Apolipoprotein E, Receptors and Modulation of Alzheimer's Disease Supplemental Information

ApoE in tauopathy

Tau is a microtubule-associated protein encoded by *MAPT* gene, which has a role in stabilizing neuronal microtubules. Hyperphosphorylation and aggregation of tau are characteristics of several neurodegenerative diseases known as tauopathies, including AD (1). Findings from clinical PET imaging, CSF, and post-mortem studies suggest that the pathological aggregation of tau is closely linked to patterns of neurodegeneration and clinical manifestations of AD (2, 3). AD patients who carry the *APOE4* genotype have greater medial temporal lobe vulnerability and post-mortem tau pathology compared to non-carriers (4-6). Preclinical studies concerning how apoE isoforms affect tauopathy in AD are limited. A transgenic animal study revealed that neuronal, but not astrocytic, apoE4 overexpression increases tau phosphorylation, suggesting a neuron-specific effect of apoE4 on tauopathy (7). An earlier study showed that tau binds avidly to apoE3, but not to apoE4, indicating that the isoform-specific interactions of apoE with tau might contribute to differential effects on tau metabolism (8).

Tauopathy in the absence of A β may reflect a pathological process that is distinct from AD. Patients with primary age-related tauopathy (PART) develop cognitive impairment that can be indistinguishable from AD, but in contrast contain none or only minimal A β deposition (9, 10). Interestingly, there is no association between PART and *APOE* genotype (10). It was also reported that the *APOE4* is associated with tau tangles in the brains with A β , whereas no such association is found in brains without A β (11). Understanding how apoE isoforms affect tauopathy in the presence and absence of amyloid pathology *in vivo* will help us to develop appropriate therapeutic strategies for AD and other tauopathies.

ApoE and apoE receptors in brain lipid transport

The brain is the most cholesterol-rich organ, and about 30% of cholesterol in the brain is metabolically active and found in membranes of glial cells and neurons, where it undergoes recycling for neuronal repair and remodeling (12, 13). ApoE is critical in redistributing cholesterol and lipids to neurons through LRP1 and LDLR to maintain the synaptic function (13, 14). It was shown that the cholesterol levels in AD brains are lower than in healthy brains (15). The association between apoE, cholesterol and AD has been intensely reviewed (14, 16-20), and it is recognized that the isoform-specific effects of apoE in AD is at least partially due to their differential ability of transporting cholesterol to neurons, with apoE4 being less efficient than apoE3 (21-23).

ApoE in glucose metabolism and mitochondrial function

Glucose is the primary energy resources for the brain, and mitochondria are recognized as subcellular organelles that are essential for generating the energy for the cells (24). Cerebral glucose hypometabolism, as assessed by FDG-PET scan, exists in pre-symptomatic AD patients long before the clinical onset of disease and has become one of the early biomarkers of AD (25-28). Multiple studies have also suggested that mitochondrial dysfunction and oxidative damage might have early and preponderant roles in AD (29-31). *APOE4* has been associated with

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glucose hypometabolism and mitochondrial dysfunction in the brain (32-34). FDG-PET studies have clearly shown that *APOE4* carriers, either as healthy adults or with dementia, have lower cerebral glucose metabolism compared to non-carriers (34-41). Moreover, *APOE4* genotype, not $A\beta$ aggregation, contributes to reduced glucose metabolism in aging (42). *APOE4* carriers also have lower mitochondrial cytochrome oxidase activity than non-carriers (43). Studies revealed that apoE4 might affect the neuronal mitochondrial respiration due to its C-terminal-truncated fragmentation or domain interaction. In addition, it has been shown that neuronal LRP1 not only modulates lipid metabolism, but also plays a critical role in regulating insulin signaling and glucose metabolism in the brain (44). Whether LRP1 maintains both lipid and glucose homeostasis in an apoE isoform-dependent manner in the brain deserves further investigation. Together, more studies are required to understand the biological mechanisms that link the glucose hypometabolism and development of AD, and how apoE isoforms modulate these events (45-47).

