

**Perkins et al 2016**

**Title:** Smith-Magenis syndrome patients often display antibody deficiency but not other immune pathologies

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**Figure E1:** A detailed account of the reactivities of SMS sera versus control sera to 337 antigens

**Figure E2:** Persistent verrucous disease of the hand (A) and foot (B) of SMSY4

**Figure E3:** Bivariate analysis of serum reactivity to tetanus toxoid by conventional laboratory testing versus antigen microarray reveals fair concordance

Tables E1-E4

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**Table E2:** History of autoimmune diseases in 76 SMS subjects (ages 6 months-37 years)

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**Figure Legends**

**Figure E1:** A detailed account of the reactivities of SMS sera versus control sera to 337 antigens. A heat map displays reactivities of sera from 18 SMS subjects 14 healthy unrelated and 3 related controls to the 337 listed antigens. Colorimetric differences on the heat map correspond.

**Figure E2:** Persistent verrucous disease of the hand (A) and foot (B) of SMSY4

**Figure E3:** Bivariate analysis of serum reactivity to tetanus toxoid by conventional laboratory testing versus antigen microarray reveals fair concordance. Linearity ( $R^2$ ) and statistical significance ( $P$ ) calculated via linear regression analysis. MFI, mean fluorescence intensity.

**Table E1-** Location, size and relevant genes contained within the deleted regions of 25 SMS blood donors

	deleted element		deleted element encompasses:			
	location	size	<i>RAI1</i>	<i>TOM1L2</i>	<i>FLCN</i>	<i>TNFRSF13B</i>
SMS1	17p11.2 <sup>a</sup>	1.5 Mb	Y	Y	Y	Y
SMS2	17p11.2 <sup>b</sup>	3.6 Mb	Y	Y	Y	Y
SMSY2	17p11.2 <sup>b</sup>	3.7 Mb	Y	Y	Y	Y
SMS4	17p11.2 <sup>a</sup>	3.4 Mb	Y	Y	Y	Y
SMS5	17p11.2 <sup>b</sup>	6.3 Mb	Y	Y	Y	Y
SMSY6	17p11.2 <sup>a</sup>	3.4 Mb	Y	Y	Y	Y
SMS7	17p11.2 <sup>a</sup>	1.5Mb	Y	Y	Y	Y
SMS8	17p11.2 <sup>a</sup>	3.1 Mb	Y	Y	Y	Y
SMS9	17p11.2 <sup>b</sup>	1.2 Mb	Y	Y	Y	Y
SMSY3	17p11.2 <sup>b</sup>	3.7 Mb	Y	Y	Y	Y
SMS10	17p11.2 <sup>b</sup>	3.4 Mb	Y	Y	Y	Y
SMS11	17p11.2 <sup>a</sup>	6.4 Mb	Y	Y	Y	Y
SMS12	17p11.2 <sup>a</sup>	3.7 Mb	Y	Y	Y	Y
SMS13	17p11.2 <sup>b</sup>	3.6 Mb	Y	Y	Y	Y
SMS14	17p11.2 <sup>a</sup>	3.6 Mb	Y	Y	Y	Y
SMSY5	17p11.2 <sup>a</sup>	6.3 Mb	Y	Y	Y	Y
SMSY7	17p11.2 <sup>a</sup>	3.3 Mb	Y	Y	Y	Y
SMS17	17p11.2 <sup>a</sup>	3.6 Mb	Y	Y	Y	Y
SMS18	17p11.2 <sup>a</sup>	3.2 Mb	Y	Y	Y	Y
SMS19	17p11.2 <sup>a</sup>	3.4 Mb	Y	Y	Y	Y
SMS20	17p11.2 <sup>a</sup>	3.7 Mb	Y	Y	Y	Y
SMS21	17p11.2 <sup>a</sup>	3.4 Mb	Y	Y	Y	Y
SMS22	17p11.2 <sup>a</sup>	2.1 Mb	Y	Y	Y	Y
SMSY4	17p11.2 <sup>a</sup>	3.5 Mb	Y	Y	Y	Y
SMS23	17p11.2 <sup>a</sup>	3.5 Mb	Y	Y	Y	Y

Y, Yes, the indicated gene is deleted

<sup>a</sup> Determined by FISH<sup>b</sup> Determined by microarray

**Table E2-** History of autoimmune diseases in 76 SMS subjects (ages 6 months-37 years)

	Percentage (n)
Autoimmune thyroiditis	2.6 (2)
Autoimmune neutropenia	1.3 (1)
Pernicious anemia	1.3 (1)

