR (160 MHz, CDCl₃) δ 29.6; ¹⁹F NMR (470 MHz, CDCl₃) δ -113.7; IR (ATR) 2979, 2932, 1619, 1587, 1566, 1422, 1371, 1258, 1141, 1113, 965, 880, 848 cm⁻¹; HRMS (DART) calcd for C₁₆H₂₁BFO₂ ([M+H]⁺) 274.15404, found 274.15335.

5g: 2.5 hr, EZ/EE = 98 : 2, 82% yield. ¹H NMR (500 MHz, CDCl₃) δ 7.62 (ddd, J = 15.5, 11.0, 1.0 Hz, 1H), 7.35 (dt, J = 12.0 Hz, 2H), 7.31 (dt, J = 8.0, 2.5 Hz, 2H), 7.00 (t, J = 12.0 Hz, 1H), 6.60 (d, J = 14.5 Hz, 1H), 5.49 (d, J = 13.5 Hz, 1H), 1.33 (s, 12H); ¹³C NMR (125 MHz, CDCl₃) δ 149.9, 135.7, 134.8, 133.5, 129.9, 128.8, 128.0, 120.7 (br), 83.2, 24.9; ¹¹B NMR (160 MHz, CDCl₃) δ 29.2; IR (ATR) 2978, 2928, 1623, 1584, 1490, 1430, 1371, 1297, 1216, 1192, 1165, 1143, 1113, 1091, 1011, 965, 876, 846 cm⁻¹; HRMS (DART) calcd for C₁₆H₂₁B³⁵ClO₂ ([M+H]⁺) 291.13231, found 291.13168.

5h: 2.5 hr, EZ/EE = 98 : 2, 76% yield. ¹H NMR (500 MHz, CDCl₃) δ 7.64 (ddd, J = 15.5, 10.5, 1.0 Hz, 1H), 7.46 (dt, J = Br B(pin) 9.5, 2.5 Hz, 2H), 7.30 (dt, J = 9.5, 2.5 Hz, 2H), 6.99 (d, J = 5h 12.0 Hz, 1H), 6.57 (d, J = 15.0 Hz, 1H), 5.50 (d, J = 13.5 Hz, 1H), 1.32 (s, 12H); ¹³C NMR (125 MHz, CDCl₃) δ 149.9, 136.2, 134.9, 131.7, 130.0, 128.3, 121.7, 120.5 (br),

81.2, 24.9; 11 B NMR (160 MHz, CDCl₃) δ 29.3; IR (ATR) 2977, 2930, 1621, 1591, 1428, 1280, 1164, 1131, 1112, 1007, 964, 876, 864 cm⁻¹; HRMS (DART) calcd for $C_{17}H_{24}B^{79}BrO_2([M+H]^+)$ 335.08180, found 335.08263.

5i: 45 min, EZ/EE > 98: 2, 80% yield. ¹H NMR (500 MHz, CDCl₃) δ 7.76-7.81 (m, 5H), 7.68 (dd, J = 8.0, 1.0 Hz, 1H), 7.42-7.48 (m, 2H), 7.08 (t, J = 12.5 Hz, 1H), 6.82 (d, J = **5i** 15.5 Hz, 1H), 5.51 (d, J = 12.0 Hz, 1H), 1.35 (s, 12H); ¹³C NMR (125 MHz, CDCl₃) δ 150.4, 136.4, 134.8, 133.6, 133.2, 129.7, 128.2, 128.1, 127.6, 127.3, 126.2, 126.0, 123.6, 119.6 (br), 83.1, 24.9; ¹¹B NMR (160 MHz, CDCl₃) δ 29.4; IR (FT-ATR) 3057, 2977, 2931, 1614, 1600, 1583, 1506, 1444, 1370, 1329, 1261, 1142, 1016, 838 cm⁻¹; HRMS (DART) calcd. for C₂₀H₂₄BO₂ ([M+H]⁺) 307.18693, found 307.18708.

5j: 45 min, EZ/EE > 98: 2, 81% yield. ¹H NMR (500 MHz, CDCl₃) δ 8.18 (d, J = 8.0 Hz, 1H), 7.86 (dd, J = 8.0, 1.5 Hz, 1H), 7.72-7.81 (m, 3H), 7.48-7.55 (m, 3H), 7.44 (d, J = 15.5 Hz, 1H), 7.18 (t, J = 12.0 Hz, 1H), 5.55 (d, J = 14.0 Hz, 1H),

1.33 (s, 12H); 13 C NMR (125 MHz, CDCl₃) δ 150.5, 134.4, 133.7, 133.0, 132.0, 131.2, 128.6, 128.3, 126.1, 125.8, 125.6, 123.9, 123.6, 120.1 (br), 83.2, 24.9; 11 B NMR (160 MHz, CDCl₃) δ 29.3; IR (FT-ATR) 3054, 2977, 2928, 1612, 1582, 1509, 1424, 1390, 1370, 1332, 1298, 1279, 1165, 1112, 965, 879 cm⁻¹; HRMS (DART) calcd. for $C_{20}H_{24}BO_{2}$ ([M+H]⁺) 307.18693, found 307.18717

5k: 30 min, EZ/EE = 97: 3, 81% yield. ¹H NMR (500 MHz, CDCl₃) δ 7.50 (dd, J = 16.5, 11.5 Hz, 1H), 7.21 (d, J = 5.5 Hz, B(pin) 1H), 7.03 (d, J = 3.5 Hz, 1H), 6.98 (t, J = 5.0 Hz, 1H), 6.95 (t, J **5k** = 12.0 Hz, 1H), 6.76 (d, J = 16.0 Hz, 1H), 5.44 (d, J = 13.5 Hz, 1H), 1.32 (s, 12H); ¹³C NMR (125 MHz, CDCl₃) δ 149.6, 142.7, 129.3, 128.8, 127.6, 126.5, 125.1, 119.3 (br), 83.1, 24.9; ¹¹B NMR (160 MHz, CDCl₃) δ 29.2; IR (ATR) 2977, 2928, 1612, 1584, 1514, 1434, 1417, 1351, 1329, 1297, 1257, 1214, 1193, 1164, 1112, 1004, 966, 954, 880, 866 cm⁻¹; HRMS (DART) calcd for C₁₄H₂₀BO₂S ([M+H]⁺) 263.12771, found 263.12744.

5l: 45 min, EZ/EE = 97: 3, 84% yield. ¹H NMR (500 MHz, CDCl₃) δ 7.51 (dd, J = 11.5, 1.5 Hz, 1H), 7.41 (d, J = 1.5 HZ, O B(pin) 1H), 6.94 (t, J = 12.0 Hz, 1H), 6.44 (d, J = 11.0 Hz, 1H), 6.39- **5l** 6.41 (m, 1H), 6.35 (d, J = 3.0 Hz, 1H), 5.44 (d, J = 13.5 Hz, 1H), 1.32 (s, 12H); ¹³C NMR (125 MHz, CDCl₃) δ 153.1, 149.8, 142.6, 127.8, 123.7, 119.8 (br), 111.7, 109.5, 83.1, 24.8; ¹¹B NMR (160 MHz, CDCl₃) δ 29.3; IR (ATR) 2978, 2929, 1599, 1546, 1481, 1389, 1259, 1185, 1112, 1013, 965, 882, 846 cm⁻¹; HRMS (DART) calcd for C₁₄H₂₀BO₂ ([M+H]⁺) 247.15055, found 247.15100.

5m: 30 min, EZ/EE = 94 : 6, 82%. ¹H NMR (500 MHz, CDCl₃) 8 6.80-6.88 (m, 2H), 5.81-5.87 (m. 1H), 5.24 (d, J = 11.0 Hz, 1H), 5.15 (q, J = 7.0 Hz, 2H), 1.36-1.43 (m, 2H), 1.27-1.34 (m, 18H), 5m 0.89 (t, J = 6.5 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) 8 151.0, 140.1, 130.7, 118.5 (br), 82.9, 32.7, 31.7, 29.0, 28.9, 24.8, 22.6, 14.1; ¹¹B NMR (160 MHz, CDCl₃) 8 29.3; IR (FT-ATR) 2977, 2954, 2929, 2857, 1597, 1471, 1462, 1452, 1420, 1389, 1371, 1289, 1254, 1212, 1143, 1089, 1005, 976, 965, 935, 863, 744 cm⁻¹; HRMS (DART) calcd. for $C_{16}H_{30}BO_{2}([M+H]^{+})$ 265.23388, found 265.23352.

5n: 30 min, EZ/EE = 97: 3, 87% yield. ¹H NMR (500 MHz, CDCl₃) δ 6.79-6.85 (m, 2H), 5.75-5.79 (m, 1H), 5.25 (d, J = 12.0 B(pin) Hz, 1H), 2.06-2.10 (m, 1H), 1.64-1.76 (m, 5H), 1.07-1.29 (m, 17H); ¹³C NMR (125 MHz, CDCl₃) δ 151.3, 145.6, 128.3, 116.5 (br), 82.9, 40.8, 32.5, 26.1, 25.9, 24.9; ¹¹B NMR (160 MHz, CDCl₃) δ 29.3; IR (FT-ATR) 2978, 2924, 2851, 1639, 1590, 1445, 1427, 1389, 1297, 1145, 1007, 966, 768 cm⁻¹; HRMS (DART) calcd. for C₁₆H₂₈BO₂ ([M+H]⁺) 263.21823, found 263.21808.

50: 30 min, EZ/EE > 98: 2, 86% yield. ¹H NMR (500 MHz, CDCl₃) δ 6.88-6.93 (m, 1H), 6.82 (t, J = 12.0 Hz, 1H), 5.80- TBSO TBSO 5.86 (m, 1H), 5.28 (d, J = 13.5, 1H), 3.69 (t, J = 7.0 Hz, 2H), 50 2.14 (q, J = 7.0 Hz, 2H), 1.28 (s, 12H), 0.88 (s, 9H), 0.05 (s, 6H); ¹³C NMR (125 MHz,

CDCl₃) δ 150.7, 135. 7, 132.5, 117.3 (br), 82.9, 62.7, 36.4, 25.9, 24.8, 18.3, -5.3; ¹¹B NMR (160 MHz, CDCl₃) δ 29.2; IR (ATR) 2979, 2954, 2929, 1643, 1590, 1471, 1424, 1300, 1257, 1215, 1145, 1097, 1006, 966, 937, 876, 835 cm⁻¹; HRMS (DART) calcd for $C_{18}H_{36}BO_{3}Si([M+H]^{+})$ 339.25268, found 339.25257.

5p: 30 min, EZ/EE = 96: 4, 91% yield. ¹H NMR (500 MHz, (H₂C)₉) CDCl₃) δ 6.82-6.86 (m, 2H), 5.81-5.87 (m, 1H), 5.24 (d, J = TBSO' B(pin) 12.0, 1H), 3.60 (t, J = 6.5 Hz, 2H), 2.14 (q, J = 7.0 Hz, 2H), 5p 1.47-1.54 (m, 2H), 1.38-1.43 (m, 2H), 1.28 (m, 22H), 0.89 (s, 9H), 0.05 (s, 6H); ¹³C NMR (125 MHz, CDCl₃) δ 151.0, 140.1, 130.7, 116.8 (br), 82.9, 63.3, 32.9, 32.7, 29.6, 29.5, 29.4, 29.2, 29.0, 26.0, 25.8, 24.8, 18.3, -5.3; ¹¹B NMR (160 MHz, CDCl₃) δ 29.2; IR (ATR) 2978, 2926, 2854, 1641, 1589, 1463, 1424, 1388, 1378, 1329, 1299, 1255, 1215, 1144, 1096, 1006, 965, 879, 773 cm⁻¹; HRMS (DART) calcd for C₂₅H₅₀BO₃Si ([M+H]⁺) 437.36223, found 437.36140.

5q: 30 min, EZ/EE = 95: 5, 93% yield. ¹H NMR (500 MHz, Me CDCl₃) δ 7.06 (dd, J = 16.0, 11.5 Hz, 1H), 6.90 (t, J = 12.0 Hz, 1H), 6.30 (d, J = 11.0 Hz, 1H), 5.31 (d, J = 13.5 Hz, 1H), 2.06 (t, J = 6.5 Hz, 2H), 1.77 (d, J = 1.0 Hz, 3H), 1.59-1.64 (m, 2H), 1.46-1.48 (m, 2H), 1.30 (s, 12H), 1.08 (s, 6H); ¹³C NMR (125 MHz, CDCl₃) δ 151.8, 137.0, 135.2, 132.9, 132.0, 116.5 (br), 82.8, 40.2, 34.0, 33.7, 28.9, 24.8, 24.5, 21.7, 19.2; ¹¹B NMR (160 MHz, CDCl₃) δ 29.3; IR (FT-ATR) 2976, 2928, 2865, 2824, 1607, 1580,

1424, 1389, 1370, 1327, 1297, 1257, 1212, 1164, 1143, 1113, 1009, 967, 846, 771 cm⁻¹;

HRMS (DART) calcd. for $C_{19}H_{32}BO_2([M+H]^+)$ 303.24953, found 235.25009.

5r²⁰ [CAS: 1192488-91-7]: 30 min, EZ/EE = 97: 3, 70% yield. ¹H NMR (500 MHz, CDCl₃) δ 6.65 (d, J = 14.5 Hz, 1H), 5.83 (t, J = 5.0 B(pin) Hz, 1H), 5.18 (d, J = 15.0 Hz, 1H), 2.24-2.27 (m, 2H), 2.11-2.15 (m, **5r** 2H), 1.54-1.66 (m, 4H), 1.30 (s, 12H); ¹³C NMR (125 MHz, CDCl₃) δ 149.7, 137.9, 132.0, 114.1 (br), 83.3,26.3, 26.0, 24.8, 22.4, 22.1; ¹¹B NMR (160 MHz, CDCl₃) δ 30.5; IR (FT-ATR) 2977, 2927, 2858, 2831, 1628, 1598, 1434, 1389, 1370, 1298, 1228, 1141,

1107, 1004, 966, 918, 845, 801, 775 cm⁻¹; HRMS (DART) calcd. for $C_{14}H_{24}BO_2$ ([M+H]⁺) 235.18693, found 235.18714.

General procedure H for catalytic hydroboration of internal 1,3-enynes

To a 5-mL vial charged with 1,3-envne 6 (0.25 mmol) and the catalyst (Pd₂dba₃/L3) solution (0.05 M in CH₂Cl₂, 0.20 mL, 0.01 mmol) was added HBCat (39 µL, 0.375 mmol). The resulting mixture was allowed to stir at room temperature until completion as monitored by TLC. At the conclusion of the reaction, the crude ¹H NMR was taken to determine ratio of the trans/cis hydroboration adducts (using chemical shift of CH next to R^1 (7a-71), or CH_2 next to CB (7m-7q) as the diagnostic tool to assign the *trans/cis* hydroboration adducts). Then, pinacol (354 mg, 3.0 mmol) in CH₂Cl₂ (3.0 mL) was introduced, and the resulting mixture was allowed to stir at room temperature 1 hr. After removal of the solvent, the residue was purified by column chromatography on silica gel with (Hex/EtOAc = 100: 1) as the eluent to afford dienyl boronates 7. The ratio of trans/cis hydroboration adducts for the Bpin products may slightly differ from the ratio originally observed for the Bcat intermediates (this latter ratio is shown in Table 3). The spectra (including the integration of both diastereomers in the inset) for the Bpin products are provided in the NMR collection. 1D-NOE experiments were performed for 7a, 7g, cis-7g (independently synthesized)²¹, 7l, and 7g as representative compounds. The 1D-NOE spectra for 7a, 7g, 7l, and 7q are consistent with *trans*-hydroboration.

7a: 1.5 hr, EE/EZ = 96: 4, 92% yield. Crystals of **7a** suitable for single crystal X-ray diffraction analysis were grown from slow evaporation of a pentane solution at -30 °C. 1 H NMR (500 MHz, 7 a CDCl₃) δ 6.79 (ddd, J = 15.5, 11.0, 1.0 Hz, 1H), 6.56 (d, J = 10.5 Hz, 1H), 5.63 (dd J = 15.5, 7.0 Hz, 2H), 1.98-2.10 (m, 1H), 1.82 (s, 3H), 1.70-1.75 (m, 4H), 1.63-1.66 (m, 1H), 1.08-1.29 (m, 17H); 13 C NMR (125 MHz, CDCl₃) δ 146.1, 142.4, 127.6, 82.9, 40.7, 32.7, 26.2, 25.9, 24.9, 22.3 (B-alkenyl carbon signal not observed); 11 B NMR (160 MHz,

CDCl₃) δ 29.7; IR (ATR) 2977, 2923, 2850, 1634, 1593, 1449, 1421, 1400, 1371, 1343, 1290, 1212, 1085, 975, 865, 672 cm⁻¹; HRMS (DART) calcd for $C_{17}H_{30}BO_2$ ([M+H]⁺) 277.23388, found 277.23525.

7b: 30 min, EE/EZ = 93: 7, 80% yield. ¹H NMR (500 MHz, CDCl₃) δ 6.82 (dd, J = 15.0, 10.5 Hz, 1H), 6.56 (d, J = 10.5 Hz, 1H), 5.69 (dt, J = 15.0, 7.0 Hz, 1H), 2.11 (q, J = 7.0 Hz, 2H), 1.82 **7b** (s, 3H), 1.36-1.42 (m, 2H), 1.23-1.32 (m, 18H), 0.89 (t, J = 7.0 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 145.8, 136.8, 130.0, 82.9, 32.7, 31.8, 29.2, 28.9, 24.9, 22.6, 22.4, 14.1 (B-alkenyl carbon signal not observed); ¹¹B NMR (160 MHz, CDCl₃) δ 29.7; IR (ATR) 2977, 2956, 2925, 2854, 1640, 1596, 1453, 1421, 1389, 1371, 1267, 1254, 1214, 1143, 1110, 1093, 975, 965, 836, 686 cm⁻¹; HRMS (DART) calcd. for C₁₇H₃₂BO₂ ([M+H]⁺) 279.24953, found 279.25033.

7c: 2 hr, EE/EZ = 82: 18, 93% yield. ¹H NMR (500 MHz, CDCl₃) 6 6.80 (dd, J = 15.5, 11.5 Hz, 1H), 6.53 (d, J = 11.5 Hz, 1H), 5.71 6 (dt, J = 15.0, 8.0 Hz, 1H), 2.08-2.17 (m, 4H), 1.37-1.43 (m, 2H), 6 (m, 18H), 1.00 (t, J = 7.0 Hz, 3H), 0.88 (t, J = 7.0 Hz, 3H); 13 C NMR (125 MHz, CDCl₃) 6 143.9, 136.9, 130.1, 82.9, 32.8, 31.8, 29.8, 29.1, 28.9, 24.8, 22.6, 14.8, 14.1; 11 B NMR (160 MHz, CDCl₃) 6 30.0; IR (ATR) 2958, 2925, 2855, 1639, 1591, 1458, 1425, 1405, 1388, 1378, 1290, 1214, 1143, 1109, 967, 833, 673 cm⁻¹; HRMS (DART) calcd for $C_{18}H_{34}BO_{2}$ ([M+H]⁺) 293.26594, found 293.26518.

7d: 2 hr, EE/EZ = 81: 19, 77% yield. ¹H NMR (500 MHz, CDCl₃) δ 6.72-6.82 (m, 1H), 6.51 (d, J = 11.5 Hz, 1H), 5.69-5.73 (m, 1H), 2.13 (t, J = 7.5 Hz, 2H), 1.78 (d, J = 7.0 Hz, 3H), 1.25-1.38 (m, 7d 16H), 0.88 (t, J = 7.5 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 144.5, 131.5, 131.2, 82.9, 34.5, 32.6, 24.8, 22.3, 18.3, 14.0 (B-alkenyl carbon signal not observed); ¹¹B NMR (160 MHz, CDCl₃) δ 29.9; IR (ATR) 2977, 2956, 2929, 2871, 2858, 1642, 1591, 1466, 1424, 1405, 1378, 1301, 1285, 1245, 1213, 1144, 1111, 978, 966, 864, 703 cm⁻¹; HRMS (DART) calcd for C₁₅H₂₈BO₂ ([M+H]⁺) 251.21823, found 251.21859.

7e: 30 min, EE/EZ = 92: 8, 84% yield. ¹H NMR (500 MHz, CDCl₃) δ 6.87 (dd, J = 15.0, 10.5 Hz, 1H), 6.56 (d, J = 11.0 Hz, 1H), 5.67 (dt, J = 15.5, 7.0 Hz, 1H), 3.66 (t, J = 11.0 Hz, 2H), 2.35 (q, J = 7.0 Hz, 2H), 1.83 (s, 3H), 1.28 (s, 12H), 0.89 (s, 9H), 0.06 (s, 6H); ¹³C NMR (125 MHz, CDCl₃) δ 145.5, 132.2, 131.9, 83.0, 63.0, 36.4, 26.9, 24.8, 22.4, 18.3, -5.3 (B-alkenyl carbon signal not observed); ¹¹B NMR (160 MHz, CDCl₃) δ 29.7; IR (ATR) 2978, 2957, 2925, 2855, 1641, 1589, 1466, 1424, 1389, 1279, 1007, 965, 879, 847 cm⁻¹; HRMS (DART) calcd for C₁₉H₃₈BO₃Si ([M+H]⁺) 353.26833, found 353.26890.

7f: 30 min, EE/EZ = 94: 6, 89% yield. ¹H NMR (500 MHz, CDCl₃) δ 6.81 (dd, J = 15.0, 10.5 Hz, 1H), 6.56 (d, J = 10.0 TBSO(H₂C)₉ 7f B(pin) Hz, 1H), 5.69 (dt, J = 15.0, 7.0 Hz, 1H), 3.59 (t, J = 7.0 Hz, 2H), 2.11 (q, J = 7.5 Hz, 2H), 1.82 (s, 3H), 1.49-1.55 (m, 2H), 1.36-1.39 (m, 2H), 1.23-1.29 (m, 22H), 0.89 (s, 9H), 0.05 (s, 6H); ¹³C NMR (125 MHz, CDCl₃) δ 145.9, 136.8, 130.0, 82.9, 63.3, 32.9, 32.7, 29.6, 29.5, 29.4, 29.2, 25.7, 25.8, 24.9, 22.4, 18.3, -5.3 (Balkenyl carbon signal not observed); ¹¹B NMR (160 MHz, CDCl₃) δ 30.0; IR (ATR) 2978, 2926, 2854, 1640, 1596, 1462, 1421, 1400, 1389, 1253, 1214, 1144, 1094, 1034, 1005, 975, 835, 812 cm⁻¹; HRMS (DART) calcd for C₂₆H₅₂BO₃Si ([M+H]⁺) 451.37788, found 451.37929.

7g: 30 min, EE/EZ = 97: 3, 79% yield. ¹H NMR (500 MHz, CDCl₃) δ Me 6.31 (s, 1H), 5.65 (s, 1H), 2.15 (br, 2H), 2.08 (br, 2H), 1.83 (s, 3H), 7g 1.54-1.63 (m, 4H), 1.28 (s, 12H); ¹³C NMR (125 MHz, CDCl₃) δ 7g 143.1, 137.2, 127.7, 83.3, 27.1, 25.7, 24.7, 23.1, 22.6, 22.2 (B-alkenyl carbon signal not observed); ¹¹B NMR (160 MHz, CDCl₃) δ 31.0; IR (ATR) 2977, 2927, 2857, 2832, 1608, 1446, 1398, 1389, 1221, 1213, 1189, 1143, 1107, 963, 920, 670 cm⁻¹; HRMS (DART) calcd for $C_{15}H_{26}BO_{2}$ ([M+H]⁺) 249.20258, found 249.20367.

7h: 15 min, EE/EZ > 98: 2, 87% yield. ¹H NMR (500 MHz, CDCl₃) δ 7.63 (dd, J = 15.5, 11.0 Hz, 1H), 7.32 (d, J = 7.5 Hz, 2H), 7.14 (d, J = 7.5 Hz, 2H), 6.77 (d, J = 11.0 Hz, Th) (1H), 6.51 (d, J = 15.5 Hz, 1H), 2.35 (s, 3H), 1.93 (s, 3H),

1.34 (s, 12H); 13 C NMR (125 MHz, CDCl₃) δ 145.6, 137.1, 135.1, 133.4, 129.2, 128.3, 126.4, 83.1, 24.9, 22.6, 21.2 (B-alkenyl carbon signal not observed); 11 B NMR (160 MHz, CDCl₃) δ 29.7; IR (FT-ATR) 2976, 2883, 1620, 1587, 1508, 1450, 1397, 1289, 1136, 1034, 1014, 964, 805 cm⁻¹; HRMS (DART) calcd for $C_{18}H_{26}BO_2$ ([M+H]⁺) 285.20258, found 285.20216.

7i: 15 min, EE/EZ > 98: 2, 87% yield. ¹H NMR (500 MHz, CDCl₃) δ 7.58 (dd, J = 1.5.5, 10.5 Hz, 1H), 7.35-7.38 (m, 2H), 7.01 (t, J = 8.5 Hz, 2H), 6.74 (d, J = 11.0 Hz, 1H), 6.48 7i (d, J = 15.5 Hz, 1H), 1.92 (s, 3H), 1.33 (s, 12H); ¹³C NMR (125 MHz, CDCl₃) δ 161.2 (d, J = 245.8 Hz), 145.2, 134.0 (d, J = 3.8 Hz), 132.0, 128.9, 127.9 (d, J = 8.5 Hz, 1H), 115.5 (d, J = 20.9 Hz), 83.2, 24. 9. 22.7 (B-alkenyl carbon signal not observed); ¹¹B NMR (160 MHz, CDCl₃) δ 29.7; ¹⁹F NMR (470 MHz, CDCl₃) δ -114.6; IR (ATR) 2978, 2933, 1621, 1596, 1581, 1506, 1451, 1396, 1318, 1245, 1229, 1140, 1110 1090, 966, 857, 817, 774, 685 cm⁻¹; HRMS (DART) calcd for C₁₇H₂₃BFO₂ ([M+H]⁺) 289.17751, found 289.17695.

7j: 15 min, EE/EZ > 98: 2, 85% yield. ¹H NMR (500 MHz, CDCl₃) δ 7.63 (dd, J = 15.5, 11.0 Hz, 1H), 7.26-7.33 (m, 4H), 6.74 (d, J = 10.5 Hz, 1H), 6.45 (d, J = 15.5 Hz, 1H), 7j
1.93 (s, 3H), 1.33 (s, 12H); ¹³C NMR (125 MHz, CDCl₃) δ 145.0, 136.4, 132.8, 131.9, 129.7, 128.7, 127.6, 83.2, 24.9, 22.7 (B-alkenyl carbon signal not observed); ¹¹B NMR (160 MHz, CDCl₃) δ 29.7; IR (ATR) 3046, 3001, 2885, 1619, 1595, 1584, 1448, 1406, 1390, 1371, 1284, 1246, 1165, 1136, 1109, 1088, 1008, 964, 864, 683 cm⁻¹; HRMS (DART) calcd for C₁₇H₂₃B³⁵ClO₂ ([M+H]⁺) 305.14796, found 305.14837.