ApoE and apoE receptors in vascular integrity and function

AD often co-exists with CAA, and is associated with microvascular dysfunction and degeneration in the brain (48). *APOE4* shows an association with CAA and CAA-related hemorrhages (49). Interestingly, *APOE2* is also a risk factor for CAA although it is protective against AD (50). ApoE4 interrupts cerebrovascular functions likely through both Aβ-independent and Aβ-dependent manner (51). ApoE4-TR mice showed an age-dependent progressive BBB breakdown by activating CypA-nuclear factor (NF) κ B-MMP-9 pathway in brain capillary pericytes (52). The accelerated pericyte loss contributing to BBB damage was also found in human *APOE4* carriers (53). ApoE4-TR mice showed compromised BBB transport

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function and cerebral vascular dysfunction and atrophy (54). Furthermore, it was shown that apoE produced by pericytes modulates A β removal and cytotoxicity near the vasculature in the brain, which might contribute to the development of CAA (55). Thus, it is possible that vascular defects in apoE4 precede neuronal dysfunction and could initiate neurodegenerative changes. However, a recent study reported no evidence of widespread BBB dysfunction in AD mouse models and apoE4-TR mice (56). Whether apoE4-mediated deficit impacts the global homeostatic capacity of the BBB which contributes to AD pathogenesis remains to be elucidated (57).

ApoE and apoE receptors in modulating neuroinflammation

Neuroinflammation is observed in both normal aging and AD (58-60). Resident immune cells, microglia and astrocytes, are activated by A β (61, 62), and joined by blood-borne monocytes that traverse the BBB and convert into activated macrophages, to release various cytokines, chemokines and proteolytic enzymes (63, 64). ApoE receptors, LRP1 and LDLR, mediate A β -induced astrocyte activation, initiating and modulating the inflammatory response induced by A β (65). By using a transgenic mouse overexpressing heparanase, an endoglucuronidase that specifically degrades HS side chains, it was found that all aspects of immune cell recruitment and activation are significantly attenuated in both lipopolysaccharide (LPS)-treated or microinjection of aggregated A β elicited inflammation models, which indicates that HSPGs are required to mediate neuroinflammatory responses (66). ApoE likely modulates the neuroinflammation in an isoform-dependent manner. It was reported that apoE4 amplifies the proinflammation (toll-like receptor 4-p38 α) pathways induced by A β , and suppresses the anti-inflammatory (IL-4R-nuclear receptor) pathway, resulting in an adverse neuroinflammatory

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phenotype that causes neuronal dysfunction (67, 68). Additionally, a specific apoE4 fragment (residues 186-299) detected in human AD brains can increase the level of inflammatory molecules in SHY5Y neuronal cell line (69). Following intracerebroventricular injection of LPS, it was shown that microglial and NF- κ B activation are more pronounced in apoE4-TR than in apoE3-TR mice, indicating that NF- κ B signaling pathway might mediate brain inflammation in apoE4-TR mice (70).

The missense mutation (R47H) of Triggering Receptor Expressed on Myeloid Cells 2 (TREM2), a protein expressed specifically in microglia in the brain, is associated with AD risk by dysregulating neuroinflammation and increasing AD pathology (71-76). Interestingly, apoE is an endogenous ligand of TREM2 (77). The uptake of apoE-A β complexes is reduced in macrophages from human subjects carrying the *TREM2* variant (78). It will be informative to further investigate whether apoE affects TREM2-mediated neuroinflammatory responses in an isoform-specific manner in the pathogenesis of AD.

Supplemental References

- 1. Wang Y, Mandelkow E (2016): Tau in physiology and pathology. *Nat Rev Neurosci*. 17:5-21.
- 2. Ossenkoppele R, Schonhaut DR, Scholl M, Lockhart SN, Ayakta N, Baker SL, et al. (2016): Tau PET patterns mirror clinical and neuroanatomical variability in Alzheimer's disease. *Brain.* 139:1551-1567.
- 3. Brier MR, Gordon B, Friedrichsen K, McCarthy J, Stern A, Christensen J, et al. (2016): Tau and Abeta imaging, CSF measures, and cognition in Alzheimer's disease. *Sci Transl Med*. 8:338ra366.
- 4. Lehmann M, Ghosh PM, Madison C, Karydas A, Coppola G, O'Neil JP, et al. (2014): Greater medial temporal hypometabolism and lower cortical amyloid burden in ApoE4-positive AD patients. *J Neurol Neurosurg Psychiatry*. 85:266-273.

