

Supplemental Table 1. Plasmids and oligonucleotides used.

Plasmids:

Name	Purpose/modification	Source
pBAD2- <i>bgaB-htrAp3</i>	pBAD2- <i>bgaB</i> driven by <i>htrAp3</i> ; Ap ^r	Klinkert, et al. 2012
pBAD2- <i>bgaB-htrAp3-ΔC</i>	pBAD2- <i>bgaB</i> driven by <i>htrAp3-ΔC</i> ; Ap ^r	Klinkert, et al. 2012
pBAD2- <i>bgaB-htrAp3-U25C</i>	pBAD2- <i>bgaB</i> driven by <i>htrAp3-U25C</i> ; Ap ^r	This work
pUC19	Backbone for full-length <i>htrA</i> SHAPE template	New England Biolabs
<i>htrAp1-cass1</i> -pUC19	Production of full-length <i>htrA</i> SHAPE template	This work

Primers and DNA oligonucleotides:

Name	Purpose	Sequence
222	5' end of template for <i>htrA</i> RNA in SHAPE cassette, top strand	5'-TTCTAATACGACTCACTATAGGCCTTCGGGCCAAAC AGCAATTTTGC GTTACCTGTTAATCGAGATTGAAAC
223	3' end of template for <i>htrA</i> RNA in SHAPE cassette, bottom strand	5'-GAACCGGACCGAAGCCCGATTGGCTTCGGCGAACC GAAGCTTTTTTCATGTGTTTCAATCTCGATTAACAGG
224	5' end of template for ΔC <i>htrA</i> RNA in SHAPE cassette, top strand	5'-TTCTAATACGACTCACTATAGGCCTTCGGGCCAAAC AGCAATTTTGGTTACCTGTTAATCGAGATTGAAAC
225	Reverse primer for SHAPE template	5'-GAACCGGACCGAAGCCCG
225-6FAM	SHAPE reaction primer, 5' 6-FAM label	5'-6-FAM-GAACCGGACCGAAGCCCG
225-VIC	SHAPE sequencing primer, 5' VIC label	5'-VIC-GAACCGGACCGAAGCCCG
226	Forward primer for SHAPE template	5'-TTCTAATACGACTCACTATAGGCCTTCGGG
272	3' end of template for <i>htrA</i> RNA in alternate SHAPE cassette, bottom strand	5'-GAACCGGACCGAAGCCCGATTGGATCCGGCGAACC GGATCGATTTTCATGTGTTTCAATCTCGATTAACAGG
319	Forward primer for creation of U25C mutation in <i>bgaB</i> plasmid	5'- ACAGCAATTTTCGCGTTACCTG
320	Reverse primer for creation of U25C mutation in <i>bgaB</i> plasmid	5'- GCTAGCGAGAAACAGTAG
323	5' end of template for U25C <i>htrA</i> RNA in SHAPE cassette, top strand	5'- TTCTAATACGACTCACTATAGGCCTTCGGGCCAAAC AGCAATTTTCGCGTTACCTGTTAATCGAGATTGAAAC
324	5' end of template for AAAAA <i>htrA</i> RNA in SHAPE cassette, top strand	5'- TTCTAATACGACTCACTATAGGCCTTCGGGCCAAAC AGCAAAAAAACGTTACCTGTTAATCGAGATTGAAAC
341	5' end of template for U23C <i>htrA</i> RNA in SHAPE cassette, top strand	5'- TTCTAATACGACTCACTATAGGCCTTCGGGCCAAAC AGCAATCTTGC GTTACCTGTTAATCGAGATTGAAAC
342	5' end of template for G28A <i>htrA</i> RNA in SHAPE cassette, top strand	5'- TTCTAATACGACTCACTATAGGCCTTCGGGCCAAAC AGCAATTTTGCATTACCTGTTAATCGAGATTGAAAC
348	5' end of template for U40C <i>htrA</i> RNA in SHAPE cassette, top strand	5'- TTCTAATACGACTCACTATAGGCCTTCGGGCCAAAC AGCAATTTTGC GTTACCTGTTAACCAGATTGAAAC
349	3' end of template for U40C <i>htrA</i> RNA in SHAPE cassette, bottom strand	5'-GAACCGGACCGAAGCCCGATTGGCTTCGGCGAACC GAAGCTTTTTTCATGTGTTTCAATCTCGGTTAACAGG
275	gBlock dsDNA used for <i>htrAp1</i> RNA in SHAPE cassette	5'-GATGCCGGGAATTCTAATACGACTCACTATAGGCCTTCGGGCCAAGTCGGTTGATTTCAGGA TTATATCAGCGGGATGACTGACCTTTACGCATGGGATGAATATCGGCGTTTGATGGCGGTTCGA ACAGTAAATGGACTTTTGTAAGATGGACAATAAATTTTACTTTTTCCAGAACTTTGTTCCG GAACTTCGCGTTATAAAATGAATCTGACGTACACAGCAATTTTGC GTTACCTGTTAATCGAGA TTGAAACACATGAAAAAGCTTCGGTTCGCCGAAGCCAAATCGGGCTTCGGTCCGGTTCGCGAG TTAACGGATCTCTAGACGCG

