

S1 Appendix Supplemental Material
**Evidence for Complex Life Cycle Constraints
on Salamander Body Form Diversification**

Ronald M. Bonett and Andrea L. Blair

Contents	page #
Supplemental Table S1 (Ancestral state reconstructions)	2
Supplemental Figure S1 (Rates of larval shape evolution)	3
Supplemental Table S2 (Fit of trunk vertebral evolution models without direct developers)	4
Supplemental Table S3 (Fit of adult body form evolution models without direct developers)	5
Supplemental Table S4 (Fit of adult body form evolution models with direct developers)	6
Supplemental Table S5 (Fit of trunk vertebral evolution models with direct developers)	7
Supplemental Table S6 (Fit of evolutionary rate matrix models to direct developers only)	8
Supplemental Table S7 (Fit of trunk vertebral evolution models to direct developers only)	8
Supplemental Table S8 (Fit of trunk form evolution models to direct developers only)	8
Supplemental Table S9 (Ecological and phenotypic data, and Genbank accession numbers)	9
Supplemental Table S10 (Partitions and models of sequence evolution)	15
Supplemental Table S11 (Divergence time calibration priors)	15
Supplemental Figure S2 (Consensus phylogeny of 516 salamander taxa)	16
Supplemental Figure S3 (Landmarking strategy for larval geometric morphometrics)	17
Supplemental references	18
Specimen voucher numbers	22
Acknowledgements for specimen access and loans	23

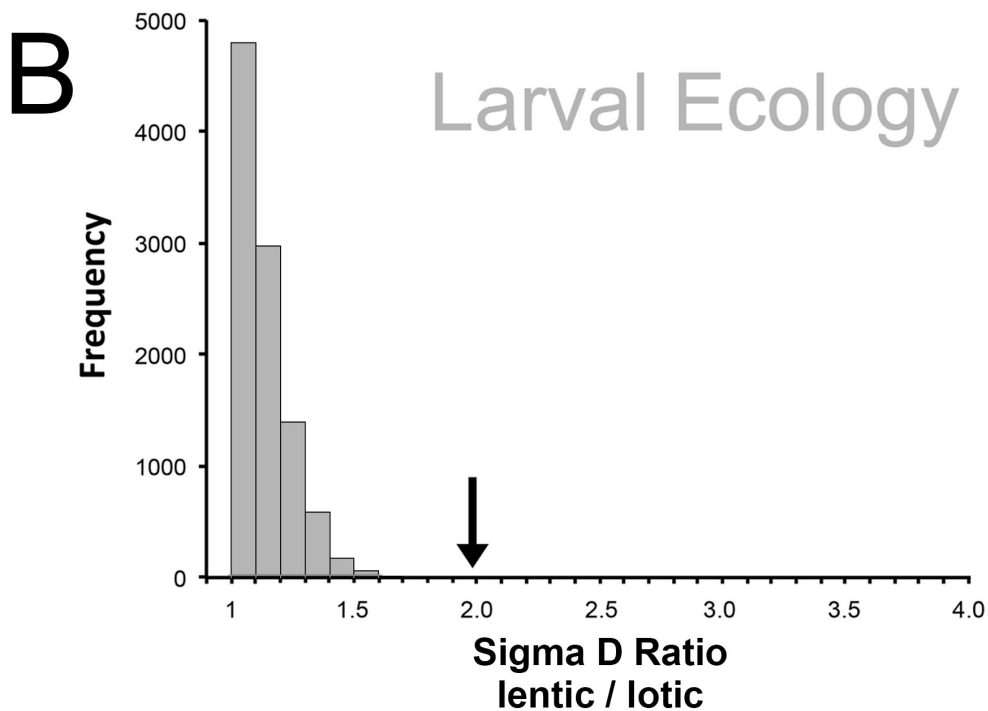
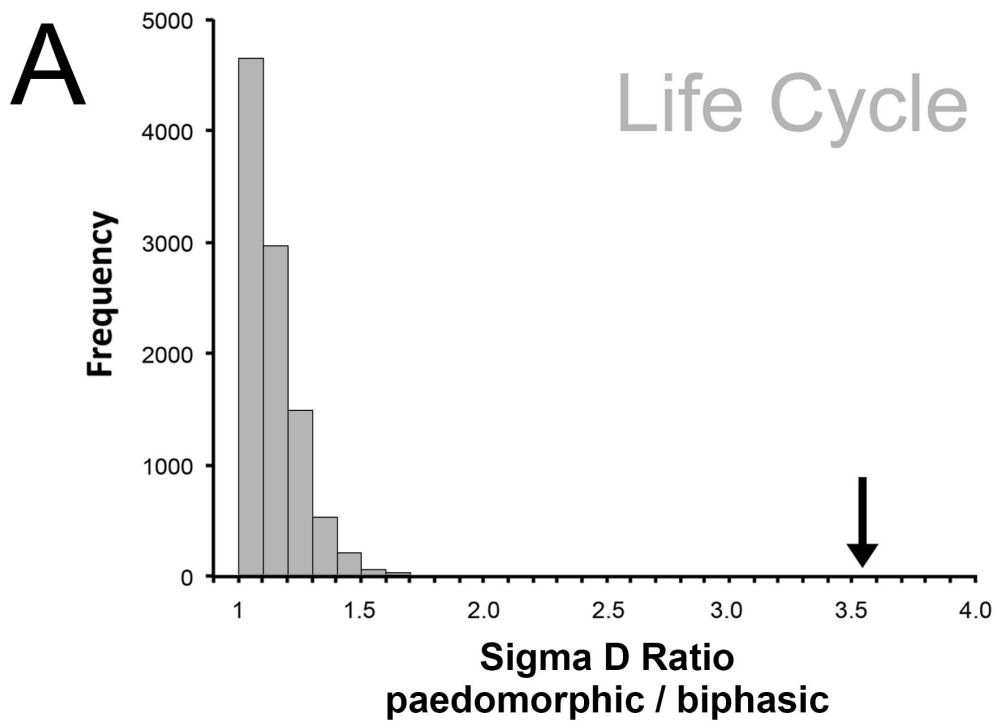
Supplemental Table S1. Bayesian ancestral state reconstructions of salamander life cycle modes and body form metrics.

Life cycle was reconstructed using BayesTraits [S1] and the three modes were treated as ordered categorical states: paedomorphic (pd), biphasic (bi), and direct development (dd). Median *Trunk Vertebral Number* (Vert) and median *Adult Body Forms* (ABF) including 95% confidence intervals were reconstructed in StableTraits [S2]. Ancestral Vert estimates were rounded to the nearest integer. The ancestral state estimates listed below include the common ancestors of all 10 extant families and internal nodes joining families. The life cycle stage with the highest probability, and median *Trunk Vertebral Number* and *Adult Body Form* estimates are shown in Fig. 1.

Node	pd	bi	dd	Vert Median	Vert 95% CI	ABF Median	ABF 95% CI
Living Salamanders (Root Node)	0.13	0.87	0.00	17	15-18	8.8	6.1-11.5
Cryptobranchoidea	0.32	0.68	0.00	17	17-18	8.7	6.0-11.3
Salamandroidea + Sirenoidea	0.21	0.79	0.00	17	15-18	9.1	6.6-11.7
Salamandroidea	0.02	0.98	0.00	17	14-18	9.1	6.9-11.3
Ambystomatidae + Dicamptodontidae + Salamandridae	0.01	0.99	0.00	14	13-17	8.7	6.6-10.9
Ambystomatidae + Dicamptodontidae	0.01	0.99	0.00	14	13-15	7.9	5.6-10.2
Proteidae + Rhyacotritonidae + Amphiumidae + Plethodontidae	0.55	0.45	0.00	17	15-18	9.5	7.4-11.8
Rhyacotritonidae + Amphiumidae + Plethodontidae	0.03	0.88	0.09	16	15-17	9.9	7.7-12.1
Amphiumidae + Plethodontidae	0.14	0.51	0.35	16	15-17	10.3	8.1-12.6
Cryptobranchidae	1.00	0.00	0.00	17	17-18	6.8	4.8-9.0
Hynobiidae	0.02	0.98	0.00	17	17-17	8.4	5.7-11.1
Sirenidae	1.00	0.00	0.00	33	31-36	14.8	12.0-23.2
Ambystomatidae	0.00	1.00	0.00	14	13-15	7.6	6.0-9.0
Dicamptodontidae	0.32	0.68	0.00	14	14-14	6.5	5.9-7.2
Salamandridae	0.00	1.00	0.00	13	13-13	7.8	6.2-9.7
Proteidae	0.99	0.01	0.00	17	15-18	9.6	6.9-12.6
Rhyacotritonidae	0.00	1.00	0.00	16	15-16	9.4	8.3-10.4
Amphiumidae	1.00	0.00	0.00	61	61-61	17.1	15.6-18.4
Plethodontidae	0.00	0.04	0.96	16	15-17	10.6	9.4-11.9

Considering potential discordance between Mesozoic fossils and reconstructions based on extant species: Our ancestral life cycle reconstructions were based entirely on data from extant species, and only obligately paedomorphic taxa were coded as paedomorphic. The reconstruction shows support for biphasic ancestors for all salamanders, salamandroids, and cryptobranchoids. This is partially supported by several fossil salamanders from the Late Jurassic through the Early Cretaceous with a metamorphosed morphology (cryptobranchoids: *Liaoxitriton daohugouensis* [S3-S5], *Laccotriton subsolanus* [S5, S6], *Pangerpeton sinense* [S4, S5, S7], *Iridotriton hechti* [S4, S5, S8], *Nuominerpeton aquilonaris* [S9]; and a salamandroid: *Valdotriton gracilis* [S4, S5, S10]). However, there are also several other fossil salamanders from the same range of time that were likely paedomorphic (salamandroids: *Beiyanerpeton jianpingensis* [S4, S5] and *Qinglongtriton gangouensis* [S11]; and cryptobranchoids: *Sinerpeton fengshanensis* [S5, S12], *Chunerpeton tianyiensis* [S4, S5, S13], *Jeholotriton paradoxus* [S5, S14]). Furthermore, the Family Karauridae is thought to have been paedomorphic (aquatic), and is often placed as sister to all extant salamanders [S4, S5, S15]. This has been used as evidence for a paedomorphic ancestor for all extant salamanders [S15]. The nature of paedomorphosis (facultative vs. obligate) in fossil taxa is difficult to evaluate. Curiously though, none of these paedomorphic Jurassic taxa had obviously aberrant body forms (or trunk vertebral numbers [S5]). Based on our trait analyses, this would suggest that these taxa were facultatively paedomorphic or recently evolved obligate paedomorphs. This notion is also supported by the suggested close relationships between many paedomorphic and metamorphic Jurassic taxa [S4, S5]. Nevertheless, we find no major difference in patterns of body form evolution when we fix the basal salamander node to paedomorphic (Tables S2 and S3).

Considering fossil taxa in trunk vertebral number reconstructions: Reconstructions of ancestral trunk vertebral numbers were based completely on data from extant species. However, our estimates (range 14 to 17) overlap well with counts from Late Jurassic through Early Cretaceous species assigned to the Salamandroidea and Cryptobranchoidea (range 13 to 16 [S5]). Note: Ascarrunz et al. [S5] counted all presacrals; our counts were from between the atlas and the sacrum (i.e., minus the atlas).



Supplemental Figure S1. Comparisons of rates of larval shape evolution. Sigma D ratio calculated using the *compare.evol.rates* function in geomorph [S16, S17] shows the differences in rates of larval body shape evolution for species partitioned by life cycle (A, paedomorphic vs. biphasic) or larval ecology (B, lentic vs. lotic). Rates of larval body shape evolution (arrows) were estimate to be 3.54 times higher for paedomorphs than biphasics, and 1.99 times higher for lentic compared to lotic dwelling larvae.

Supplemental Table S2. Comparison of the fit of BM and OU models to salamander *Trunk Vertebral Number* with differential coding of larval ecologies and life cycles, and with alternative coding of facultatively paedomorphic taxa.

Three alternative partitions were tested: *Life Cycle* (one-part vs. two-part), *Larval Ecology* (lotic vs. lentic), and *Facultative* (facultative paedomorphs coded the same as obligate paedomorphs). Throughout this study we code facultatively paedomorphic taxa as biphasic, because we were primarily interested the consequences of long-term life cycle transitions. Fitting models with facultative paedomorphs coded the same as obligate paedomorphs allowed us to test whether our results (life cycle vs. larval ecology) were robust regardless of how we coded taxa with variable life cycles (i.e. facultative paedomorphs). It also allows us to determine whether the vertebral columns of facultative paedomorphs evolve more similar to obligate paedomorphs or biphasics. Models were fit using OUwie [S18] and parameters allowed to vary among groups included rates of evolution (σ^2) and optima (θ). Model fit was based on Δ AIC and AIC Weights (w_i). OU-models with multiple selection parameters (α : $OU_{\theta\alpha}$ and $OU_{\theta\sigma^2\alpha}$) were not included, because they were poorly fit or did not improve upon simpler models (based on preliminary analyses). BM-models with multiple rates and a single optimum (BM_{σ^2}) were also examined, but were never better than the best $BM_{\theta\sigma^2}$ model. Parameter estimates and 95% confidence intervals for the overall best-fit model (Life Cycle $BM_{\theta\sigma^2}$) are listed below. Life cycle models with different rates and optima for paedomorphs vs. biphasics (including facultative paedomorphs) were substantially better than all models based on larval ecology (Δ AIC > 114). Facultative paedomorphs were a better fit when combined with biphasics than with obligate paedomorphs. However, even when facultative species were coded as paedomorphic, models with rate and optimum differences separated by life cycle (facultative + obligate vs. biphasic) were still a better fit than larval ecology (Δ AIC > 49).

Model	-lnL	AIC	Δ AIC	w_i
Life Cycle $BM_{\theta\sigma^2}$	-268.45	544.91	0.00	0.6995
Life Cycle $OU_{\theta\sigma^2}$	-268.30	546.60	1.69	0.3004
Facultative $BM_{\theta\sigma^2}$	-300.73	609.47	64.56	< 0.0001
Facultative $OU_{\theta\sigma^2}$	-300.82	611.64	66.73	< 0.0001
Larval Ecol $BM_{\theta\sigma^2}$	-326.72	661.44	116.53	< 0.0001
Larval Ecol $OU_{\theta\sigma^2}$	-326.47	662.95	118.04	< 0.0001
Life Cycle OU_{θ}	-333.77	675.55	130.64	< 0.0001
Facultative OU_{θ}	-336.50	680.99	136.08	< 0.0001
BM1	-339.68	683.36	138.45	< 0.0001
Larval Ecol OU_{θ}	-338.48	684.97	140.06	< 0.0001
OU1	-339.68	685.36	140.45	< 0.0001

θ : bi = 15.69±0.04; pd = 40.52±0.55

σ^2 : bi = 0.040±0.0005; pd = 1.994±0.108

Outliers: Analyses performed without *Amphiuma*. Salamanders of the Family Amphiumidae have approximately 60 trunk vertebrae (Fig. 1), which is two to four times as many as nearly all other extant salamanders. Analyses performed without the three extant species of *Amphiuma*, also recovered Life Cycle $BM_{\theta\sigma^2}$ as the best-fit model. Paedomorphic lineages still show a higher rate of trunk vertebral evolution than biphasic lineages (by >11 times; σ^2 : bi = 0.041±0.0005; pd = 0.472±0.0065), but with a lower optimum (θ : bi = 15.93±0.0266; pd = 29.99±0.2960). Multi-rate life cycle models were still a better fit than all models based on larval ecology (Δ AIC > 72).