7k: 15 min, EE/EZ > 98: 2, 90% yield. ¹H NMR (500 MHz, CDCl₃) δ 7.55 (dd, J = 15.5, 10.5 Hz, 1H), 7.36 (d, J = 8.5 Hz, 2H), 6.87 (d, J = 8.5 Hz, 2H), 6.75 (d, J = 11.0 Hz, 1H), 6.48 (d, J = 15.5 Hz, 1H), 3.82 (s, 3H), 1.92 (s, 3H), 1.34 (s,

12H); ¹³C NMR (125 MHz, CDCl₃) δ 159.1, 141. 8, 133.0, 130.7, 127.7, 127.2, 114.0, 83.1, 55.2, 24.9, 22.6 (B-alkenyl carbon signal not observed); ¹¹B NMR (160 MHz, CDCl₃) δ 29.8; IR (ATR) 2974, 2928, 2885, 2836, 1620, 1605, 1588, 1447, 1371, 1297, 1267, 1252, 1211, 1166, 1136, 1109, 1091, 1047, 977, 684 cm⁻¹; HRMS (DART) calcd for C₁₈H₂₆BO₃ ([M+H]⁺) 301.19750, found 301.19769.

7I: 15 min, EE/EZ > 98: 2, 88% yield. ¹H NMR (500 MHz, CDCl₃) δ 7.66 (dd, J = 16.0, 11.5 Hz, 1H), 7.41 (d, J = 7.5 Hz, 2H, 7.32 (t, J = 7.5 Hz, 2H), 7.21 (t, J = 7.5 Hz, 1H), 6.76 (d, J = 71 11.0 Hz, 1H), 6.52 (d, J = 15.5 Hz, 1H), 1.92 (s, 3H), 1.33 (s, 12H); ¹³C NMR (125 MHz, CDCl₃) δ 145.4, 137.8, 133.4, 129.2, 128.5, 127.3, 126.5, 83.2, 24.9, 22.7 (B-alkenyl carbon signal not observed); ¹¹B NMR (160 MHz, CDCl₃) δ 29.7; IR (FT-ATR) 2982, 2942, 2886, 1618, 1597, 1586, 1452, 1422, 1403, 1391, 1372, 1353, 1316, 1290, 1277, 1246, 1206, 1167, 1136, 1112, 1088, 1073, 976, 965, 869, 755, 682 cm⁻¹; HRMS (DART) calcd. for C₁₇H₂₄BO₂ ([M+H]⁺) 271.18693, found 271.18705.

7m: 3.5 hr, EE/EZ = 95 : 5, 96% yield. ¹H NMR (500 MHz, CDCl₃) δ 7.64 (dd, J = 15.5, 11.0 Hz, 1H), 7.40 (d, J = 7.0 Hz, Tm B(pin) 2H), 7.31 (t, J = 7.0 Hz, 2H), 7.21 (t, J = 7.5 Hz, 1H), 6.73 (d, J = 11.0 Hz, 1H), 6.54 (d, J = 16.0 Hz, 1H), 2.25 (q, J = 7.0 Hz, 2H), 1.34 (s, 12H), 1.05 (t, J = 7.5 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 143.5, 137.9, 133.5, 129.3, 128.5, 127.3, 126.5, 83.1, 30.0, 24.9, 14.7 (B-alkenyl carbon signal not observed); ¹¹B NMR (160 MHz, CDCl₃) δ 30.0; IR (ATR) 2975, 2930, 2870, 1622, 1598, 1448, 1404, 1370, 1349, 1282, 1269, 1213, 1140, 1109, 1043, 968, 866, 705 cm⁻¹; HRMS (DART) calcd for C₁₈H₂₆BO₂ ([M+H]⁺) 282.20258, found 282.20342.

7n: 3.5 hr, EE/EZ = 93: 7, 88% yield. ¹H NMR (500 MHz, CDCl₃) δ 7.62 (dd, J = 15.5, 11.0 Hz, 1H), 7.40 (d, J = 7.5 Hz, Hz, 2H), 7.31 (t, J = 7.5 Hz, 2H), 7.21 (t, J = 7.5 Hz, 1H), 6.71 (d, J = 11.5 Hz, 1H), 6.52 (d, J = 16.5 Hz, 1H), 2.23 (t, J = 7.5 Hz, 2H), 1.38-1.43 (m, 2H), 1.27-1.35 (m, 14H), 0.91 (t, J = 7.0 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 144.2, 137.9, 133.4, 129.2, 128.5, 127.2, 126.5, 83.1, 36.7, 32.5, 24.9, 24. 7, 14.0 (B-alkenyl carbon signal not observed); ¹¹B NMR (160 MHz, CDCl₃) δ 30.0; IR (ATR) 2976, 2957, 2929, 2871, 1714, 1680, 1597, 1449, 1404, 1390, 1371, 1326, 1245, 1214, 1140, 1008, 973, 748, 690 cm⁻¹; HRMS (DART) calcd for C₂₀H₃₀BO₂ ([M+H]⁺) 313.23388, found 313.23470.

70: 3.5 hr, EE/EZ = 93: 7, 90% yield. ¹H NMR (500 MHz, CDCl₃) δ 7.60 (dd, J = 15.5, 10.5 Hz, 1H), 7.38 (dd, J = 7.0, 1.0 Hz, 2H), 7.29 (t, J = 7.5 Hz, 2H), 7.19 (t, J = 6.5 Hz, 1H), 6.69 (d, J = 11.0 Hz, 1H), 6.51 (d, J = 15.5 Hz, 1H), 2.21 (t, J = 7.5 Hz, 2H), 1.39-1.42 (m, 2H), 1.32 (m, 12H), 1.26 (br, 10H), 0.87 (t, J = 6.5 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 144.2, 137.9, 133.4, 129.2, 128.5, 127.3, 126.5, 83.1, 37.1, 31.9, 30.3, 29.5, 29.4, 29.3, 24.9, 22.7, 14.1 (B-alkenyl carbon signal not observed); ¹¹B NMR (160 MHz, CDCl₃) δ 29.9; IR (ATR) 2976, 2955, 2924, 2853, 1587, 1449, 1423, 1404, 1389, 1371, 1350, 1292, 1270, 1142, 1109, 868, 690 cm⁻¹; HRMS (DART) calcd for C₂₄H₃₈BO₂ ([M+H]⁺) 369.29648, found 369.29744.

7p: 3.5 hr, EE/EZ = 86: 14, 86% yield. ¹H NMR (500 MHz, CDCl₃) δ 7.63 (dd, J = 15.0, 10.5 Hz, 1H), 7.41 (d, J = 8.0 Hz, 2H), 7.32 (t, J = 7.5 Hz, 2H), 7.21 (t, J = 7.5 Hz, 1H), **7p** 6.68 (d, J = 11.0 Hz 1H), 6.54 (d, J = 16.0 Hz, 1H), 2.12 (d, J = 6.5 Hz, 2H), 1.71-1.77 (m, 1H), 1.33 (s, 12H), 0.89 (d, J = 7.0 Hz, 6H); ¹³C NMR (125 MHz, CDCl₃) δ 145.4, 137.9, 133.5, 129.1, 128.5, 127.3, 126.5, 83.1, 46.4, 29.1, 24.9, 22.5 (B-alkenyl carbon signal not observed); ¹¹B NMR (160 MHz, CDCl₃) δ 30.0; IR (ATR) 2976, 2953, 2929, 2867, 1621, 1597, 1586, 1464, 1449, 1424, 1404, 1389, 1371, 1292, 1281, 1212, 1165,

1141, 1108, 1029, 966, 865, 749, 691 cm⁻¹; HRMS (DART) calcd for $C_{20}H_{30}BO_2$ ($[M+H]^+$) 313.23388, found 313.23389.

Gram-scale trans-hydroboration of 7l

To a 20-mL vial charged with ligand L3 (34.5 mg, 0.0800 mmol) and Pd_2dba_3 (36.6 mg, 0.0400 mmol) was added CH_2Cl_2 (1.4 mL). The resulting mixture was allowed to stir at room temperature for 16 hr to generate the Pd/L complex. Then, 1,3-enyne **61** (1.138 g, 8.000 mmol) in CH_2Cl_2 (5.0 mL) was added to the catalyst solution, followed by slow addition (ca. 5 min) of HBCat (1.25 mL, 12.0 mmol) via syringe. The resulting mixture was allowed to stir at room temperature for 0.5 hr. At the conclusion of the reaction, the crude NMR was then taken to determine the stereoselectivity. The crude mixture was then poured into a 500-mL flask charged with a pinacol solution (11.3 g, 96.0 mmol) in 50 mL CH_2Cl_2 . The resulting mixture was then allowed to stir at room temperature for 1 hr. The mixture was then diluted with water (200 mL) and extracted with CH_2Cl_2 (3 × 50 mL). The combined organic layer was dried over anhydrous Na_2SO_4 . After removal of the solvent, the residue was purified by column chromatography on silica gel with hexanes/EtOAc (100: 1) as the eluent to afford **71** as a white solid (1.819 g, 84%, EE/EZ > 98: 2). The characterization data are consistent with those described using the general procedure H.

Suzuki-Miyaura coupling of 71 with bromobenzene (eq 2)

The protocol for the Suzuki-Miyaura coupling was adapted from literature procedures.²² To a 20-mL reaction flask charged with S-Phos (3.2 mg, 0.0080 mmol), Pd₂dba₃ (3.6 mg, 0.0040 mmol), dienyl boronate **7I** (51.2 mg, 0.190 mmol), and bromobenzene (47.0 mg, 0.300 mmol) was added THF (1.8 mL) followed by 3.0 M aq. NaOH (0.60 mL, 1.80

mmol). After degasing the reaction mixture via freeze-pump-thaw cycles (3 times), the reaction mixture was allowed to stir at 60 °C for 16 hr. At the conclusion of the reaction, the reaction mixture was allowed to cool to the room temperature. Water (2.0 mL) and ether (2.0 mL) were then added to quench the reaction. The organic layer was separated, and the aqueous layer was extracted 3 times with ether. The combined organic phase was dried over Na₂SO₄. After removal of the solvent, the crude residue was purified by column chromatography on silica gel using hexanes as the eluent to afford **8** as colorless oil (36.4 mg, 87%).

EZ/EE > 98: 2. ¹H NMR (500 MHz, CDCl₃) δ 7.38-7.42 (m, 2H), 7.24-7.33 (m, 7H), 7.17-7.19 (m, 1H), 6.88 (dd, J = 15.5, 11.5 Hz, 1H), 6.55 (d, J = 15.5 Hz, 1H), 6.33 (d, J = 10.5 Hz, 1H), 2.21 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 141.5, 139.5, 137.8, 131.3, 128.5, 128.4, 128.2, 127.6, 127.1, 126.7, 126.2, 25.6; IR (ATR) 3077, 3027, 2929, 1594, 1572, 1492, 1442, 1433, 1371, 1073, 1026, 1000, 963, 765, 747, 700, 691 cm⁻¹; HRMS (DART) calcd for $C_{17}H_{17}([M+H]^+)$ 221.13303, found 221.13331.

Diels-Alder Reaction of 7l with N-methyl maleimide (eq 3)

The Diels-Alder reaction was adapted from literature procedures.²³ To a J-Y tube charged with dienyl boronate **7l** (51.2 mg, 0.190 mmol) and *N*-Me maleimide (22.2 mg, 0.200 mmol) was added mestiylene (1.0 mL). The resulting mixture was heated at 170 °C for 3 days. At the conclusion of the reaction, the mixture was allowed to cool to room temperature. The mixture was then directly subjected to the column chromatography on silica gel with hexanes/EtOAc (7: 1) as the eluent to afford **9** as white solid (49.2 mg, 67%). Crystals of **9** suitable for single crystal X-ray diffraction analysis were grown from slow evaporation of a pentane solution at -30 °C.

endo/exo >98: 2. ¹H NMR (500 MHz, CDCl₃) δ 7.33 (t, J = 7.5 Hz, 2H), 7.21-7.28 (m, 3H), 6.08 (dd, J = 10.0, 4.0 Hz, 1H), 5.92 (dd, J = 14.5, 3.0 Hz, 1H), 3.58-3.61 (m, 1H), 3.44 (t, J = 7.5 Hz, 1H), 3.29 (d, J = 8.0 Hz, 1H), 2.75 (s, 3H), 1.54 (s, 3H), 1.29 (s, 6H),

1.25 (s, 6H); 13 C NMR (125 MHz, CD₂Cl₃) δ 178.1, 176.9, 141.0, 138.1, 129.4, 128.6, 128.2, 127.2, 84.8, 48.7, 46.7, 41.9, 25.2, 24.8, 24.6, 20.6 (B-allyl carbon signal not observed); 11 B NMR (160 MHz, CD₂Cl₂) δ 32.6; IR (ATR) 3030, 2976, 2930, 2873, 1772, 1696, 1453, 1431, 1372, 1318, 1285, 1213, 1166, 1105, 966, 914, 732 cm⁻¹; HRMS (DART) calcd for C₂₂H₂₉BNO₄ ([M+H]⁺) 382.21896, found 382.22021.

Homologation of 7l (eq 4)

The homologation reaction was adapted from literature procedures.²⁴ To a 20-mL vial charged with racemic carbamate (125 mg, 0.500 mmol) and anhydrous Et₂O (2.0 mL) was slowly added ^sBuLi (0.38 mL, 1.4 M in cyclohexane, 0.53 mmol) in a dropwise fashion within 2 min at -78 °C. The resulting mixture was allowed to stir at -78 °C for 30 min, and then dienyl boronate ester 71 (140 mg, 0.518 mmol) in Et₂O (1.0 mL) was added dropwise within 2 min by vigorous stirring at -78 °C. The reaction mixture was allowed to stir at -78 °C for 30 min, and then at room temperature for 2 hr. At the conclusion of the reaction, the reaction mixture was guenched with H₂O (0.10 mL). After removal of the solvent, the mixture was passed through a pad of silica gel with hexanes/EtOAc (7: 1) as the eluent to afford the crude homologated product. This crude material was then dissolved in anhydrous THF (2.0 mL) containing BHT (2.5 mg), and the mixture was cooled to 0 °C. Then, an ice-cold mixture of 3.0 M NaOH (1.4 mL) and 30% ag. H₂O₂ (0.7 mL) was added all at once at 0 °C. The resulting mixture was then allowed to stir at room temperature for 0.5 hr. Then the mixture was diluted with H₂O (10 mL) and extracted with Et₂O (3 X 10 mL). The combined organic layer was then dried over Na₂SO₄. After removal of the solvent, the residue was purified by column chromatography on silica gel with hexanes/EtOAc (15: 1) as the eluent to afford 11 as a colorless oil (82.0 mg, 62%).

¹H NMR (500 MHz, CD₂Cl₂) δ 7.52-7.54 (m, 2H), 7.35-7.38 (m, 2H), 7.22-7.28 (m, 3H), 7.13-7.19 (m, 4H), 6.29 (d, J = 15.0 Hz, 1H), 6.09 (d, J = 11.5 Hz, 1H), 2.10 (s, 1H), 1.93 (s, 3H), 1.77 (s, 3H); ¹³C NMR (160 MHz, CD₂Cl₂) δ 148.8, 144.4, 138.4 132.1, 129.0,

128.8, 128.4, 127.7, 127.6, 127.3, 126.8, 126.0, 78.0, 30.0, 23.8; IR (ATR) 3559 3447, 3057, 3022, 2972, 2936, 1595, 1491, 1446, 1369, 1308, 1268, 1066, 999, 907, 748, 692 cm⁻¹; HRMS calcd for $C_{19}H_{19}([M+H-H_2O]^+)$ 247.14868, found 247.14847.

Preparation of complex 12 and its use in trans-hydroboration

To a 5-ml vial charged 1,4-azaborine **L4** (21 mg, 0.047 mmol) and Pd₂dba₃ (21 mg, 0.023 mmol) was added CH₂Cl₂ (0.90 ml). The mixture was stirred at room temperature for 4 hours. Then, CH₂Cl₂ was removed under vacuum. The resulting mixture was recrystallized from benzene/pentane to afford complex **12** as reddish crystal suitable for single crystal X-ray diffraction analysis (27 mg, 73% yield). Due to conformational dynamics the ¹H and ¹³C NMR signals are broad. ¹¹B NMR (160 MHz, CD₂Cl₂) δ 32.4 (br); ³¹P NMR (202 MHz, CD₂Cl₂) δ 29.2 (br); IR (ATR) 3051, 2949, 2919, 2859, 1638, 1620, 1572, 1447, 1434, 1366, 1331, 1181, 1094, 1076, 953, 758, 744, 694, 640 cm⁻¹.

Hydroboration of 4a with complex 12

A 5-mL vial was charged with complex **12** (7.9 mg, 0.01 mmol) and CH₂Cl₂ (1.0 ml), and the mixture was stirred for 10 min to produce a homogeneous solution. Then, HBCat (45 mg, 0.375 mmol) and 1,3-enyne **4a** (32 mg, 0.25 mmol) were added. The resulting mixture was allowed to stir at room temperature for 30 min. Then, pinacol (354 mg, 3.0 mmol) in CH₂Cl₂ (3.0 mL) was introduced, and the mixture was allowed to stir at room temperature for 1 hr. After removal of the solvent, the crude residue was purified by column chromatography on silica gel with (Hex/EtOAc = 100: 1) as the eluent to afford **5a** as light yellow oil (52 mg, 81%, >98:2 *trans*-hydroboration selectivity). The characterization data is identical to those reported under general procedure G (Table 2).

Synthesis of CC-L3 and its performance in hydroboration reactions

To a 20-mL round bottom flask containing 1-bromo-2-ethylnaphthalene 25 (94 mg, 0.40 mmol) and magnesium (23 mg, 0.96 mmol) were added THF (5.0 mL). The reaction mixture was then heated to 65 °C in an oil bath and 1,2-dibromoethane (2.0 $\mu L)$ was added dropwise via syringe to initiate the reaction. After heating the mixture at 65 °C for 60 min, 1-bromo-2-chloro-benzene (84 mg,

0.44 mmol) was added at the same temperature slowly over one hour. After an additional 1 hour of stirring at 65 °C, the reaction mixture was allowed to cool to room temperature, and the reaction vessel was moved into the glove box. Anhydrous copper(I) chloride (8.0 mg, 0.08 mmol) was added into the mixture. Then, ClPPh₂ (88 mg, 0.40 mmol) was added via syringe. The resulting mixture was stirred at room temperature for 12 hours. The reaction was quenched with H₂O (10 mL), and the resulting mixture was extracted with Et₂O (2 times, 10 mL each). The combined organic layers were dried over MgSO₄, concentrated and purified by column chromatography on silica gel with hexanes/EtOAc (100/1) as the eluent to afford a white solid (50 mg, 30% yield).

¹H NMR (500 MHz, CD₂Cl₂) δ 7.84 (d, J = 8.5 Hz, 1H, Ar**H**), 7.81 (dt, J = 8.2, 0.9 Hz, 1H, Ar**H**), 7.49 (td, J = 7.4, 1.4 Hz, 1H, Ar**H**), 7.45 – 7.38 (m, 2H, Ar**H**), 7.37 – 7.13 (m, 12H, Ar**H**), 7.09 – 7.01 (m, 3H, Ar**H**), 2.31 – 2.11 (m, 2H, C**H**₂CH₃), 0.99 (t, J = 7.6 Hz, 3H, CH₂C**H**₃); ¹³C NMR (126 MHz, CD₂Cl₂) δ 146.14 (d, J = 33.5 Hz), 140.37 (d, J = 1.9 Hz), 138.63 (d, J = 11.8 Hz), 137.95 (d, J = 12.9 Hz), 137.48 (d, J = 12.9 Hz), 137.17 (d, J = 7.3 Hz), 134.87 (d, J = 1.9 Hz), 134.20 (d, J = 20.5 Hz), 133.81 (d, J = 20.0 Hz), 133.46 (d, J = 2.1 Hz), 132.07, 131.34 (d, J = 6.2 Hz), 129.46, 128.83, 128.74, 128.68, 128.60, 128.57, 128.22, 127.99 (d, J = 12.4 Hz), 126.94, 126.88, 125.90, 125.03, 27.16 (d, J = 1.7 Hz), 15.30; ³¹P NMR (202 MHz, CD₂Cl₂) δ -15.39; IR (ATR) 3050, 2964, 1476, 1457, 1432, 1374, 1088, 1067, 1025, 951, 817, 763, 741, 711, 695, 678, 621, 504, 429, 417; HRMS (DART) calcd for C₃₀H₂₆P ([M+H]⁺) 417.17721, found 417.17894.

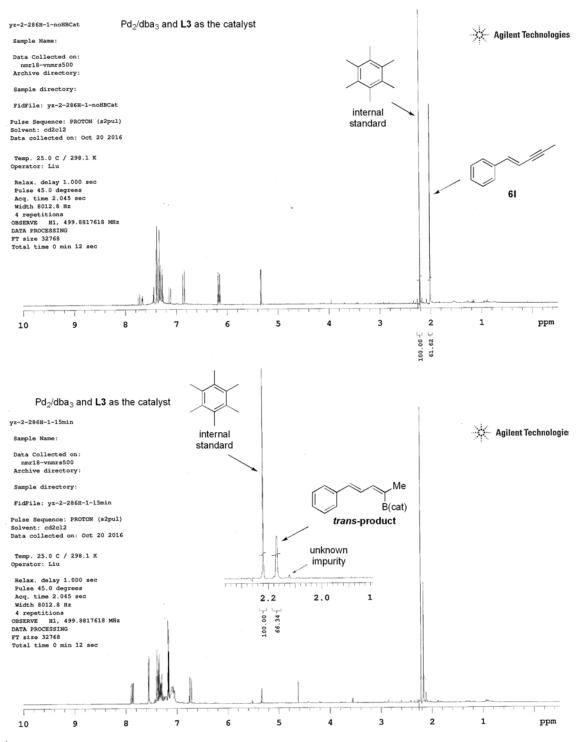
Comparative Trans-Hydroboration Catalysis Pd/L3 vs. Pd/CC-L3

To a 4-mL vial charged with ligand L3 (21.6 mg, 0.050 mmol) or CC-L3 (20.8 mg, 0.050 mmol) and Pd_2dba_3 (22.9 mg, 0.025 mmol) was added CH_2Cl_2 (1.0 mL). The resulting mixture was allowed to stir at room temperature for 16 hours to generate the Pd/L complexes. The resulting two stock solutions (0.05 M in CH_2Cl_2) were used for the catalytic reactions.

To a 4-mL vial charged with stock solution of catalyst (200 μL, 0.05 M in CH₂Cl₂, 0.01 mmol), internal standard hexamethylbenzene, **6l** (35.5mg, 0.25 mmol), was added catecholborane (45.0 mg, 0.375 mmol). The resulting mixture was allowed to stir at room temperature for 15 min. After removal of the solvent, the conversion, the percentage of *trans*-, *cis*-hydroboration and allene products were determined by ¹H NMR.

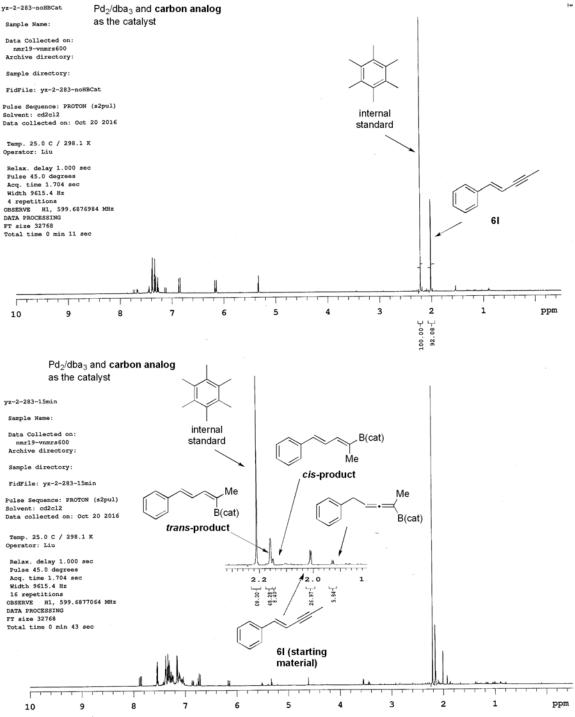
catalyst	conversion (%)	trans-hydroboration (%)	cis-hydroboration (%)	allene (%)
Pd/L3	100	>98	0	0
Pd/CC-	71	52	9	6
L3				

¹H NMR before addition of H-BCat for Pd/L3 system.



¹H NMR at the conclusion of the reaction.

¹H NMR before addition of H-BCat for Pd/CC-L3 system.



Crystallographic data for 3a (Liu160)

Data / restraints / parameters

Final R indices [I>2sigma(I)]

Largest diff. peak and hole

Goodness-of-fit on F²

R indices (all data)

Identification code	liu160
Empirical formula	C10 H11 B Cl N
Formula weight	191.46
Temperature	193(2) K
Wavelength	0.71073 Å
Crystal system	Triclinic
Space group	P-1
Unit cell dimensions	$a = 7.6350(13) \text{ Å}$ $\alpha = 89.640(3)^{\circ}$.
	$b = 9.2691(16) \text{ Å}$ $\beta = 79.774(3)^{\circ}$.
	$c = 14.887(3) \text{ Å}$ $\gamma = 70.461(3)^{\circ}$.
Volume	975.5(3) Å ³
Z	4
Density (calculated)	1.304 Mg/m^3
Absorption coefficient	0.339 mm ⁻¹
F(000)	400
Crystal size	$0.27 \times 0.19 \times 0.12 \text{ mm}^3$
Theta range for data collection	2.34 to 25.00°.
Index ranges	-9<=h<=9, -11<=k<=11, -17<=l<=17
Reflections collected	9361
Independent reflections	3416 [R(int) = 0.0550]
Completeness to theta = 25.00°	99.7 %
Absorption correction	Semi-empirical from equivalents
Max. and min. transmission	0.9605 and 0.9141
Refinement method	Full-matrix least-squares on F ²

3416 / 0 / 323

R1 = 0.0488, wR2 = 0.1344R1 = 0.0548, wR2 = 0.1426

0.405 and -0.257 e.Å-3

0.980

Crystallographic data for 5a

Empirical formula C16 H21BO2

Formula weight 256.14
Temperature 100(2) K

Wavelength 0.71073 ° Crystal system Monoclinic

Space group C c

Unit cell dimensions a = 8.2753(8) Å $\alpha = 90^{\circ}$.

b = 23.637(3) Å $\beta = 91.4540(18) ^{\circ}.$

c = 15.5395(16) Å $\gamma = 90^{\circ}$.

Volume 3038.5(5) Å ³

Z 8

Density (calculated) 1.120 Mg/m³
Absorption coefficient 0.071 mm⁻¹

F(000) 1104

Crystal size $0.600 \times 0.450 \times 0.380 \text{ mm}^3$

Theta range for data collection 1.723 to 28.360°.

Index ranges -10 <= h <= 11, -31 <= k <= 31, -20 <= l <= 20

Reflections collected 21673

Independent reflections 7208 [R(int) = 0.0213]

Completeness to theta = 25.242∞ 100.0 %

Absorption correction Semi-empirical from equivalents

Max. and min. transmission 0.7457 and 0.7003

Refinement method Full-matrix least-squares on F²

Data / restraints / parameters 7208 / 2 / 351

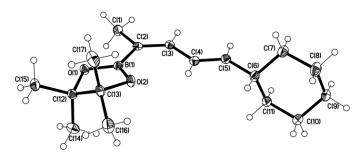
Goodness-of-fit on F^2 1.033

Final R indices [I>2sigma(I)] R1 = 0.0352, wR2 = 0.0890 R indices (all data) R1 = 0.0378, wR2 = 0.0907

Absolute structure parameter 0.1(2)
Extinction coefficient na

Largest diff. peak and hole 0.274 and -0.164 e. Å -3

Crystallographic data for 7a



Empirical formula C17 H29 B O2

Formula weight 276.21
Temperature 100(2) K
Wavelength 0.71073 Å
Crystal system Monoclinic

Space group $P2_1/c$

Unit cell dimensions a = 11.7894(15) Å $\alpha = 90^{\circ}$

b = 6.7732(9) Å $\beta = 96.611(2)^{\circ}$

c = 21.252(3) Å $\gamma = 90^{\circ}$

Volume 1685.7(4) Å³

Z 4

Density (calculated) 1.088 Mg/m³
Absorption coefficient 0.068 mm⁻¹

F(000) 608

Crystal size $0.350 \times 0.160 \times 0.120 \text{ mm}^3$

Theta range for data collection 1.739 to 28.268°.