- 5. van der Flier WM, Pijnenburg YA, Fox NC, Scheltens P (2011): Early-onset versus lateonset Alzheimer's disease: the case of the missing APOE varepsilon4 allele. *Lancet Neurol*. 10:280-288.
- 6. Tiraboschi P, Hansen LA, Masliah E, Alford M, Thal LJ, Corey-Bloom J (2004): Impact of APOE genotype on neuropathologic and neurochemical markers of Alzheimer disease. *Neurology*. 62:1977-1983.
- 7. Brecht WJ, Harris FM, Chang S, Tesseur I, Yu GQ, Xu Q, et al. (2004): Neuron-specific apolipoprotein e4 proteolysis is associated with increased tau phosphorylation in brains of transgenic mice. *J Neurosci*. 24:2527-2534.
- 8. Strittmatter WJ, Saunders AM, Goedert M, Weisgraber KH, Dong LM, Jakes R, et al. (1994): Isoform-specific interactions of apolipoprotein E with microtubule-associated protein tau: implications for Alzheimer disease. *Proc Natl Acad Sci U S A*. 91:11183-11186.
- 9. Duyckaerts C, Braak H, Brion JP, Buee L, Del Tredici K, Goedert M, et al. (2015): PART is part of Alzheimer disease. *Acta Neuropathol*. 129:749-756.
- 10. Crary JF, Trojanowski JQ, Schneider JA, Abisambra JF, Abner EL, Alafuzoff I, et al. (2014): Primary age-related tauopathy (PART): a common pathology associated with human aging. *Acta Neuropathol.* 128:755-766.
- 11. Farfel JM, Yu L, De Jager PL, Schneider JA, Bennett DA (2016): Association of APOE with tau-tangle pathology with and without beta-amyloid. *Neurobiol Aging*. 37:19-25.
- 12. Dietschy JM, Turley SD (2001): Cholesterol metabolism in the brain. *Curr Opin Lipidol*. 12:105-112.
- 13. Dietschy JM, Turley SD (2004): Thematic review series: brain Lipids. Cholesterol metabolism in the central nervous system during early development and in the mature animal. *J Lipid Res.* 45:1375-1397.
- 14. Holtzman DM, Herz J, Bu G (2012): Apolipoprotein E and apolipoprotein E receptors: normal biology and roles in Alzheimer disease. *Cold Spring Harb Perspect Med.* 2:a006312.
- 15. Shobab LA, Hsiung GY, Feldman HH (2005): Cholesterol in Alzheimer's disease. *Lancet Neurol*. 4:841-852.
- 16. de Chaves EP, Narayanaswami V (2008): Apolipoprotein E and cholesterol in aging and disease in the brain. *Future Lipidol*. 3:505-530.
- 17. Bu G (2009): Apolipoprotein E and its receptors in Alzheimer's disease: pathways, pathogenesis and therapy. *Nat Rev Neurosci*. 10:333-344.
- 18. Liu CC, Kanekiyo T, Xu H, Bu G (2013): Apolipoprotein E and Alzheimer disease: risk, mechanisms and therapy. *Nat Rev Neurol*. 9:106-118.
- 19. Kanekiyo T, Xu H, Bu G (2014): ApoE and Abeta in Alzheimer's disease: accidental encounters or partners? *Neuron*. 81:740-754.
- 20. Mahley RW (2016): Central Nervous System Lipoproteins: ApoE and Regulation of Cholesterol Metabolism. *Arterioscler Thromb Vasc Biol.* 36:1305-1315.