A

S_Enteritidis str. P125109	ACAGCAATTTTTCGCGTTACCTGTTAATCGAGATTGAAACACATGAAA
S_Typhimurium str. LT2	ACAGCAATTTTTCGCGTTACCTGTTAATCGAGATTGAAACACATGAAA
S_Typhi str. CT18	ACAGCAATTTTTCGCGTTACCTGTTAATCGAGATTGAAACACATGAAA
S_Paratyphi_A str. ATCC 9150	ACAGCAATTTTTCGCGTTACCTGTTAATCGAGATTGAAACACATGAAA
S_Paratyphi_B str. SPB7	ACAGCAATTTTTCGCGTTACCTGTTAATCGAGATTGAAACACATGAAA
S_Paratyphi_C str. RKS4594	ACAGCAATTTTTCGCGTTACCTGTTAATCGAGATTGAAACACATGAAA
S_diarizonae str. 11-01853	ACAGCAATTTTTCGCGTTACCTGTTAATCGAGATTGAAACACATGAAA
S_arizonae ser. 62:z4,z23:--	ACAGCAATTTTTCGCGTTACCTGTTAATCGAGATTGAAACACATGAAA
S_bongori str. NCTC 12419	ACAGCAATTTTTCGCGTTACCTGTTAATCGAGACTGAAACACATGAAA
E_coli_K12 str. K-12, MG1655	ACAGCAATTTTTCGCGTTATCTGTTAATCGAGACTGAAATACATGAAA
E_coli_O157_H7 str. EDL933	ACAGCAATTTTTCGCGTTATCTGTTAATCGAGACTGAAATACATGAAA