Index ranges -15 <= h <= 15, -9 <= k <= 8, -28 <= l <= 28

Reflections collected 31882

Independent reflections 4162 [R(int) = 0.0457]

Completeness to theta = 25.242° 100.0 %

Absorption correction Semi-empirical from equivalents

Max. and min. transmission 0.7457 and 0.7082

Refinement method Full-matrix least-squares on F²

Data / restraints / parameters 4162 / 0 / 186

Goodness-of-fit on F^2 1.027

Final R indices [I>2sigma(I)] R1 = 0.0420, wR2 = 0.0965 R indices (all data) R1 = 0.0584, wR2 = 0.1055

Extinction coefficient na

Largest diff. peak and hole 0.341 and -0.229 e.Å-3

Crystallographic data for 9

Identification code C22H28BNO4

Empirical formula C22H28BNO4
Formula weight 381.26
Temperature 100(2) K
Wavelength 1.54178 Å
Crystal system Monoclinic

Space group P2₁/c

Unit cell dimensions a = 7.1599(3) Å $\alpha = 90^{\circ}$.

b = 17.6140(8) Å $\beta = 90.955(2)^{\circ}.$

c = 33.0261(15) Å $\gamma = 90^{\circ}$.

Volume 4164.5(3) Å ³

Z 8

Density (calculated) 1.216 Mg/m³
Absorption coefficient 0.658 mm⁻¹

F(000) 1632

Crystal size $0.300 \times 0.060 \times 0.050 \text{ mm}^3$

Theta range for data collection 2.676 to 66.712°.

Index ranges -8 <= h <= 8, -20 <= k <= 20, -39 <= l <= 39

Reflections collected 45317

Independent reflections 7370 [R(int) = 0.0244]

Completeness to theta = 66.750∞ 99.7 %

Absorption correction Semi-empirical from equivalents

Max. and min. transmission 0.7528 and 0.6635

Refinement method Full-matrix least-squares on F²

Data / restraints / parameters 7370 / 0 / 517

Goodness-of-fit on F^2 1.008

Final R indices [I>2sigma(I)] R1 = 0.0344, wR2 = 0.0863 R indices (all data) R1 = 0.0360, wR2 = 0.0878

Extinction coefficient na

Largest diff. peak and hole 0.319 and -0.198 e. Å -3

Crystallographic data for 12

Empirical formula C54 H51 B N O P Pd

Formula weight 878.14
Temperature 100(2) K
Wavelength 1.54178 Å
Crystal system Triclinic

Space group P-1

Unit cell dimensions a = 12.0431(9) Å $\alpha = 85.720(3)^{\circ}$.

 $b = 15.5970(12) \text{ Å} \qquad \beta = 89.462(3)^{\circ}.$ $c = 23.7704(17) \text{ Å} \qquad \gamma = 87.941(4)^{\circ}.$

Volume 4449.5(6) Å³

Z 4

Density (calculated) 1.311 Mg/m³
Absorption coefficient 3.999 mm⁻¹

F(000) 1824

Crystal size $0.220 \times 0.130 \times 0.050 \text{ mm}^3$

Theta range for data collection 2.843 to 66.771°.

Index ranges -14 <= h <= 14, -18 <= k <= 18, 0 <= l <= 28

Reflections collected 15665

Independent reflections 15665 [R(int) = ?]

Completeness to theta = 67.679° 97.3 %

Absorption correction Semi-empirical from equivalents

Max. and min. transmission 0.7528 and 0.4399

Refinement method Full-matrix least-squares on F²

Data / restraints / parameters 15665 / 45 / 1137

Goodness-of-fit on F² 1.060

Final R indices [I>2sigma(I)] R1 = 0.0617, wR2 = 0.1714 R indices (all data) R1 = 0.0697, wR2 = 0.1838

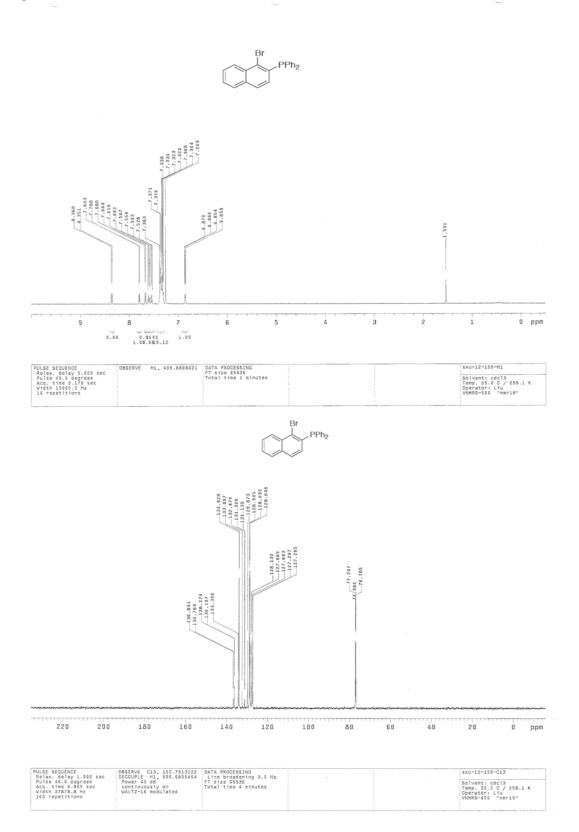
Extinction coefficient na

Largest diff. peak and hole 2.032 and -1.121 e.Å-3

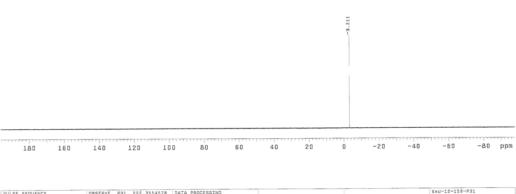
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NMR spectra of all compounds Compound S1

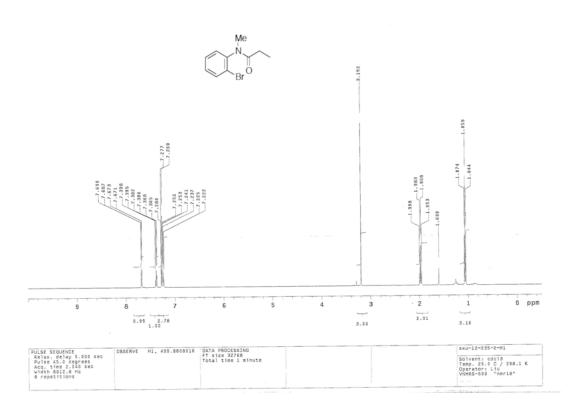




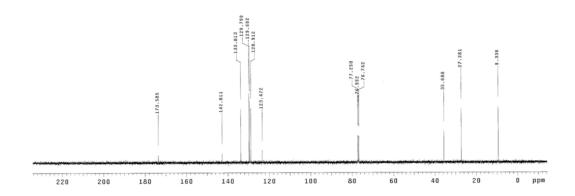


Aug. time 5.5 sc or during acquisition Total time I minute Temp. 25.0 C / 26 Temp. 25.0 C / 26 Temp. 25.0 C / 26 Temp. 25.0 C / 26	Pulse 45.0 degrees Power 40 dB Acq. time 0.551 sec on during acquisition Vidth 59523.8 Hz off during delay	E15 Line broadening 0.5 Hz FT size 65538 Total time 1 minute	Solvent: cdcl3 Temp. 25.0 / 298.1 K Operator: Liu VNMRS-590 "nnr18"
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Compound 1a

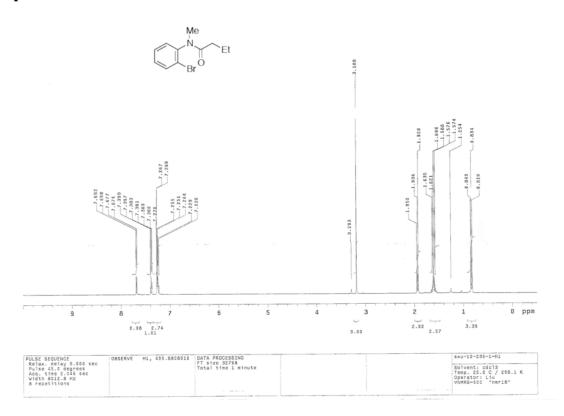




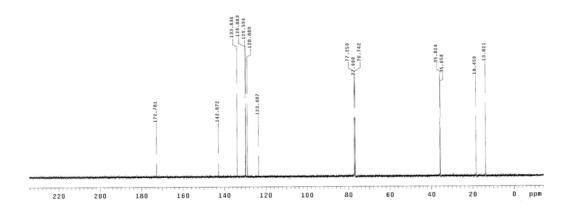


PULSE SEQUENCE	OBSERVE C13, 125.6951338 DECOUPLE H1, 499.8833015		sxu-12-235-2-C13
Relax. delay 1.000 sec Pulse 45.0 degrees Acq. time 1.049 sec Width 31250.0 Hz 76 repetitions	Power 40 dB continuously on WALTZ-16 modulated	Line grounding 0.5 MZ FT size 58536 Total time 2 minutes	Solvent: cdcl3 Tepp. 25.0 C / 298.1 K Operator: Liu VNMRS-500 "nmr18"

Compound 1b

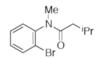


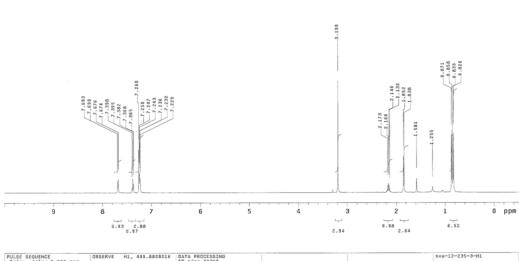




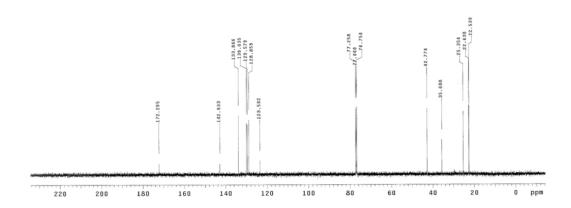
Pulse 45.0 degrees : Power 40 dB	DATA PROCESSING Line broadening 0.5 Hz FF size 5535 Total time 3 minutes	SXU-12-235-1-CL3 Solvent 2 dd(13 Solvent 2 dd(13 Solvent 2 dd(13 Operator: L1 VMMS-500 "nmr18"
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Compound 1c



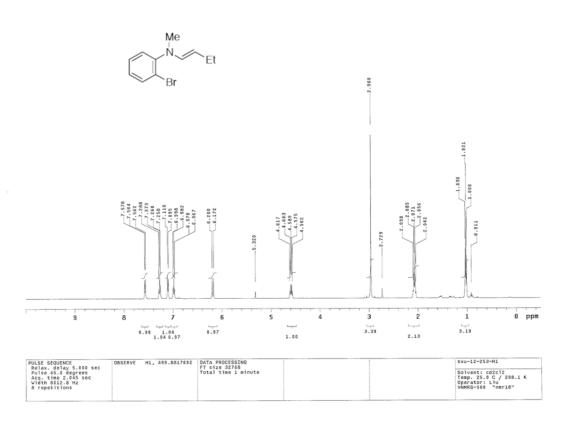




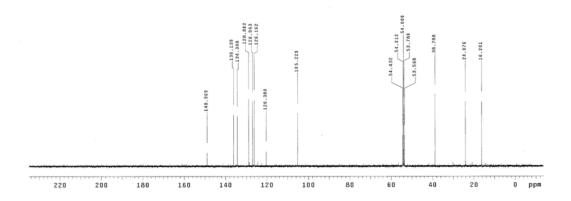


PULSE SECURINCE OBSERVE C13, 125.6951290 DATA PROCESSING Relax. delay 1.00 sec DECOUPLE H1, 493.8033015 Line broadening 6.5 Hz Pulse 45.0 degrees Power 40 dB 7 f size 65536 Acq. time 1.645 sec Continuously on Total time 3 minutes Water Continuously on Total time 3 minutes 38 repetitions	\$xu-12-255-3-Cl3 Solvent: cl3 Tesp. 25.0 C / 298.1 K O
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Compound 2b

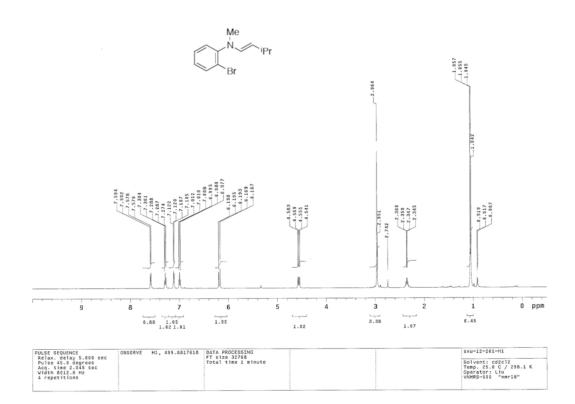




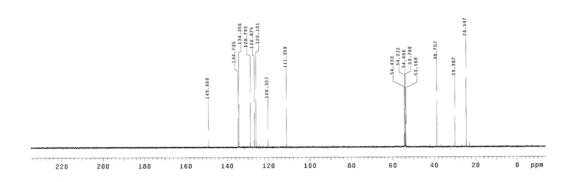


PULSE SEQUENCE		DATA PROCESSING	1	sxu-12-253-C13
Relax. delay 1.000 sec Pulse 45.0 degrees	DECOUPLE H1, 499.8842612 Power 40 dB	Line broadening 0.5 Hz FT size 65536		Solvent: cd2c12
Acq. time 1.049 sec	continuously on	Total time 5 minutes		Temp. 25.0 C / 298.1 K
Vidth 31250.0 Hz 148 repetitions	VALTZ-16 modulated			Operator: Liu VNMRS-500 "nmr18"
140 repetitions				VIANO-300 IIII 20

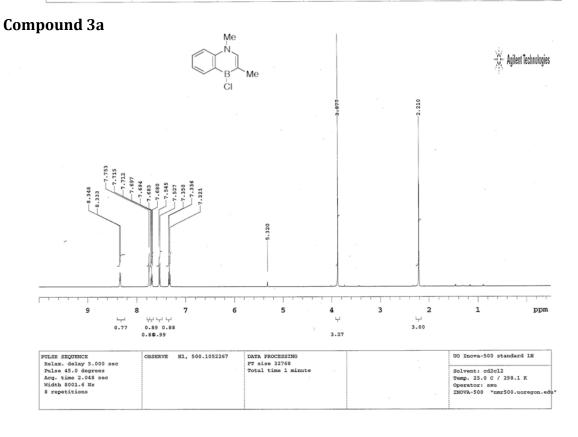
Compound 2c

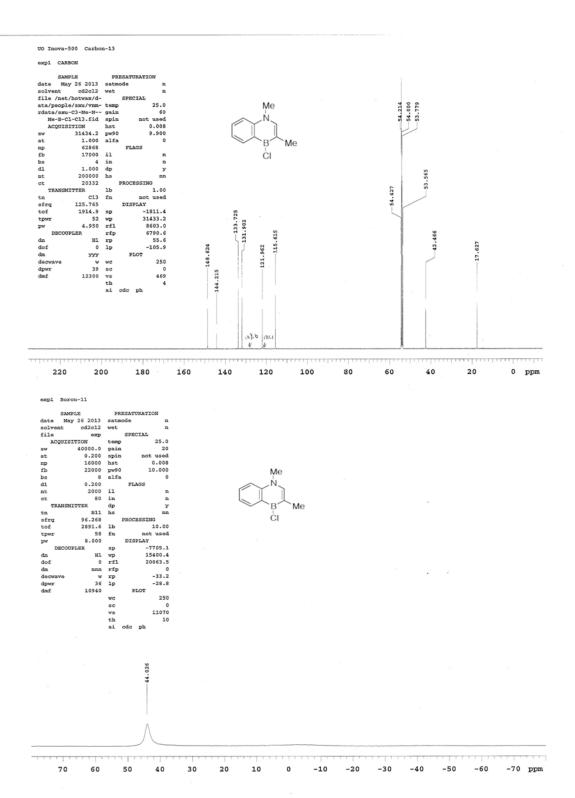




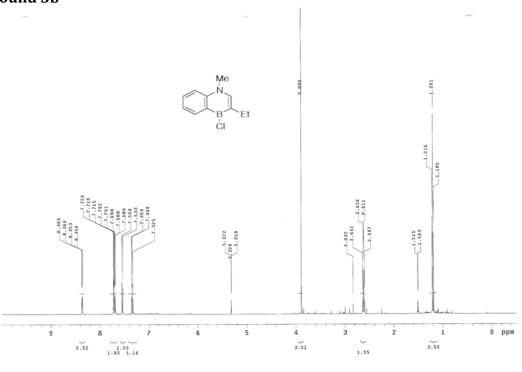


PULSE SEQUENCE	OBSERVE C13, 125.6952907	DATA PROCESSING	sxu-12-261-C13
Relax, delay 1,000 sec	DECOUPLE H1, 499.8842612	Line broadening 0.5 Hz	
Pulse 45.0 degrees Acq. time 1.049 sec Width 31250.0 Hz 170 repetitions	Power 40 dB continuously on VALTZ-16 modulated	Tr size 65536 Total time 5 minutes	Solvent: cd2cl2 Tenp. 25.0 C / 258.1 K Operator: Liu VMMRS-300 "mari8"

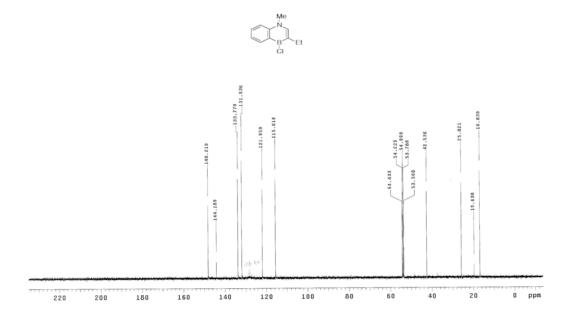




Compound 3b

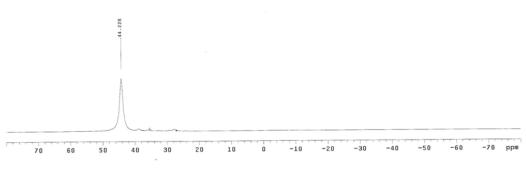


PULSE SEQUENCE Relax. delay 5.000 sec Pulse 45.0 degrees Acq. time 2.175 sec Vidth 15050.2 Mz 18 repetitions	OBSERVE	H1, 499.8817682	DATA PROCESSING FT size 65536 Total time 1 minutes		sxu-11-93-H1 Solvent: cd2c12 Temp. 25.0 C / 298.1 K Operator: Liu VNMRS-500 "nmr18"
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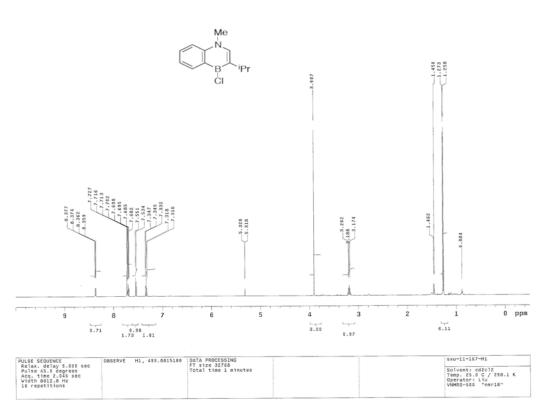
PULSE SEQUENCE Relax. delay 1.000 sec	OBSERVE C13, 125.6952926 DECOUPLE H1, 499.8842612			sxu-11-93-C13
Pulse 45.0 degrees Acq. time 1.049 sec Vidth 31250.0 Hz 358 repetitions	Power 40 dB continuously on VALTZ-16 modulated	FT size 65536 Total time 12 minutes		Solvent: cd2cl2 Temp. 25.0 C / 298.1 K Operator: Liu VNMRS-500 "nmr18"



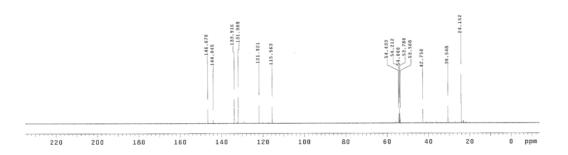


	1, 160.3819248 DATA PROCESSING	sxu-11-93-811	7
Relax. delay 0.010 sec Pulse 90.0 degrees Acq. time 0.020 sec Vidth 32051.3 Hz 1000 repetitions	Line broadening 10.0 Hz FT size 32788 Total time I minute	Solvent: cd2cl2 Temp. Z5.9 G / 286.1 K Operator: L1 WMMRS-389 mar18"	

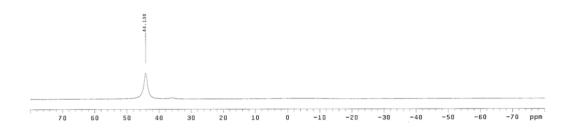
Compound 3c



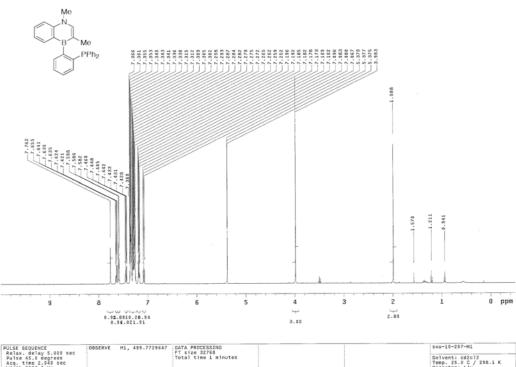




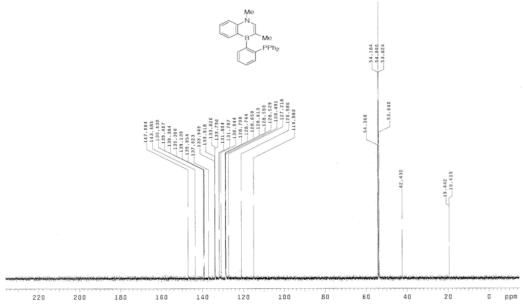
PULSE SEQUENCE Relax. delay 1.000 sec	OBSERVE C13, 125.6352926 DECOUPLE H1. 493.8842612	DATA PROCESSING Line broadening 0.5 Hz	sxu-11-167-C13
Pulse 45.0 degrees Acq. time 1.049 sec Vidth 31250.0 Hz 268 repetitions	Power 40 d8 continuously on VALTZ-16 modulated	FT size 65536 Total time S minutes	Solvent: cd2cl2 Temp. 25.0 C / 298.1 K Operator: Liu VNMKS-500 "mmrl8"



PULSE SEQUENCE Relax. delay 0.010 sec Pulse 50.0 degrees Acq. time 0.020 sec Vidth 32051.3 Hz 1000 repetitions	OBSERVE B11, 160.3819092	DATA PROCESSING Line broadening 10.0 Hz FT size 32768 Total time 1 minute		Solvent: cd2c12 Temp. 25.0 C / 298.1 K Operator: Liu VNMRS-500 "nmr18"

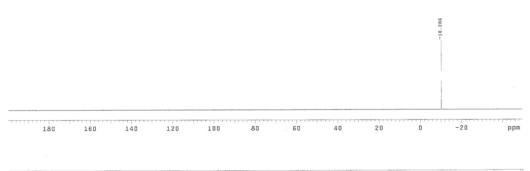


PULSE SEQUENCE Relax. delay 5.000 s Pulse 45.0 degrees Acq. time 2.049 sec Vidth 7396.0 Hz 16 repetitions	ec	H1, 499.7729647	DATA PROCESSING FT size 32768 Total time 1 minutes		Solvent: cd2cl2 Temp. 25.0 C / 298.1 K Operator: Liu INOVA-500 "nmrl1"
			Me		



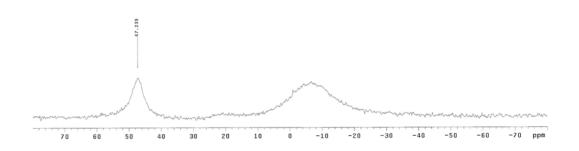
PULSE SEQUENCE Relax. delav 1.000 sec	OBSERVE C13, 150.7914992 DECOUPLE H1, 599.6906368	DATA PROCESSING Line broadening 0.5 Hz		 sxu-1@-297-C13
Pulse 45.0 degrees Acq. time 0.885 sec Width 37878.8 Hz 848 repetitions	Power 45 d8 continuously on WALTZ-16 modulated	FT size 65536 Total time 26 minutes		Solvent: cd2cl2 Temp. 25.0 C / 298.1 K Operator: Liu VNNRS-500 "omr19"



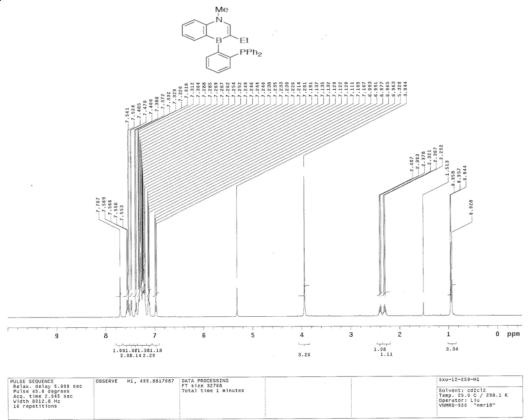


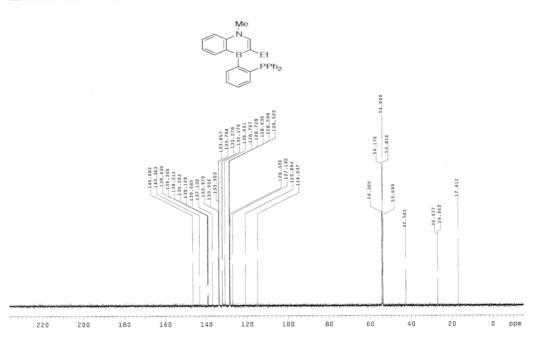
PULSE SEQUENCE OBSERVE P31, 181,8286588 DATA PROCESSING SUM-11-237-P31 Relax. delay. 100 sec DECOUPLE H1, 385.789432 Line proadening 0.5 Hz Solvent Line 1 Solvent Carlot Line 1 Solvent Carlot Line 2 Solvent Carlot Line 2 Solvent Carlot Line 3 Solvent Carlot Line 3 Solvent Lin	
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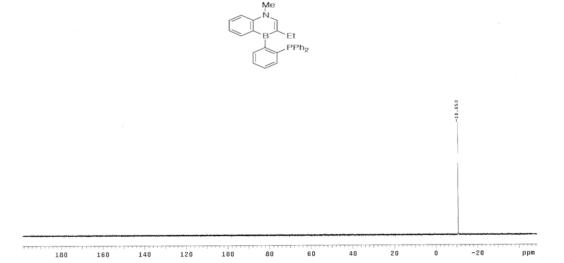