- 21. Gong JS, Kobayashi M, Hayashi H, Zou K, Sawamura N, Fujita SC, et al. (2002): Apolipoprotein E (ApoE) isoform-dependent lipid release from astrocytes prepared from human ApoE3 and ApoE4 knock-in mice. *J Biol Chem*. 277:29919-29926.
- 22. Xu Q, Brecht WJ, Weisgraber KH, Mahley RW, Huang Y (2004): Apolipoprotein E4 domain interaction occurs in living neuronal cells as determined by fluorescence resonance energy transfer. *J Biol Chem*. 279:25511-25516.
- 23. Michikawa M, Fan QW, Isobe I, Yanagisawa K (2000): Apolipoprotein E exhibits isoformspecific promotion of lipid efflux from astrocytes and neurons in culture. *J Neurochem*. 74:1008-1016.
- 24. McCall AL (2004): Cerebral glucose metabolism in diabetes mellitus. *Eur J Pharmacol.* 490:147-158.
- 25. Willette AA, Bendlin BB, Starks EJ, Birdsill AC, Johnson SC, Christian BT, et al. (2015): Association of Insulin Resistance With Cerebral Glucose Uptake in Late Middle-Aged Adults at Risk for Alzheimer Disease. *JAMA Neurol.* 72:1013-1020.
- 26. Bertens D, Knol DL, Scheltens P, Visser PJ (2015): Temporal evolution of biomarkers and cognitive markers in the asymptomatic, MCI, and dementia stage of Alzheimer's disease. *Alzheimers Dement*. 11:511-522.
- 27. Jack CR, Jr., Holtzman DM (2013): Biomarker modeling of Alzheimer's disease. *Neuron*. 80:1347-1358.
- 28. Petersen RC, Wiste HJ, Weigand SD, Rocca WA, Roberts RO, Mielke MM, et al. (2015): Association of Elevated Amyloid Levels With Cognition and Biomarkers in Cognitively Normal People From the Community. *JAMA Neurol*.1-8.
- 29. Moreira PI, Carvalho C, Zhu X, Smith MA, Perry G (2010): Mitochondrial dysfunction is a trigger of Alzheimer's disease pathophysiology. *Biochim Biophys Acta*. 1802:2-10.
- 30. Moreira PI, Cardoso SM, Santos MS, Oliveira CR (2006): The key role of mitochondria in Alzheimer's disease. *J Alzheimers Dis*. 9:101-110.
- 31. Su B, Wang X, Nunomura A, Moreira PI, Lee HG, Perry G, et al. (2008): Oxidative stress signaling in Alzheimer's disease. *Curr Alzheimer Res.* 5:525-532.
- 32. Reiman EM, Caselli RJ, Chen K, Alexander GE, Bandy D, Frost J (2001): Declining brain activity in cognitively normal apolipoprotein E epsilon 4 heterozygotes: A foundation for using positron emission tomography to efficiently test treatments to prevent Alzheimer's disease. *Proc Natl Acad Sci U S A*. 98:3334-3339.
- 33. Reiman EM, Chen K, Alexander GE, Caselli RJ, Bandy D, Osborne D, et al. (2004): Functional brain abnormalities in young adults at genetic risk for late-onset Alzheimer's dementia. *Proc Natl Acad Sci U S A*. 101:284-289.
- 34. Reiman EM, Chen K, Alexander GE, Caselli RJ, Bandy D, Osborne D, et al. (2005): Correlations between apolipoprotein E epsilon4 gene dose and brain-imaging measurements of regional hypometabolism. *Proc Natl Acad Sci U S A*. 102:8299-8302.

- 35. Reiman EM, Caselli RJ, Yun LS, Chen K, Bandy D, Minoshima S, et al. (1996): Preclinical evidence of Alzheimer's disease in persons homozygous for the epsilon 4 allele for apolipoprotein E. *N Engl J Med.* 334:752-758.
- 36. Small GW, Ercoli LM, Silverman DH, Huang SC, Komo S, Bookheimer SY, et al. (2000): Cerebral metabolic and cognitive decline in persons at genetic risk for Alzheimer's disease. *Proc Natl Acad Sci U S A*. 97:6037-6042.
- 37. Small GW, Mazziotta JC, Collins MT, Baxter LR, Phelps ME, Mandelkern MA, et al. (1995): Apolipoprotein E type 4 allele and cerebral glucose metabolism in relatives at risk for familial Alzheimer disease. *JAMA*. 273:942-947.
- 38. Mosconi L, Perani D, Sorbi S, Herholz K, Nacmias B, Holthoff V, et al. (2004): MCI conversion to dementia and the APOE genotype: a prediction study with FDG-PET. *Neurology*. 63:2332-2340.
- 39. Mosconi L, Sorbi S, Nacmias B, De Cristofaro MT, Fayyaz M, Bracco L, et al. (2004): Age and ApoE genotype interaction in Alzheimer's disease: an FDG-PET study. *Psychiatry Res.* 130:141-151.
- 40. Drzezga A, Riemenschneider M, Strassner B, Grimmer T, Peller M, Knoll A, et al. (2005): Cerebral glucose metabolism in patients with AD and different APOE genotypes. *Neurology*. 64:102-107.
- 41. Mosconi L, Nacmias B, Sorbi S, De Cristofaro MT, Fayazz M, Tedde A, et al. (2004): Brain metabolic decreases related to the dose of the ApoE e4 allele in Alzheimer's disease. *J Neurol Neurosurg Psychiatry*. 75:370-376.
- 42. Jagust WJ, Landau SM (2012): Apolipoprotein E, not fibrillar beta-amyloid, reduces cerebral glucose metabolism in normal aging. *J Neurosci*. 32:18227-18233.
- 43. Valla J, Yaari R, Wolf AB, Kusne Y, Beach TG, Roher AE, et al. (2010): Reduced posterior cingulate mitochondrial activity in expired young adult carriers of the APOE epsilon4 allele, the major late-onset Alzheimer's susceptibility gene. *J Alzheimers Dis*. 22:307-313.
- 44. Liu CC, Hu J, Tsai CW, Yue M, Melrose HL, Kanekiyo T, et al. (2015): Neuronal LRP1 regulates glucose metabolism and insulin signaling in the brain. *J Neurosci*. 35:5851-5859.
- 45. Chang S, ran Ma T, Miranda RD, Balestra ME, Mahley RW, Huang Y (2005): Lipid- and receptor-binding regions of apolipoprotein E4 fragments act in concert to cause mitochondrial dysfunction and neurotoxicity. *Proc Natl Acad Sci U S A*. 102:18694-18699.
- 46. Nakamura T, Watanabe A, Fujino T, Hosono T, Michikawa M (2009): Apolipoprotein E4 (1-272) fragment is associated with mitochondrial proteins and affects mitochondrial function in neuronal cells. *Mol Neurodegener*. 4:35.
- 47. Chen HK, Ji ZS, Dodson SE, Miranda RD, Rosenblum CI, Reynolds IJ, et al. (2011): Apolipoprotein E4 domain interaction mediates detrimental effects on mitochondria and is a potential therapeutic target for Alzheimer disease. *J Biol Chem.* 286:5215-5221.
- 48. Zlokovic BV (2011): Neurovascular pathways to neurodegeneration in Alzheimer's disease and other disorders. *Nat Rev Neurosci*. 12:723-738.