fourU SD start codon

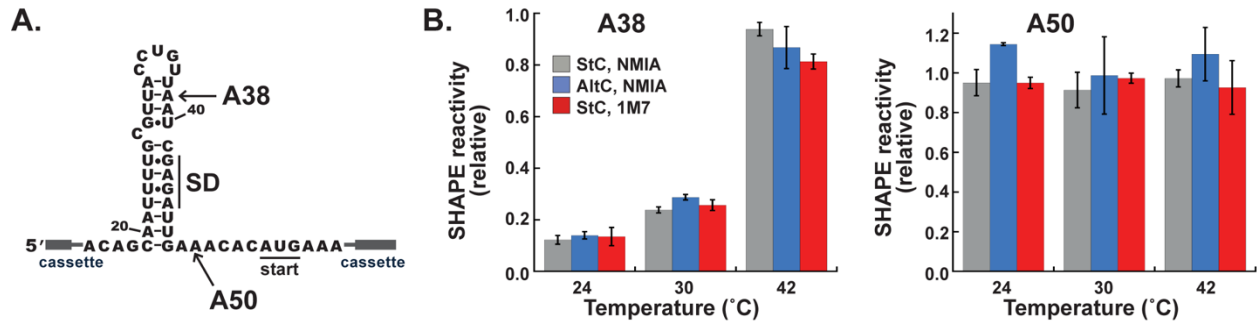
stem loop stem

B

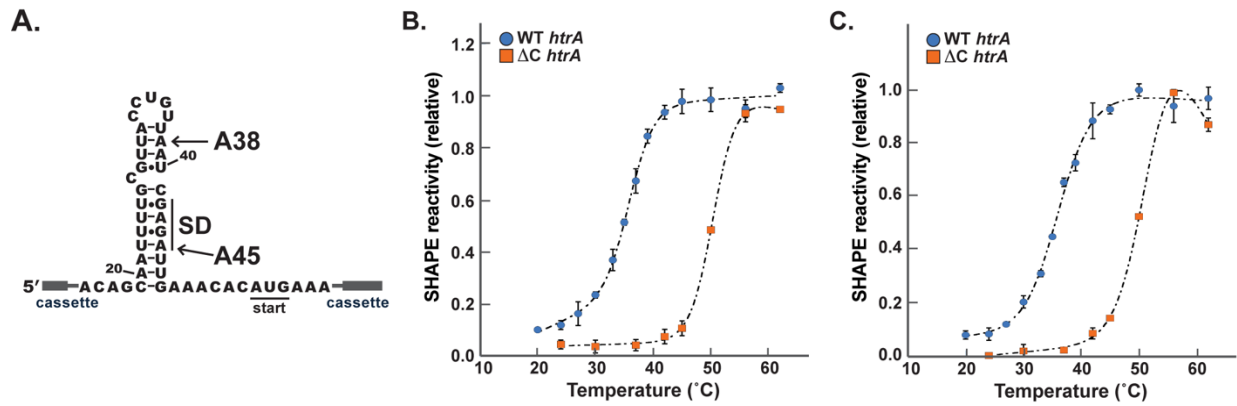
		<i>htrAp1</i>	
		↗	
S_Enteritidis	1	GTCGGTTGATTTCAGGATTATATCAGCGGGATGACTGACCTTTACGCATGGGATGAATATCGGCGTTTGATGGCGGTCGAA	
S_Typhimurium	1	GTCGGTTGATTTCAGGATTATATCAGCGGGATGACTGACCTTTACGCATGGGATGAATATCGGCGTTTGATGGCGGTCGAA	
S_Typhi	1	GTCGGTTGATTTCAGGATTATATCAGCGGGATGACTGACCTTTACGCATGGGATGAATATCGGCGTTTGATGGCGGTCGAA	
S_ParatyphiB	1	GTCGGTTGATTTCAGGATTATATCAGCGGGATGACTGACCTTTACGCATGGGATGAATATCGGCGTTTGATGGCGGTCGAA	
Ecoli_K-12	1	GCCGCTGCTGCAGGATTATATCAGCGGGATGACCGGCTTATGCGTGGGATGAATACCGACGCTGATGGCGGTAGAA	
		<i>dgt</i> stop codon	
		↓	
S_Enteritidis	81	CAGTAAATGGACTTTTGTAAAGATGGACAATAAATTTTACTTTTTCCAGAACTTTGTTCCGGAACCTTCGCGTTATAAA	
S_Typhimurium	81	CAGTAAATGGACTTTTGTAAAGATGGACAATAAATTTTACTTTTTCCAGAACTTTTATCCGGAACCTTCGCGTTATAAA	
S_Typhi	81	CAGTAAATGGACTTTTGTAAAGATGGACAATAAATTTTACTTTTTCCAGAACTTTTATCCGGAACCTTCGCGTTATAAA	
S_ParatyphiB	81	CAGTAAATGGACTTTTGTAAAGATGGACAATAAATTTTACTTTTTCCAGAACTTTGTTCCGGAACCTTCGCGTTATAAA	
Ecoli_K-12	81	CAATAACCAGGCTTTTGTAAAGACGAACAATAAATTTTACTTTTTCCAGAACTTTAGTTCCGGAACCTTCAGGCTATAAA	
		<i>htrAp2</i> <i>htrAp3</i> thermometer <i>htrA</i> start codon	
		↗ ↗ ↓	
S_Enteritidis	161	ATGAATCTGACGTACACACCAATTTTTCGCGTTACCTGTTAATCGAGATTGAAACACATGAAAAAACC	
S_Typhimurium	161	ATGAATCTGACGTACACACCAATTTTTCGCGTTACCTGTTAATCGAGATTGAAACACATGAAAAAACC	
S_Typhi	161	ATGAATCTGACGTACACACCAATTTTTCGCGTTACCTGTTAATCGAGATTGAAACACATGAAAAAACC	
S_ParatyphiB	161	ATGAATCTGACGTACACACCAATTTTTCGCGTTACCTGTTAATCGAGATTGAAACACATGAAAAAACC	
Ecoli_K-12	161	ACGAATCTGAAGAACACACCAATTTTTCGCGTTACTGTTAATCGAGACTGAAATACATGAAAAAACC	