Transformation: Why we did not transform vertebral count data. Throughout the study we did not transform vertebral count data because there was no correlation between numbers of trunk vertebrae and their variance. For example, paedomorphic clades with the highest numbers of trunk vertebrae (e.g. Amphiumidae) do not necessarily have more variation among species than paedomorphic clades with fewer trunk vertebrae (e.g. plethodontids of the genus *Eurycea* from central Texas). To evaluate this we tested for a correlation between the independent contrasts and ancestral state estimates [S19] of numbers of trunk vertebrae. These variables were not correlated ($R^2 = 0.0570$). Nevertheless, analyses performed on Log10-transformed trunk vertebral numbers yielded the same results as those presented. In the case above, Life Cycle $BM_{\theta\sigma^2}$ was still a better-fit model (Δ AIC > 68) than every model based on larval ecology. Parameter estimates for analyses of Log10-transformed data under the best-fit model (Life Cycle $BM_{\theta\sigma^2}$) were also relatively the same: a higher rate of evolution (by >13 times; σ^2 : bi = 0.000033±4.13e⁻⁰⁷; pd = 0.00043±8.47e⁻⁰⁶) and optimum number of trunk vertebrae (antilog θ : bi = 15.67±1.00; pd = 40.36±1.02) in paedomorphic compared to biphasic lineages.

Ancestral Salamanders Paedomorphic? Fossil evidence suggests that ancestral salamanders may have been paedomorphic [S15]. We set the ancestral salamander node to paedomorphic and recovered the same patterns of model fit for trunk vertebrae as when this node was metamorphic. The only difference was that the optimum vertebral number of paedomorphs was reduced (best fit model: Life Cycle $BM_{\theta\sigma^2}$; θ : bi = 15.07±0.09; pd = 26.85±0.44; σ^2 : bi = 0.041±0.0005; pd = 1.48±0.072).

Supplemental Table S3. Comparison of the fit of BM and OU models to salamander *Adult Body Form* data with differential coding of larval ecologies and life cycles, and with alternative coding of facultatively paedomorphic taxa.

Three alternative partitions were tested: *Life Cycle* (one-part vs. two-part), *Larval Ecology* (lotic vs. lentic), and *Facultative* (facultative paedomorphs coded the same as obligate paedomorphs). Throughout this study we code facultatively paedomorphic taxa as biphasic, because we were primarily interested the consequences of long-term life cycle transitions. Fitting models with facultative paedomorphs coded the same as obligate paedomorphs allowed us to test whether our results (life cycle vs. larval ecology) were robust, regardless of how we coded taxa with variable life cycles (i.e. facultatively paedomorphs). It also allows us to determine whether the adult body forms of facultative paedomorphs evolve more similar to obligate paedomorphs or biphasics. Models were fit using OUwie [S18] and parameters allowed to vary among groups included rates of evolution (σ^2) and optima (θ). Model fit was based on Δ AIC and AIC Weights (w_i). OU-models with multiple selection parameters (α : $OU_{\theta\alpha}$ and $OU_{\theta\sigma^2\alpha}$) were not included, because they were poorly fit or did not improve upon simpler models (based on preliminary analyses). BM-models with multiple rates and a single optimum (BM_{σ^2}) were also examined, but were never better than the best $BM_{\theta\sigma^2}$ model. Parameter estimates and 95% confidence intervals for the overall best-fit model (Life Cycle $OU_{\theta\sigma^2}$) are listed below. Life cycle models with different rates and optima for paedomorphs vs. biphasics (including facultative paedomorphs) were substantially better than all models based on larval ecology (Δ AIC > 74). Facultative paedomorphs were better fit when combined with biphasics than with obligate paedomorphs. However, even when facultative species were coded as paedomorphic, models with rate differences separated by life cycle (facultative + obligate vs. biphasic) were still a better fit than larval ecology (Δ AIC > 40).

Model	-lnL	AIC	Δ AIC	w_i
Life Cycle $OU_{\theta\sigma^2}$	-220.63	451.25	0.00	0.9888
Life Cycle $BM_{\theta\sigma^2}$	-226.11	460.22	8.97	0.0115
Facultative $OU_{\theta\sigma^2}$	-237.06	484.11	32.86	< 0.0001
Facultative $BM_{\theta\sigma^2}$	-242.81	493.61	42.36	< 0.0001
Life Cycle OU_{θ}	-262.28	532.57	81.32	< 0.0001
Larval Ecol $OU_{\theta\sigma^2}$	-262.21	534.42	83.17	< 0.0001
OU1	-264.40	534.80	83.55	< 0.0001
Facultative OU_{θ}	-263.67	535.33	84.08	< 0.0001
BM1	-265.92	535.85	84.33	< 0.0001
Larval Ecol OU_{θ}	-263.79	535.58	84.60	< 0.0001
Larval Ecol $BM_{\theta\sigma^2}$	-264.01	536.01	84.76	< 0.0001

θ : bi = 8.27±0.01; pd = 15.57±0.09

σ^2 : bi = 0.083±0.001; pd = 1.538±0.023

α : 0.013±0.0003

Transformation: Why we did not transform “*Adult Body Form*” data. Our metric of *Adult Body Form* was a ratio of body length divided by body width and therefore was not log transformed.

Outliers: Removing the most elongate salamanders from “*Adult Body Form*” analyses was unwarranted. Numbers of trunk vertebrae are dramatically higher for one clade (Amphiumidae), and therefore we performed analyses with and without this family (see Table S2). However, even though amphiumids have over 20 more trunk vertebrae than other families, they are not equivalently more elongate than all other elongate salamanders. In fact, the salamanders with the most elongate *Adult Body Form* (highest body length to body width ratio) were actually species from four different families (Amphiumidae, Plethodontidae, Proteidae, and Sirenidae). Given the repeated evolution of extreme body elongation, it did not seem reasonable to remove all of these taxa from our analyses.

Ancestral Salamanders Paedomorphic? Fossil evidence suggests that ancestral salamanders may have been paedomorphic [S15]. When we set the ancestral node of salamanders to paedomorphic we recovered the same patterns of model fit for adult body form as when this node was primarily metamorphic. The only difference was that the estimated body form optima were reduced (best fit model: Life Cycle $OU_{\theta\sigma^2}$; θ : bi = 7.86±0.018; pd = 13.47±0.089; σ^2 : bi = 0.091±0.0014; pd = 1.44±0.017).

Supplemental Table S4. Comparison of the fit of BM and OU models to salamander *Adult Body Form* data including direct developing lineages.

Five alternative partitions were tested: *Adult Ecology* (aquatic vs. terrestrial), *2 Life Cycles* (simple vs. complex), *3 Life Cycles* (paedomorphic, biphasic, direct development), *Partial Metamorph*, and *Larval Ecology* (lotic, lentic, direct development). For *Adult Ecology*, direct developers and biphasics were merged to create groups with terrestrial vs. aquatic (paedomorphic) adults. In the *2 Life Cycles* model, paedomorphs and direct developers were merged to create simple vs. complex (biphasic) life cycle groups. For the *Partial Metamorph* model, two families that partially metamorphose (Amphiumidae and Cryptobranchidae) were coded the same as groups that fully transform (biphasics and direct developers) vs. obligate paedomorphs (as in all other analyses). *Larval Ecology* separated taxa into three groups based on their “larval environment”: lotic, lentic or direct development. Models were fit using OUwie [S18] and parameters allowed to vary among groups included rates of evolution (σ^2) and/or optima (θ). Model fit was based on ΔAIC and AIC Weights (w_i). OU-models with multiple selection parameters (α : $OU_{\theta\alpha}$ and $OU_{\theta\sigma^2\alpha}$) were not included, because they were poorly fit or did not improve upon simpler models (based on preliminary analyses). BM-models with multiple rates and a single optimum (BM_{σ^2}) were also examined, but were never better than the best $BM_{\theta\sigma^2}$ model. Parameter estimates and 95% confidence intervals for the best-fit model are listed below. The life cycle model with different rates and optima for paedomorphs, biphasics, and direct developers (3 Life Cycles $OU_{\theta\sigma^2}$) was substantially better than all other models. Parameters for direct developers were more similar to biphasics than paedomorphs. Consistent with this, the second best fit model based on adult ecology (Adult Ecology $OU_{\theta\sigma^2}$) treated biphasic and direct developing taxa as a single group. Partial metamorphs were better fit when combined with permanently aquatic obligate paedomorphs than with fully transformed taxa with terrestrial adults (biphasics and direct developers; compare Adult Ecology $OU_{\theta\sigma^2}$ to Partial Metamorph $OU_{\theta\sigma^2}$: $\Delta AIC > 38$). Similar to analyses without direct developers, models based on larval ecology were poorly fit compared to all multi-rate models based on life cycle, adult ecology, or partial metamorphosis.

Model	-lnL	AIC	ΔAIC	w_i
3 Life Cycles $OU_{\theta\sigma^2}$	-386.81	787.63	0.00	0.9578
Adult Ecology $OU_{\theta\sigma^2}$	-391.97	793.94	6.31	0.0408
3 Life Cycles $BM_{\theta\sigma^2}$	-394.52	801.05	13.42	0.0012
Adult Ecology $BM_{\theta\sigma^2}$	-398.26	804.53	16.90	0.0002
Partial Metamorph $OU_{\theta\sigma^2}$	-411.13	832.26	44.63	< 0.0001
Partial Metamorph $BM_{\theta\sigma^2}$	-414.01	836.02	48.39	< 0.0001
2 Life Cycles $OU_{\theta\sigma^2}$	-414.38	838.76	51.13	< 0.0001
2 Life Cycles $BM_{\theta\sigma^2}$	-422.17	852.34	64.71	< 0.0001
Larval Ecology $OU_{\theta\sigma^2}$	-429.49	872.98	85.35	< 0.0001
2 Life Cycles OU_{θ}	-434.16	876.32	88.69	< 0.0001
Larval Ecology $BM_{\theta\sigma^2}$	-433.21	878.43	90.80	< 0.0001
Adult Ecology OU_{θ}	-436.02	880.04	92.41	< 0.0001
Partial Metamorph OU_{θ}	-436.25	880.51	92.88	< 0.0001
3 Life Cycles OU_{θ}	-435.69	881.38	93.75	< 0.0001
OU1	-438.61	883.23	95.60	< 0.0001
Larval Ecology OU_{θ}	-437.78	885.57	97.94	< 0.0001
BM1	-440.82	885.65	98.02	< 0.0001

θ : bi = 8.17±0.02; pd = 15.39±0.08; dd = 10.64±0.04

σ^2 : bi = 0.084±0.0013; pd = 1.549±0.026; dd = 0.135±0.0011

α : 0.0138±0.0002

Different plethodontid divergence time estimates (70 mya vs. 85 mya) produce the same results. All direct developers are in the Family Plethodontidae. Divergence time estimates for the deepest node within the family have varied: 127 [S20], 85 [S21], and 66 [S22] mya. The analyses presented were a based on an average plethodontid divergence of 70 mya. However, we also performed this (and all) analyses with a plethodontid divergence of ~85 mya. The model ranks and patterns of significance were the same, as were the rates and optima among the 3 life cycle modes.

Removing all bolitoglossines with 14 trunk vertebrae provides even stronger support for this analysis. Nearly all bolitoglossines (except *Oedipina*) have 14 trunk vertebrae [S23, S24]. When these taxa were removed the 3 Life Cycles $OU_{\theta\sigma^2}$ model was an even better fit (compared to all other models, $\Delta AIC > 10$). Parameter estimates were also similar.

Treating ancestral plethodontids as biphasic (vs. direct developers) provides even stronger support for this analysis. Ancestral plethodontids were likely direct developers [S25-S27]. To evaluate the influence of the ancestral plethodontid life cycle on our results, we “fixed” this node as biphasic by adding an adjacent tip with and infinitely small branch coded as biphasic. Parameter estimates were the same, and the 3 Life Cycles $OU_{\theta\sigma^2}$ model was best fit compared to all other models ($\Delta AIC > 18$).

Supplemental Table S5. Comparison of the fit of BM and OU models to salamander *Trunk Vertebral Number* including direct developing lineages. Five alternative partitions were tested: *Adult Ecology* (aquatic vs. terrestrial), *2 Life Cycles* (simple vs. complex), and *3 Life Cycles* (paedomorphic, biphasic, direct development), *Partial Metamorph*, and *Larval Ecology* (lotic, lentic, direct development). For *Adult Ecology*, direct developers and biphasics were merged to create groups with terrestrial vs. aquatic (paedomorphic) adults. In the *2 Life Cycles* model, paedomorphs and direct developers were merged to create simple vs. complex (biphasic) life cycle groups. For the *Partial Metamorph* model, two families that partially metamorphose (Amphiumidae and Cryptobranchidae) were coded the same as groups that fully transform (biphasics and direct developers) vs. obligate paedomorphs (as in all other analyses). *Larval Ecology* separated taxa into three groups based on their “larval environment”: lotic, lentic or direct development. Models were fit using OUwie [S18] and parameters allowed to vary among groups included rates of evolution (σ^2) and/or optima (θ). Model fit was based on Δ AIC and AIC Weights (w_i). OU-models with multiple selection parameters (α : $OU_{\theta\alpha}$ and $OU_{\theta\sigma^2\alpha}$) were not included, because they were poorly fit or did not improve upon simpler models (based on preliminary analyses). BM-models with multiple rates and a single optimum (BM_{σ^2}) were also examined, but were never better than the best $BM_{\theta\sigma^2}$ model. Parameter estimates and 95% confidence intervals for the best-fit model are listed below. Models with different rates and optima based on adult ecology or three different life cycle modes were substantially better than all other models (Δ AIC > 118). The simplest best-fit model was based on adult ecology (Adult Ecology $BM_{\theta\sigma^2}$). Partial metamorphs were better fit when combined with permanently aquatic obligate paedomorphs than with fully transformed taxa with terrestrial adults (biphasics and direct developers; compare Adult Ecology $BM_{\theta\sigma^2}$ to Partial Metamorph $OU_{\theta\sigma^2}$: Δ AIC > 188). Similar to analyses without direct developers, models based on larval ecology were a poor-fit compared to all multi-rate models based on adult ecology or life cycle.

Model	-lnL	AIC	Δ AIC	w_i
Adult Ecology $BM_{\theta\sigma^2}$	-466.60	941.20	0.00	0.5695
Adult Ecology $OU_{\theta\sigma^2}$	-466.60	943.20	2.00	0.2095
3 Life Cycles $BM_{\theta\sigma^2}$	-465.85	943.70	2.50	0.1631
3 Life Cycles $OU_{\theta\sigma^2}$	-465.89	945.78	4.58	0.0577
Larval Ecology $BM_{\theta\sigma^2}$	-526.04	1064.07	122.87	< 0.0001
Larval Ecology $OU_{\theta\sigma^2}$	-526.82	1067.63	126.43	< 0.0001
2 Life Cycles $BM_{\theta\sigma^2}$	-534.06	1076.12	134.92	< 0.0001
2 Life Cycles $OU_{\theta\sigma^2}$	-534.06	1078.12	136.92	< 0.0001
Partial Metamorph $BM_{\theta\sigma^2}$	-560.72	1129.43	188.23	< 0.0001
Partial Metamorph $OU_{\theta\sigma^2}$	-560.72	1131.43	190.23	< 0.0001
3 Life Cycles OU_{θ}	-562.33	1134.66	193.46	< 0.0001
Adult Ecology OU_{θ}	-563.53	1135.06	193.86	< 0.0001
2 Life Cycles OU_{θ}	-568.41	1144.88	203.68	< 0.0001
Larval Ecology OU_{θ}	-569.68	1149.37	208.17	< 0.0001
BM1	-572.92	1149.84	208.64	< 0.0001
Partial Metamorph OU_{θ}	-571.36	1150.71	209.51	< 0.0001
OU1	-572.92	1151.84	210.64	< 0.0001

θ : bi+dd = 15.71±0.04; pd = 40.90±0.44

σ^2 : bi+dd = 0.046±0.0026; pd = 1.948±0.0921

Different plethodontid divergence time estimates (70 mya vs. 85 mya) produce the same results. The analyses presented in the paper are based on an average plethodontid divergence of 70 mya. However, we also performed this (and all) analyses with a plethodontid divergence of ~85 mya. Both divergence dates produce similar parameter estimates, and show that the Adult Ecology $BM_{\theta\sigma^2}$ and 3 Life Cycles $BM_{\theta\sigma^2}$ models are statistically the same.