- 49. McCarron MO, Nicoll JA (2000): Apolipoprotein E genotype and cerebral amyloid angiopathy-related hemorrhage. *Ann N Y Acad Sci.* 903:176-179.
- 50. Nelson PT, Pious NM, Jicha GA, Wilcock DM, Fardo DW, Estus S, et al. (2013): APOEepsilon2 and APOE-epsilon4 correlate with increased amyloid accumulation in cerebral vasculature. *J Neuropathol Exp Neurol*. 72:708-715.
- 51. Zlokovic BV (2013): Cerebrovascular effects of apolipoprotein E: implications for Alzheimer disease. *JAMA Neurol*. 70:440-444.
- 52. Bell RD, Winkler EA, Singh I, Sagare AP, Deane R, Wu Z, et al. (2012): Apolipoprotein E controls cerebrovascular integrity via cyclophilin A. *Nature*. 485:512-516.
- 53. Halliday MR, Rege SV, Ma Q, Zhao Z, Miller CA, Winkler EA, et al. (2016): Accelerated pericyte degeneration and blood-brain barrier breakdown in apolipoprotein E4 carriers with Alzheimer's disease. *J Cereb Blood Flow Metab.* 36:216-227.
- 54. Alata W, Ye Y, St-Amour I, Vandal M, Calon F (2015): Human apolipoprotein E varepsilon4 expression impairs cerebral vascularization and blood-brain barrier function in mice. *J Cereb Blood Flow Metab.* 35:86-94.
- 55. Bruinsma IB, Wilhelmus MM, Kox M, Veerhuis R, de Waal RM, Verbeek MM (2010): Apolipoprotein E protects cultured pericytes and astrocytes from D-Abeta(1-40)-mediated cell death. *Brain Res.* 1315:169-180.
- 56. Bien-Ly N, Boswell CA, Jeet S, Beach TG, Hoyte K, Luk W, et al. (2015): Lack of Widespread BBB Disruption in Alzheimer's Disease Models: Focus on Therapeutic Antibodies. *Neuron*. 88:289-297.
- 57. Ulrich JD, Huynh TP, Holtzman DM (2015): Re-evaluation of the Blood-Brain Barrier in the Presence of Alzheimer's Disease Pathology. *Neuron*. 88:237-239.
- 58. Akiyama H, Barger S, Barnum S, Bradt B, Bauer J, Cole GM, et al. (2000): Inflammation and Alzheimer's disease. *Neurobiol Aging*. 21:383-421.
- 59. Alcolea D, Martinez-Lage P, Sanchez-Juan P, Olazaran J, Antunez C, Izagirre A, et al. (2015): Amyloid precursor protein metabolism and inflammation markers in preclinical Alzheimer disease. *Neurology*. 85:626-633.
- 60. Malik M, Parikh I, Vasquez JB, Smith C, Tai L, Bu G, et al. (2015): Genetics ignite focus on microglial inflammation in Alzheimer's disease. *Mol Neurodegener*. 10:52.
- 61. Zine A (2003): Molecular mechanisms that regulate auditory hair-cell differentiation in the mammalian cochlea. *Mol Neurobiol*. 27:223-238.
- 62. Town T, Nikolic V, Tan J (2005): The microglial "activation" continuum: from innate to adaptive responses. *J Neuroinflammation*. 2:24.
- 63. Rezai-Zadeh K, Gate D, Town T (2009): CNS infiltration of peripheral immune cells: D-Day for neurodegenerative disease? *J Neuroimmune Pharmacol*. 4:462-475.
- 64. Gate D, Rezai-Zadeh K, Jodry D, Rentsendorj A, Town T (2010): Macrophages in Alzheimer's disease: the blood-borne identity. *J Neural Transm (Vienna)*. 117:961-970.