PULSE SEQUENCE OBSERVE 811, 160.3471555 DATA PROCESSING SXW-10-27 Relax, delay 0.010 sec Fisize 2048 Fisize 2048 Acq. time 0.020 sec Total time 1 minute 1000 reportitions Total time 1 minute 1000 reportitions Total time 1 minute 1000 reportitions 1000 report	cd2c12 0 C / 298.1 K
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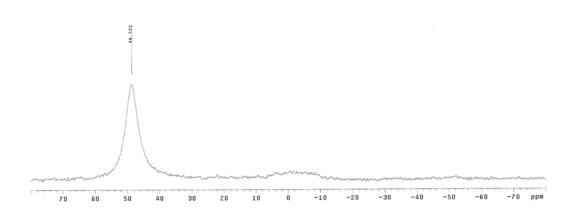


PULSE SEQUENCE Relax. delay 1.000 sec	OBSERVE C13, 150.7915004 DECOUPLE H1, 599.6906968	Line broadening 0.5 Hz	sxu-12-259-C13
Pulse 45.0 degrees Acq. time 0.865 sec Width 37878.8 Hz 442 repetitions	Power 45 d8 continuously on WALTZ-16 modulated	FT size 85538 Total time 13 minutes	Solvent: cd2cl2 Temp. 25.0 C / 298.1 K Operator: Liu VNMRS-800 "nmr15"

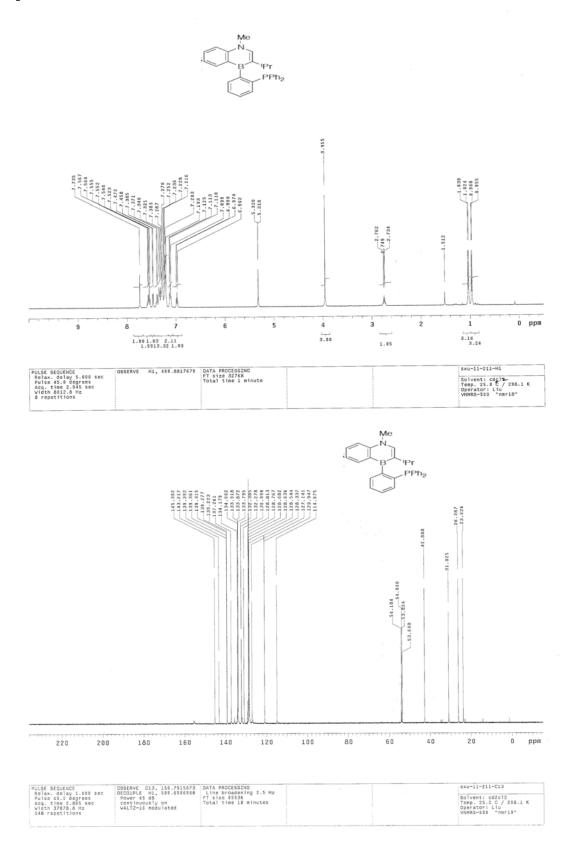


Relax. delay 1.000 sec DECOU Pulse 45.0 degrees Pow Acq. time 0.655 sec on 6 Vidth 50000.0 Hz off	ver 40 dB FT size	oadening 0.5 Hz	\$xu-12-259-P31 SOlvent: cd2cl2 Temp. 25.0 C / 298.1 K Opprator: Li WHRG-309 "mnr18"

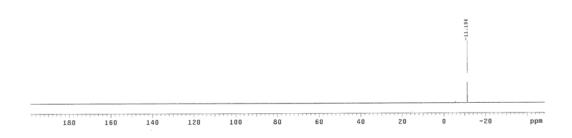




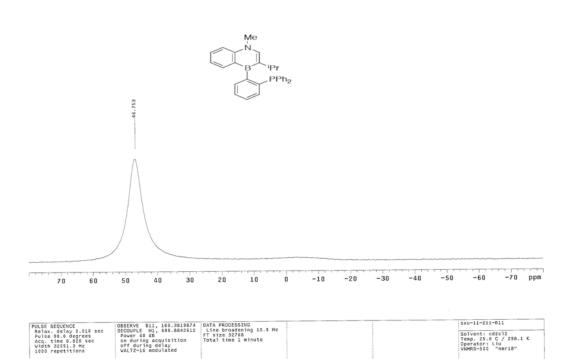
PULSE SEQUENCE Relax, delay 0.010 sec	OBSERVE B11, 160.3817077 DECOUPLE H1, 499.8842612		sxu-12-259-811
Pulse 90.0 degrees Acq. time 0.020 sec Width 32051.3 Hz 1000 repetitions	Power 40 dB on during acquisition off during delay WALTZ-16 modulated	FT size 32768 Total time 1 minute	Solvent: cd2cl2 Temp. 25.0 C / 298.1 K Operator: Liu VNMRS-500 "nmr18"

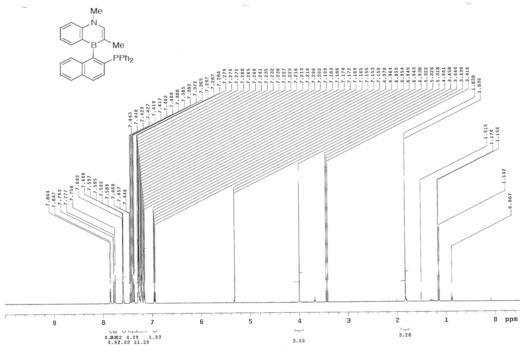






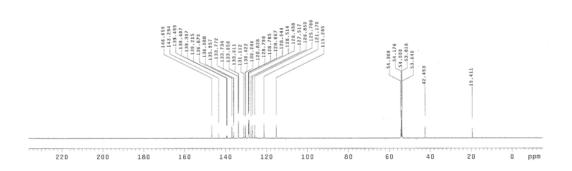
PULSE SEQUENCE Relax. delay 1.000 sec Pulse 45.0 degrees Acq. time 0.655 sec Width 50000.0 Hz 30 repetitions	OBSERVE P31, 202.3558463 DECOUPLE H1, 499.8842612 Power 40 dB on during acquisition off during delay VALTZ-16 modulated	DATA PROCESSING Line broadening 0.5 Hz FT size 65536 Total time 1 minute		SOlvent: cd2cl2 Temp. 25.0 C / 258.1 K Operator: Liu VNMRS-500 "mmr18"
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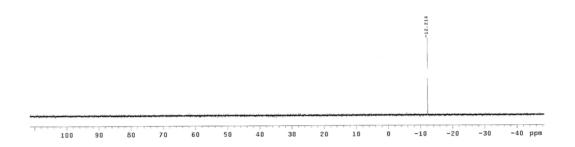
PULSE SEQUENCE Relax. delay 5.000 sec Pulse 45.0 degrees Acq. time 2.045 sec Width 8012.8 Hz 16 repetitions	OBSERVE H1, 4	FT :	A PROCESSING size 32768 al time 1 minutes	,	sxu-10-33-H1 Solvent: cd2cl2 Temp. 25.0 C / 298.1 K Operator: Lfu VNMRS-500 "nmr18"



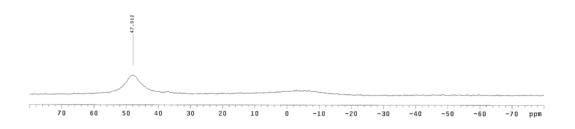


PULSE SEQUENCE Relax. delay 1.000 sec Pulse 45.0 degrees Acq. time 0.855 sec Width 37878.8 Hz 712 repetitions	OBSERVE C13, 150.7915015 DECOUPLE HI, 593.5908988 Power 45 dB continuously on WALTZ-16 modulated				SXU-10-33-C13 Solvent: cd2c12 Yeap. 25.0 C / 298.1 K Operator: Liu VNNRS-600 "nmr18"
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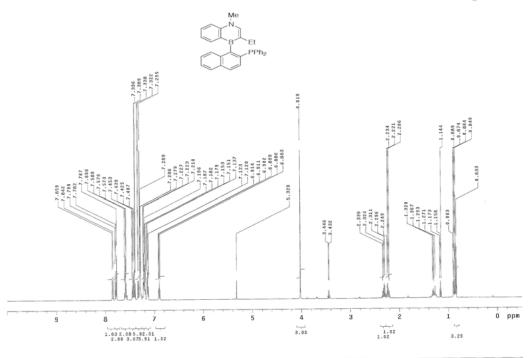




PULSE SEQUENCE CREAK PAIL 222.3558462 PAIL 222.3558462 PAIL 222.3558462 PAIL 223.356462 PAIL 2		sxu-11-33-p31 Solvent: cd2c12 Temp. 25.0 C / 296.1 K Operator: Lu VMMKS-558 "mar10"
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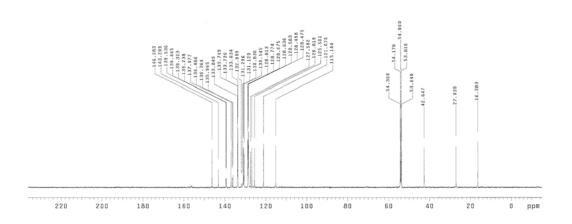


PULSE SEQUENCE Relax delay 0.010 sec Pose 10.0 secs	sxu-11-33-Bii Solvent: cd2c12 Temp. 25.0 C / 298.1 K Operator: Liu VNMRS-500 "mmrla"
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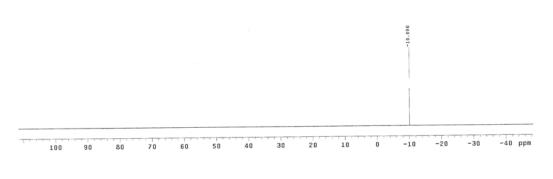
Relay, delay 5,000 sec	DATA PROCESSING FF size 32768 Total time 1 minutes	Solvent: cd2cl2 Temp. 25.0 C / 298.1 K Operator: Liu VNMRS-500 "mmr18"
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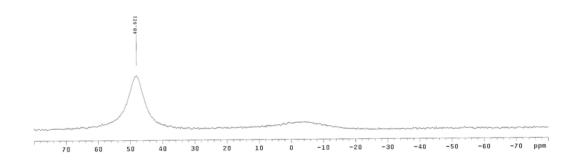
PULSE SEQUENCE Relax. delay 1.000 sec Pulse 45.0 degrees Acq. time 0.865 sec Vidth 37878.8 Hz 466 repetitions	OBSERVE C13, 150.7915027 DECOUPLE H1, 599.6906968 Power 45 d5 continuously on WALTZ-16 modulated	DATA PROCESSING Line broadening 3.0 Hz FT size 85538 Total time 14 minutes		Solvent: cd2cl2 Temp. 25.0 C / 298.1 K Operator: Liu VNMRS-500 "nmr13"





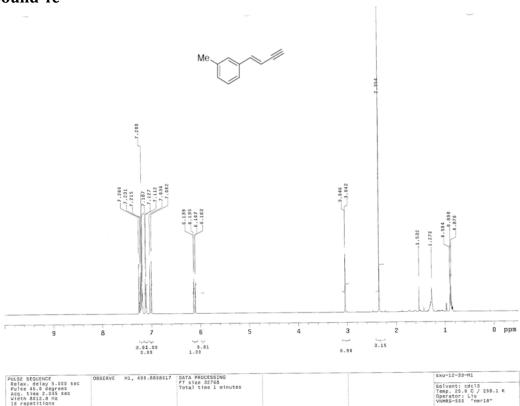
PULSE SEQUENCE OBSERVE P31, 202.3534578 DATA PROCESSING SEQUENCE Relax. delay 1.010 sec Secoupil H1, 491.8633913 Fine broadering 0.5 Hz Solvent cdc13 Fr size 5558 Sec on during acquisition off during delay of during delay of during delay VMHS-368 USA CF. size 555 Sec Of during delay VMHS-368 USA CF. size 555 Sec Of during delay VMHS-368 USA CF. size 555 Sec Of during delay VMHS-368 "mari8" USA CF. size 555 Sec Of during delay VMHS-368 USA CF. size 555 Sec Of during delay VMHS-368 "mari8" USA CF. size 555 Sec Of during delay VMHS-368 "mari8" USA CF. size 555 Sec Of during delay VMHS-368 "mari8" USA CF. size 555 Sec Of during delay VMHS-368 "mari8" USA CF. size 555 Sec Of during delay VMHS-368 "mari8" USA CF. size 555 Sec Of during delay VMHS-368 USA

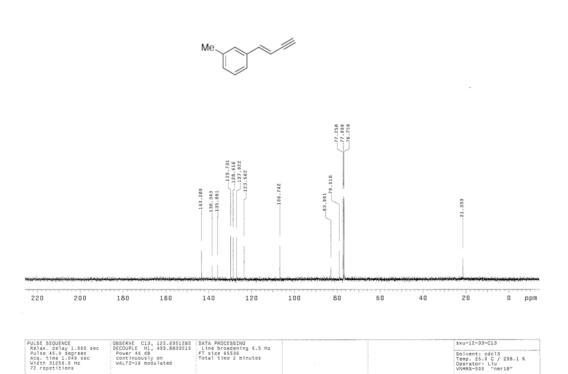




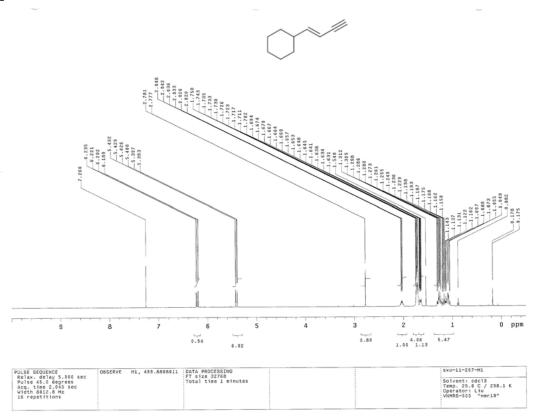
| PULSE SEQUENCE | DREAM | DATA PROCESSING | SECURITY |

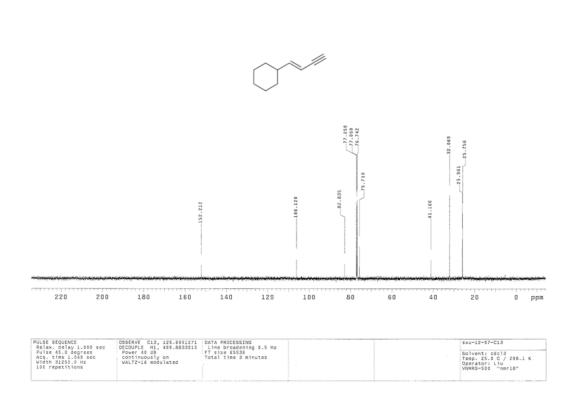




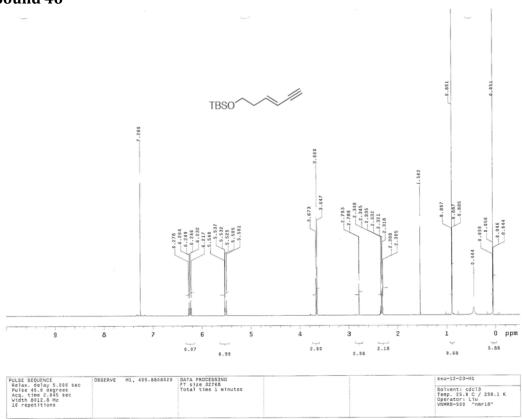


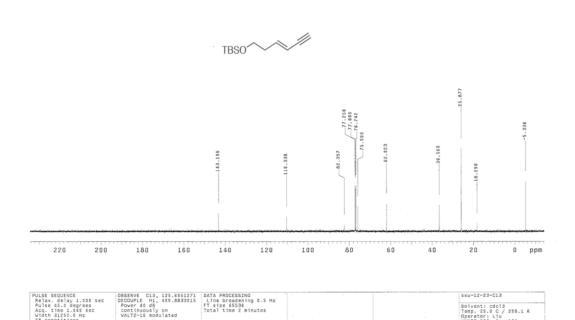
Compound 4n



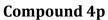


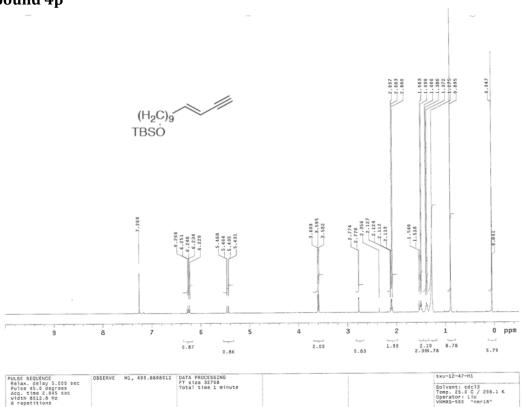
Compound 4o

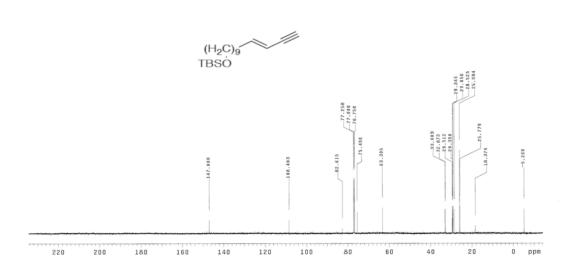




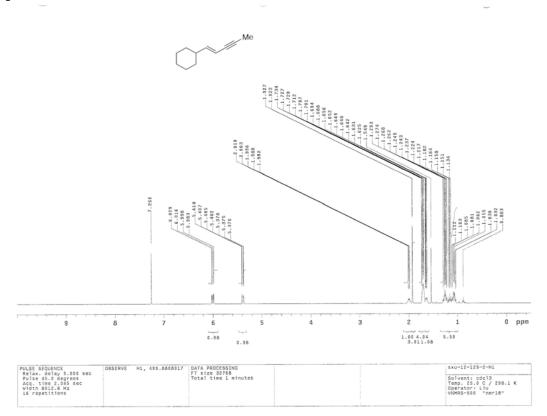
Solvent: cdcl3 Temp. 25.0 C / 298.1 K Operator: Liu VNMRS-500 "nmr18"

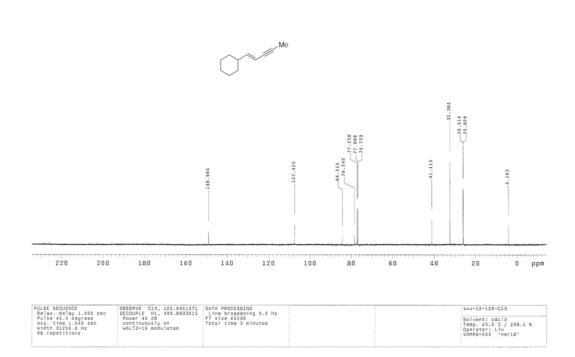




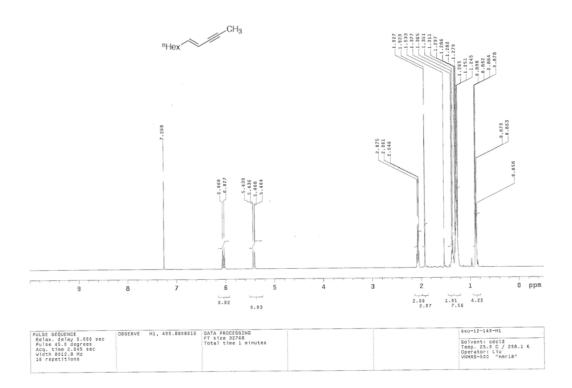


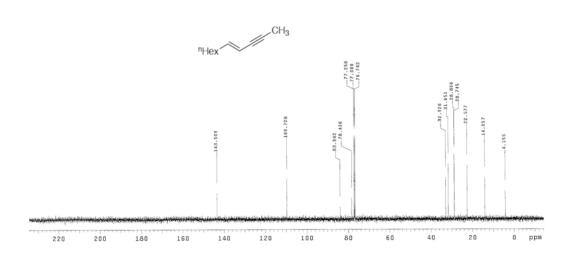
Compound 6a





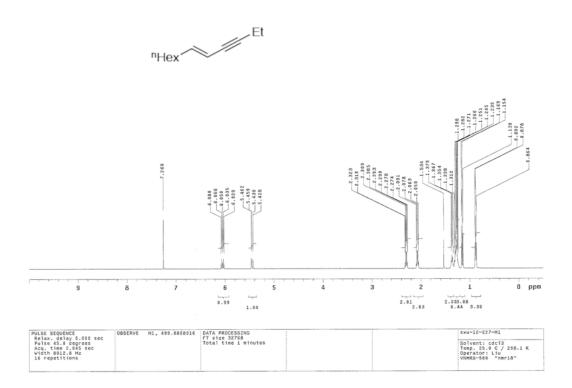
Compound 6b

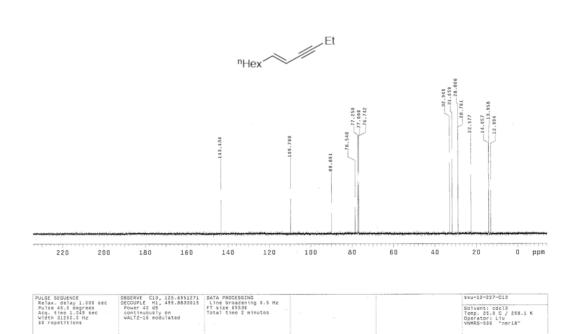




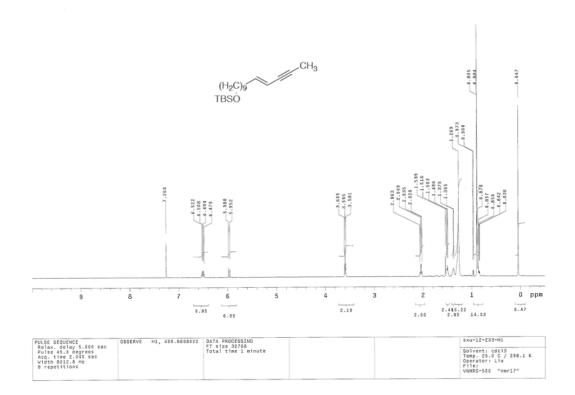
PULSE SEQUENCE OBSERVE C13, 125.691271 DATA PROCESSING Relax delsy 1.960 sec CECQUPLE H., 489.8833015 Line proadening 0.5 Hz Pulse 45.0 degrees Power 48 db Fr size 65536 Acq. tine 1.043 sec Continuously on Total time 1 minutes Vidth 31259.0 Hz VALTZ-16 modulated	sxu-12-149-C13 Solvent: cdcl3 Temp. 25,6 C / 238,1 K Operator: Liu VNMRS-500 "nnr18"
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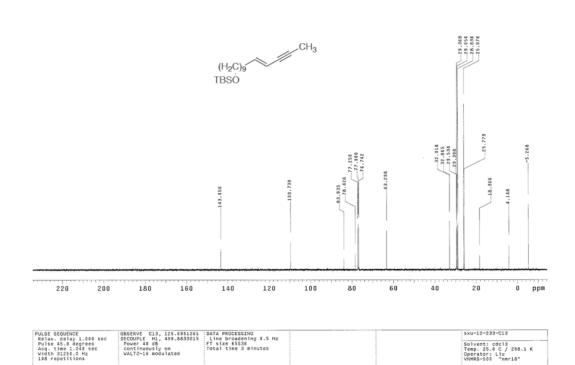
Compound 6c



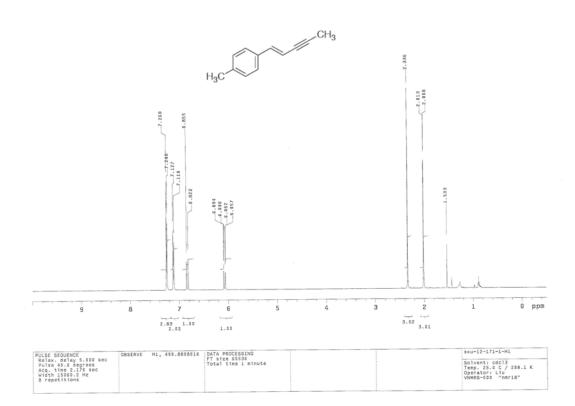


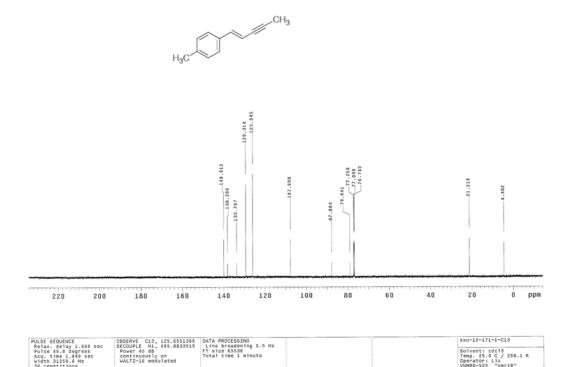
Compound 6f





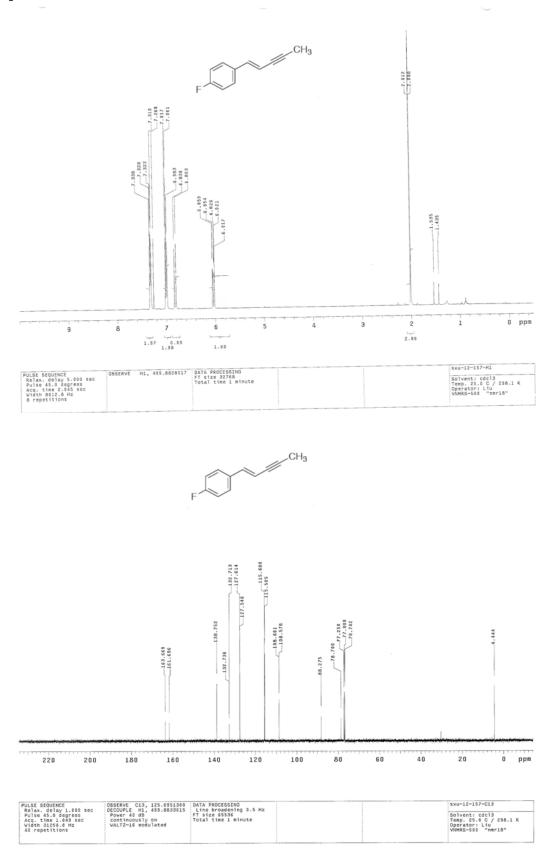
Compound 6h

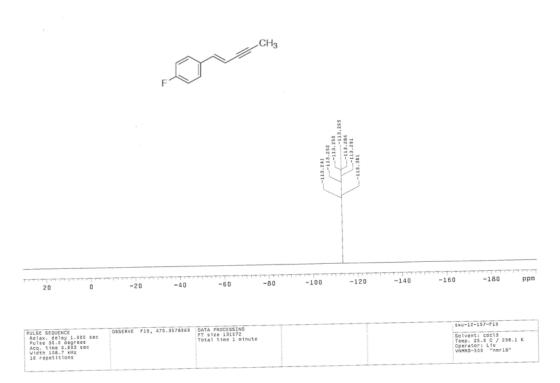




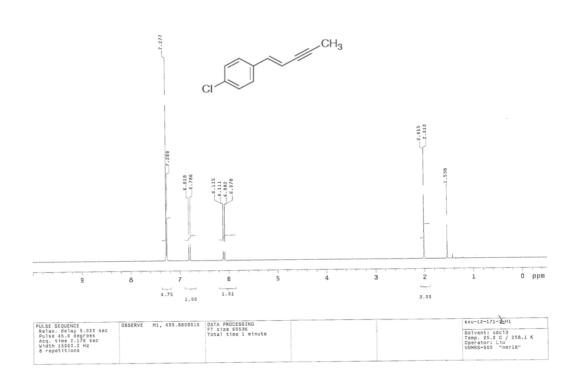
Solvent: cdcl3 Temp. 25.0 C / 298.1 K Operator: Liu VNMRS-500 "nmr18"