Removing all bolitoglossines with 14 trunk vertebrae has only a small (but not substantial) effect on this result. Nearly all bolitoglossines (except *Oedipina*) have 14 trunk vertebrae [S23, S24]. When we remove these taxa the 3 Life Cycles $BM_{\theta\sigma^2}$ model was favored slightly over the Adult Ecology $BM_{\theta\sigma^2}$ model, but this was not substantial (Δ AIC = 4.5). The optimum for dd increased to 18 trunk vertebrae, but the rate differences among the three groups did not change.

Treating ancestral plethodontids as biphasic (vs. direct developers) has no effect on this analysis. Ancestral plethodontids were likely direct developers [S25-S27]. To evaluate the influence of the ancestral plethodontid life cycle on our results, we “fixed” this node as biphasic by adding an adjacent tip with an infinitely small branch and coded it as biphasic. Analyses with the ancestral plethodontid as biphasic produced equivalent parameters as the results shown above. The best-fit models (3 Life Cycles $BM_{\theta\sigma^2}$ and Adult Ecology $BM_{\theta\sigma^2}$) were also equivalent (Δ AIC = 0.2).

Supplemental Tables S6, S7 and S8. Comparison of the fit of BM and/or OU models to *Trunk Vertebral Number* and *Adult Trunk Form*, or both variables for direct developers only. These analyses test whether there are different rates of *Adult Trunk Form* and *Trunk Vertebral Number* evolution in bolitoglossines (minus *Oedipina*) compared to the rest of direct developers (other dds). Bolitoglossines are the most species rich clade of salamanders and nearly all (except the genus *Oedipina*) have 14 trunk vertebrae [S23, S24]. These analyses tests whether the fixed number of trunk vertebrae in this group is concomitant with strong constrains on the rate of evolution of adult trunk form. Alternatively, bolitoglossines may have evolved adult trunk form at a rate similar to other direct developers, despite static trunk vertebral numbers. BM bivariate evolutionary rate matrices for the two groups were compared to evaluate whether they shared common rates or common correlations (Table S6). Also, models were fit using OUwie [S18] and parameters allowed to vary among groups included rates of evolution (σ^2) and/or optima (θ ; Tables S7 and S8). Model fit was based on Δ AIC and AIC Weights (w_i). OU-models with multiple selection parameters (α : $OU_{\theta\alpha}$ and $OU_{\theta\sigma^2\alpha}$) were not included, because they were poorly fit or did not improve upon simpler models. Below each table are parameter estimates and 95% confidence intervals for the best-fit models.

Table S6. Bivariate evolutionary rate matrix analysis. The best-fit model was “no common structure” between the bivariate matrices (*Trunk Vertebral Number* and *Adult Trunk Form*) of bolitoglossines (minus *Oedipina*) compared to other direct developers. Univariate and bivariate rates of the best-fit model for each trait and group are listed below. Note: evolvcv.lite will seemingly not execute if any variable is identical across all taxa. All of the bolitoglossines (minus *Oedipina*) that we included have 14 trunk vertebrae, so we arbitrarily set one taxon (*Aquiloerycea cephalica*) to 14.001 trunk vertebrae for this analysis only.

Model	-lnL	AIC	Δ AIC	w_i
no common structure (between groups)	-33.09	82.18	0.00	0.9999
different rates, common correlation	-47.99	109.99	27.81	< 0.0001
common rates, common correlation	-218.36	446.73	364.55	< 0.0001
common rates, different correlation	-217.77	447.56	365.38	< 0.0001
	bolitoglossines	other dds		
σ^2 trunk vertebral number:	1.00e-10±4.59e-18	0.187±0.0703		
σ^2 adult trunk form:	0.059±0.0071	0.066±0.0180		
σ^2 bivariate:	4.68e-9±4.59e-9	0.093±0.0355		

Table S7. Evolution of *Trunk Vertebral Number* for direct developers. As expected, multi-rate models were substantially better than single rate models. The rate of trunk vertebral evolution in bolitoglossines (minus *Oedipina*) was dramatically slower than all other direct developers.

Model	-lnL	AIC	Δ AIC	w_i
$OU_{\theta\sigma^2}$	228.30	-446.60	0.00	1.0000
$BM_{\theta\sigma^2}$	133.08	-258.17	188.43	< 0.0001
BM_{σ^2}	133.72	-257.43	189.17	< 0.0001
OU_{θ}	-112.72	233.44	680.04	< 0.0001
BM1	-119.00	242.00	688.60	< 0.0001
OU1	-119.00	244.00	690.60	< 0.0001

θ : bolitoglossines = 14.02±0.01; other dds = 17.51±0.03
 σ^2 : bolitoglossines = 0.0003±0.0001; other dds = 49.45±0.60
 α : 4.834±0.0586

Table S8. Evolution of *Adult Trunk Form* for direct developers. Allowing for differences in rate among the groups does not substantially improve BM or OU models. In other words, the rate of adult trunk form evolution in bolitoglossines (minus *Oedipina*) is similar to other direct developers. Albeit not substantial, the best-fit models allowed for differences in optima between the groups.

Model	-lnL	AIC	Δ AIC	w_i
$BM_{\theta\sigma^2}$	-109.94	227.88	0.00	0.4484
OU_{θ}	-110.49	228.98	1.10	0.2587
$OU_{\theta\sigma^2}$	-109.93	229.87	1.99	0.1658
BM1	-114.01	232.03	4.15	0.0563
BM_{σ^2}	-113.16	232.32	4.44	0.0487
OU1	-113.95	233.90	6.02	0.0221

θ : bolitoglossines = 7.21±0.04; other dds = 10.41±0.02
 σ^2 : bolitoglossines = 0.065±0.0005; other dds = 0.041±0.0002

Supplemental Table S9. Regimes, body form metrics, and Genbank accession numbers. Alternative life cycles include: biphasic (bi), direct development (dd), and paedomorphic (pd). Only species that are obligately paedomorphic in nature were coded as paedomorphic [S28]. Facultative paedomorphs (f-bi) were coded as biphasic unless otherwise indicated (see Facultative models in the Results section). Larval ecologies were coded based on the primary habitat utilized: lotic (lo) or lentic (le). Direct developing and viviparous species do not have a free-living larval stage, so their larval ecology is listed as na. Two closely related salamandrid genera (*Lyciasalamandra* and *Salamandra*) exhibit viviparity (vi) or ovoviviparity (ovi) [S29]. These taxa may also represent simplified life cycle modes (depending on the duration of offspring retention and developmental progression), but we pruned these taxa from our analyses due to the limited phylogenetic distribution of these reproductive modes in salamanders and their lability within some taxa. Furthermore, many “newts” in the Family Salamandridae have a three-part life cycle (aquatic larva - terrestrial eft - aquatic adult). All non-viviparous and non-ovoviviparous salamandrids were coded as biphasic. This is because all of these species minimally have a larval stage and post-metamorphic stage (the latter being potentially facultative). We acknowledge that there are potentially some species that metamorphose but remain aquatic (e.g. some, but not all, species in the genus *Pachytriton* [S30]). Terrestrial juveniles are known in *Pachytriton* (e.g. *P. feii*) and their terrestrial stage may be extensive [S30-S32]. There is generally limited life history information on wild populations for most species of *Pachytriton*, especially for juvenile stages [S30, S31]. We found that these taxa had no impact on our results (when omitted or re-coded). However, fine-scale trait analyses of such groups may prove enlightening for some characteristics [S33]. *Trunk Vertebral Number* was based on modal numbers from the literature or data collected by the authors. In species with sexually dimorphic trunk vertebral numbers, an average of male and female mode numbers was used. *Adult Body Form* was snout-to-vent length divided by body width (data from Wiens and Hoverman [S34]). *Adult Trunk Form*, which was used to analyze direct developers was snout-to-vent length minus head length divided by body width, data also from Wiens and Hoverman [S34]. Taxa included in larval shape analyses are indicated with an asterisk. Genes used for the phylogeny include three mitochondrial genes: Cytochrome oxidase 1 (*Co1*), Cytochrome b (*Cytb*), NADH dehydrogenase 2 (*ND2*), and four nuclear genes: Brain Derived Neurotrophic Factor (*BDNF*), Proopiomelanocortin (*Pomc*), Recombination Activating Gene 1 (*RAG1*), Solute carrier family 8 member A3 (*Slc8a3*).

Species	Life Cycle	Larval Eco	Trunk Vert	Vert Refs	Adult Body Form	Adult Trunk Form	Co1 Accession	Cytb Accession	ND2 Accession	BDNF Accession	Pomc Accession	RAG1 Accession	Slc8a3 Accession
Ambystomatidae													
<i>Ambystoma andersoni</i>	pd	lo	15	S35			AY659993	AY659993	AY659993				
<i>Ambystoma annulatum*</i>	bi	le	16	S36	8.98		KU986254		KC870849				
<i>Ambystoma barbouri</i>	bi	lo			9.25		NC014568	NC014568	NC014568	KJ610094	KJ610471		
<i>Ambystoma bishopi</i>	bi	le			9.14		NC027501	NC027501	NC027501				
<i>Ambystoma californiense</i>	bi	le			6.51		AY659995	AY659995	AY659995				
<i>Ambystoma cingulatum</i>	bi	le							KC870856				
<i>Ambystoma dumerilii</i>	pd	le	15	S37			AY659994	AY659994	AY659994				
<i>Ambystoma gracile</i>	f-bi	le			6.54		AY691729		KC870857			AY650131	
<i>Ambystoma jeffersonianum</i>	bi	le	15	S38,S39	9.36		EF036687		KC870859	KJ610128	KJ610490		
<i>Ambystoma laterale</i>	bi	le			8.58		AY728218	AY728218	AY728218				
<i>Ambystoma mabeei</i>	bi	le			8.42		KU985600	EF036632	KC870863				
<i>Ambystoma macrodactylum</i>	bi	le	15	S38,S39	8.73			EF036634	KC870864				
<i>Ambystoma maculatum*</i>	bi	le	14	S38,S39	7.80			EF036637	KC870867				
<i>Ambystoma mexicanum*</i>	pd	le	15	S37			AY659991	AY659991	AY659991	EF195175		EF551561	EF107367
<i>Ambystoma opacum</i>	bi	le	14	S38,S39	7.02		KU986081	AY691730	KC870871			AY650130	
<i>Ambystoma ordinarium</i>	f-bi	lo			5.32				KC870872			AY583345	
<i>Ambystoma rosaceum</i>	f-bi	lo								EU275887	EU275841		
<i>Ambystoma talpoideum</i>	f-bi	le	13	S39	5.93		KU985959	EF036640	KC870874				
<i>Ambystoma texanum*</i>	bi	le	16	S39	8.04		GU078471	GU078471	GU078471	KJ610104	KJ610445		
<i>Ambystoma tigrinum</i>	f-bi	le	15	S38,S39	5.88		AY659992	AY659992	AY659992				
Amphiumidae													
<i>Amphiuma means</i>	pd	le	61	S36,S37	17.43		FJ951301	AY691722	AY916037		FJ951366	AY650127	JX145021
<i>Amphiuma pholeter</i>	pd	le	60	S36	25.25		FJ951302	AY691723	AY916035		FJ951367	AY650128	JX145022
<i>Amphiuma tridactylum*</i>	pd	le	61	S36	17.03		FJ951300	FJ951359	AY916036	EU275863	FJ951368	FJ951369	JX145023
Cryptobranchidae													
<i>Andrias davidianus</i>	pd	lo	17	S36	6.85		NC004926	NC004926	NC004926	EU275889	EU275843	AY650142	AY948911
<i>Andrias japonicus</i>	pd	lo					AB208679	AB208679	AB208679			AY583346	
<i>Cryptobranchus alleganiensis*</i>	pd	lo	18	S36,S37	6.18		GQ368662	GQ368662	GQ368662			AY650141	
Dicamptodontidae													
<i>Dicamptodon copei*</i>	pd	lo	15	S36	7.30			AY734609			KC295575	AY691695	JX145020
<i>Dicamptodon ensatus</i>	f-bi	lo	14	S36	6.36		KU985936	AY734622				EF107335	
<i>Dicamptodon tenebrosus*</i>	f-bi	lo	14	S36	6.15			AY734613	AY916018	EU275870	EU275824	AY650132	
Hynobiidae													
<i>Batrachuperus karlschmidti</i>	bi	lo					KF748919	KF748917	KF748919			KJ715360	
<i>Batrachuperus longdongensis</i>	bi	lo					DQ333809	DQ333809	DQ333809	HM037737	HM037762	HM037712	
<i>Batrachuperus pinchonii</i>	bi	lo	17	S40	6.87		DQ333815	DQ333815	DQ333815	HM037738	HM037763	HM037713	EF107362
<i>Batrachuperus tibetanus</i>	bi	lo					DQ333817	DQ333817	DQ333817	HM037739	HM037764	HM037714	
<i>Batrachuperus yenyuanensis</i>	bi	lo			7.05		DQ333818	DQ333818	DQ333818	HM037740	HM037765	HQ902535	
<i>Hynobius abei</i>	bi	le			7.16			AY915939					
<i>Hynobius amjiensis</i>	bi	le					DQ333808	DQ333808	DQ333808	HM037741	HM037766	HM037716	
<i>Hynobius baulengeri</i>	bi	lo						AY915936					
<i>Hynobius chinensis</i>	bi	le					DQ333819	DQ333819	DQ333819	HM037742	HM037767	HM037717	
<i>Hynobius dunni</i>	bi	le			7.88			LC003308	AY915926				
<i>Hynobius formosanus</i>	bi	lo					DQ333816	DQ333816	DQ333816			DQ347285	EF107400
<i>Hynobius guabangshanensis</i>	bi	le					GU384690	GU384690	GU384690	HM037743	HM037768	HM037718	
<i>Hynobius hidamontanus</i>	bi	le					JQ929919	JQ929919	JQ929919				
<i>Hynobius kimurae</i>	bi	lo					JQ929920	JQ929920	JQ929920			KJ715369	
<i>Hynobius leechii*</i>	bi	le	16	S40	7.30		DQ333811	DQ333811	DQ333811	HM037744	HM037769	HM037719	
<i>Hynobius lichenatus</i>	bi	le			7.64		JQ929921	JQ929921	JQ929921				
<i>Hynobius maershanensis</i>	bi	le					NC023789	NC023789	NC023789	HM037745	HM037770	HM037720	
<i>Hynobius naevius</i>	bi	lo	17	S40	7.48			AB266672	AY915937				
<i>Hynobius nebulosus</i>	bi	le	16	S40	7.96		HM036356	HM036356	HM036356	HM037746	HM037771	AY650144	
<i>Hynobius nigrescens</i>	bi	le	15	S40	7.39		JQ929922	JQ929922	JQ929922			KJ715356	
<i>Hynobius quoelpaertensis</i>	bi	lo					EF201847	EF201847	EF201847	HM037747	HM037772	HM037722	
<i>Hynobius retardatus</i>	f-bi	le					HM036351	HM036351	HM036351	HM037748	HM037773	HM037723	
<i>Hynobius sonani</i>	bi	lo						AY915945	EU275864	EU275818			
<i>Hynobius stejnegeri</i>	bi	lo			8.87			AB921162	AY915938				
<i>Hynobius takedai</i>	bi	le						AY915942					
<i>Hynobius tokyoensis</i>	bi	le					HM036357	HM036357	HM036357	HM037749	HM037774	HM037724	
<i>Hynobius tsuensis</i>	bi	le			7.61		JQ929923	JQ929923	JQ929923				
<i>Hynobius yangi</i>	bi	lo					JN415127	JN415127	JN415127				
<i>Hynobius yiwuensis</i>	bi	le					HM036354	HM036354	HM036354	HM037750	HM037775	HM037725	
<i>Liaa shihi</i>	bi	lo	16.5	S40,S41	7.13		DQ333810	DQ333810	DQ333810	HM037751	HM037776	HM037726	
<i>Liaa tsinpaensis</i>	bi	lo					KP233806	KP233806	KP233806	HM037752	HM037777	HM037727	
<i>Onychodactylus fischeri*</i>	bi	lo	20	S40			DQ333820	DQ333820	DQ333820	HM037753	HM037778	KC165594	
<i>Onychodactylus japonicus</i>	bi	lo	18	S40	9.21		JX158189	AB452955	AY915923			AY650143	
<i>Onychodactylus zhangyapingi</i>	bi	lo					NC026853	NC026853	NC026853			KJ715373	