Supplemental Figure 1. The *htrA* thermometer and upstream sequence is conserved across *Salmonella enterica*, but differs in more diverse prokaryotes. A. Alignment of the *htrA* thermometer sequence from diverse strains of *S. enterica* with the corresponding sequence from *S. bongori* and *E. coli*. The thermometer sequence is absolutely conserved in the *S. enterica* strains. Sequence differences in *S. bongori* and *E. coli* would be predicted to destabilize the hairpin significantly, as they disrupt a base pair observed in the *S. enterica htrA* hairpin. Sequences were retrieved from GenBank. *S. enterica* subsp. *enterica* serovars include: *S. Enteritidis* str. P125109 (NC_011294.1), *S. Typhimurium* str. LT2 (NC_003197.2), *S. Typhi* str. CT18 (NC_003198.1), *S. Paratyphi A* str. ATCC 9150 (NC_006511.1), *S. Paratyphi B* str. SPB7 (CP000886.1), and *S. Paratyphi C* str. RKS4594 (NC_012125.1). Other *S. enterica* subspecies include: *S. enterica* subsp. *diarizonae* str. 11-01853 (CP011289.1) and *S. enterica* subsp. *arizonae* serovar 62:z4,z23:-- (CP000880.1). The *S. bongori* sequence comes from NCTC 12419, culture collection SGSC SARC11 (FR877557.1), while the *E. coli* sequences come from str. K-12 MG1655 (U00096.3) and O157:H7 str. EDL933 (AE005174.2). Strains for the alignment were chosen based on the phylogenetic trees from Desai PT, Porwollik S, Long F, Cheng P, Wollam A, Clifton S, Weinstock GM, McClelland M. 2013. Evolutionary genomics of the *Salmonella enterica* subspecies. mBio 4(2):e00579-12. doi:10.1128/mBio.00579-12.

B. Alignment of the extended region upstream of the *htrA* coding region of several *S. enterica* strains with that of *E. coli*. Transcription start sites are noted, with the two upstream start sites in the coding region of the gene *dgt* (stop codon marked). The *S. Enteritidis* sequence was used for the extended sequence in Figure 5.

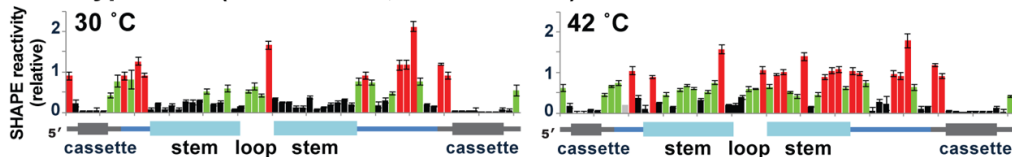


Supplemental Figure 2. Trends in reactivity with respect to temperature are independent of the structural cassette or of the SHAPE reagent used. **A.** Position of the nucleotides A38 and A50, shown on the secondary structure of the *htrA* thermometer. **B.** SHAPE results for A38 and A50. A38 is part of the upper stem of the thermometer hairpin, and it is thus predicted to have increased reactivity at elevated temperatures. A50 is predicted to be unpaired at all temperatures. The standard cassette (StC) is the sequence used throughout the rest of the work. Alternate cassette (AltC, shown in blue) used a different sequence in the first hairpin of the 3' end of the structure cassette. Experiments testing the effect of using the SHAPE reagent 1M7 are shown in red. Error bars represent the standard deviation of two to six experiments.

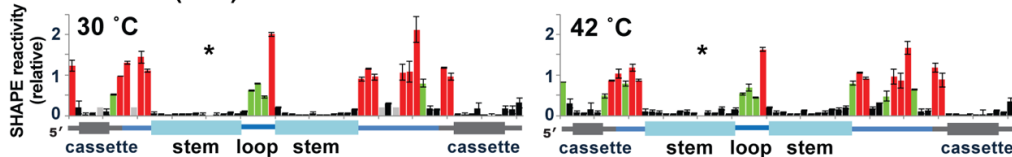


Supplemental Figure 3. Representative SHAPE melting curves, with error shown. A. Position of the nucleotides A38 and A45, shown on the secondary structure of the *htrA* thermometer. **B., C.** SHAPE melting curves for A38 (**B**) and A45 (**C**) for the wild-type thermometer (blue circles) and for the ΔC mutant (orange squares). Error bars represent the standard deviation of two to six experiments.