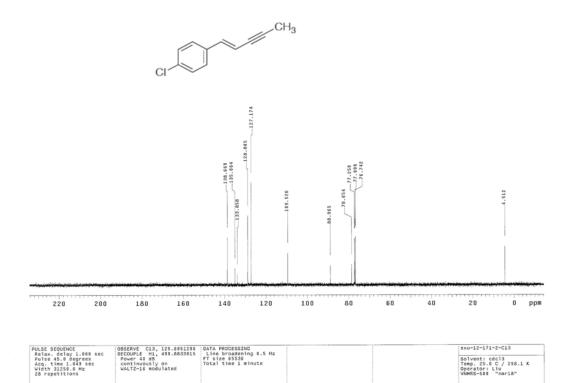
Compound 6i



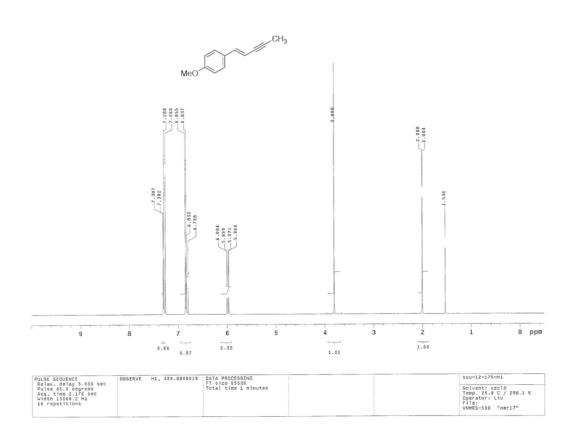


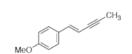
Compound 6j

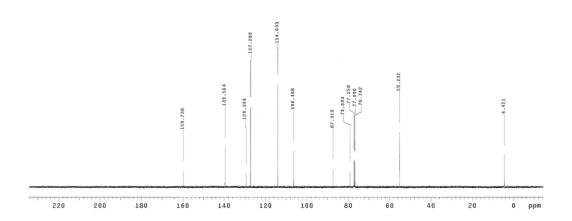




Compound 6k

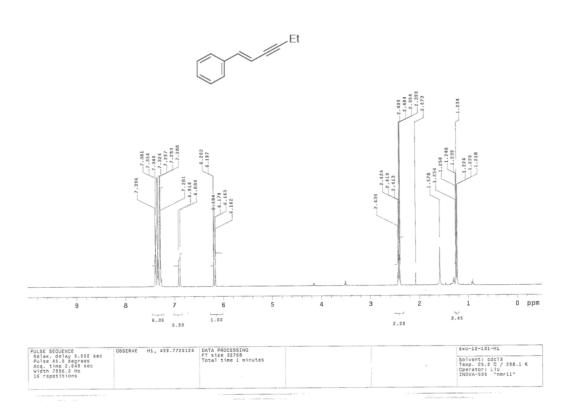


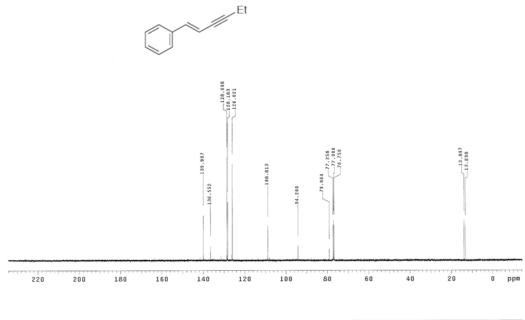




PULSE SEQUENCE OBSERVE C13, 125.6551228 DATA PROCESSING Relax. 648 pl 1.000 sec DECOUPLE H1, 439.8633015 Line broadening 0.5 Hz Pulse 45.0 degrees Power 40 d8 FT size 65536 FT size 65536 Pulse	SOlvent: cdc13 SOlvent: cdc13 Tesp. 25.0 C / 298.1 K Operator L I UNRS-509 "mnr18"
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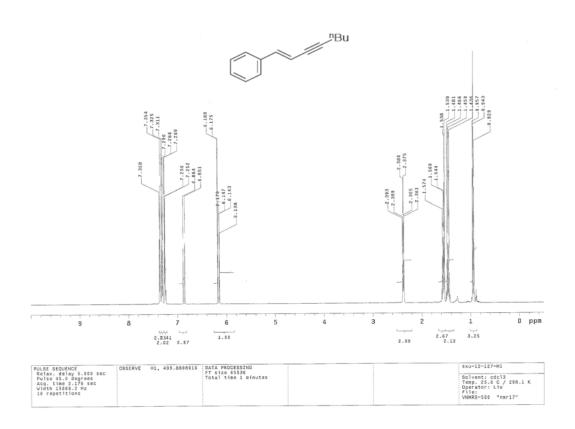
Compound 6m

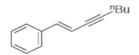


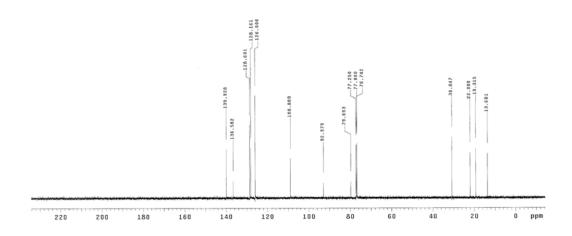


PULSE SEQUENCE Relax. delay 1.000 sec Pulse 45.0 degrees Acq. time 1.049 sec Vidth 31250.0 Hz 42 repetitions	OBSERVE C13, 125.6951309 DECOUPLE H1, 499.8833015 Power 40 dB continuously on WALTZ-16 modulated	\$xu-12-131-C13 Solvent: cdc13 Temp. 25.0 C / 298.1 K Operator: L1u VMM8-520 "mar18"

Compound 6n

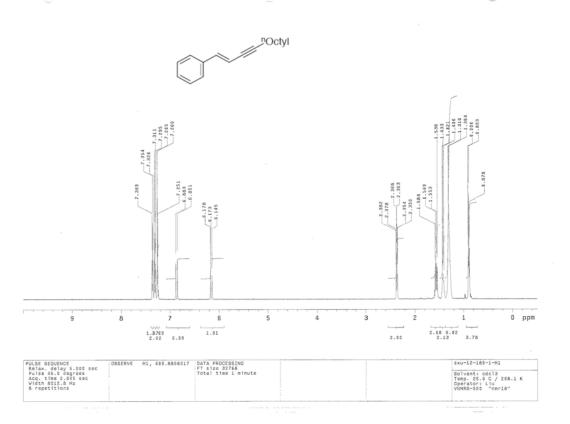




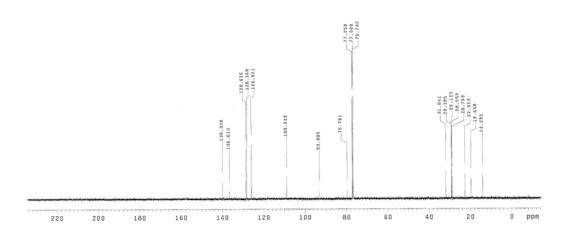


PULSE SEQUENCE Relax. delay 1.000 sec Pulse 45.0 degrees Acq. time 1.049 sec Vidth 31250.0 Hz 38 repetitions	OBSERVE C13, 125.6951300 DECOUPLE H1, 499.8833015 Power 40 dB continuously on WALTZ-16 modulated			Sxu-12-127-C13 Solvent: cdc13 Temp. 25.0 C / 298.1 K Operator: Liu VNMRS-500 "nmr18"
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Compound 60

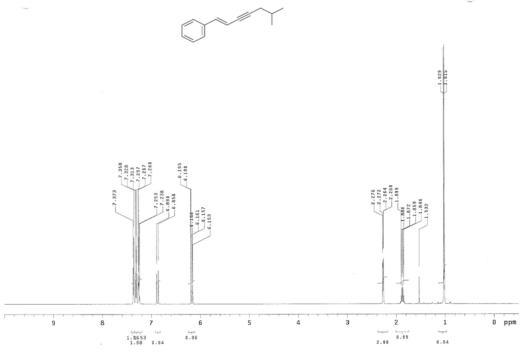




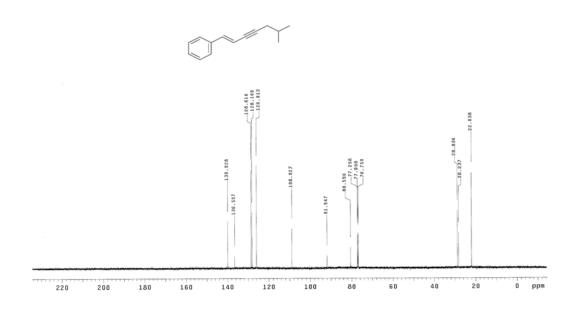


PULSE SEQUENCE Relax. delay 1.000 sec Pulse 45.0 degrees Acq. time 1.049 sec Vidth 31250.0 Hz 208 repetitions	OBSERVE C13, 125.6951271 DECOUPLE H1, 499.8833015 Power 40 dB continuously on WALTZ-16 modulated	DATA PROCESSING Line broadening 0.5 Hz FT size 85336 Total time # minutes	SNU-12-189-1-C13 Solvent: cdc13 Tesp. 25.0 c / 258.1 K Operator: Lu VNNRS-910 "mari8"

Compound 6p

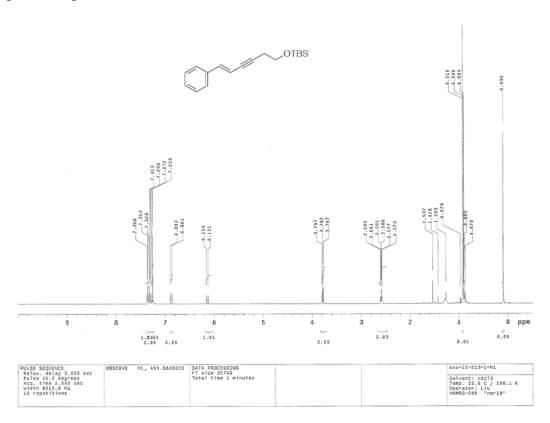


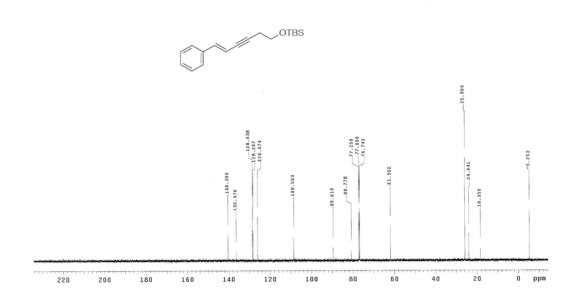
PULSE SEQUENCE Relax. delay 5.000 sec Pulse 45.0 degrees Acq. time 2.065 sec 1 for pulse 45.0 degrees Acq. time	SKU-12-201-2-M1 Solvent: Cdc13 Temp. 25.6 C/ 258.1 K UNMRS-580 "mar18"
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	OBSERVE C13, 125.6951290 DECOUPLE H1, 499.8833915 Power 40 dB continuously on VALTZ-16 modulated	DATA PROCESSING Line broadening 0.5 Hz FT size 65536 Total time 1 minute	\$xu-12-201-3-C13 Solvent: cdc13 Temp. 25.0 C / 298.1 K Operator: Liu VNMRS-500 "nmr18"	

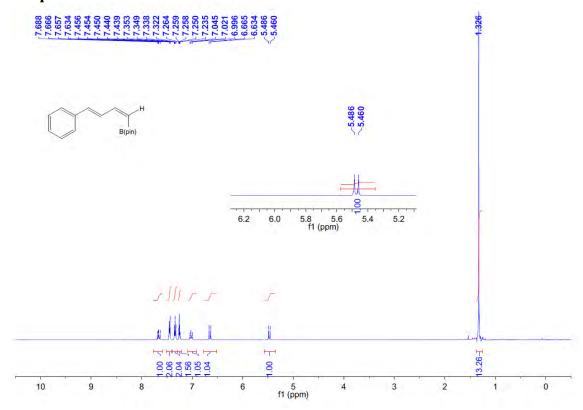
Compound 6q

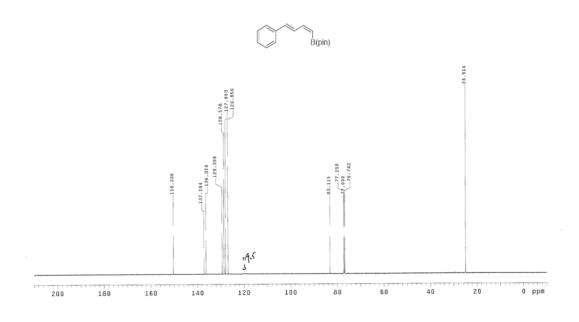




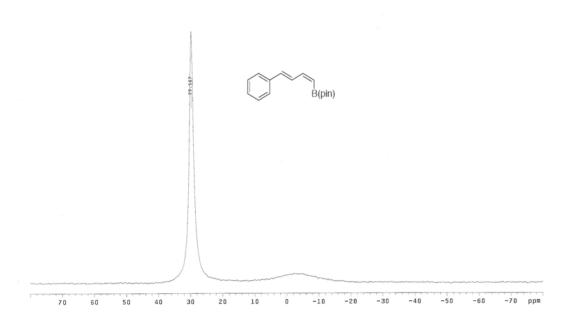
PULSE SEQUENCE Relax. delay 1.000 sec Pulse 45.0 degrees Acq. time 1.045 sec Vidth 31750.3 Hz 44 repetitions VALTZ-16 modulated			Solvent: cdcl3 Temp. 25.0 C / 298.1 K Operator: Liu VNMRS-500 "nmr18"
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Compound 5a



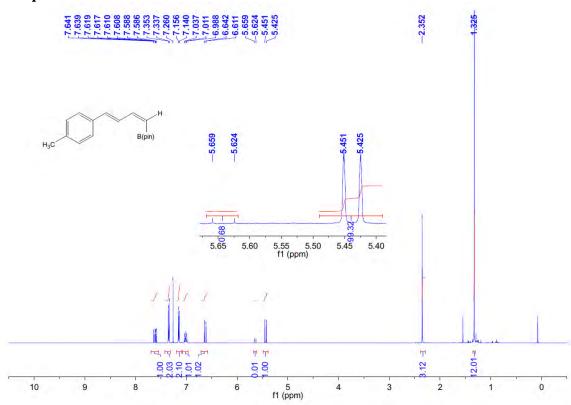


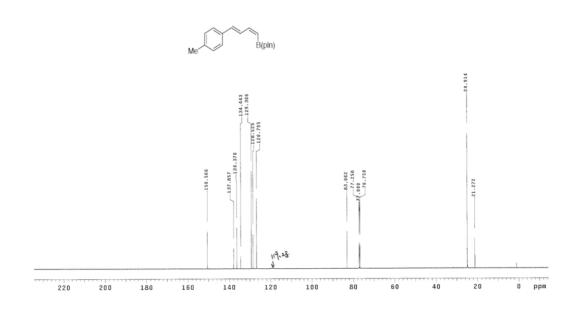
PULSE SEQUENCE CB.2.1.225.0951306 Relax. delay 1.800 sec DECOUPLE M1. 893.8833915 Power 40 dB. Cq. time 1.045 sec continuously on VALTZ-15 modulated		swu-1-241-C13 Solvent: cdc13 Temp. 25.0 C / 298.1 K Operator: Liu File: VMMRS-500 "nmr17"
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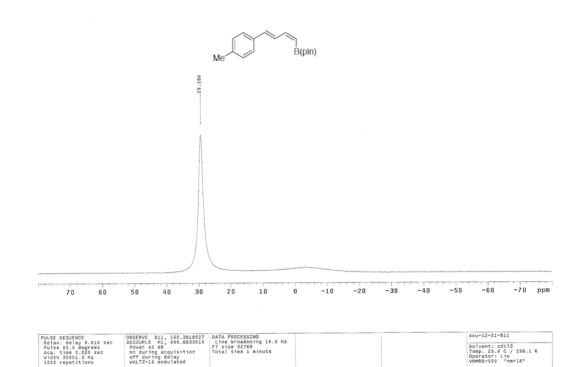
PULSE SEQUENCE	OBSERVE B11, 160.3816266 DECOUPLE H1, 499.8833015	DATA PROCESSING	sxu-11-241-H1
Relax. delay 0.010 sec Pulse 90.0 degrees Acq. time 0.020 sec Width 32051.3 Hz 1024 repetitions	Power 40 dB on during acquisition off during delay WALTZ-16 modulated	FT size 32768 Total time 1 minute	Solvent: cdcl3 Temp. 25.0 C / 298.1 K Operator: Liu File: VNMRS-500 "nmr17"

Compound 5b

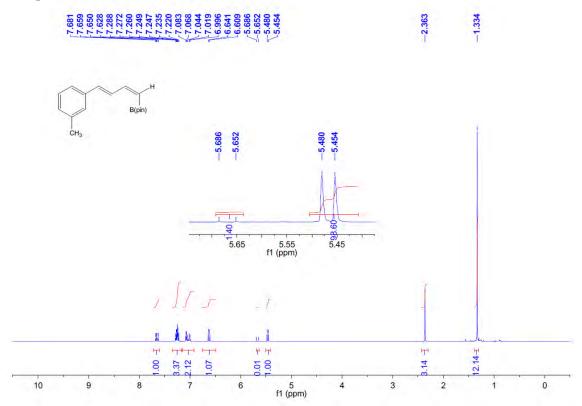


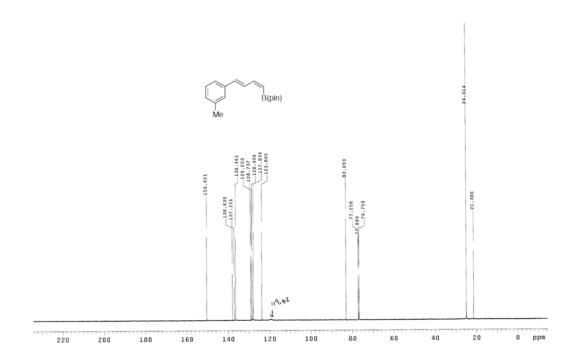


PULSE SEQUENCE	SXU-12-21-C13 Solvent: cdc13 Teap. 25.0 C / 298.1 K Operator: Lfu VNMRS-500 "nmr18"
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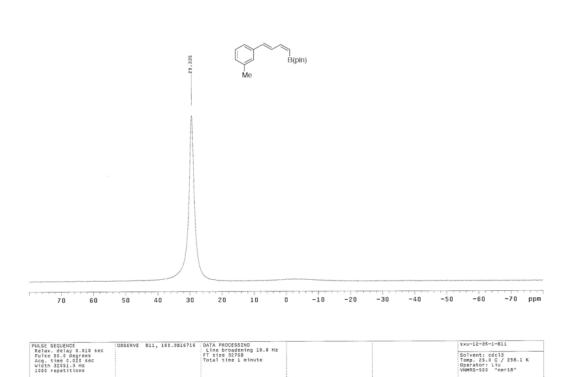


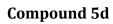
Compound 5c

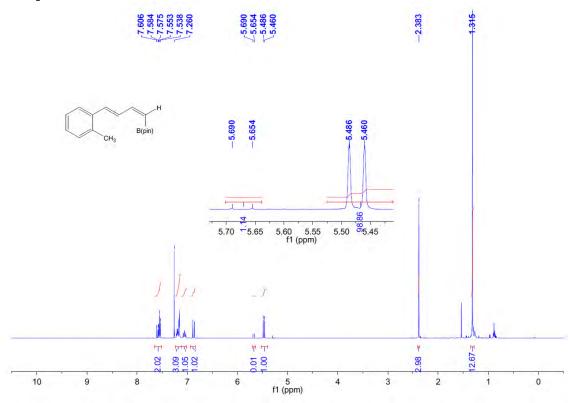


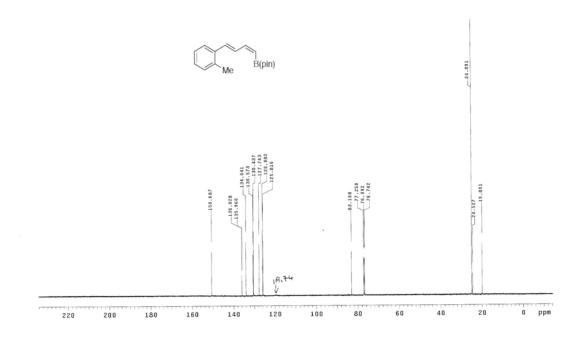


PULSE SEQUENCE Relax. delay 1.000 sec Pulse 45.0 degrees Acq. time 1.049 sec Width 31250.0 Hz 436 repetitions	OBSERVE C13, 125.6951309 DECOUPLE H1, 499.8833015 Power 40 dB continuously on WALTZ-16 modulated	DATA PROCESSING Line broadening 0.5 Hz FT size 65538 Total time 14 minutes	sxu-12-51-C013 Solvent: cdc13 Temp. 25.0 C / 298.1 K Operator: Liu VNNR-500 "mer18"



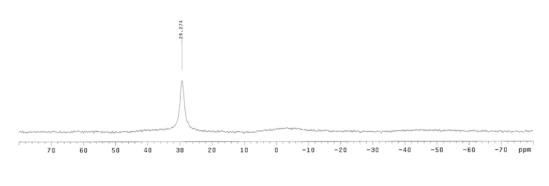






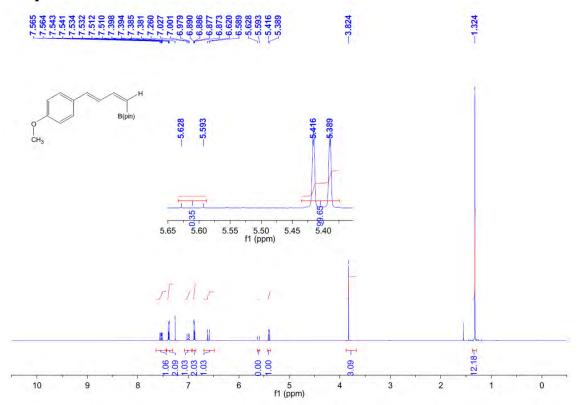
PULSE SEQUENCE Relax. delay 1.000 sec Pulse 45.0 degrees Acq. time 1.048 sec Vidth 3125.0 %tz 230 repetitions OBSERVE C13, 125.6951309 DECOUPLE H1, 439.8033915 Power 40 d6 continuously or VALTZ-16 modulated	DATA PROCESSING Line broadening 0.5 Hz Ff size 6558 Total time 7 minutes	Sxu-12-81-1-C13 Solvent: cdc18 Temp. 25.0 C / 258.1 K Operator: Liu VANMS-500 "ner18"
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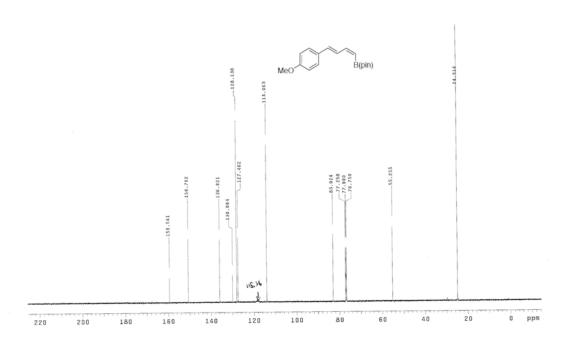




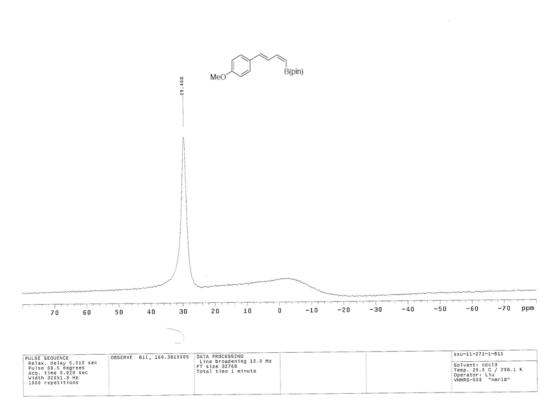
PULSE SEQUENCE G08SERVE 811, 180.3818798 GDATA PROCESSING Relax delay 0.010 sec DECOUPLE H1, 458.683915 interpretation 10.0 Mz Pulse 90.0 degrees Power 40 dB F5 size 32788 Acq. time 0.000 sec on during acquisition Total time 1 minute Viola 2001.3 Hz Valle-18 modulated Viola 2001.3 Hz Godding Company	Sxu-12-81-1-811 Solvent: cdcl3 Temp. 25.0 C / 298.1 K Operator: Ltu VNMRS-500 "nmr18"
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Compound 5e

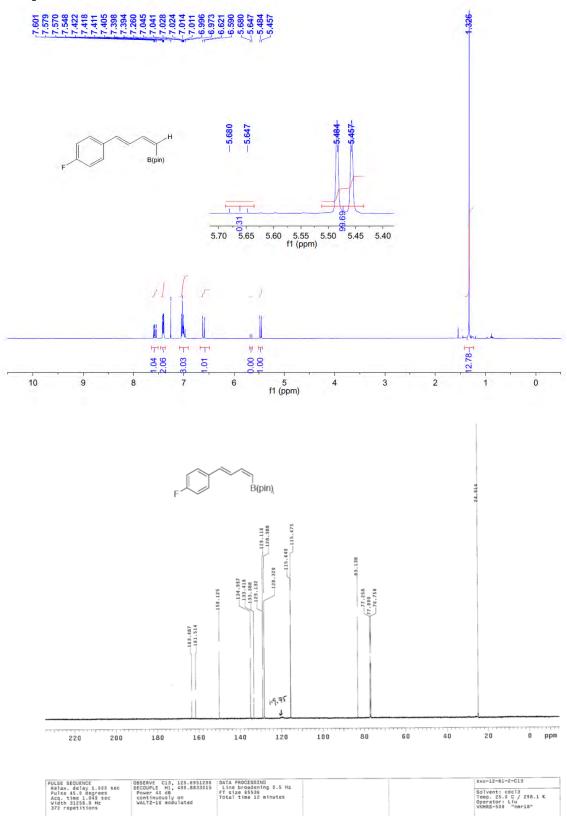


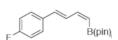


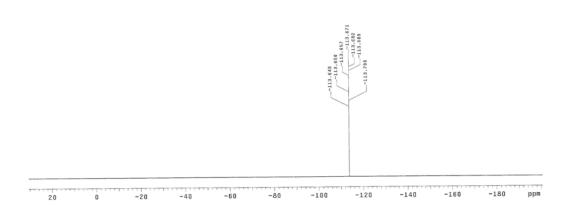
PULSE SEQUENCE Relax. delay 1.000 sec Pulse 45.0 degrees Acq. time 1.043 sec Width 31250.0 Hz 210 repetitions	OBSERVE C13, 125.6851309 OECOUPLE HI, 499.8833315 Power 40 dB continuously on WALTZ-16 modulated	DATA PROCESSING Line broadening 0.5 Hz FT size \$5535 Total time 7 minutes	sku-11-271-CL3 Solvent: cdc13 Temp. 25.0 C / 258.1 K Operator: Liu VMRRS-500 "mnr18"
			 y and a solution of the y