Species	Life Cycle	Larval Eco	Trunk Vert	Vert Refs	Adult Body Form	Adult Trunk Form	CoI Accession	Cytb Accession	ND2 Accession	BDNF Accession	Pomc Accession	RAG1 Accession	Sic8a3 Accession
<i>Pachyhynobius shangchengensis</i>	bi	lo			7.46		DQ333812	DQ333812	DQ333812	HM037754	HM037779	HM037729	
<i>Paradactylodon gorganensis</i>	bi	lo					DQ333822	DQ333822	DQ333822	HM037755	HM037780	HM037730	
<i>Paradactylodon mustersi</i>	bi	lo	17	540	6.87		DQ333821	DQ333821	DQ333821	HM037756	HM037781	HM037731	
<i>Pseudohynobius flavamaculatus</i>	bi	lo					NC020635	NC020635	NC020635	HM037758	HM037783	HM037733	
<i>Pseudohynobius jinfa</i>	bi	le					NC026698	NC026698	NC026698				
<i>Pseudohynobius puxiongensis</i>	bi	lo	17	542			NC020634	NC020634	NC020634	HM037757	HM037782	HM037732	
<i>Pseudohynobius shuichengensis</i>	bi	le					NC021001	NC021001	NC021001	HM037759	HM037784	HM037734	
<i>Ranodon sibiricus</i>	bi	lo	15	540	7.40		NC004021	NC004021	NC004021	HM037760	HM037785	HM037735	
<i>Salamandrella keysringii</i>	bi	le	17	540	6.92		DQ333814	DQ333814	DQ333814	HM037761	HM037786	AY650145	
<i>Salamandrella tridactyla</i>	bi	le					NC021106	NC021106	NC_021106	KJ855094		KJ855096	
Plethodontidae													
<i>Aneides aeneus</i>	dd	na	16	536	9.95	7.8		AY691742		EU275890	EU275844	AY691701	
<i>Aneides ferreus</i>	dd	na	17	536			DQ105339	KF781793	DQ105339	EU275891	EU275845	EU275805	
<i>Aneides flavipunctatus</i>	dd	na	16	536	9.89	7.9	AY728214	AY728214	AY728214	EU275894	EU275848	EU275809	
<i>Aneides hardii</i>	dd	na	16	536	9.89	8	AY728226	AY728226	AY728226	EU275857	EU275811	EU275790	
<i>Aneides lugubris</i>	dd	na	16	536	7.41	5.8	DQ105329	AY691758		EU275893	EU275847	AY650118	
<i>Aneides vagrans</i>	dd	na			10.51		DQ105343	KF781800		EU275892	EU275846	EU275806	
<i>Aquiloerycea cephalica</i>	dd	na	14	523	9.68	7.7	KP886919	KP900066			KP900108	KP900152	KP900198
<i>Aquiloerycea galeanae</i>	dd	na			8.88		KP886904	KP900051			KP900093	KP900137	KP900184
<i>Aquiloerycea quetzalensis</i>	dd	na					KP900055	KP900055			KP900097	KP900141	KP900188
<i>Batrachoseps altiserae</i>	dd	na	18.5	543			KM203060				KM202828	KM203006	
<i>Batrachoseps attenuatus</i>	dd	na			13.77		EU117188		EU117188	JF449372	KM202699	KM202873	KP900217
<i>Batrachoseps bramei</i>	dd	na	18.5	543				JQ035768			KM202847	KM203025	
<i>Batrachoseps campi</i>	dd	na	18	544			EU117193	KM203094	EU117193		KM202860	KM203039	
<i>Batrachoseps diabolicus</i>	dd	na	20	S44,S45			EU117190	KM203054	EU117190		KM202783	KM202961	
<i>Batrachoseps gabrieli</i>	dd	na	20	S44,S46			EU117195	KM203056	EU117195		KM202763	KM202941	
<i>Batrachoseps gowlandensis</i>	dd	na	20	S44,S47			EU117191	KM203065	EU117191		KM202725	KM202901	
<i>Batrachoseps gregarius</i>	dd	na	21	S44,S45			EU117192	KM203079	EU117192		KM202790	KM202968	
<i>Batrachoseps incognitus</i>	dd	na						KM203058			KM202737	KM202913	
<i>Batrachoseps kawia</i>	dd	na	20	S44,S45				KM203066			KM202822	KM203000	
<i>Batrachoseps luciae</i>	dd	na						KM203053			KM202713	KM202889	
<i>Batrachoseps major</i>	dd	na	20.5	544	14.72	12.7	EU117194	JQ250228	EU117194	EU275901	KM202771	KM202949	
<i>Batrachoseps minor</i>	dd	na						JQ250327			KM202738	KM202914	
<i>Batrachoseps nigriventris</i>	dd	na	21	544			NC028184	NC028184	NC028184		KM202745	KM202927	
<i>Batrachoseps pacificus</i>	dd	na						JQ250330			KM202758	KM202935	
<i>Batrachoseps pacificus</i>	dd	na	20	544				JQ250330			KM202758	KM202935	
<i>Batrachoseps regius</i>	dd	na	19.5	S44,S45				KM203071			KM202811	KM202989	
<i>Batrachoseps relictus</i>	dd	na	18.5	544				KM203093			KM202852	KM203030	
<i>Batrachoseps robustus</i>	dd	na						KM203064			KM202862	KM203040	
<i>Batrachoseps simatus</i>	dd	na	20.5	543				JQ035754			KM202851	KM203029	
<i>Batrachoseps wrighti</i>	dd	na	17	544				KM203062		JF449369	KM202865	KM203043	
<i>Batrachoseps wrightorum</i>	dd	na					AY728221	AY728221	AY728221			EU020165	
<i>Bolitoglossa adspersa</i>	dd	na	14	S44,S45	9.82	8.1		AF212984					
<i>Bolitoglossa alberchi</i>	dd	na					KP886900	KP735278		KP735258	KP735288	KP735306	KP735323
<i>Bolitoglossa alvaradoi</i>	dd	na						AY526194					
<i>Bolitoglossa aureogularis</i>	dd	na						JQ899182					
<i>Bolitoglossa biseriata</i>	dd	na					KM527317	AY526161				KC614436	
<i>Bolitoglossa borburata</i>	dd	na											
<i>Bolitoglossa bramei</i>	dd	na						JQ899189					
<i>Bolitoglossa carri</i>	dd	na						AY526176				KC614458	
<i>Bolitoglossa cataguana</i>	dd	na						KJ628090					
<i>Bolitoglossa celaque</i>	dd	na						AY526177					
<i>Bolitoglossa cerroensis</i>	dd	na	14	523				AF212096				KC614459	
<i>Bolitoglossa chinanteca</i>	dd	na						KC288079			KC288041		
<i>Bolitoglossa chucantiensis</i>	dd	na					KM527308						
<i>Bolitoglossa colonnea</i>	dd	na	14	523			FJ766578	AY526162					
<i>Bolitoglossa compacta</i>	dd	na						JQ899193					
<i>Bolitoglossa cananti</i>	dd	na						GU725458				KC699924	
<i>Bolitoglossa cuchumatana</i>	dd	na						GU725467					
<i>Bolitoglossa cuna</i>	dd	na					KM527307						
<i>Bolitoglossa decora</i>	dd	na						AY526180					
<i>Bolitoglossa diaphora</i>	dd	na						GU725460					
<i>Bolitoglossa dofleini</i>	dd	na	14	536	6.67	5.5		KP900047		KP900089	KP900133	KP900180	
<i>Bolitoglossa dunni</i>	dd	na	14	523				GU725459				KC614438	
<i>Bolitoglossa engelhardti</i>	dd	na	14	523	10.89	8.6		GU725461				KC699925	
<i>Bolitoglossa epimela</i>	dd	na						AF212097					
<i>Bolitoglossa equitoriana</i>	dd	na						DQ353842				KC614451	
<i>Bolitoglossa eremia</i>	dd	na						HQ009998					
<i>Bolitoglossa flavimembris</i>	dd	na						GU725462		KP900087	KP900132	KP900178	
<i>Bolitoglossa flaviventris</i>	dd	na	14	523				AF212983					
<i>Bolitoglossa franklini</i>	dd	na						AY526184				KC614439	
<i>Bolitoglossa gomezi</i>	dd	na						JQ899171					
<i>Bolitoglossa gracilis</i>	dd	na						AF212067					
<i>Bolitoglossa hartwegi</i>	dd	na					KP886897	KC288103		KC288057	KP900131		
<i>Bolitoglossa heirorelas</i>	dd	na						HQ010010					
<i>Bolitoglossa helmrichi</i>	dd	na	14	523				AY691755				AY650124	
<i>Bolitoglossa hermosa</i>	dd	na						AF416678					
<i>Bolitoglossa kamuk</i>	dd	na						JQ899175					
<i>Bolitoglossa kaqchikelorum</i>	dd	na						HQ010020					
<i>Bolitoglossa lincolni</i>	dd	na						GU725464				KC614440	
<i>Bolitoglossa longissima</i>	dd	na						AY526186				KC614441	
<i>Bolitoglossa macrinii</i>	dd	na						AF416679					
<i>Bolitoglossa marmorea</i>	dd	na	14	523				U89627					
<i>Bolitoglossa medemi</i>	dd	na					KM527309	AY526163				KC614437	
<i>Bolitoglossa meliana</i>	dd	na						KJ175105					
<i>Bolitoglossa mexicana</i>	dd	na	14	523	10.49	8.6		KC288104		KC288058	EF018055	EF107384	
<i>Bolitoglossa minutula</i>	dd	na						AF212098				KC614434	
<i>Bolitoglossa mambachoensis</i>	dd	na						AY133485					
<i>Bolitoglossa mario</i>	dd	na	14	S23,S36	8.13	6.6		KJ787752				KC699926	
<i>Bolitoglossa mucuyensis</i>	dd	na						JQ665278					
<i>Bolitoglossa mulleri</i>	dd	na			9.52			HQ010012					
<i>Bolitoglossa nigrescens</i>	dd	na	14	523				JQ899194					
<i>Bolitoglossa nymphea</i>	dd	na					KP886896	KC288068		KC288021	KP900130	KP900176	
<i>Bolitoglossa oaxacensis</i>	dd	na						AF416681		KP900088	KP900137	KP900179	
<i>Bolitoglossa occidentalis</i>	dd	na	14	523	10.40	8.1		AY526158		KC288030	KC614435		
<i>Bolitoglossa odonnelli</i>	dd	na						HQ009993				KC699922	
<i>Bolitoglossa orestes</i>	dd	na	14	523				JQ665281					
<i>Bolitoglossa pacaya</i>	dd	na						KJ787751					
<i>Bolitoglossa palmata</i>	dd	na			9.38			AY526164					
<i>Bolitoglossa paraensis</i>	dd	na						AY526166					
<i>Bolitoglossa peruviana</i>	dd	na						DQ353815				KC614443	