**Table E3-** Atopic history in 76 SMS subjects  
(ages 6 months-37 years)

	Percentage (n)
Allergic Rhinitis	15.8 (12)
Atopic Dermatitis	9.2 (7)
Food Allergy	6.6 (5)
Asthma	7.9 (6)
Drug Allergy	2.6 (2)

**Table E4-** Relative frequencies of lymphocyte subsets in the peripheral blood of 19 SMS subjects

Reference	Age <sup>a</sup>	Sex	CD3 <sup>+</sup> (% PBLs)	CD3 <sup>+</sup> CD4 <sup>+</sup> (% PBLs)	CD3 <sup>+</sup> CD8 <sup>+</sup> (% PBLs)	CD3 <sup>+</sup> CD4 <sup>+</sup> CD25 <sup>hi</sup> FOXP3 <sup>+</sup> CD127 <sup>-</sup> (% CD4 <sup>+</sup> T cells)	CD19 <sup>+</sup> (% PBLs)	CD19 <sup>+</sup> CD27 <sup>+</sup> (% B cells)	CD19 <sup>+</sup> CD27 <sup>+</sup> IgM <sup>+</sup> (% B cells)	CD3 <sup>+</sup> CD16 <sup>+</sup> CD56 <sup>+</sup> (% PBLs)
			<sub>-b</sub>	<sub>-b</sub>	<sub>-b</sub>	<sub>-b</sub>	<sub>-b</sub>	<sub>-b</sub>	<sub>-b</sub>	<sub>-b</sub>
SMS2	6	M	33.5 (L)	14.4 (L)	18.9	4.6	41.8 (H)	13.6	1.2 (L)	3.4 (L)
SMS4	7	M	56.7	38.3	15.6	2.7 (L)	15.5	23.9	2.5 (L)	19.1
SMS7	10	M	68.4	28.7	18.0	5.5	19.5	11.8 (L)	1.7 (L)	4.7
SMS8	10	F	48.0	29.4	17.4	5.4	36.1 (H)	41.2 (H)	12.0	11.1
SMS9	11	M	63.4	31.1	25.4	5.9	18.6	16.3	3.9 (L)	8.1
SMS10	13	F	68.6	28.6	28.0	ND	30.5 (H)	9.5 (L)	2.8 (L)	6.2
SMS11	15	F	54.1	35.9	14.7	4.1	48.0 (H)	22.8	4.9 (L)	3.6
SMS12	16	F	61.0	37.5	19.1	4.9	26.7	20.1	4.1 (L)	4.7
SMS13	16	M	58.9	34.2	21.9	4.5	26.2	29.3	3.5 (L)	8.2
SMS14	20	F	63.4	38.8	23.0	5.7	25.3	27.6	7.6 (L)	5.2
SMSY5	20	F	56.8	22.2	32.1	6.5 <sup>c</sup>	20.1	11.8 (L)	1.7 (L) <sup>c</sup>	15.7
SMSY7	20	F	52.2	36.5	14.9	4.0 <sup>c</sup>	31.6 (H)	20.6	3.5 (L) <sup>c</sup>	9.4
SMS17	20	F	57.1	33.8	16.8	4.5	33.0 (H)	21.6	3.3 (L)	5.3
SMS18	21	F	67.9	45.1	19.1	5.4	7.8	5.1 (L)	2.6 (L)	9.7
SMS19	22	M	69.9	50.3	18.7	4	9.2	20.2	7.0 (L)	3.7
SMS20	22	F	65.9	37.4	25.9	4.1	21.5	22.6	4.9 (L)	8.6
SMS21	23	M	48.2	22.8	22.1	8.6	39.7 (H)	18.1	3.8 (L)	11.6
SMSY4	26	F	58.9	33.4	19.6	5.4 <sup>c</sup>	20.6	22.5	12.1 <sup>c</sup>	5.7
SMS23	27	M	51.1	30.1	19.4	4.8	32.7 (H)	24.3	6.5 (L)	12.8

F, female; H, higher than normal range; L, below normal range; M, male; ND, not done; PBL, peripheral blood lymphocyte

<sup>a</sup> Age (years) at time of testing

<sup>b</sup> Institutional reference ranges vary by age

<sup>c</sup> Value previously published<sup>3</sup>