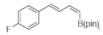
Compound 5f

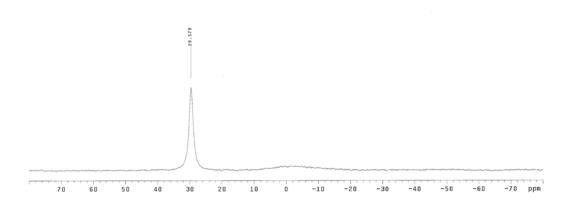




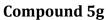


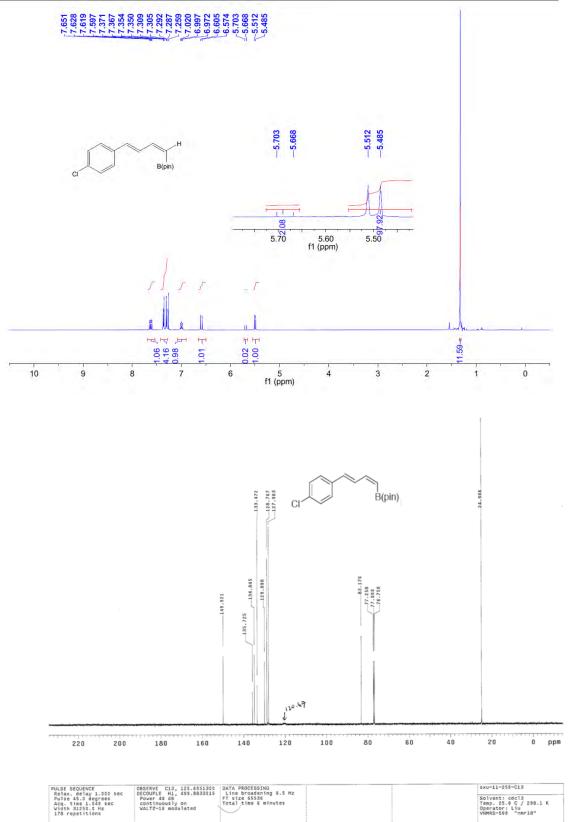
PULSE SEQUENCE Relax. delay 1.000 sec Pulse 30.0 degrees Acq. time 0.693 sec Width 108.7 kHz 14 repetitions	OBSERVE	F19, 470.3578969	DATA PROCESSING FT size 131072 Total time 1 minute		SCHUZZ-81-Z-F15 SCHURT: CGC13 Temp. 25.6 C / 296.1 K Operator: Lit VMMRS-500 "nmr18"
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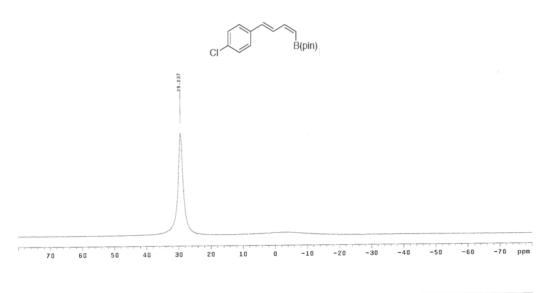




PULSE SEQUENCE Relax, delay 0.010 sec	OBSERVE B11, 160.3816266 DECOUPLE H1, 499.8833015	DATA PROCESSING Line broadening 10.0 Hz		sxu-12-81-2-B11
Pulse 90.0 degrees Acq. time 0.020 sec Vidth 32051.3 Hz 100 repetitions	Power 40 dB on during acquisition off during delay VALTZ-16 modulated	FT size 32768 Total time 1 minute		Solvent: cdcl3 Temp. 25.0 C / 298.1 K Operator: Liu VNMRS-500 "nmr18"

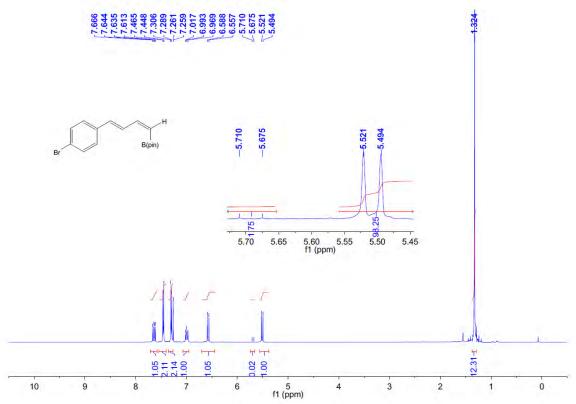


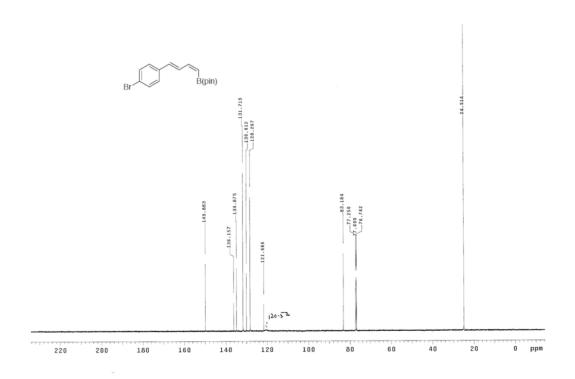




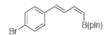
PULSE SEQUENCE Relax. delay 0.010 sec	OBSERVE	B11, 1	160.3819005	Line broadening 10.0 Hz	Т		sxu-11-259-81	
Pulse 30.0 degrees Acq. time 0.020 sec Width 32051.3 Hz 1000 repetitions				FT size 32768 Total time 1 minute			Temp. 25.0 C . Operator: Liu	/ 298.1 K

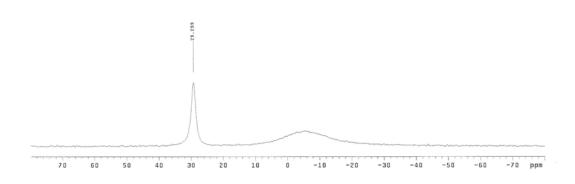
Compound 5h



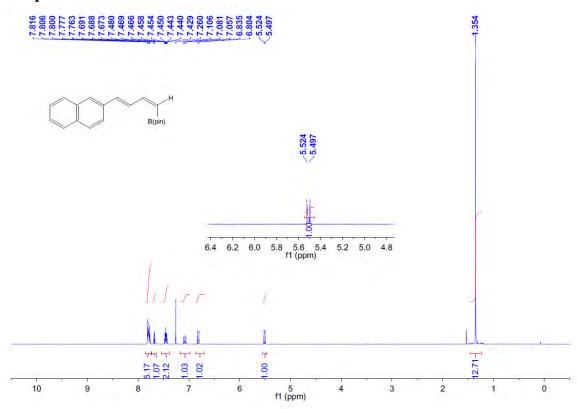


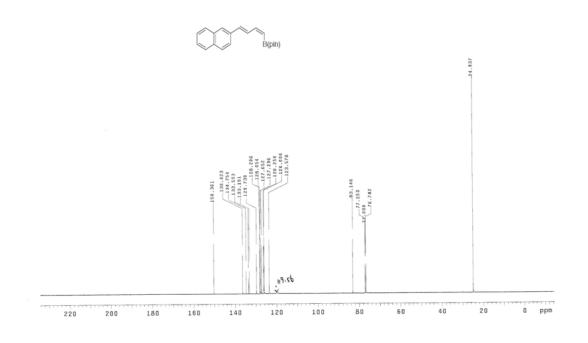
PULSE SEQUENCE Relax. delay 1.000 sec Pulse 45.0 degrees Acq. time 1.049 sec Width 31250.0 Hz 270 repetitions				Sxu-12-87-C13 Solvent: cdc13 Temp. 25.0 C / 298.1 K Operator: Liu vNMRS-500 "nmr18"
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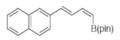


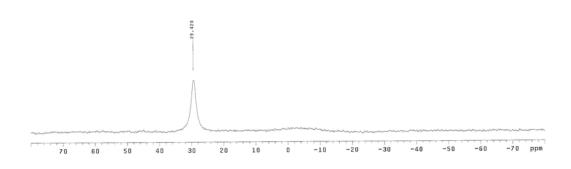
Compound 5i





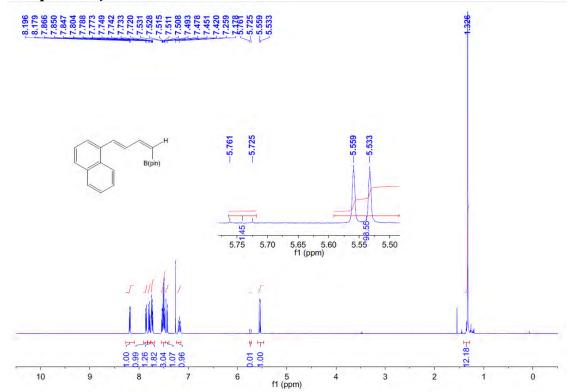
PULSS SEQUENCE DBSERVE C.13, 125.6951338 PULSS 45.0 degrees Power 49 dB	DATA PROCESSING Line broadening 0.5 Hz FT size 65538 Total time 24 minutes	sxu-12-79-2-Cl3 Solvent: Cd13 Tesp. 25.8 C / 298.1 K Operator: L1u VMM85-590 "mer18"

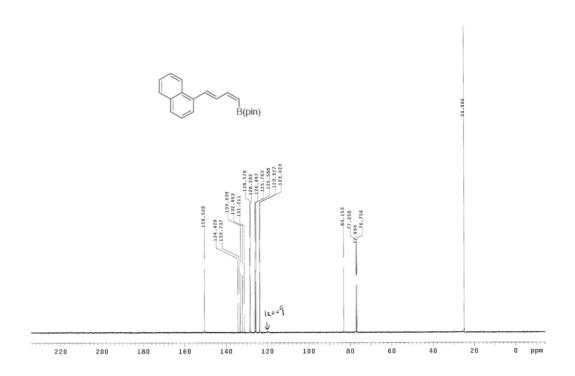




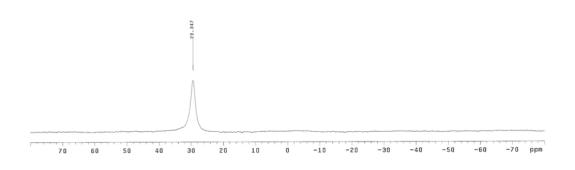
PULSE SEQUENCE Relax. delay 8.010 sec Pulse 36.0 degrees Acq. time 0.020 sec Vice 100 repetitions VALTZ-18 modulated		sxu-12-79-2-811 Solvent: cdc13 Solvent: cdc13
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Compound 5j



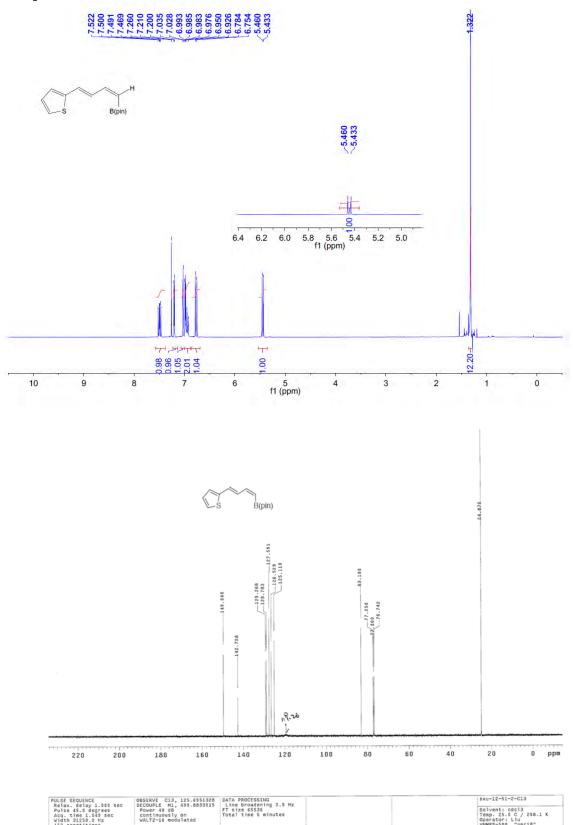


PULSE SEQUENCE	OBSERVE C13, 125.6951319 DECOUPLE H1, 499.8833015	DATA PROCESSING Line broadening 0.5 Hz		sxu-12-79-3-C13
Relax. delay 1.000 sec Pulse 45.0 degrees Acq. time 1.049 sec Width 31250.0 Hz 308 repetitions	Power 40 dB continuously on WALTZ-16 modulated	FT size 65536 Total time 10 minutes		Solvent: cdcl3 Temp. 25.0 C / 298.1 K Operator: Liu vNMRS-500 "nmr18"

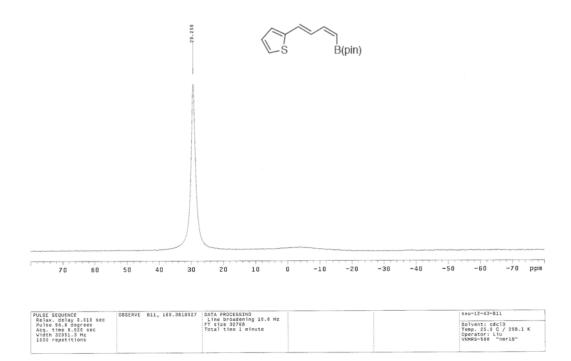


PULSE SEQUENCE	OBSERVE B11, 160.3816736		sxu=12=79=3=B11
Relax. delay 0.010 sec Pulse 30.0 degrees Acq. time 0.020 sec Width 32051.3 Hz 100 repetitions	DECOUPLE H1, 499.8833015 Power 40 dB on during acquisition off during delay VALTZ-16 modulated	Line broadening 10.0 Hz FT size 32768 Total time 1 minute	Solvent: cdcl3 Temp. 25.0 C / 298.1 K Operator: Liu VMMRS-500 "nmr18"

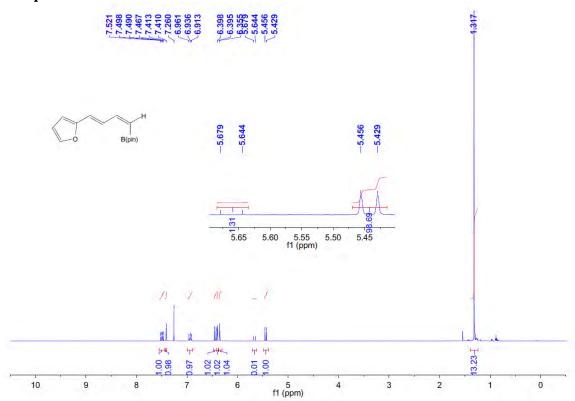
Compound 5k

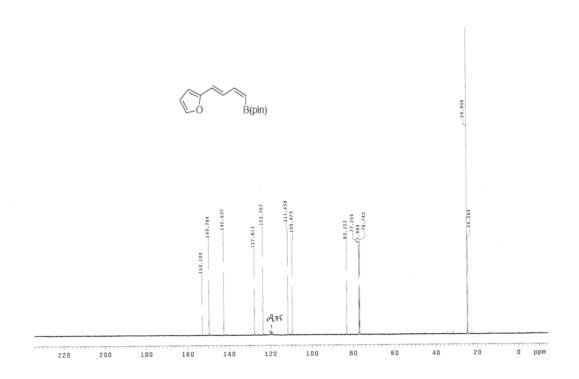


Solvent: cdcl3 Temp. 25.0 C / 298.1 K Operator: Liu VNMRS-500 "nmr18"

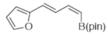


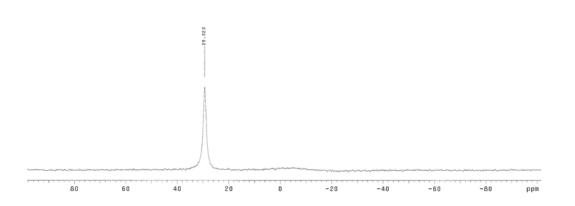
Compound 51





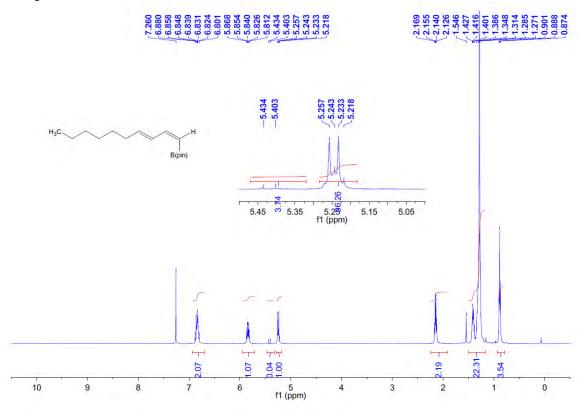
PULSE SEQUENCE Relaw. delay 1:000 sec Plase 4.5 sec Action 1:000 sec Width 81259.0 Hz 332 repetitions DESCUPIE: C:3, 125.8551300 SECUPIE: C:3, 125.	DATA PROCESSING Line broadening 0.5 Hz FT size 65536 Total time 11 minutes	sxu-12-79-1-013 Solvent: cdc13 Tesp. 25.0 C / 238.1 K Operator: 1 tu VKMRS-509 "nmr18"

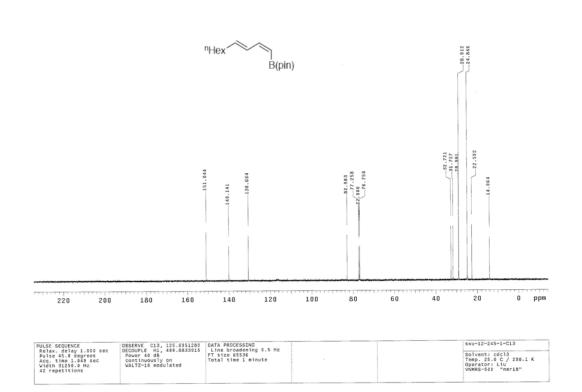


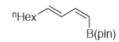


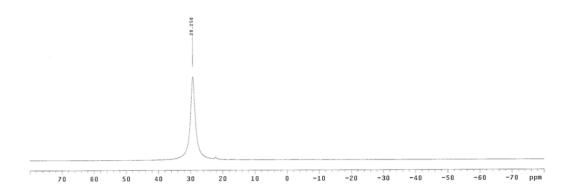
PULSE SEQUENCE Relax. delay 0.010 sec Pulse 90.0 degrees Acq. time 0.020 sec Vidth 32051.3 Hz 190 repetitions	OBSERVE 811, 160.3818887 DECOUPLE HI, 499.8833915 Power 40 dB on during acquisition off during delay VALTZ-16 modulated	OATA PROCESSING Line broadening 10.0 Hz FT size 32768 Total time 1 minute	SNU-12-79-1-011 SNU-011 cdc13 Temper 25-9 C J 298.1 K Operator: Liu VANKES-500 "ner10"
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Compound 5m



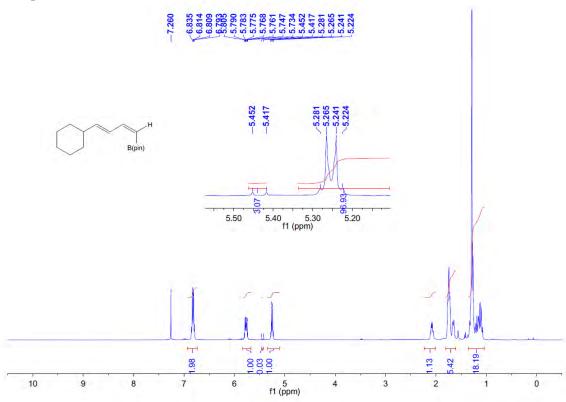


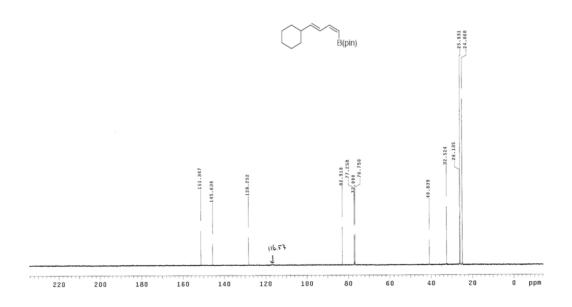




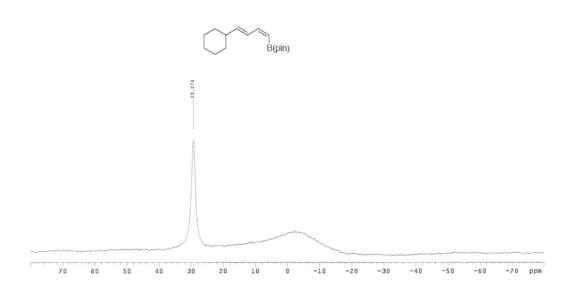
Relax. delay 0.010 sec DE Pulse 90.0 degrees P. Acq. time 0.020 sec O Vidth 32051.3 Hz 0	COUPLE H1, 499.8833015 Power 40 dB	DATA PROCESSING Line broadening 10.0 Hz FT size 32768 Total time 1 minute		Sxu-12-245-1-B11 Solvent: cdcl3 Temp. 25.0 C / 298.1 K Operator: Liu vNMRS-500 "nmr18"

Compound 5n



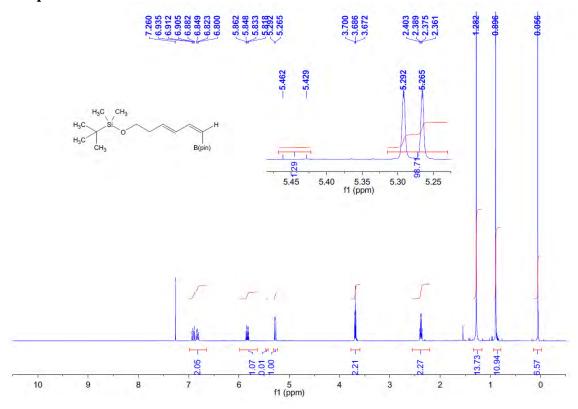


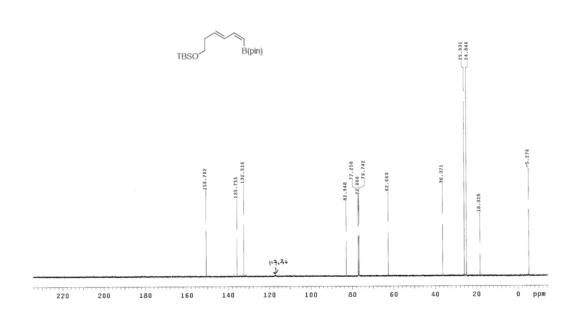
PULSE SEQUENCE Relax. delay 1.000 sec Pulse 45.0 degrees Power 46 ds Power 45 ds VALTZ-16 modulated VALTZ-18 modulated VALTZ-18 modulated Total time 3 minutes	SU-11-277-C13 Solvent: GGC 238.1 K Temp. 22.6 C / 238.1 K Operator: L'u VMMRS-500 "nnr18"
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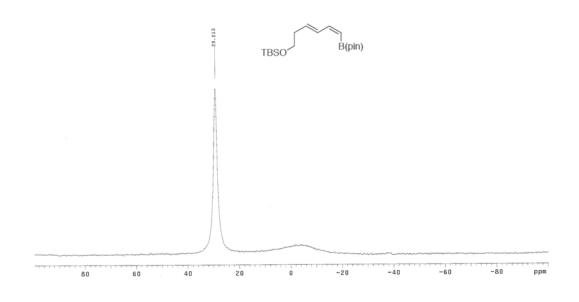


PULSE SEQUENCE Relax, delay 0,010 sec	OBSERVE B11, 160.3818966	DATA PROCESSING Line broadening 10.0 Hz	sxu-11-277-811	
Pulse 60.5 degrees Acq. time 0.020 sec Width 32051.3 Hz 1024 repetitions			FT size 32768 Total time 1 minute	Solvent: cdcl3 Temp. 25.0 C / 298.1 K Operator: Liu VAMAS-500 "maria"

Compound 50

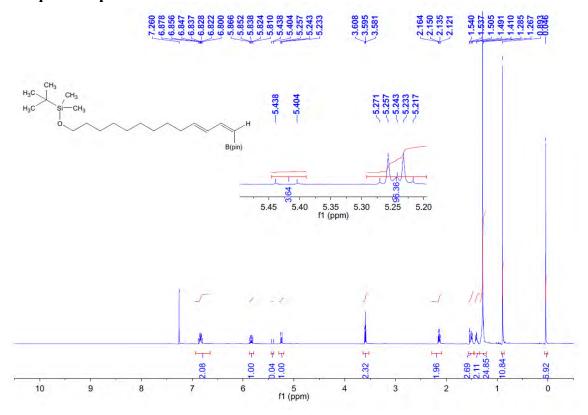


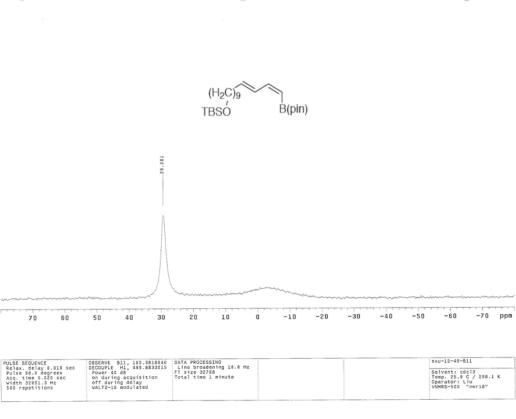




	OBSERVE 811, 160.3816716	DATA PROCESSING	sxu-12-35-2-811
Relax. delay 0.010 sec Pulse 90.0 degrees Acq. time 0.020 sec Vidth 32051.3 Hz 1000 repetitions		Line broadening 10.0 Hz FT size 32768 Total time 1 minute	Solvent: cdc13 Temp. 25.0 C / 298.1 K Operator: Liu VNMRS-500 "nmr18"