Species	Life Cycle	Larval Eco	Trunk Vert	Vert Refs	Adult Body Form	Adult Trunk Form	Co1 Accession	Cytb Accession	ND2 Accession	BDNF Accession	Pomc Accession	RAG1 Accession	Sic8a3 Accession
<i>Bolitoglossa pesrubra</i>	dd	na						AF212070					
<i>Bolitoglossa platyductyla</i>	dd	na	14	S23	8.93	7.4		AY133484				KC699923	
<i>Bolitoglossa porrasorum</i>	dd	na						AY526188					
<i>Bolitoglossa rietti</i>	dd	na						AF416682					
<i>Bolitoglossa robinsoni</i>	dd	na						JQ899191					
<i>Bolitoglossa robusta</i>	dd	na	14	S23	8.04	6.7		EU448110					
<i>Bolitoglossa rostrata</i>	dd	na	14	S23,S36	10.52	8.6		KJ175107					
<i>Bolitoglossa rufescens</i>	dd	na	14	S23	8.77	7.1	KR736078	KC288065			KC288019	KF911887	
<i>Bolitoglossa schizodactyla</i>	dd	na	14	S23			FJ766579	AY526171					
<i>Bolitoglossa sombra</i>	dd	na								EU275897	EU275851		KP900181
<i>Bolitoglossa splendida</i>	dd	na						JQ899181					
<i>Bolitoglossa subpalmata</i>	dd	na	14	S23,S36	8.80	7.3		AF212092					
<i>Bolitoglossa suchitanensis</i>	dd	na						HQ010001					
<i>Bolitoglossa synoria</i>	dd	na						AY526193					
<i>Bolitoglossa tica</i>	dd	na						JQ899192					
<i>Bolitoglossa yucatanana</i>	dd	na			9.98			AF212980					
<i>Bolitoglossa zapoteca</i>	dd	na						AF416683					
<i>Bradytriton silus</i>	dd	na	14	S48	7.75	6.3	KP886934	KP337342		KP735274	KP735303	KP735308	KP735326
<i>Chiroptertriton arboreus</i>	dd	na	14	S23				KP900083			KP900124	KP900170	KP900212
<i>Chiroptertriton chondrostega</i>	dd	na			10.93			KT820699					
<i>Chiroptertriton cracens</i>	dd	na						KT820700					
<i>Chiroptertriton dimidiatus</i>	dd	na	14	S23	11.95	9.5		KT820701					
<i>Chiroptertriton lavae</i>	dd	na			11.54			KT820702					
<i>Chiroptertriton magnipes</i>	dd	na						KP900085			KP900126	KP900172	KP900214
<i>Chiroptertriton miquihuanus</i>	dd	na						KT820713					
<i>Chiroptertriton mosaueri</i>	dd	na						KT820703					
<i>Chiroptertriton multidentatus</i>	dd	na	14	S23,S36	12.33	9.6		KT820704					
<i>Chiroptertriton priscus</i>	dd	na	14	S23	9.39	7.6		KT820707					
<i>Cryptotriton alvarezdeltoroi</i>	dd	na						KP735279		KP735269	KP735293	KP735318	KP735332
<i>Cryptotriton manzoni</i>	dd	na						KJ547609		KP735262	KP735290	KP735314	KP735328
<i>Cryptotriton nasalis</i>	dd	na	14	S24	10.96	8.7	KP886935	KJ563294		KP735264	KP735291	KP735310	KP735335
<i>Cryptotriton necopinus</i>	dd	na						KJ547610		KP735268	KP735296	KP735309	
<i>Cryptotriton sierraminensis</i>	dd	na						KJ547605		KP735267	KP735295	KP735312	KP735336
<i>Cryptotriton varaeopacis</i>	dd	na						KJ547606		KP735266	KP735297	KP735317	KP735329
<i>Dendrotriton bromeliocius</i>	dd	na	14	S48			KP886939	JN559990		KP735259	JN560007	JN560024	JN560041
<i>Dendrotriton chujorum</i>	dd	na						JN559993			JN560010	JN560026	JN560044
<i>Dendrotriton cuchumatanus</i>	dd	na	14	S48				JN559995			JN560012	JN560028	JN560046
<i>Dendrotriton kekchiourum</i>	dd	na						JN559996			JN560013	JN560030	JN560047
<i>Dendrotriton megarhinus</i>	dd	na	14	S48				JN559998			JN560015	JN560031	JN560049
<i>Dendrotriton rabbi</i>	dd	na	14	S48				AF199194			JN560016	JN560033	JN560050
<i>Dendrotriton sanctibarbarus</i>	dd	na					KP886940	JN560000			JN560017	JN560034	JN560051
<i>Dendrotriton xoloccalcae</i>	dd	na	14	S23	12.92	10.1		JN560004			JN560021	JN560037	JN560055
<i>Desmognathus abditus</i>	bi	lo							KR732330		KR732341	KR732365	
<i>Desmognathus aeneus</i>	dd	na	15	S23	10.07	8	KU985866	AY691736	AY612342		KR732342	AY691696	
<i>Desmognathus apalachicola</i>	bi	lo						EU311666	AY612373		KR732343	KR732367	
<i>Desmognathus auriculatus</i>	bi	le	15	S23	9.35			EU311650	AY612414		KR732344	KR732368	
<i>Desmognathus brimleyorum*</i>	bi	lo	15	S36	8.34			KU986036	AY691737		AY612422	EU275819	AY691697
<i>Desmognathus carolinensis</i>	bi	lo	15	S36				EU311642	AF442540		AY612368	KR732345	KR732369
<i>Desmognathus conanti</i>	bi	lo						EU311667	KF242415		AY612383	KR732346	KR732370
<i>Desmognathus folkertsi</i>	bi	lo						EU311714	AY612351		AY612351	KM107913	
<i>Desmognathus fuscus</i>	bi	lo	15	S23	8.03			AY728227	AY728227		AY728227	EU275812	KR827007
<i>Desmognathus imitator</i>	bi	lo			9.60			AF437504	AY612343		AY612343	KR732349	KR732373
<i>Desmognathus marmoratus</i>	bi	lo	15	S23,S36	7.73			EU311718	AY612344		AY612344	KR732350	KR732374
<i>Desmognathus manticola</i>	bi	lo	15	S23,S36	7.70				AY691738		AY612374	KR732352	AY691698
<i>Desmognathus ochrophaeus</i>	bi	lo	15	S23	8.91			KU986074	AF442525		AY612366	KR732353	KR732377
<i>Desmognathus ocoee</i>	bi	lo	15	S23,S36					AF442541		AY612352	KR732354	KR827010
<i>Desmognathus orestes</i>	bi	lo						EU311703	AF442536		AY612363	KR732356	KR827015
<i>Desmognathus organi</i>	dd	na							KF242417		KR827001	KR732357	KR732381
<i>Desmognathus planiceps</i>	bi	lo							EF028646		KR732337	KR732358	KR732382
<i>Desmognathus quadramaculatus*</i>	bi	lo	15	S23,S36	7.08			EU311700	AY691739		AY612347	KR732359	AY650117
<i>Desmognathus santeeah</i>	bi	lo						EU311676	KF242410		AY612391	KR732360	KR732384
<i>Desmognathus weltersi</i>	bi	lo			8.47			EU311675	AY612416		AY612416	KR732362	KR732386
<i>Desmognathus wrightii</i>	dd	na	15	S23	10.71	8.5	KU986239	AY691740	KR732339		KR732363	KR732363	AY691699
<i>Ensatina eschscholtzii</i>	dd	na	14	S36	9.17	7.4	AY728216	AY728216			AY728216	EU275862	AY650119
<i>Eurycea aquatica</i>	bi	lo							KF562543		FJ785624		KF562645
<i>Eurycea bislineata*</i>	bi	lo	15	S36	11.13		AY728217	AY528402			JQ920804	EU275861	AY691706
<i>Eurycea chamberlaini</i>	bi	le	17	S36					KF562544		JQ920881	JQ920752	KF562646
<i>Eurycea chisholmensis</i>	pd	lo	18	S36					KF562545				KF562647
<i>Eurycea cirrigera</i>	f-bi	lo	15	S36	11.04				KF562548		DQ018556	JQ920728	KF562650
<i>Eurycea guttolineata</i>	bi	lo	15	S36					KF562549		DQ018388	JQ920731	KF562651
<i>Eurycea junaluska</i>	bi	lo			10.05				KF562550		DQ018655		FJ750246
<i>Eurycea latitans</i>	pd	lo	18	S36	9.23				KF562551		JQ920812		KF562652
<i>Eurycea longicauda</i>	bi	lo	15	S36	10.84				AY528403		DQ018386	JQ920730	AY650121
<i>Eurycea longicauda melanopleura</i>	bi	lo	15	S36					KF562552		FJ866452		KF562653
<i>Eurycea lucifuga</i>	bi	lo	15	S36	9.86				KF562553		JQ920807	JQ920729	KF562654
<i>Eurycea multiplicata</i>	bi	lo	20	S36	12.49				AY528339		JQ920803	JQ920725	AY691707
<i>Eurycea nana</i>	pd	lo	17	S36	12.75				KF562554		JQ920814	JQ920735	KF562656
<i>Eurycea naufragia</i>	pd	lo	17	S36					KF562555		JQ920811	JQ920733	KF562657
<i>Eurycea neotenes*</i>	pd	lo	17	S36	10.87				AY528400		JQ920817		AY650122
<i>Eurycea paludicola*</i>	bi	le	17	S36	12.62				AY528401		JQ920823	JQ920740	KF562662
<i>Eurycea pterophila</i>	pd	lo	17	S36					KF562556		JQ920816		KF562658
<i>Eurycea quadridigitata</i>	bi	le	17	S36					KF562560		JQ920863	JQ920750	KF562658
<i>Eurycea rathbuni*</i>	pd	lo	13	S36	9.20				KF562561				AY691708
<i>Eurycea sosorum</i>	pd	lo	16	S36	11.29				KF562562		JQ920815		KF562664
<i>Eurycea spelaea</i>	bi	lo	19	S36	9.39				AY528397				KF562667
<i>Eurycea subfluvicola</i>	pd	lo	21	S36					KJ372372				KJ372331
<i>Eurycea tonkawae</i>	pd	lo	17	S36					KF562564		JQ920810	JQ920732	AY691709
<i>Eurycea tridentifera</i>	pd	le	14	S36					KF562565				KF562669
<i>Eurycea troglodytes Metamorphic</i>	f-bi	lo							KF562567				KF562671
<i>Eurycea troglodytes Paedomorphic</i>	f-bi	lo							KF562568		JQ920813	JQ920734	KF562672
<i>Eurycea tynerensis Metamorphic*</i>	f-bi	lo	20	S36					AY528367				KF562676
<i>Eurycea tynerensis Paedomorphic*</i>	pd	lo	21	S36	13.41				AY528374				KF562675
<i>Eurycea wallacei</i>	pd	le	14	S23,S36	16.97				KF562583				KF562693
<i>Eurycea waterlooensis</i>	pd	le	13	S36					KF562569				KF562679
<i>Eurycea wilderae</i>	bi	lo	15	S36	11.01				KF562570		DQ018650	JQ920727	KF562680
<i>Gyrinophilus gulolineatus</i>	pd	lo							KF562571				KF562681
<i>Gyrinophilus pallucus</i>	pd	lo	19	S50			NC028297		KF562574				KF562684
<i>Gyrinophilus porphyriticus*</i>	bi	lo	18	S36	10.26				KF562581		EU275899	EU275853	KF562691
<i>Gyrinophilus subterraneus</i>	f-bi	lo							KF562582				KF562692
<i>Hemidactylum scutatum</i>	bi	le	15	S36	9.62		AY728231	AY728231	AY728231		EU275898	EU275852	AY691711
<i>Hydromantes ambroii</i>	dd	na	13	S51					FJ602258		FJ602195		FJ602321
<i>Hydromantes brunus</i>	dd	na	14	S36			AY728234	AY728234	AY728234		EU275871	EU275825	AY887134

Species	Life Cycle	Larval Eco	Trunk Vert	Vert Refs	Adult Body Form	Adult Trunk Form	CoI Accession	Cytb Accession	ND2 Accession	BDWF Accession	Pomc Accession	RAG1 Accession	Sic8a3 Accession
<i>Hydromantes flavus</i>	dd	na	13	S51						FJ602199		FJ602327	
<i>Hydromantes genei</i>	dd	na			12.39					FJ602205	EU275840	FJ602333	
<i>Hydromantes imperialis</i>	dd	na	13	S51				FJ602288	KJ834057	FJ602219		FJ602359	
<i>Hydromantes italicus</i>	dd	na	13	S51	11.48	9.1	AY728215	AY728215	AY728215	EU275872	EU275826	EU275792	EF107415
<i>Hydromantes platycephalus</i>	dd	na			9.48		KU985943			EU275874	EU275828	EU275874	HM797672
<i>Hydromantes shostae</i>	dd	na						HSU89610		EU275875	EU275829	HM797671	HM797759
<i>Hydromantes strinatii</i>	dd	na	13	S51				FJ602305	KJ834039	FJ602245		FJ602367	
<i>Hydromantes supramontis</i>	dd	na	13	S51					KJ834058	FJ602248		FJ602370	
<i>Isthmura bellii</i>	dd	na	14	S23	8.77	7.1		AY854681					
<i>Ialotriton parvus</i>	dd	na									KP900117		KP900205
<i>Ialotriton niger</i>	dd	na									KP900118		KP900206
<i>Karsenia koreana</i>	dd	na	16	S52				JF449367	JF449367	EU275868	EU275822	AY887135	
<i>Nototriton abscondens</i>	dd	na					AY728229	AY728229		JF449371	JF449371	JF449379	
<i>Nototriton barboursi</i>	dd	na	14	S48			JN377401	AF199138					
<i>Nototriton bradei</i>	dd	na					KP886938	KP735285		KP735276	KP735305	KP735322	KP735325
<i>Nototriton limnospectator</i>	dd	na					KP886937	JQ899197			KP735304	KP735321	KP735324
<i>Nototriton picadoi</i>	dd	na	14	S48	6.89	5.5		AF199145					
<i>Nototriton picucha</i>	dd	na					JN377404			AF199147			
<i>Nototriton richardi</i>	dd	na	14	S48									
<i>Nototriton saslaya</i>	dd	na					JN377406						
<i>Nyctanolis pernix</i>	dd	na			11.40								
<i>Oedipina carablanca</i>	dd	na	18	S53				FJ196869		KP735277	JN560023	AY691714	JN560057
<i>Oedipina complex</i>	dd	na	18	S53	15.70	13.1		AF199157					
<i>Oedipina cyclocauda</i>	dd	na			14.94			AF199158					
<i>Oedipina elongata</i>	dd	na	18	S53				AF199160					
<i>Oedipina gephyra</i>	dd	na						AF199161					
<i>Oedipina gracilis</i>	dd	na						AF199163					
<i>Oedipina grandis</i>	dd	na						AF199165					
<i>Oedipina maritima</i>	dd	na	18	S24				AF199166					
<i>Oedipina nica</i>	dd	na	18	S53	13.95	11.7	KP886941	JN560005		KP735261	JN560022	JN560039	JN560056
<i>Oedipina parvipis</i>	dd	na	18	S53			FJ766760	AF199155					
<i>Oedipina poelzi</i>	dd	na					NC006326	NC_006326	NC006326				
<i>Oedipina pseudouniformis</i>	dd	na						AF199178					
<i>Oedipina savagei</i>	dd	na	18	S24				AF199153					
<i>Oedipina taylori</i>	dd	na	22.5	S36,S53				KP900080			KP900121	KP900167	KP900209
<i>Oedipina tomasi</i>	dd	na					KP886942	KP900079			KP900120	KP900166	KP900208
<i>Oedipina uniformis</i>	dd	na	20.5	S53	15.09	13		AF199188					
<i>Parvimolge townsendi</i>	dd	na	14	S23	9.09	7.1	KP886932	KP900078	AY916024		KP900119	KP900164	KP900207
<i>Phaeognathus hubrichti</i>	dd	na	22	S23,S54	12.13	10.5		AY728233	AY728233		EU275814	AY691700	
<i>Plethodon albagula</i>	dd	na			9.15		KU986279	JF504320	AY874996			DQ995008	
<i>Plethodon amplus</i>	dd	na						DQ994912	AY874880			DQ995010	
<i>Plethodon angusticlavius</i>	dd	na	19	S55	11.72	9.8		DQ994913	DQ018677			DQ995011	
<i>Plethodon asupak</i>	dd	na						AY688299				KF911898	
<i>Plethodon aureolus</i>	dd	na						DQ994914	AY874997			DQ995012	
<i>Plethodon caddoensis</i>	dd	na	17	S55	10.56	8.3	KU985859	FJ266743	AY874875			DQ995013	
<i>Plethodon chattahoochee</i>	dd	na						DQ994919	AY874999			DQ995014	
<i>Plethodon cheach</i>	dd	na							AY875045			DQ995015	
<i>Plethodon chlorobryonis</i>	dd	na			8.30			DQ994923	AY875010				EF107404
<i>Plethodon cinereus</i>	dd	na	20	S55	12.68	10.7	AY728232	AY728232	AY728232	JF449368	FJ951365	AY691703	KM884463
<i>Plethodon cylindraceus</i>	dd	na			9.74			DQ994928	AY875011			DQ995022	
<i>Plethodon dorsalis</i>	dd	na	19	S55	12.34	10.3		GQ464404	DQ018679			DQ995023	
<i>Plethodon dunni</i>	dd	na	16	S36	10.80	9.0		AY183763	DQ018662			KF911900	
<i>Plethodon electromorphus</i>	dd	na						AY378065	DQ018665			DQ995024	
<i>Plethodon elongatus</i>	dd	na	19	S36,S55	11.52	9.4		AY728223	AY728223	EU275882	EU275836	AY650120	
<i>Plethodon fourchensis</i>	dd	na					KU986243	FJ611346	AY874876	EU275884	EU275838	DQ995026	
<i>Plethodon glutinosus</i>	dd	na	17	S55	9.53	7.9	KU986027	DQ994937	AY875024			DQ995027	
<i>Plethodon grabmani</i>	dd	na			9.69		KU986107	DQ994941	AY875026			DQ995028	
<i>Plethodon hoffmani</i>	dd	na			13.56		KU985922	AY378048	DQ018666	EU275883	EU275837	DQ995029	
<i>Plethodon hubrichti</i>	dd	na	20	S55	12.94	11.0	JF731302	AY378056	DQ018668			DQ995030	
<i>Plethodon idahoensis</i>	dd	na	15	S55	9.93	7.9		AY572059				GQ247792	
<i>Plethodon jordani</i>	dd	na	17	S55	9.74	8.1	KU986146	DQ994947	AY874897	EU275881	EU275835	DQ995032	
<i>Plethodon kentucki</i>	dd	na			10.64			DQ994948	AY875027			DQ995033	
<i>Plethodon kiamichi</i>	dd	na						FJ266739	DQ018679			DQ995034	
<i>Plethodon kisatchi</i>	dd	na						DQ994951	DQ018699			DQ995035	
<i>Plethodon larselli</i>	dd	na	16	S36,S55						EU275885	EU275839	DQ995036	
<i>Plethodon longicrus</i>	dd	na						DQ994952				DQ995037	
<i>Plethodon meridianus</i>	dd	na						DQ994954	AY874901			DQ995038	
<i>Plethodon metcalfi</i>	dd	na						DQ994956	AY874995			DQ995040	
<i>Plethodon mississippi</i>	dd	na			10.11			DQ994958	DQ018701				
<i>Plethodon montanus</i>	dd	na					KU985917	DQ994962	AY874994			DQ995041	
<i>Plethodon neomexicanus</i>	dd	na	20	S36,S55	12.95	10.9						DQ995044	
<i>Plethodon nettingi</i>	dd	na	19	S36,S55	13.17	11.0		AY378059	DQ018669			DQ995045	
<i>Plethodon ocmulgee</i>	dd	na						DQ994967	AY875028			DQ995048	
<i>Plethodon oconaluftee</i>	dd	na						DQ994966				DQ995047	
<i>Plethodon ouachitae</i>	dd	na	17	S55	10.51	8.4	KU986090	FJ266744	AY874877	EU275877	EU275831	AY691704	
<i>Plethodon petraeus</i>	dd	na					AY728222	AY728222	AY728222			DQ995049	
<i>Plethodon punctatus</i>	dd	na			10.70			AY378078	DQ018685			DQ995050	
<i>Plethodon richmondi</i>	dd	na	22	S55	14.10	12.2		AY378071	DQ018670			DQ995054	
<i>Plethodon savannah</i>	dd	na						DQ994978	AY875042			DQ995055	
<i>Plethodon sequoyah</i>	dd	na						DQ994979	DQ018705			DQ995056	
<i>Plethodon serratus</i>	dd	na			12.13			AY691748	DQ018673	EU275876	EU275830	AY691705	
<i>Plethodon shenandoah</i>	dd	na			11.96			AY378043	DQ018674			DQ995062	
<i>Plethodon shermani</i>	dd	na						DQ994987	AY875050			DQ995063	
<i>Plethodon stormi</i>	dd	na	18	S36				AY183825					
<i>Plethodon tephalee</i>	dd	na			11.44		KU985764	DQ994990	AY875030	EU275880	EU275834	DQ995068	
<i>Plethodon vandykei</i>	dd	na	15	S36,S55	10.67	8.9		AY691759		EU275879	EU275833	AY691715	
<i>Plethodon variolatus</i>	dd	na						AY691759	AY875043			DQ995069	
<i>Plethodon vehiculum</i>	dd	na	17	S36,S55	11.56	9.8		AY691760	DQ018661			AY691716	
<i>Plethodon ventralis</i>	dd	na			12.84			DQ994994	DQ018681			DQ995071	
<i>Plethodon virginia</i>	dd	na						AY378049	DQ018675			DQ995072	
<i>Plethodon websteri</i>	dd	na						AY378076	DQ018682			DQ995073	
<i>Plethodon wehrlei</i>	dd	na	18	S55	11.54	9.6		AY378079	DQ018687			DQ995075	
<i>Plethodon welleri</i>	dd	na	17	S55	11.30	9.3		AY691761	DQ018684			AY691717	
<i>Plethodon yanahlassee</i>	dd	na	17	S55	9.63	7.9	KU985784	AY691762	AY874879	EU275878	EU275832	AY691718	
<i>Pseudoeurycea altamontana</i>	dd	na					KP886917	KP900064			KP900106	KP900150	KP900196
<i>Pseudoeurycea aurantia</i>	dd	na					KP886901	KP900048			KP900090	KP900134	
<i>Pseudoeurycea cochraniae</i>	dd	na					KP886920	KP900067			KP900109	KP900153	KP900199
<i>Pseudoeurycea firscheini</i>	dd	na					KP886921	KP900068			KP900110	KP900154	
<i>Pseudoeurycea gadovii</i>	dd	na	14	S23	9.00	7.1	KP886903	KP900050			KP900092	KP900136	KP900183
<i>Pseudoeurycea juarezi</i>	dd	na					KP886905	KP900052			KP900094	KP900138	KP900185
<i>Pseudoeurycea leprosa</i>	dd	na	14	S23	10.76	9.2	KP886922	KP900069				KP900155	
<i>Pseudoeurycea lineola</i>	dd	na	14	S24			KP886923	KP900070			KP900111	KP900156	KP900200