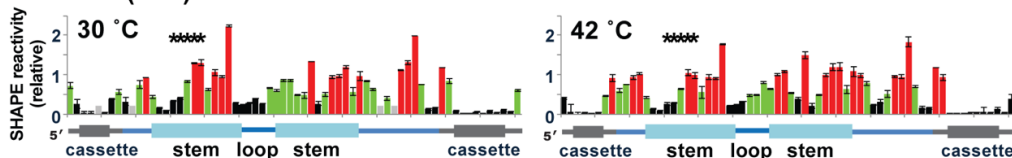
A. wild-type *htrA* (n=6 for 30 °C; n=4 for 42 °C)



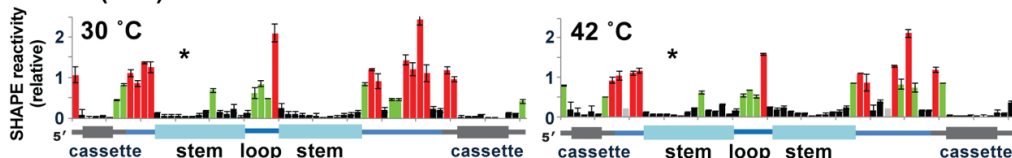
B. ΔC control (n=2)



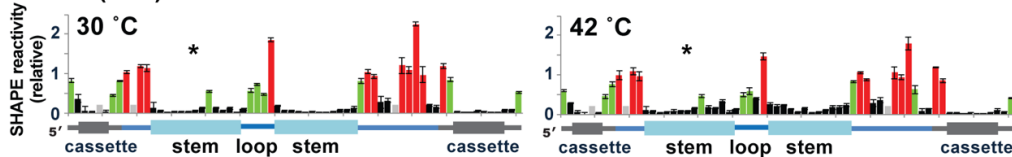
C. AAAAA (n=2)



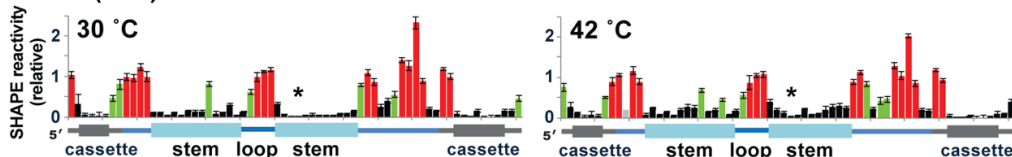
D. U23C (n=2)



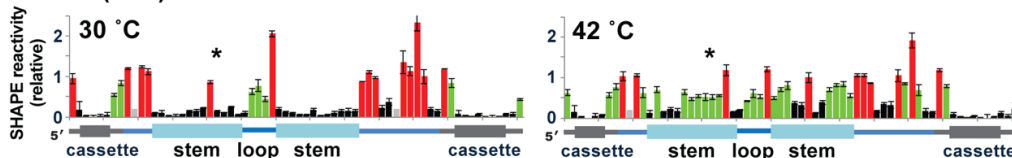
E. U25C (n=3)



F. U40C (n=4)



G. G28A (n=3)



Supplemental Figure 4. SHAPE data for the *htrA* stem mutants at 30 °C and 42 °C. Average values for the wild-type *htrA* thermometer and the ΔC control are shown in panels **A** and **B**, respectively, for comparison, for comparison with results from the AAAAA mutant (**C**), U23C (**D**), U25C (**E**), U40C (**F**), and G28A (**G**). Positions of mutation(s) have been indicated with an asterisk (for the ΔC control, the asterisk marks the site of the missing nucleotide). Coloring as in Figure 2. Positions for which SHAPE values could not be determined due to high background signal are shown in grey. Values are the average of several independent experiments, as marked, with standard deviation indicated with error bars. Negative values are set to zero.