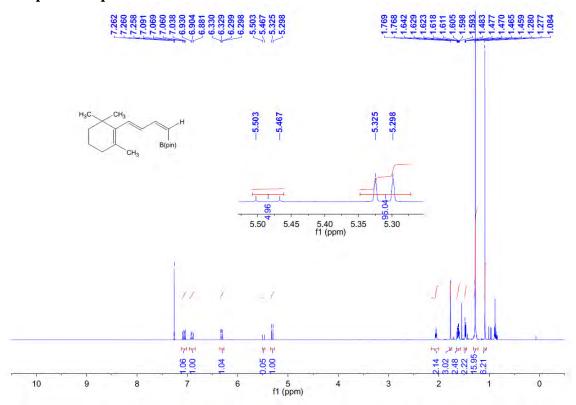
Compound 5p

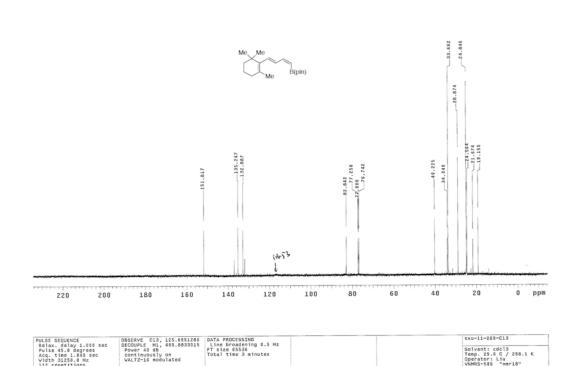


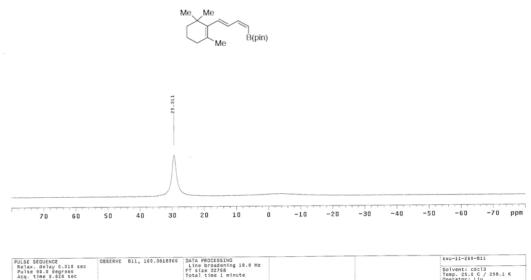




Compound 5q

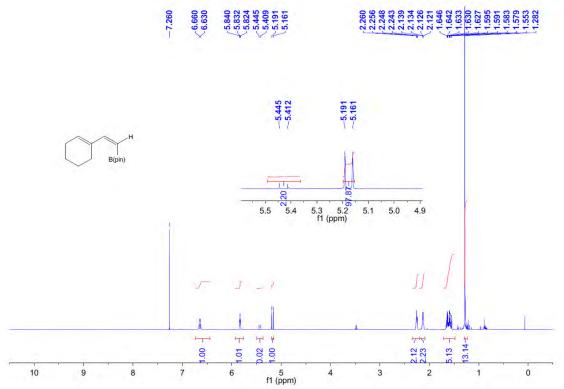


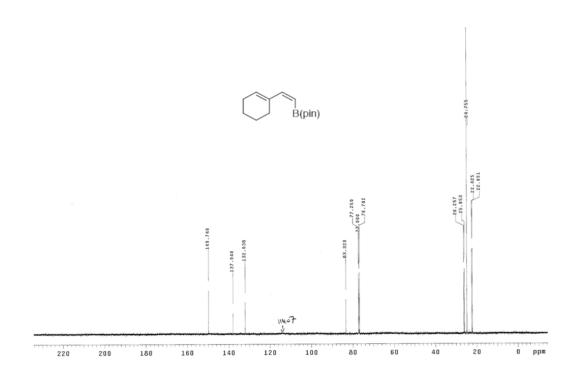




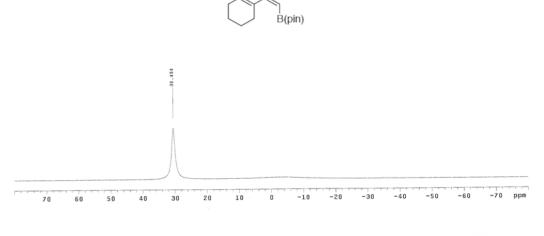
PULS SECUENCE Relax - Gelay 0.310 sec Pulse 56.0 degrees Fine broadening 10.0 Hz Fine broadening 10.0

Compound 5r



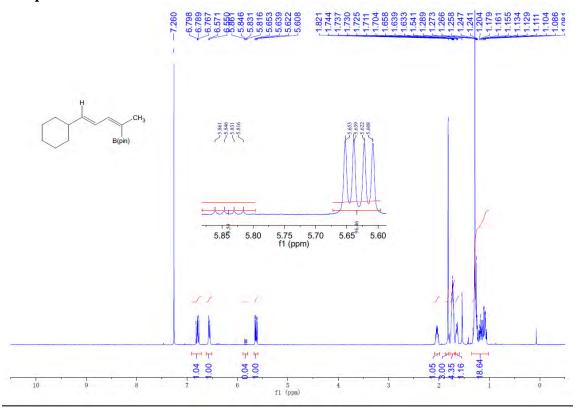


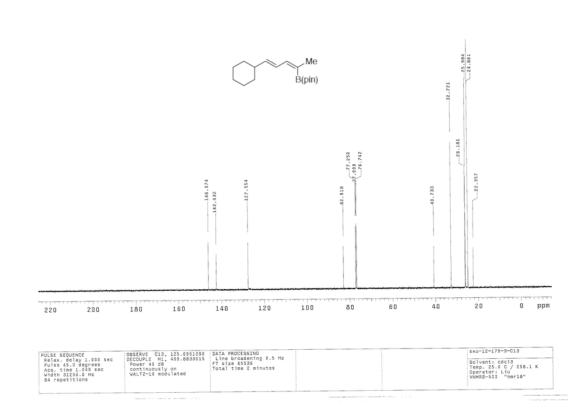
PULSE SEQUENCE Relax. delay 1.000 sec Pulse 45.6 degrees Acq. time 1.849 sec Victin 312.89 de Las repetitions OBSERVE C13, 125.695128 DECOUPLE H1, 439.88331 DECOUPLE H1, 439.8831 DEC			Sxu-11-257-C13 Solvent: cdcl3 Temp. 25.0 C / 298.1 K Operator: Liu VNMRS-500
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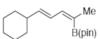


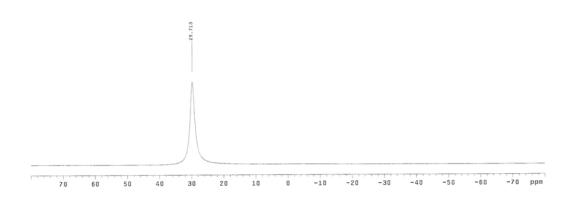
PULSE SEQUENCE CREATER SEQUENCE CONSERVE B11, 168.381 BECOUPLE H1, 498.883 Poles CREATER SEQUENCE CREATER SE	3015 Line broadening 10.0 Hz FT size 32768	SXU-11-257-811 Solvent: cdc13 Tep. 25.0 C / 286.1 K Operator: Liu VMMRS-560 "nmr18"

Compound 7a









PULSE SEQUENCE Relax, delay 0.010 sec	OBSERVE B11, 160.3816755 DECOUPLE H1, 499.8833015	DATA PROCESSING		sxu-12-179-3-811
Pulse S0.0 degrees Acq. time 0.020 sec Width 32051.3 Hz 1000 repetitions	Power 40 dB on during acquisition off during delay VALTZ-16 nodulated	FT size 32768 Total time 1 minute		Solvent: cdcl3 Temp. 25.0 C / 298.1 K Operator: Liu VNMRS-500 "nmr18"
1000 100011110110				

H_c H_a

Нь

7a

Me

B(pin)

yz-2-203H-1 Selective band center: 1.82 (ppm); width: 8.8 (Hz)

Sample Name:

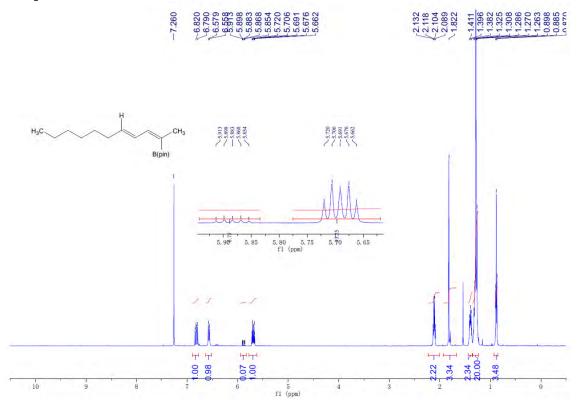
Data Collected on: nmr19-vnmrs600 Archive directory:

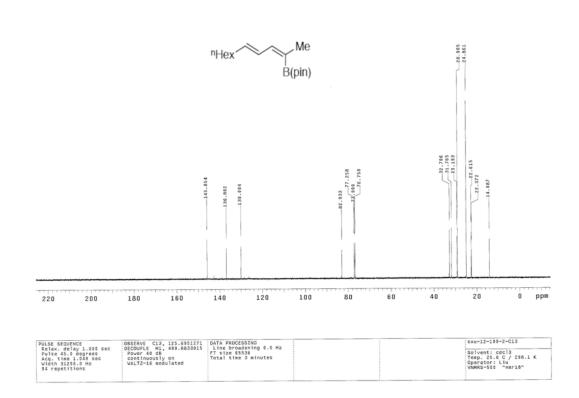
Sample directory:

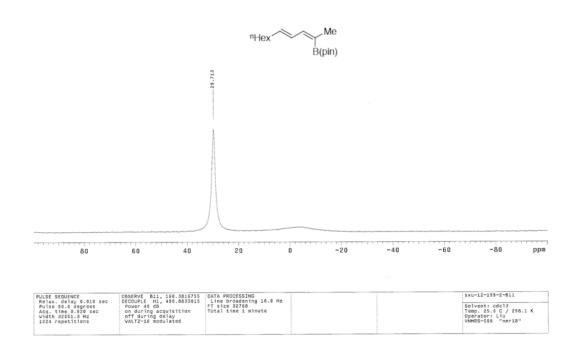
FidFile: yz-2-283H-1-noesy1

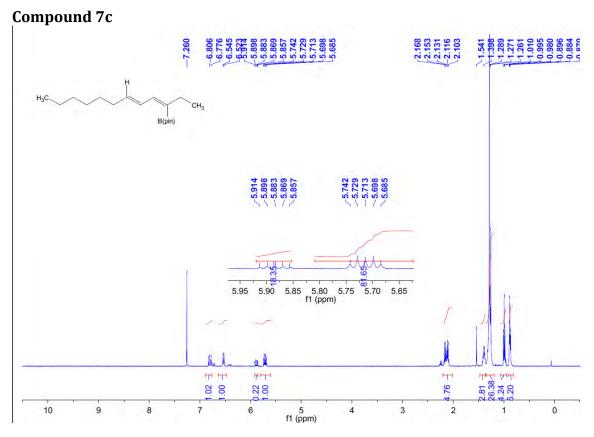
Pulse Sequence: NOESY1D Solvent: cdcl3 Data collected on: Oct 18 2016 $-\mathsf{H}_{\mathsf{a}}$ -Ме 10 6 5 ppm 5.53-€

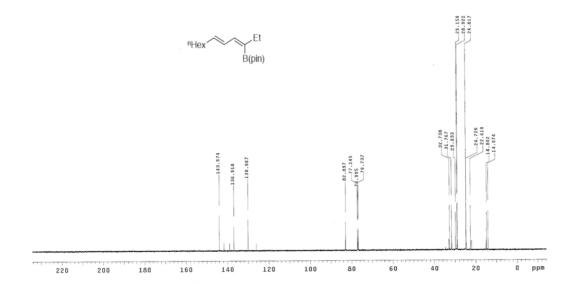
Compound 7b



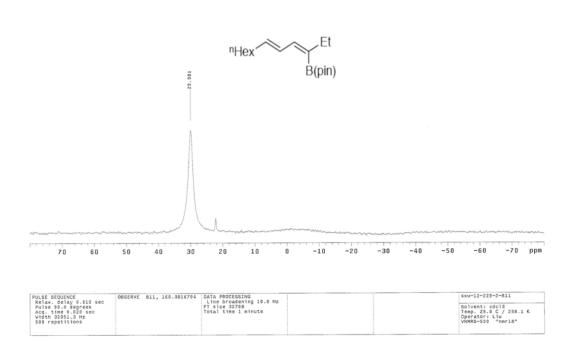




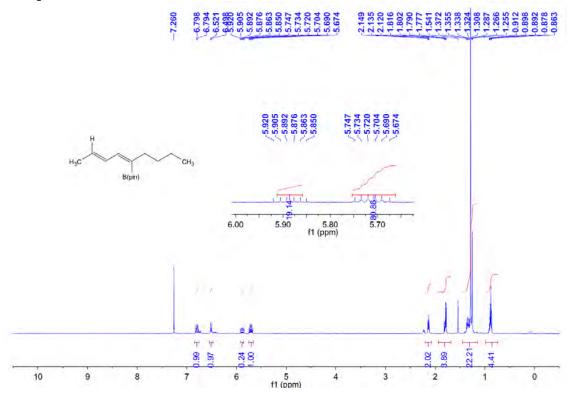


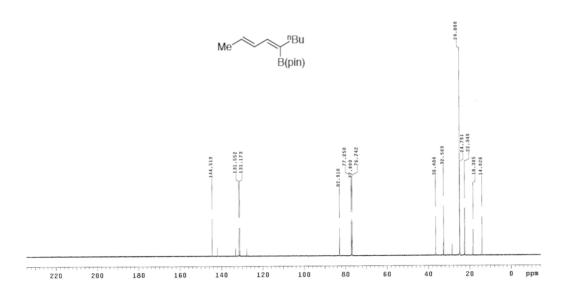


Relax. delay 1.000 sec DE Pulse 45.0 degrees Po Acg. time 1.049 sec G	ISERVE C13, 125.5951268 COUPLE H1, 499.8833615 Ower 46 dB Ontinuously on MALTZ-16 modulated	OATA PROCESSING Line broadening 0.5 Hz FT size 68536 Total time 3 minutes		Sxu-12-228-2-C13 Solvent: cdc13 Temp. 25.0 C / 298.1 K Operator: Liu vNMRS-500 "ner18"
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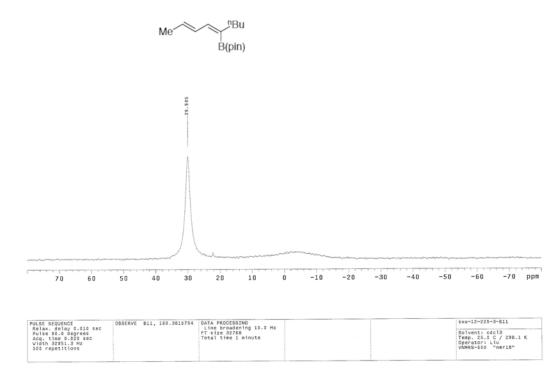


Compound 7d

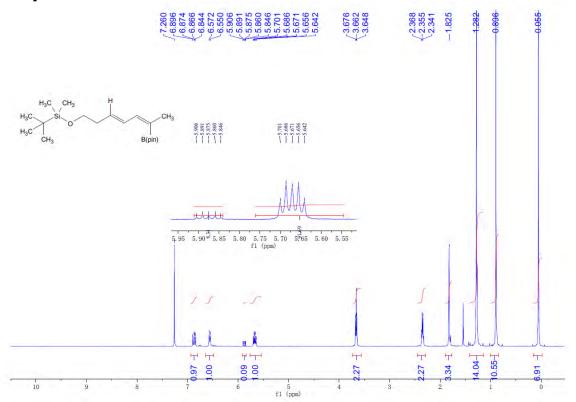


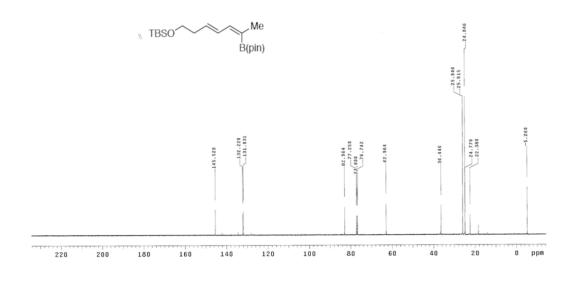


	OBSERVE C13, 125.6951271	DATA PROCESSING Line broadening 0.5 Hz	sxu-12-229-3-C13
Relax. delay 1.000 sec Pulse 45.0 degrees Acq. time 1.048 sec Width 31250.0 Hz 324 repetitions	DECOUPLE H1, 499.8833015 Power 40 dB continuously on WALTZ-15 modulated	time broadening w.s nz Ff size 6536 Total time 11 minutes	Solvent: cdc13 Temp. 25.8 C / 298.1 K Operator: Liu VNMRS-509 "nmr18"

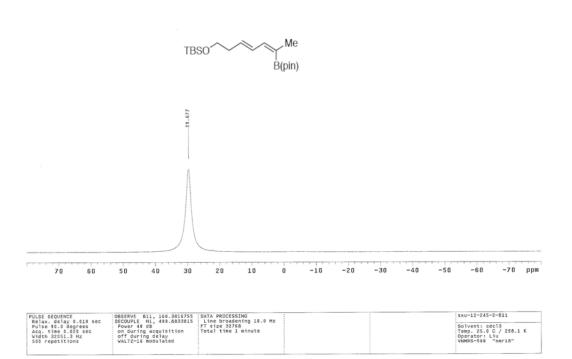


Compound 7e

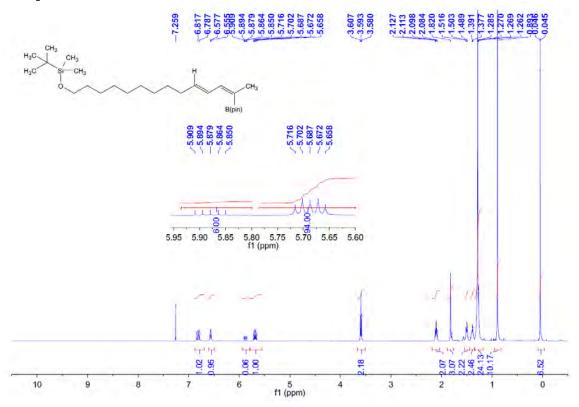


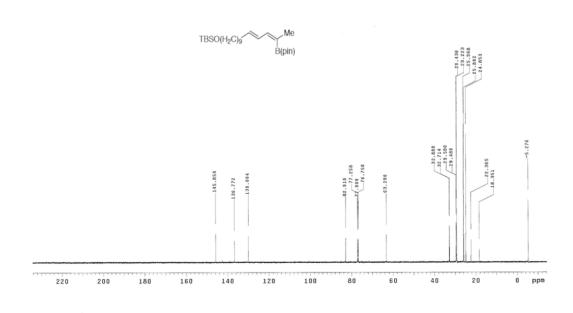


Relax. delay 1.000 sec DECOU	UPLE H1, 499.8833015	DATA PROCESSING Line broadening 0.5 Hz FT size 65536		sxu-12-245-2-C13 Solvent: cdc13
Acq. time 1.049 sec cont		Total time 2 minutes		Temp. 25.0 C / 298.1 K Operator: Liu VNMRS-500 "nmr18"

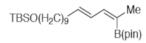


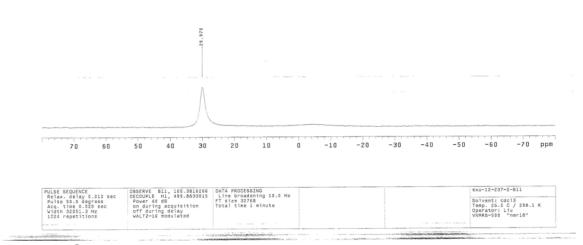
Compound 7f



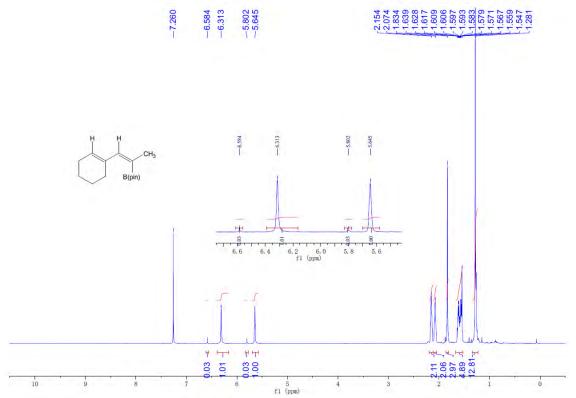


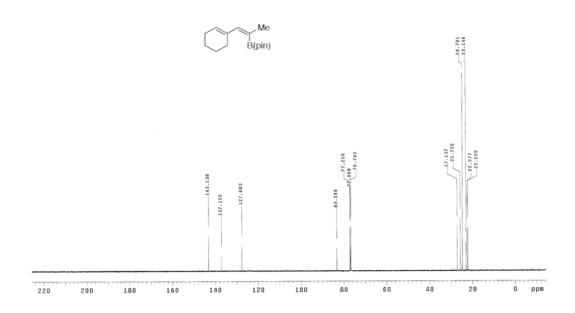
PULSE SEQUENCE Relax, delay 1,000 sec	OBSERVE C13, 125.6951280 DECOUPLE H1, 499.8833015		sxu-12-237-2-C13
Pulse 45.0 degrees Acq. tine 1.049 sec Width 31250.0 Hz 46 repetitions	Power 40 dB continuously on WALTZ-16 modulated	FT size 65536 Fotal time 1 minutes	Solvent: cdc13 Temp, 25: 0 C / 298.1 K Operator: Liu VHMRS-500 "mmr18"





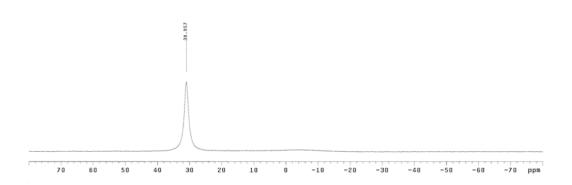
Compound 7g



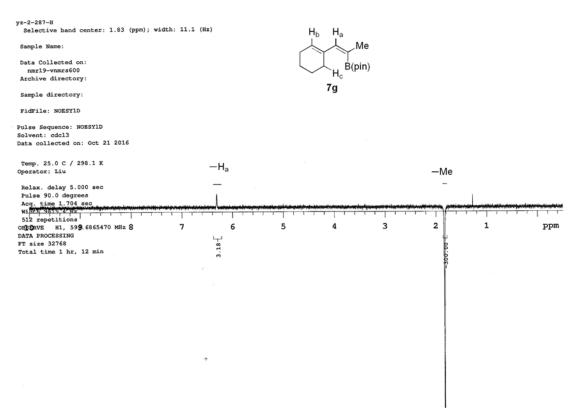


PULSE SEQUENCE Relax. delay 1.000 sec Pulse 45.0 degrees	OBSERVE C13, 125-6951280 DECOUPLE H1, 499-8833015 Power 40 dB		sxu-12-241-C13 Solvent: cdcl3
Acq. time 1.049 sec Width 31250.0 Hz 210 repetitions	continuously on WALTZ-16 modulated	Total time 7 minutes	Temp. 25.0 C / 298.1 K Operator: Liu VNMRS-500 "nmr18"

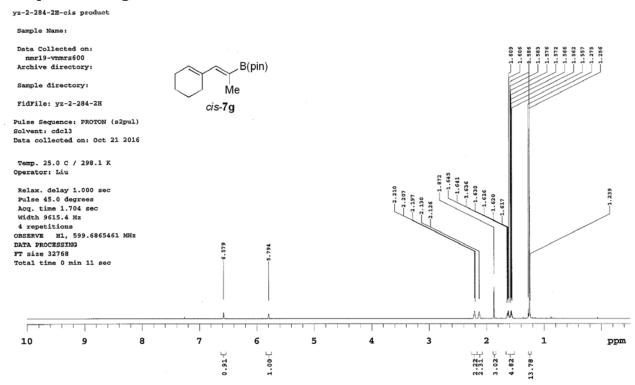




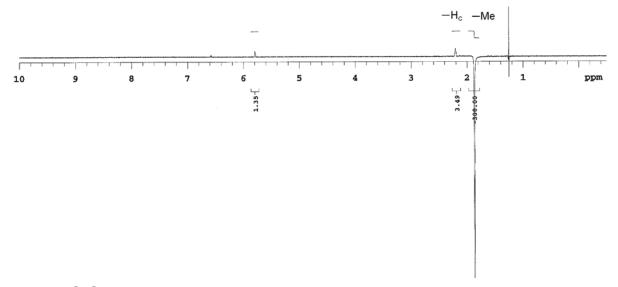
PULSE SEQUENCE Relax, delay 0.010 sec	OBSERVE B11, 160.3816775 DECOUPLE H1. 499.8833015		tsxu-12-237-1-B11
Pulse 90.0 degrees Acq. time 0.020 sec Vidth 32051.3 Hz 1000 repetitions	Power 40 dB on during acquisition off during delay WALTZ-16 modulated	FT size 32768 Total time 1 minute	Solvent: cdcl3 Temp. 25.0 C / 298.1 K Operator: Liu VNNRS-500



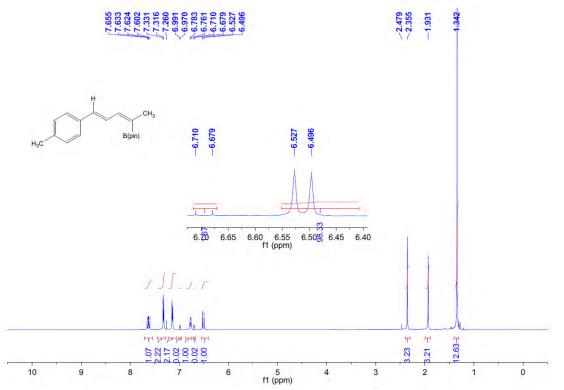
Compound cis-7g

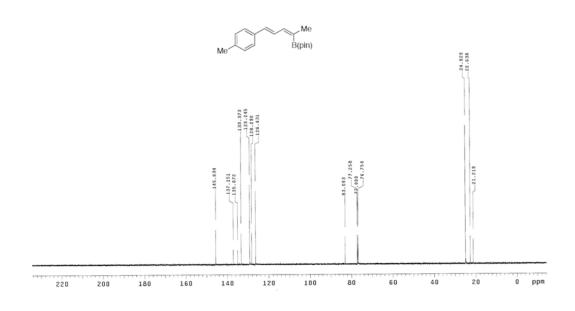




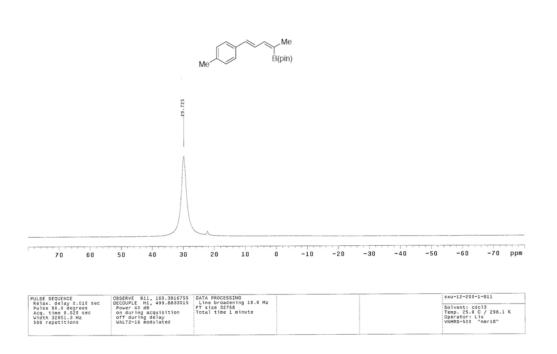


Compound 7h

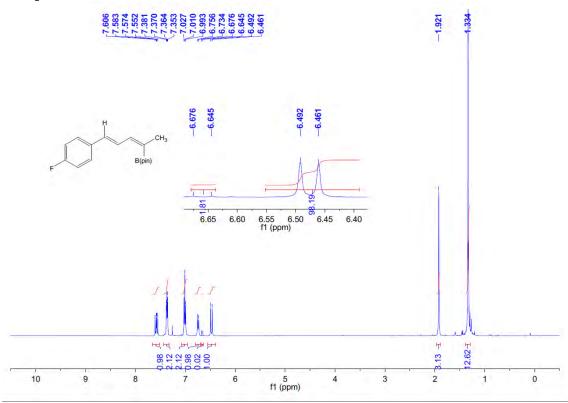


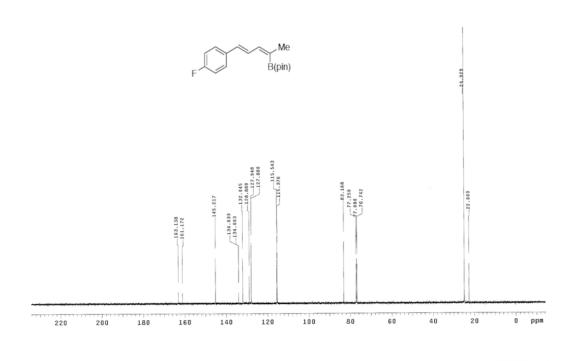


PULSE SEQUENCE	sxu=12-259=1-CL3 Solvent: cd13 Temp. 25.0 C / 258.1 K Operator: Liu VMMRS-500 "ner18"
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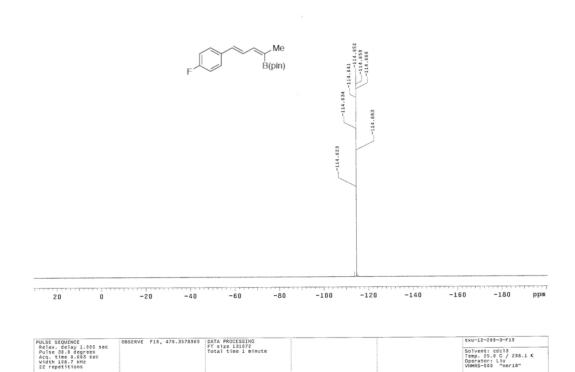