Species	Life Cycle	Larval Eco	Trunk Vert	Vert Refs	Adult Body Form	Adult Trunk Form	Co1 Accession	Cytb Accession	ND2 Accession	BDNF Accession	Pomc Accession	RAG1 Accession	Sic8a3 Accession
<i>Pseudoeurycea langicauda</i>	dd	na					KP886906	KP900053			KP900095	KP900139	KP900186
<i>Pseudoeurycea melanomolga</i>	dd	na					KP886924	KP900071			KP900112	KP900157	KP900201
<i>Pseudoeurycea mixcaatl</i>	dd	na					KP886925	KP900072			KP900113	KP900158	KP900202
<i>Pseudoeurycea obesa</i>	dd	na					KP886926	KP900073			KP900114	KP900159	KP900203
<i>Pseudoeurycea archileucus</i>	dd	na									KP900104	KP900148	KP900195
<i>Pseudoeurycea archimelae</i>	dd	na	14	S24	13.68	11.4	KP886916	KP900063			KP900105	KP900149	
<i>Pseudoeurycea papenfussi</i>	dd	na					KP886907	KP900054			KP900096	KP900140	KP900187
<i>Pseudoeurycea rex</i>	dd	na	14	S23	10.58	8.2	KP886908	KP900056	AY916025		KP900098	AY650125	KP900189
<i>Pseudoeurycea robertsi</i>	dd	na	14	S23,S36	9.00	7.2	KP886909	KP900057			KP900099	KP900143	KP900190
<i>Pseudoeurycea ruficauda</i>	dd	na					KP886927	KP900074			KP900115	KP900160	KP900204
<i>Pseudoeurycea saltator</i>	dd	na					KP886910	KP900058			KP900100	KP900144	KP900191
<i>Pseudoeurycea smithi</i>	dd	na	14	S23			KP886911	KP900059			KP900101	KP900145	KP900192
<i>Pseudoeurycea tenchalli</i>	dd	na					KP886912	KP900060			KP900102	KP900146	KP900193
<i>Pseudoeurycea unguidentis</i>	dd	na						AF380774					
<i>Pseudoeurycea werleri</i>	dd	na	14	S23	8.01	6.1	KP886928	KP900075			KP900116	KP900161	
<i>Pseudotriton montanus</i>	bi	lo	17	S36	9.56			KF562585	AY916021		KF562695		EF107443
<i>Pseudotriton ruber*</i>	bi	lo	17	S36	9.31		AY728220	AY728220	AY728220	EU275900	EU275854		
<i>Stereochilus marginatus</i>	bi	le	19	S36	14.79		AY728212	AY728212	AY728212	EU275859	EU275813	AY691713	
<i>Thorius arboreus</i>	dd	na						KC884119			KC884221		
<i>Thorius aureus</i>	dd	na	15	S56				KC884065			KC884197		
<i>Thorius boreas</i>	dd	na						KC884067			KC884198		
<i>Thorius dubitus</i>	dd	na	14	S57				DQ640019			KC884200		
<i>Thorius grandis</i>	dd	na	14	S58				KC884115			KC884181		
<i>Thorius lunaris</i>	dd	na	14	S57				KC884069			KC884201		
<i>Thorius macdougalli</i>	dd	na	14	S23	10.24	8.2		KC884072			KC884202		
<i>Thorius magnipes</i>	dd	na	14	S57				KC884122			KC884191		
<i>Thorius maxillibrochus</i>	dd	na						KC884103			KC884182		
<i>Thorius minutissimus</i>	dd	na						DQ640021			KC884194		
<i>Thorius myndemus</i>	dd	na	14	S57				KC884100					
<i>Thorius munificus</i>	dd	na	14	S57			KP886944	KP900081			KP900122	KP900168	KP900210
<i>Thorius narisovalis</i>	dd	na						KC884077			KC884206		
<i>Thorius omiltemi</i>	dd	na	14	S58				KC884079			KC884208		
<i>Thorius papaloe</i>	dd	na	14	S59				KC884081			KC884214		
<i>Thorius pennatulus</i>	dd	na	14	S23,S57	10.83	8.6		KC884082			KC884210		
<i>Thorius pulmonaris</i>	dd	na	14	S23	11.63	9.5		KC884101					
<i>Thorius schmidti</i>	dd	na	14	S57				KC884084			KC884195		
<i>Thorius spilogaster</i>	dd	na	14	S57				KC884087			KC884218		
<i>Thorius troglodytes</i>	dd	na	14	S57			KP886945	KC884087			KP900123	KF911904	KP900211
<i>Urselperpes brucei</i>	bi	lo	15.5	S60				FJ917634	JQ920802		JQ920724	FJ917630	
Proteidae													
<i>Necturus alabamensis</i>	pd	lo			9.79		KU985890	AY691725	DQ517763		JX144997	JX145009	JX145025
<i>Necturus beyeri*</i>	pd	lo	18	S37	9.40		GQ368658	GQ368658	GQ368658		JX144998	JX145010	JX145026
<i>Necturus lewisi</i>	pd	lo							AY916042		JX144999	JX145011	JX145027
<i>Necturus maculosus</i>	pd	lo	18	S37	8.28		KU985645	AY691724			JX145001	JX145013	JX145029
<i>Necturus punctatus</i>	pd	lo	18	S37	8.58		KU985769				JX145002	JX145014	JX145030
<i>Proteus anguinus</i>	pd	lo	29	S61	22.94		GQ368659	GQ368659	GQ368659		KC295576	AY650138	
Rhyacotritonidae													
<i>Rhyacotriton cascadae</i>	bi	lo	16	S62	9.76			AY691727				AY691694	
<i>Rhyacotriton kezeri</i>	bi	lo	15	S62	9.27			AY691728				AY650129	
<i>Rhyacotriton olympicus*</i>	bi	lo	16	S62	9.49		KU985758	EF036689					
<i>Rhyacotriton variegatus*</i>	bi	lo	15	S62	8.36		AY728219	AY728219	AY728219	EU275869	EU275823	AY691693	
Salamandridae													
<i>Calotriton arnoldi</i>	bi	lo	14	S61,S63			EU880307	EU880307	EU880307			KC665966	
<i>Calotriton asper</i>	f-bi	lo	14	S61,S64	7.55			KC665957	DQ517766			AY583348	EF107346
<i>Chioglossa lusitana</i>	bi	lo	13	S61,S64	9.17		EU880308	EU880308	EU880308			AY583347	
<i>Cynops cyanurus*</i>	bi	le	13	S65,S66			EU880309	EU880309	EU880309			AY583347	
<i>Cynops ensicauda</i>	bi	le	13	S66	6.74		EU880310	EU880310	EU880310			AB574756	
<i>Cynops orientalis</i>	bi	le	13	S65,S66			EU880311	EU880311	EU880311			KC165590	
<i>Cynops orphicus</i>	bi	le	13	S66			EU880312	EU880312	EU880312				
<i>Cynops pyrrhogaster</i>	bi	le	13	S66	6.58		EU880313	EU880313	EU880313		AB572298	AB754794	
<i>Echinotriton andersoni</i>	bi	le	12	S67	4.77		NC017870	NC017870	NC017870		HM462065	AB856892	
<i>Echinotriton chinhalensis</i>	bi	le	13	S68			EU880315	EU880315	EU880315				
<i>Euproctus montanus</i>	bi	lo	13	S61,S64			EU880316	EU880316	EU880316				
<i>Euproctus platycephalus</i>	f-bi	lo	14	S61,S64			EU880317	EU880317	EU880317				
<i>Lissotriton boscai</i>	f-bi	le	12	S69	9.10		JN379840	DQ821219	DQ821219		DQ517831		
<i>Lissotriton helveticus</i>	f-bi	le	12	S69	6.39		JF812747	DQ821239	AY951504		DQ821239		
<i>Lissotriton italicus</i>	f-bi	le	12	S69				DQ821243	JN788173				
<i>Lissotriton montandoni</i>	bi	le	12	S69	8.28		EF526010	DQ821259	DQ517841				
<i>Lissotriton vulgaris</i>	f-bi	le	13	S69	7.54		EU880339	EU880339	EU880339				
<i>Lyciasalamandra antalyana</i>	vi	na							DQ517778				
<i>Lyciasalamandra atiji</i>	vi	na	16	S61,S64			AF154053	AF154053	AF154053	KF645905	KF645786	KF645737	KF645521
<i>Lyciasalamandra billae</i>	vi	na	16	S61,S64					DQ517781				
<i>Lyciasalamandra fazilae</i>	vi	na	16	S61,S64					DQ517782	KF645908	KF645788	KF645740	KF645524
<i>Lyciasalamandra flavimembris</i>	vi	na	16	S61,S64			EU880318	EU880318	EU880318				
<i>Lyciasalamandra helverseni</i>	vi	na	16	S61,S64				DQ473603	DQ517785				
<i>Lyciasalamandra luschani</i>	vi	na	16	S61,S64			KF645998	KF645995	DQ517783	KF645907	KF645787	AY323753	KF645523
<i>Mertensiella caucasica</i>	bi	lo	15	S61,S64	7.25		EU880319	EU880319	EU880319				
<i>Mesotriton alpestris</i>	f-bi	le	12	S61,S64	6.15		EU880335	EU880335	EU880335				
<i>Neurergus crocatus</i>	bi	lo	13	S61,S64	7.23			AY336661	DQ517788				
<i>Neurergus kaiseri</i>	bi	lo					EU880320	EU880320	EU880320				
<i>Neurergus micropilotus</i>	bi	lo							DQ517790				
<i>Neurergus strauchii</i>	bi	lo	13	S61,S64	5.94		EU880321	EU880321	EU880321				
<i>Notophthalmus meridionalis</i>	f-bi	le	13	S61	7.20		EU880322	EU880322	EU880322				
<i>Notophthalmus perstriatus</i>	f-bi	le			8.74		NC028278	NC028278	NC028278				
<i>Notophthalmus viridescens*</i>	f-bi	le	13	S61	6.46		EU880323	EU880323	EU880323				
<i>Ommatriton ophryticus</i>	f-bi	le					EF526046	DQ821267	DQ517844			AY650134	
<i>Ommatriton vittatus</i>	f-bi	le	12	S70	9.80		EU880338	EU880338	EU880338				
<i>Pachytriton archosopus</i>	bi	lo					JN700858	KU374978	JX907837		KU375036	GQ303708	
<i>Pachytriton brevipes</i>	bi	lo	12	S65,S66	7.53		EU880324	EU880324	EU880324		KU375039	GQ303700	
<i>Pachytriton feii</i>	bi	lo	12	S61			NC029345	NC029345	NC029345		KU375037	JX907980	
<i>Pachytriton granulatus</i>	bi	lo	12	S61				KU374999	JX907843		KU375042	JX907975	
<i>Pachytriton inexpectatus</i>	bi	lo						KU375002	JX907834		KU375044	JX907960	
<i>Pachytriton labiatus</i>	bi	lo					EU880325	EU880325	EU880325			AY583351	
<i>Pachytriton moi</i>	bi	lo	12	S61				KU375004	JX237746				
<i>Pachytriton wuguanfui</i>	bi	lo						KU374985			KU375046		
<i>Pachytriton xanthospilos</i>	bi	lo						JX237762	JX237730		KU375047	JX907997	
<i>Paramesotriton caudopunctatus</i>	bi	lo	12	S61,S64	7.22		EU880326	EU880326	EU880326				
<i>Paramesotriton chinensis</i>	bi	lo	12	S61,S64	7.26			AB638712	DQ517800				
<i>Paramesotriton deloustali</i>	bi	lo	12	S61,S64			EU880327	EU880327	EU880327			GQ303710	