- 65. LaDu MJ, Shah JA, Reardon CA, Getz GS, Bu G, Hu J, et al. (2000): Apolipoprotein E receptors mediate the effects of beta-amyloid on astrocyte cultures. *J Biol Chem*. 275:33974-33980.
- 66. Zhang X, Wang B, O'Callaghan P, Hjertstrom E, Jia J, Gong F, et al. (2012): Heparanase overexpression impairs inflammatory response and macrophage-mediated clearance of amyloid-beta in murine brain. *Acta Neuropathol*. 124:465-478.
- 67. Tai LM, Ghura S, Koster KP, Liakaite V, Maienschein-Cline M, Kanabar P, et al. (2015): APOE-modulated Abeta-induced neuroinflammation in Alzheimer's disease: current landscape, novel data, and future perspective. *J Neurochem*. 133:465-488.
- 68. Rodriguez GA, Tai LM, LaDu MJ, Rebeck GW (2014): Human APOE4 increases microglia reactivity at Abeta plaques in a mouse model of Abeta deposition. *J Neuroinflammation*. 11:111.
- 69. Dafnis I, Tzinia AK, Tsilibary EC, Zannis VI, Chroni A (2012): An apolipoprotein E4 fragment affects matrix metalloproteinase 9, tissue inhibitor of metalloproteinase 1 and cytokine levels in brain cell lines. *Neuroscience*. 210:21-32.
- 70. Ophir G, Amariglio N, Jacob-Hirsch J, Elkon R, Rechavi G, Michaelson DM (2005): Apolipoprotein E4 enhances brain inflammation by modulation of the NF-kappaB signaling cascade. *Neurobiol Dis.* 20:709-718.
- Jonsson T, Stefansson H, Steinberg S, Jonsdottir I, Jonsson PV, Snaedal J, et al. (2013): Variant of TREM2 associated with the risk of Alzheimer's disease. *N Engl J Med.* 368:107-116.
- 72. Guerreiro R, Wojtas A, Bras J, Carrasquillo M, Rogaeva E, Majounie E, et al. (2013): TREM2 variants in Alzheimer's disease. *N Engl J Med.* 368:117-127.
- 73. Roussos P, Katsel P, Fam P, Tan W, Purohit DP, Haroutunian V (2015): The triggering receptor expressed on myeloid cells 2 (TREM2) is associated with enhanced inflammation, neuropathological lesions and increased risk for Alzheimer's dementia. *Alzheimers Dement*. 11:1163-1170.
- 74. Painter MM, Atagi Y, Liu CC, Rademakers R, Xu H, Fryer JD, et al. (2015): TREM2 in CNS homeostasis and neurodegenerative disease. *Mol Neurodegener*. 10:43.
- 75. Heslegrave A, Heywood W, Paterson R, Magdalinou N, Svensson J, Johansson P, et al. (2016): Increased cerebrospinal fluid soluble TREM2 concentration in Alzheimer's disease. *Mol Neurodegener*. 11:3.
- 76. Jin SC, Carrasquillo MM, Benitez BA, Skorupa T, Carrell D, Patel D, et al. (2015): TREM2 is associated with increased risk for Alzheimer's disease in African Americans. *Mol Neurodegener*. 10:19.
- 77. Atagi Y, Liu CC, Painter MM, Chen XF, Verbeeck C, Zheng H, et al. (2015): Apolipoprotein E Is a Ligand for Triggering Receptor Expressed on Myeloid Cells 2 (TREM2). *J Biol Chem.* 290:26043-26050.
- 78. Yeh FL, Wang Y, Tom I, Gonzalez LC, Sheng M (2016): TREM2 Binds to Apolipoproteins, Including APOE and CLU/APOJ, and Thereby Facilitates Uptake of Amyloid-Beta by Microglia. *Neuron.* 91:328-340.