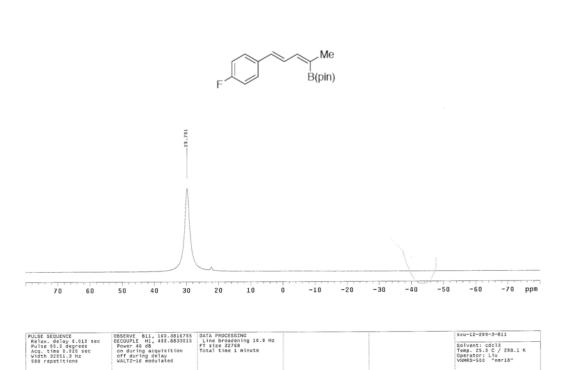
Compound 7i

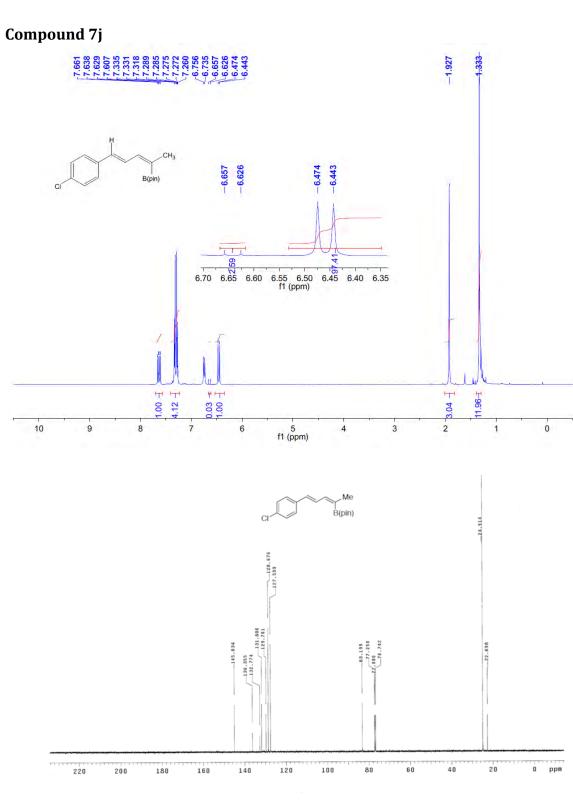




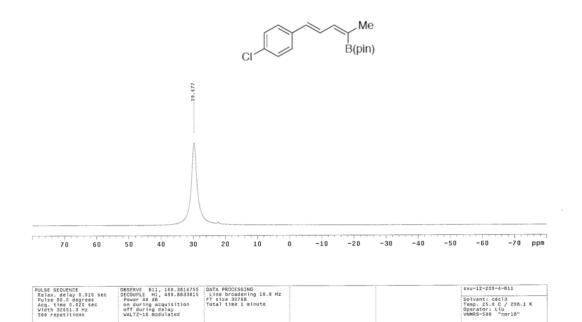
PULSE SEQUENCE Relax. delay 1.000 sec Pulse 45.0 degrees Acq. time 1.049 sec Width 31250.0 Hz 92 repetitions	OBSERVE C13, 125.6951290 DECOUPLE H1, 495.88833015 Power 40 d8 continuously on VALTZ-16 modulated	DATA PROCESSING Line broadening 0.5 Hz FT size 65386 Total time 3 minutes	Solvent: cdcl3 Temp. 25.0 C / 298.1 K Operator: Liu VNMRS-500 "nmr18"



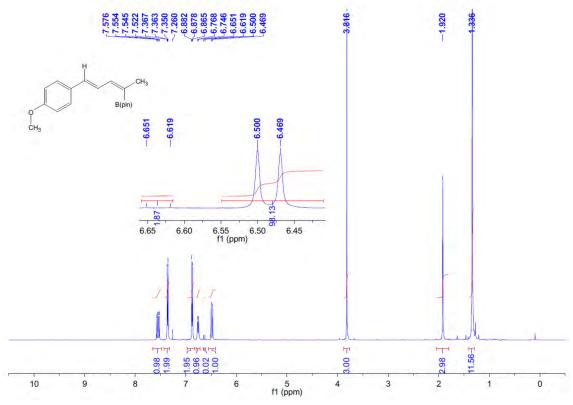


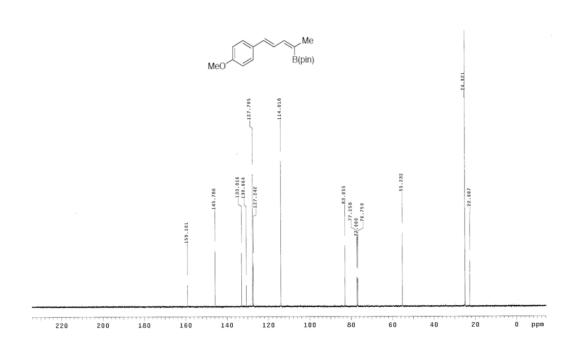


SE SEQUENCE		OBSERVE C13. 1	25.6951309	DATA PROCESSING		 	1	 sxu=12=209=	4-C13
lax. delay 1.00 lse 45.0 degree: q. time 1.049 s dth 31250.0 Hz	0 sec s ec	OBSERVE C13, 1 DECOUPLE H1, 4 Power 40 dB continuously of WALTZ-15 modul	n	Line broadenin FT size 65536 Total time 1 mi	g 0.5 Hz			Solvent: cd Temp. 25.0 Operator: L	C / 298.1
repetitions								VNMRS-500	"nmr18"

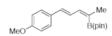


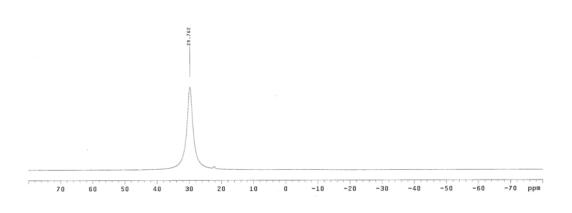
Compound 7k



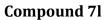


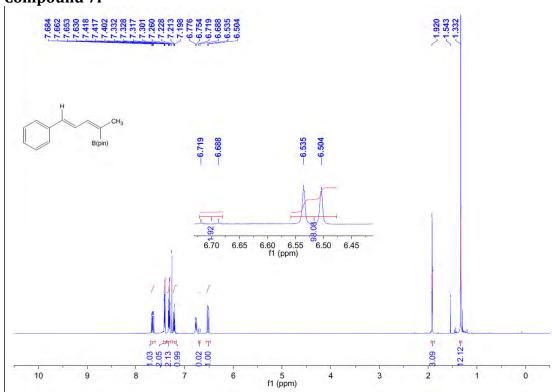
PULSE SEQUENCE	
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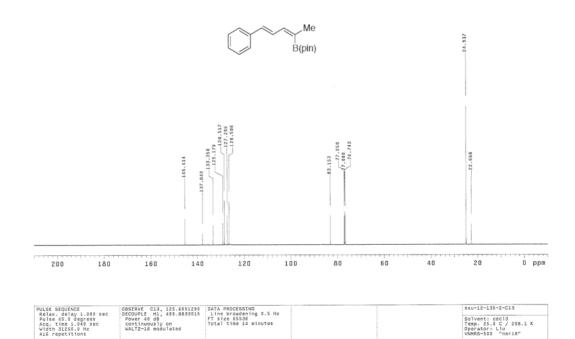




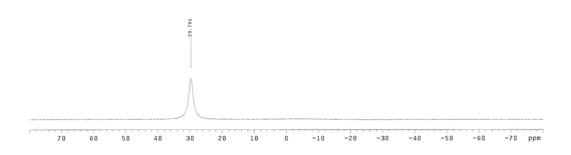
PULSE SEQUENCE Relax. delay 0. Pulse 90.0 degr Acq. time 0.020 Width 32051.3 H 500 repetitions	ees Power 40 dB sec on during acquisition z off during delay				sxu-12-209-2-811 Solvent: cdcl3 Temp. 25.0 C / 298.1 K Operator: Liu vNMRS-500 "nmr18"	
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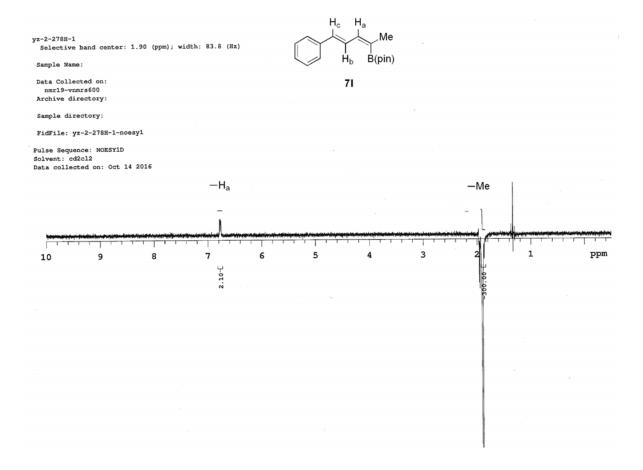




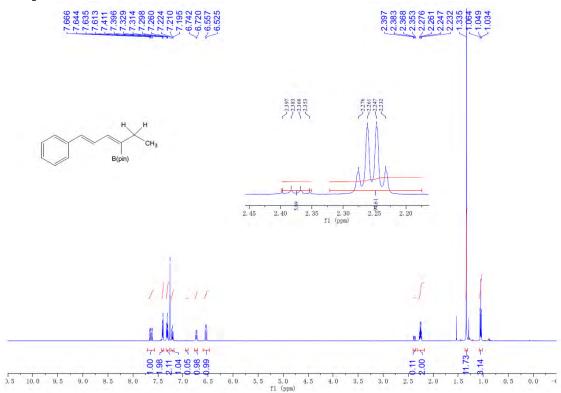


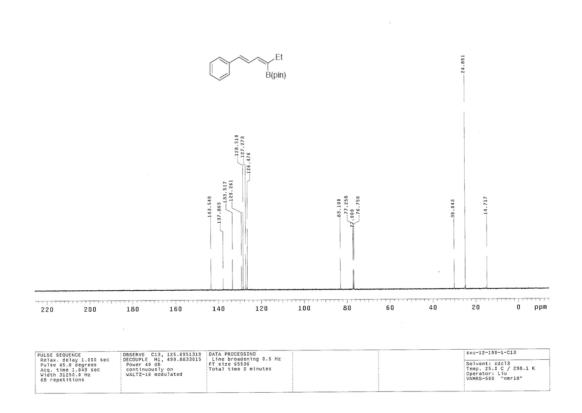


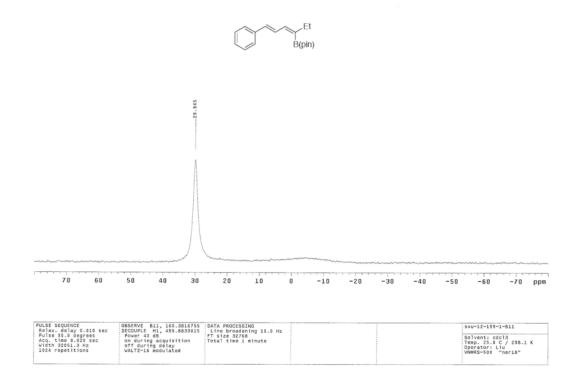
PULSE SEQUENCE Relax. delay 0.010 sec	OBSERVE	811, 160.3816794	DATA PROCESSING	sxu-12-135-2-811
Pulse 90.0 degrees Acq. time 0.020 sec Vidth 32051.3 Hz 1000 repetitions			FT size 32768 Total time 1 minute	Solvent: cdc13 Temp. 25.0 C / 298.1 K Operator: Liu VNMRS-500 "nmr18"



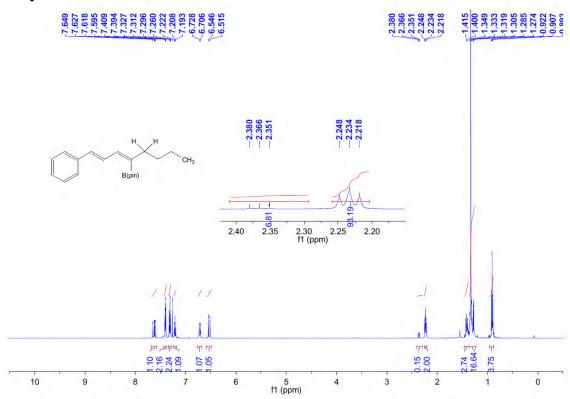
Compound 7m

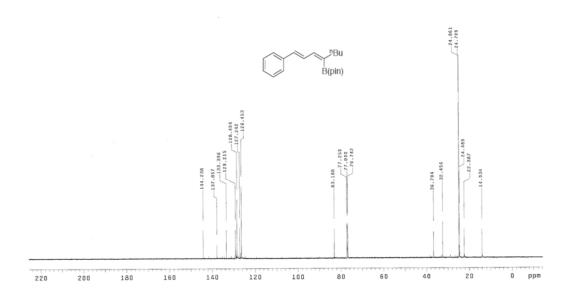






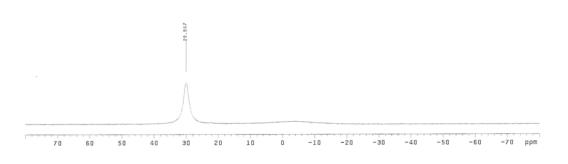
Compound 7n





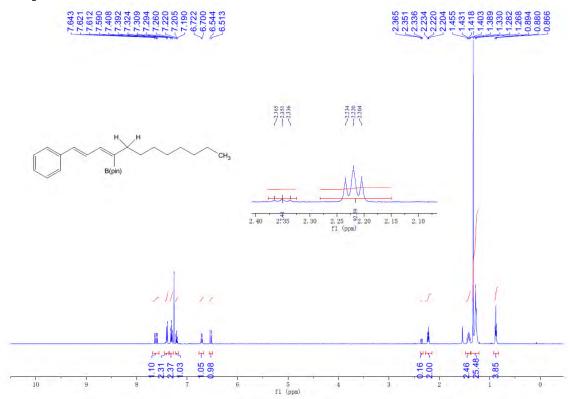


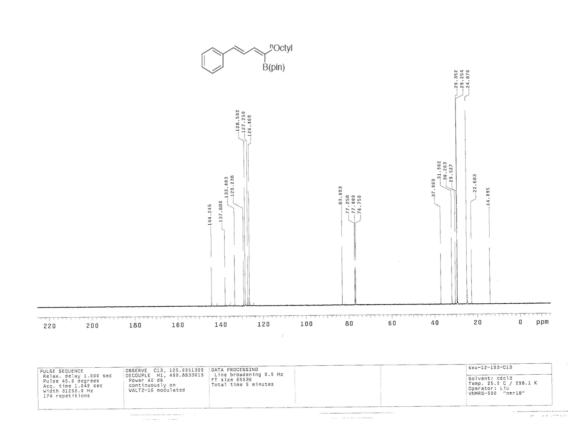




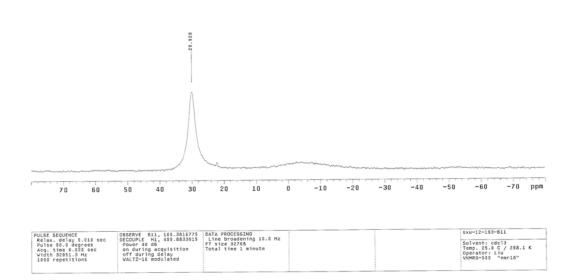
PULSE SEQUENCE Relax. delay 0.010 sec Pulse \$0.0 degrees Acq. time 0.020 sec Width 32051.3 Hz 1000 repetitions	OBSERVE	811, 160.3818848	DATA PROCESSING Line broadening 13.3 Hz FT size 32768 Total time 1 minute	Sxu-12-185-811 Solvent: cdcl3 Temp. 25.0 C / 298.1 K Operator: Liu VWM88-500 "nmr18"

Compound 7o

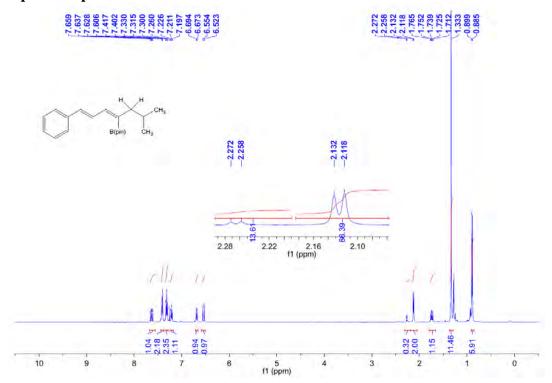


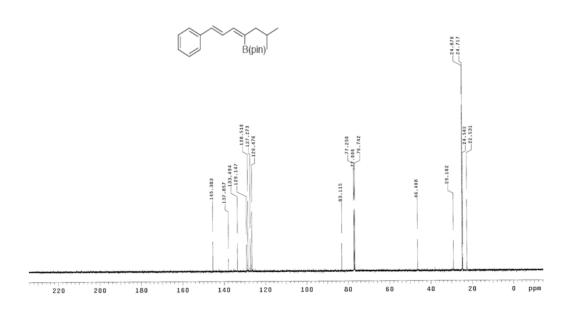




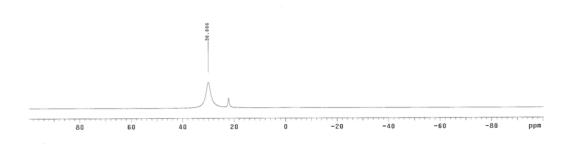


Compound 7p



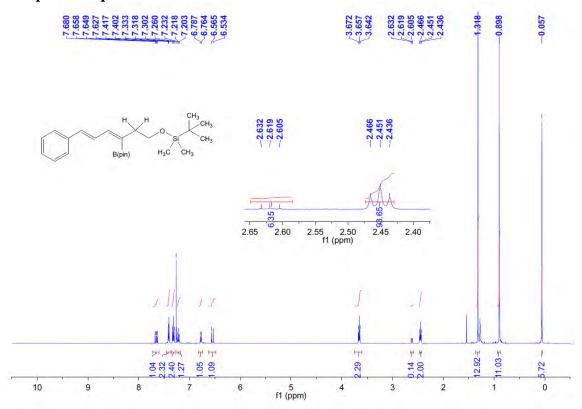


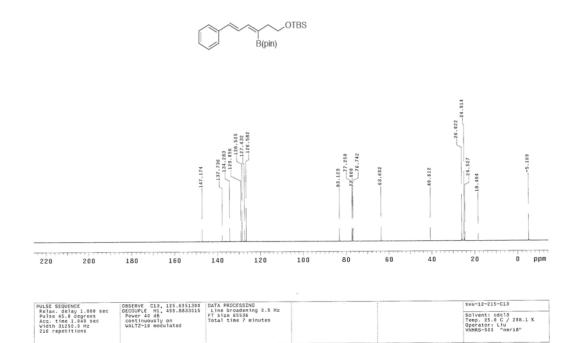
	OBSERVE C13, 125.6951290 DECOUPLE HI, 493.8833015 Power 40 dB continuously on WALTZ-16 modulated	DATA PROCESSING Line broadening 0.5 Hz FT size 65536 Total time 3 minutes		Solvent: Cdc13 Solvent: Cdc13 Teap. 25.0 C / 298.1 K Operator: Lfu VNMRS-500 "nmr18"
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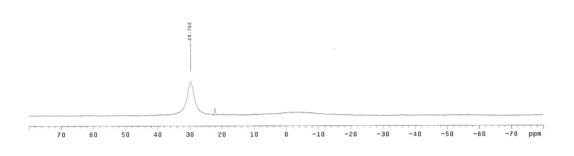
PULSE SEQUENCE Relax. delay 0.010 sec Pulse 90.0 degrees Acq. time 0.020 sec Width 32051.3 Hz 1000 repetitions	OBSERVE B11, 160.3816716 DECOUPLE H1, 499.8833015 Power 40 dB on during acquisition off during delay VALTZ-16 modulated	DATA PROCESSING Line broadening 10.0 Hz FT size 32765 Total time 1 minute			Solvent: cdcl3 Temp. 25.0 C / 298.1 K Operator: Liu VNMRS-500 "nmr18"	
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Compound 7q

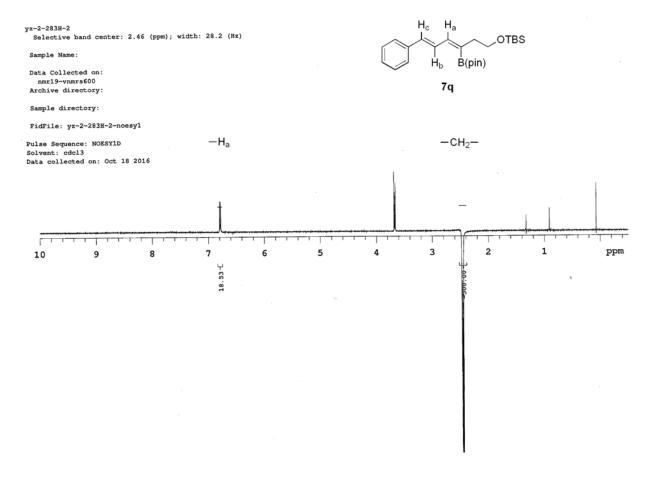




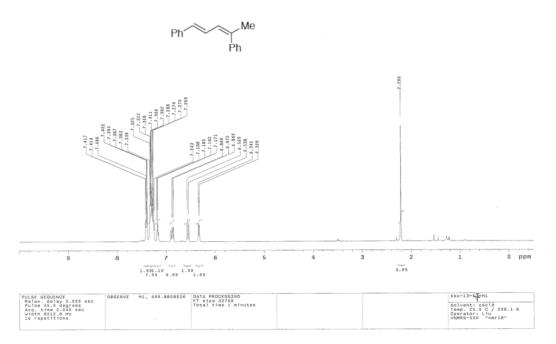




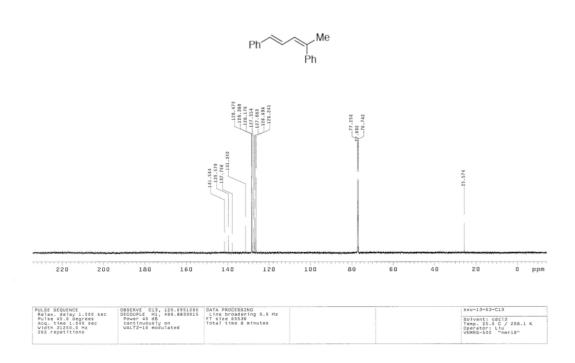
PULSE SEQUENCE Relax. delay 0.010 sec Pulse 90.0 degrees Acq. time 0.020 sec Width 32051.3 Hz 1000 repetitions	OBSERVE B11, 160.3818731 OECOUPLE H1, 499.8833015 Power 40 dB on during acquisition off during delay VALTZ-16 modulated	DATA PROCESSING Line broadening 10.0 Hz FT size 32768 Total time 1 minute		SXU-12-215-811 Solvent: cdcl3 Temp. 25. 8 C / 298.1 K UNMRS-500 "nmr18"
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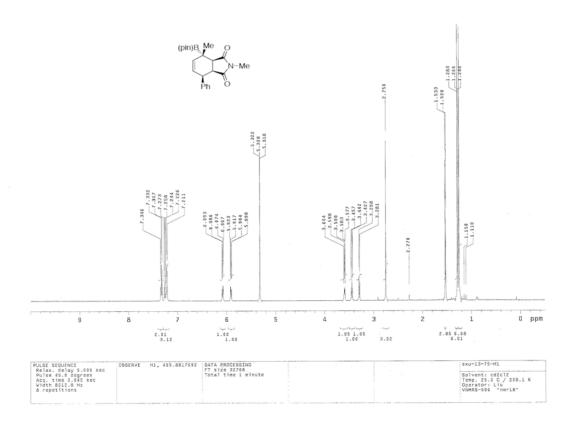


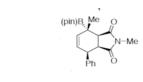
Compound 8

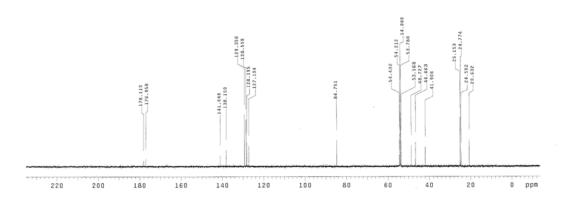


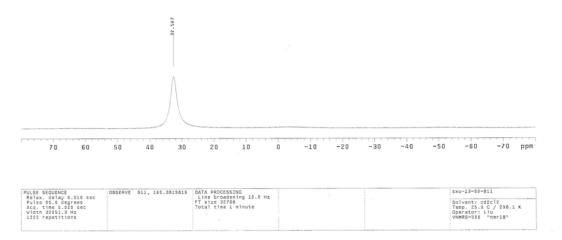
Compound 9



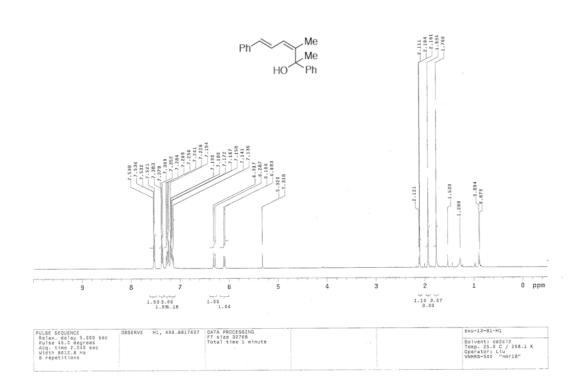


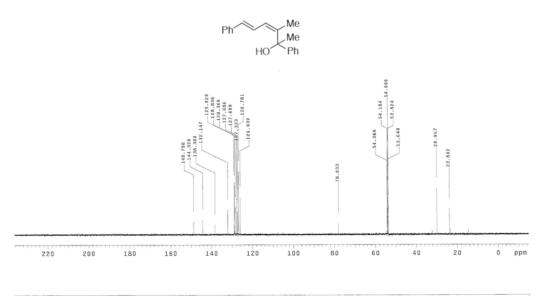






Compound 11





PULSE SEQUENCE Relax. dclay 1.000 sec Pulse 45.0 degrees Acq. time 0.855 sec Vidth 37878.8 Hz 118 repetitions	OBSERVE C13, 150.7915004 DECOUPLE H1, 593.6806968 Power 45 dB continuously on VALTZ-16 modulated		Sxu-13-81-C13 Solvent: cd2c12 Temp. 25.0 C / 298.1 K Operator: Liu VNMRS-600 "nmr13"	

Ligand CC-L3