Species	Life Cycle	Larval Eco	Trunk Vert	Vert Refs	Adult Body Form	Adult Trunk Form	Co1 Accession	Cytb Accession	ND2 Accession	BDNF Accession	Pomc Accession	RAG1 Accession	Sic8a3 Accession
<i>Paramesotriton ermizhai</i>	bi	lo						GQ303671	FJ744601			GQ303711	
<i>Paramesotriton fuzhangensis</i>	bi	lo	12	S61,564			JN700852	JX480893	DQ517803				
<i>Paramesotriton guanxiensis</i>	bi	lo	12	S61,564			JN700851	KU375006	DQ517804				
<i>Paramesotriton hongkongensis</i>	bi	lo	12	S71	6.11		AY458597	AY458597	AY458597				
<i>Paramesotriton laoenis</i>	bi	lo	12	S61,564			EU880328	EU880328	EU880328				
<i>Paramesotriton longliensis</i>	bi	le						JX480885	FJ169608				
<i>Paramesotriton wulingensis</i>	bi	lo							KJ650056				
<i>Paramesotriton yunwuensis</i>	bi	lo							GU980577				
<i>Paramesotriton zhijinensis</i>	f-bi	le							FJ169609				
<i>Pleurodeles poireti</i>	bi	le	13	S72			EU880329	EU880329	EU880329	EF453368	EU275820	EU275787	
<i>Pleurodeles waltl</i>	f-bi	le	14	S72	4.66		EU880330	EU880330	EU880330			AY523736	AY948856
<i>Salamandra atra</i>	ovi	lo	14	S61,564	8.79		KF645992	KF645930	KF645992	KF645900	KF645790	KF645961	KF645520
<i>Salamandra atra</i>	ovi	na	14	S61,564			KP697931	AY042786	DQ517816	KF645878	KF645798	KF645939	KF645496
<i>Salamandra corsica</i>	ovi	le/lo	14	S61,564			KT365336	KT365391	KF645971	KF645882	KF645817	KF645943	KF645499
<i>Salamandra infraimmaculata</i>	ovi	le	15	S61,564			KF645976	KF645919	DQ517819	KF645887	KF645819	KF645949	KF645503
<i>Salamandra lanzai</i>	vi	na	14	S61,564			KF645974	AF356699	DQ517820	KF645884	KF645822	KF645946	KF645500
<i>Salamandra salamandra</i>	ovi	lo	14	S61,564	5.56		EU880331	EU880331	EU880331	EF453369	KF645824	AY650135	EF107368
<i>Salamandrina perspicillata</i>	bi	lo	13	S61,564				HQ915638			HQ915029	HQ915231	
<i>Salamandrina terdigitata</i>	bi	lo	13	S61,564	9.71			KF431264	DQ517823		HQ915067	HQ915372	
<i>Taricha granulosa</i>	f-bi	le	12	S61	7.19		EU880333	EU880333	EU880333		HM462064		
<i>Taricha rivularis</i>	bi	lo	12	S61	6.77		EU880334	EU880334	EU880334		KF550381	AY650133	
<i>Taricha sierrae</i>	bi	lo	12	S61				DQ196282					
<i>Taricha tarosa</i>	bi	le	12	S61	6.60		KU986086	DQ196271	DQ517826			EF107340	EF107444
<i>Triturus carnifex</i>	f-bi	le	14	S61,564			NC015788	NC015788	NC015788				
<i>Triturus cristatus</i>	f-bi	le	15	S61,564	9.68		HQ697273	HQ697273	HQ697273				
<i>Triturus dobragicus</i>	f-bi	le	16	S61,564			HQ697274	HQ697274	HQ697274				
<i>Triturus karelinii</i>	bi	le	13	S61,564			HQ697277	HQ697277	HQ697277				
<i>Triturus macedonicus</i>	f-bi	le	14	S61,564			HQ697278	HQ697278	HQ697278				
<i>Triturus marmoratus</i>	f-bi	le	12	S61,564			HQ697279	HQ697279	HQ697279			AY583354	EF107350
<i>Triturus pygmaeus</i>	f-bi	le	12	S61,564	6.56		HQ697280	HQ697280	HQ697280				
<i>Tylotriton anguliceps</i>	bi	lo						LC017832					
<i>Tylotriton asperrimus</i>	bi	le	13	S73			EU880340	EU880340	EU880340		HM462069	KC165601	
<i>Tylotriton hainanensis</i>	bi	le						DQ517850			HM462070		
<i>Tylotriton himalayensis</i>	bi	le						KT765173					
<i>Tylotriton kweichowensis</i>	bi	le	15	S73	5.52		NC029231	NC029231	NC029231		KC733384		
<i>Tylotriton lizhenchangii</i>	bi	le						AB769532			AB856873	AB856886	
<i>Tylotriton liuyangensis</i>	bi	le						KJ205598					
<i>Tylotriton notialis</i>	bi	lo						AB769535					
<i>Tylotriton panhai</i>	bi	le	13	S74				KR154461	AB856889	KT304305	AB856876	AB856889	
<i>Tylotriton shanjing</i>	bi	le					KR154461	KR154461	KR154461		HM462066	AB856888	
<i>Tylotriton shanorum</i>	bi	le						AB922822					
<i>Tylotriton taliangensis</i>	bi	le	15	S73			KP979646	KP979646	KP979646	AF497712	AB856878	AB856891	EF107437
<i>Tylotriton uyenoi</i>	bi	le	13	S74				AB856882	AB830729		AB856868	AB856882	
<i>Tylotriton verrucosus</i>	bi	le	13	S73	5.94		NC017871	NC017871	NC017871		HM462067	AB856884	
<i>Tylotriton vietnamensis</i>	bi	le	13	S75,S76			JQ046329	AB856883	HM770088		HM770091	AB856883	
<i>Tylotriton wenxiensis</i>	bi	le					NC027507	NC027507	NC027507	EU275867	HM462071	EU275788	
<i>Tylotriton yangi</i>	bi	le						AB830739			AB856872	AB856885	
<i>Tylotriton zieglerei</i>	bi	le						AB769539			AB856874	AB856887	
Sirenidae													
<i>Pseudobranchius axanthus*</i>	pd	le	35	S36,S37	23.97			AY713226			JX145003	JX145015	JX145031
<i>Pseudobranchius striatus</i>	pd	le	36	S36,S37				AY713120			JX145004	JX145016	JX145032
<i>Siren intermedia intermedia</i>	pd	le	31	S36,S37	15.90						JX145005		JX145033
<i>Siren intermedia nettingi</i>	pd	le									JX145006	JX145017	JX145034
<i>Siren lacertina*</i>	pd	le	31	S36,S37	14.07						JX145008	JX145019	JX145036

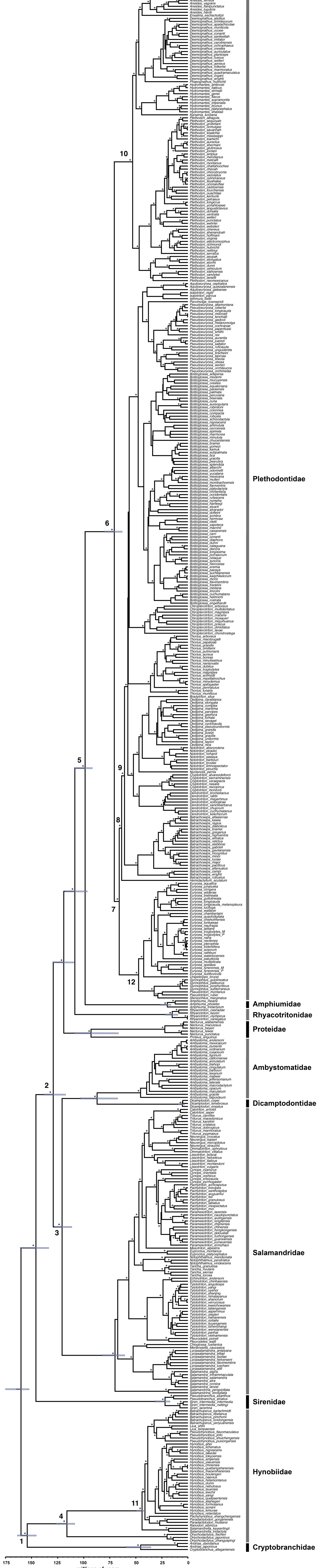
Supplemental Table S10. Partitions and models of sequence evolution were estimated with Partitionfinder [S77], based on the greedy algorithm and AIC model selection. Partitions were reduced from 21 (3 codon position of 7 genes) to 13. p# = codon position number (i.e., p1, p2, and p3 for each gene). The total number of nucleotide positions in each partition is also noted.

Partitions	Models	# pos in partition
<i>BDNF</i> p1, <i>Slc8a3</i> p1, <i>Rag1</i> p1, <i>POMC</i> p1, <i>POMC</i> p2	GTR+I+G	1452
<i>BDNF</i> p2, <i>Rag1</i> p2, <i>Slc8a3</i> p2	HKY+I+G	1086
<i>BDNF</i> p3	K80+G	235
<i>Co1</i> p1	GTR+I+G	511
<i>Co1</i> p2	TRN+I+G	511
<i>Co1</i> p3	GTR+I+G	511
<i>ND2</i> p1	GTR+I+G	337
<i>ND2</i> p2	GTR+I+G	337
<i>ND2</i> p3	GTR+G	337
<i>POMC</i> p3, <i>Slc8a3</i> p3, <i>Rag1</i> p3	SYM+I+G	1034
<i>Cytb</i> p1	GTR+I+G	350
<i>Cytb</i> p2	HKY+I+G	350
<i>Cytb</i> p3	GTR+I+G	350

Supplemental Table S11. Calibration priors used for Bayesian phylogenetic analysis in BEAST [S78]. Calibration priors were set as normal distributions and mostly based on Shen et al. [S22]. Their study was based on 50 nuclear loci and currently provides the most robust estimates of divergence times for salamanders. Hynobiid calibrations were based on Chen et al. [S79], which used 29 nuclear genes. Node numbers are plotted on the phylogeny (Figure S2).

Node Number	Inclusive Taxa	Mean (mya)	Sigma (mya)
1	Cryptobranchoidea	159	4
2	Salamandroidea	160	10
3	Ambystomatidae + Dicamptodontidae + Salamandridae	137	10.5
4	Hynobiidae	126	8
5	Amphiumidae + Plethodontidae	110	8
6	Plethodontidae	66	5
7	Hemidactyliinae	58	4
8	Bolitoglossini + Batrachosepini + Hemidactyliini	54	4
9	Bolitoglossini + Batrachosepini	51	4
10	Plethodontinae	43	3.5
11	Hynobiidae except <i>Onychodactylus</i>	41	3
12	Spelerpini	37	4

Comments on divergence time estimates: Even though we based our tree calibrations on previous molecular estimates [S22, S79], the date ranges of Late Jurassic and Early Cretaceous fossils assigned to the Salamandroidea or Cryptobranchoidea [S3-S14] either fall within these calibration ranges or are younger. Late Jurassic crown group salamander fossils indicate that the divergence between the two main clades (salamandroids and cryptobranchoidea) was at least 160 to 185 mya [S4, S5, S80]. Our estimated deepest divergence among extant salamanders is at the upper end of this range (average 160 mya; range 152 to 175 mya), and still accommodates the Late Jurassic fossil salamanders assigned to crown groups. It is important to note that the most recent molecular estimates found this date to be somewhat older (average 180 to 225, depending on the method [S22]). In our analysis, the relatively large number of shallow node calibrations may have drawn the distribution of divergence time estimates for the deepest nodes toward younger dates. Nevertheless, the trait evolution analyses presented here are based on relative (not absolute) divergence.



Supplemental Figure S2. Bayesian chronogram of 516 salamander taxa. Topology and divergence time estimates from BEAST were based on three mitochondrial (Co1, Cytb, ND2) and four nuclear (BDNF, Pomc, RAG1, Slc8a3) protein-coding genes (totaling 7401 character sets; S1 Appendix, Table S9). Asterisks indicate Bayesian posterior probability ≥ 0.90 , and bars on major nodes (for clarity) represent 95% highest posterior density intervals on divergence dates. The 12 large numbers placed at major nodes indicate calibration priors based on Shen et al. [522] or Chen et al. [579] and are listed in Tables S11.



Supplemental Figure S3. Landmarks and methods for larval body shape analyses. Seven homologous landmarks were placed on images of 27 taxa ($n = 3$ to 5 individuals per taxon; Table S9 see species with asterisk). Specimens were anesthetized by submerging with MS-222 and laid in a set of parallel tracks to ensure body straightness. We used anesthetized live larvae in order avoid preservation variation. No larvae had yolk and all had developed hind limbs (except Sirenidae, which never develop hind limbs). Larvae did not show signs of metamorphosis such as head shape changes or external gill reabsorption (except Amphiumidae, which rapidly resorb external gills after hatching, but otherwise maintain a larval morphology).

Dorsal body images were taken with a Canon G9 12 Mega pixel camera from a fixed stand. The landmarks were placed on images as follows: 1) at the most medial position on the snout, 2) anterior to the 1st gill ramus on left side of the head, 3) anterior of the forelimb on left side of the body, 4) on the left side of the body parallel to the posterior margin of the cloaca, 5) on the right side of the body parallel to the posterior margin of the cloaca, 6) anterior of the forelimb on right side of the body, 7) anterior of the 1st gill ramus on right side of the head. We placed the posterior margin of the cloaca over the grid line to mark its position. We did not use hind limb landmarks because one family (Sirenidae) lacks hind limbs. Landmarks 8 and 9 were placed on the scale bar for initial landmarking in tpsDig [S81], but these landmarks were discarded prior to analyses in geomorph [S16]. For this study we were interested in capturing overall body shape. Analyses based on more detailed landmarks on larval salamander heads will be presented elsewhere.

Supplemental References and Voucher Specimen Numbers

- S1. Pagel M, Meade A (2013) BayesTraits v. 2.0. (University of Reading, Reading, UK) (<http://www.evolution.rdg.ac.uk>)
- S2. Elliot MG, Mooers AØ (2014) Inferring ancestral states without assuming neutrality or gradualism using a stable model of continuous character evolution. *BMC Evol Biol* 14:226.
- S3. Wang Y (2004) A new Mesozoic caudate (*Liaoxitriton daohugouensis* sp. nov.) from Inner Mongolia, China. *Chinese Science Bulletin* 49(8):858–860.
- S4. Gao K-Q, Shubin NH (2012) Late Jurassic salamandroid from western Liaoning, China. *Proc Natl Acad Sci USA* 109(15):5767–5772.
- S5. Ascarrunz E, Rage J-C, Legreneur P, Laurin M (2016) *Triadobatrachus massinoti*, the earliest known lissamphibian (Vertebrata: Tetrapoda) re-examined by μ CT scan, and the evolution of trunk length in batrachians. *Contrib Zool* 85(2):201–234.
- S6. Gao K, Cheng Z, and Xu X (1998) First report of a Mesozoic urodele from China. *Chinese Geology* 250:40–41.
- S7. Wang Y, Evans SE (2006) A new short-bodied salamander from the Upper Jurassic/Lower Cretaceous of China. *Acta Palaeontologica Polonica*, 51(1):127–130.
- S8. Evans SE, Lally C, Chure DC, Elder A, Maisano JA (2005) A Late Jurassic salamander (Amphibia: Caudata) from the Morrison Formation of North America. *Zool J Linnean Soc* 143(4):599–616.
- S9. Jia J, Gao K (2016) A new hynobiid-like salamander (Amphibia, Urodela) from Inner Mongolia, China, provides a rare case study of developmental features in an Early Cretaceous fossil urodele. *PeerJ* 4:e2499.
- S10. Evans S, Milner A (1996) A metamorphosed salamander from the Early Cretaceous of Las Hoyas, Spain. *Philos Trans R Soc Lond, Biol Sci* 351(1340): 627–646.
- S11. Jia J, Gao K-Q (2016) A new basal salamandroid (Amphibia, Urodela) from the Late Jurassic of Qinglong, Hebei Province, China. *PLoS ONE* 11(5):e0153834.
- S12. Gao K-Q, Shubin NH (2001) Late Jurassic salamanders from northern China. *Nature* 410:574–577.
- S13. Gao K-Q, Shubin NH (2003) Earliest known crown-group salamanders. *Nature* 422:424–428.
- S14. Wang Y, Rose C (2005) *Jeholotriton paradoxus* (Amphibia: Caudata) from the Lower Cretaceous of southeastern Inner Mongolia, China. *J Vertebr Paleontol* 25(3): 523–532.
- S15. Skutschas P, Martin T (2011) Cranial anatomy of the stem salamander *Kokartus honorarius* (Amphibia: Caudata) from the Middle Jurassic of Kyrgyzstan. *Zool J Linnean Soc* 161(4):816–838.
- S16. Adams DC, Otarola-Castillo E (2013) Geomorph: an R package for the collection and analysis of geometric morphometric shape data. *Methods Ecol Evol* 4(4):393–399.
- S17. Adams DC (2014) Quantifying and comparing phylogenetic evolutionary rates for shape and other high-dimensional phenotypic data. *Syst Biol* 63(2):166–177.
- S18. Beaulieu JM, Jhwueng DC, Boettiger C, O'Meara BC (2012) Modeling stabilizing selection: expanding the Ornstein-Uhlenbeck model of adaptive evolution. *Evolution* 66(8):2369–2383.
- S19. Soul LC, Benson RBJ (2017) Developmental mechanisms of macroevolutionary change in the tetrapod axis: A case study of Sauropterygia. *Evolution* 71(5):1164–1177.
- S20. Mueller RL (2006) Evolutionary rates, divergence dates, and the performance of mitochondrial genes in Bayesian phylogenetic analysis. *Syst Biol* 55(2):289–300
- S21. Vieites DR, Min M-S, Wake DB (2007) Rapid diversification and dispersal during periods of global warming by plethodontid salamanders. *Proc Natl Acad Sci USA* 104(50):19903–19907.
- S22. Shen XX et al. (2016) Enlarged multilocus data set provides surprisingly younger time of origin for the Plethodontidae, the largest family of salamanders. *Syst Biol* 65(1):66–81.
- S23. Wake DB (1966) Comparative osteology and evolution of the lungless salamanders, Family Plethodontidae. *Mem So Cal Acad* 4:1–111.

- S24. Parra-Olea G, Wake DB (2001) Extreme morphological and ecological homoplasy in tropical salamanders *Proc Natl Acad Sci USA* 98(14):7888–7891.
- S25. Mueller RL, Macey JR, Jaekel M, Wake DB, Boore JL (2004) Morphological homoplasy, life history evolution, and historical biogeography of plethodontid salamanders inferred from complete mitochondrial genomes. *Proc Natl Acad Sci USA* 101(38):13820–13825.
- S26. Chippindale PT, Bonett RM, Baldwin AS, Wiens JJ (2004) Phylogenetic evidence for a major reversal of life-history evolution in plethodontid salamanders. *Evolution* 58(12):2809–2822.
- S27. Bonett RM, Steffen MA, Robison GA (2014) Heterochrony repolarized: a phylogenetic analysis of developmental timing in plethodontid salamanders. *EvoDevo* 5(1):27.
- S28. AmphibiaWeb (2017) Information on amphibian biology and conservation. <http://amphibiaweb.org/>. Berkeley, California.
- S29. Buckley D (2012) Evolution of viviparity in salamanders (Amphibia, Caudata). *eLS* 2012:1–13. 9780470015902.a0022851
- S30. Sparreboom M (2014) Salamanders of the Old World. KNNV Publishing, Zeist, Netherlands.
- S31. Nishikawa K, Jiang J-P, Matsui M, Chen C-S (2009) Morphological variation in *Pachytriton labiatus* and a re-assessment of the taxonomic status of *P. granulosus* (Amphibia: Urodela: Salamandridae). *Curr Herpetol* 28(2):49–64.
- S32. Nishikawa K, Jiang J-P, Matsui M (2011) Two new species of *Pachytriton* from Anhui and Guangxi, China (Amphibia: Urodela: Salamandridae). *Curr Herpetol* 30(1):15–30.
- S33. Wu Y, Wang Y, Hanken J (2012) Comparative osteology of the genus *Pachytriton* (Caudata: Salamandridae) from southeastern China. *Asian Herpetol Res* 3(2):83–102.
- S34. Wiens JJ, Hoverman JT (2008) Digit reduction, body size, and paedomorphosis in salamanders. *Evol Dev* 10(4):449–463.
- S35. Krebs SL, Brandon R (1984) A new species of salamander (Family Ambystomatidae) from Michoacan, Mexico. *Herpetologica* 40(3):238–245.
- S36. *Vertebral data were based on specimens from the University of Tulsa field collections RMB (Ronald M. Bonett), MAS (Michael A. Steffen), and specimens housed at the following museums: Biodiversity Research and Teaching Collection at Texas A&M University (BRTC), Florida Museum of Natural History (FLMNH), Museum of Natural Science at Louisiana State University (LSUMNS), Museum of Vertebrate Zoology at the University of California Berkeley (MVZ), Texas Natural History Collection (Texas Memorial Museum) at the University of Texas, Austin (TNHC), Tulane Museum of Natural History, New Orleans LA (TU), and University of Michigan Museum of Zoology (UMMZ).
- S37. Hilton WA (1948) The vertebrae of salamanders. *J Entom Zool* 40(3):47–65.
- S38. Worthington RD, Wake D (1972) Patterns of regional variation in the vertebral column of terrestrial salamanders. *J Morphol* 137(3):257–277.
- S39. Stokely PS, Holle P (1953) Variation in the vertebral axis of the Ambystomidae. *Herpetologica* 9(3):133–138.
- S40. Litvinchuk SN, Borkin L (2003) Variation in number of trunk vertebrae and in count of costal grooves in salamanders of the family Hynobiidae. *Contrib Zool* 72(4):195–209.
- S41. Zhang F (1985) On anatomy of the skeletal system of *Liua shihi* (Liu) (Amphibia, Hynobiidae). *Acta Herpet Sinica* 4(1):17–24.
- S42. Fei L, Ye C (2000) A new hynobiid subfamily with a new genus and new species of Hynobiidae (Amphibia: Caudata) from West China. *Cultum Herpetol Sinica* 8:64–70.
- S43. Jockusch EL, Martínez-Solano I, Hansen R, Wake D (2012) Molecular and morphological diversification of slender salamanders (Caudata: Plethodontidae: *Batrachoseps*) in the southern Sierra Nevada with descriptions of two new species. *Zootaxa* 3190:1–30.
- S44. Jockusch EL (1997) Geographic variation and phenotypic plasticity of number of trunk vertebrae in slender salamanders, *Batrachoseps* (Caudata: Plethodontidae). *Evolution* 51(6):1966–1982.

- S45. Jockusch EL, Wake DB, Yanev KP (1998) New species of slender salamanders, *Batrachoseps* (Amphibia: Plethodontidae), from the Sierra Nevada of California. *Nat Hist Mus Los Angeles Co Contrib Sci* 472:1–17.
- S46. Wake DB (1996) A new species of *Batrachoseps* (Amphibia: Plethodontidae) from the San Gabriel Mountains, southern California. *Nat Hist Mus Los Angeles Co Contrib Sci* 463:1–12.
- S47. Jockusch EL, Yanev KP, Wake DB (2001) Molecular phylogenetic analysis of slender salamanders, genus *Batrachoseps* (Amphibia: Plethodontidae), from central coastal California with descriptions of four new species. *Herpetol Monogr* 15(1):54–99.
- S48. Wake DB, Elias P (1983) New genera and a new species of Central American salamanders, with a review of the tropical genera (Amphibia, Caudata, Plethodontidae). *Nat Hist Mus Los Angeles Co Contrib Sci* 345:1–19.
- S49. Wake DB, Lawson R (1973) Developmental and adult morphology of the vertebral column in the plethodontid salamander *Eurycea bislineata*, with comments on vertebral evolution in the Amphibia. *J Morphol* 139:251–300.
- S50. Brandon RA (1966) Systematics of the salamander Genus *Gyrinophilus*. *Illinois Biological Monographs* 35:1–86.
- S51. Lanza B, Caputo V, Nascetti G, Bullini L (1995) Morphologic and genetic studies of the European plethodontid salamanders: taxonomic inferences (Genus *Hydromantes*). *Monografie Museo Regionale di Scienze Naturali Torino* 16:1–366.
- S52. Min MS, Yang SY, Bonett RM, Vieites DR, Brandon RA, Wake DB (2005) Discovery of the first Asian plethodontid salamander. *Nature* 435:87–90.
- S53. Brame A (1968) Systematics and evolution of the Mesoamerican salamander genus *Oedipina*. *J Herpetol* 2(1/2):1–64.
- S54. Brandon R (1965) Morphological variation and ecology of the salamander *Phaeognathus hubrichti*. *Copeia* 1965(1):67–71.
- S55. Highton R (1962) Revision of North American salamanders of the genus *Plethodon*. *B Fla State Mus* 6:235–367.
- S56. Hanken J, Wake D (1994) Five new species of minute salamanders, Genus *Thorius* (Caudata: Plethodontidae), from northern Oaxaca, Mexico. *Copeia* 1994(3):573–590.
- S57. Hanken J, Wake D (1998) Biology of tiny animals: systematics of the minute salamanders (*Thorius*: Plethodontidae) from Veracruz and Puebla, Mexico, with descriptions of five new species. *Copeia* 1998(2):312–345.
- S58. Hanken J, Wake D, Freeman H (1999) Three new species of minute salamanders (*Thorius*: Plethodontidae) from Guerrero, México, including the report of a novel dental polymorphism in Urodeles. *Copeia* 1999(4):917–931.
- S59. Hanken, J, Wake D (2001) A seventh species of minute salamander (*Thorius*: Plethodontidae) from the sierra de Juarez, Oaxaca, Mexico. *Herpetologica* 57(4):515–523.
- S60. Camp CD, Peterman W, Milanovich J, Lamb T, Maerz J, Wake D (2009) A new genus and species of lungless salamander (Family Plethodontidae) from the Appalachian highlands of the south-eastern United States. *J Zool* 279(1):86–94.
- S61. Arntzen JW, Beukema W, Galis F, Ivanović A (2015) Vertebral number is highly evolvable in salamanders and newts (Family Salamandridae) and variably associated with climatic parameters. *Contrib Zool* 84(2):85–113.
- S62. Good DA, Wake D (1992) Geographic variation and speciation in the torrent salamanders of the genus *Rhyacotriton* (Caudata: Rhyacotritonidae). *U Calif Publ Zool* 126:1–91.
- S63. Carranza S, Amat F (2005) Taxonomy, biogeography and evolution of *Euproctus* (Amphibia: Salamandridae), with the resurrection of the genus *Calotriton* and the description of a new endemic species from the Iberian Peninsula. *Zool J Linn Soc Lond* 145(4):555–582.
- S64. Lanza B, Arntzen JW, Gentile E (2010) Vertebral numbers in the Caudata of the western palaearctic (Amphibia). *Atti del Mus Civico di Storia Naturale di Trieste* 54:3–114.

- S65. Zhao E, Hu Q, Jiang Y, Yang Y (1988) Studies on chinese salamanders. Society for the Study of Amphibians and Reptiles in cooperation with the Herpetological Society of Japan and Japan Wildlife Research Center, Oxford, Ohio, USA.
- S66. Chan LM, Zamudio KR, Wake DB (2001) Relationships of the salamandrid genera *Paramesotriton*, *Pachytriton*, and *Cynops* based on mitochondrial DNA sequences. *Copeia* 2001(4):997–1009.
- S67. Nussbaum RA, Brodie E (1982) Partitioning of the salamandrid genus *Tylostotriton* Anderson (Amphibia: Caudata) with a description of a new genus. *Herpetologica* 38(2):320–332.
- S68. Chang MLY (1932) Notes on two salamanders from Chekiang. *Contrib Biol Lab Sci Soc China Zool* 8:201–212.
- S69. Skorinov DB, Litvinchuk S (2012) Axial skeleton variation in newts of the genus *Lissotriton*: comparative-evolutionary study. Pp. 285–288 in: The Problems of Herpetology. Proceedings of the 5th Congress of the Alexander M. Nikolsky Herpetological Society. Russian Academy of Sciences. Minsk, Russia.
- S70. Litvinchuk SN, Zuiderwijk A, Borkin LJ, Rosanov M (2005) Taxonomic status of *Triturus vittatus* (Amphibia: Salamandridae) in western Turkey: trunk count, genome size and allozyme data. *Amphibia-Reptilia* 26(3):305–323.
- S71. Stuart BL, Papenfuss T (2002) A new salamander of the genus *Paramesotriton* (Caudata: Salamandridae) from Laos. *J Herpetol* 36:145–148.
- S72. Carranza S, Wade E (2004) Taxonomic revision of Algero-Tunisian *Pleurodeles* (Caudata: Salamandridae) using molecular and morphological data. Revalidation of the taxon *Pleurodeles nebulosus* (Guichenot, 1850). *Zootaxa* 488:1–24.
- S73. Liu C-C (1950) Amphibians of western China. (Natural History Museum, Chicago).
- S74. Nishikawa K, Khonsue W, Pomchote P, Matsui M (2013) Two new species of *Tylostotriton* from Thailand (Amphibia: Urodela: Salamandridae). *Zootaxa* 3737:261–279.
- S75. Stuart BL, Phimmachak S, Sivongxay N, Robichaud W (2010) A new species in the *Tylostotriton asperrimus* group (Caudata: Salamandridae) from central Laos. *Zootaxa* 2650:19–32.
- S76. Böhme W, Schöttler T, Nguyen Q, Köhler J (2005) A new species of salamander, Genus *Tylostotriton* (Urodela: Salamandridae), from northern Vietnam. *Salamandra* 41(4):215–220.
- S77. Lanfear R, Calcott B, Ho SY, Guignon S (2012) Partitionfinder: combined selection of partitioning schemes and substitution models for phylogenetic analyses. *Mol Biol Evol* 29(6):1695–1701.
- S78. Bouckaert R et al. (2014). BEAST 2: A software platform for Bayesian evolutionary analysis. *PLoS Comput Biol* 10(4): e1003537.
- S79. Chen MY, Mao RL, Liang D, Kuro-o M, Zeng XM, Zhang P (2015) A reinvestigation of phylogeny and divergence times of Hynobiidae (Amphibia, Caudata) based on 29 nuclear genes. *Mol Phylogenet Evol* 83:1–6.
- S80. Anderson JS (2012) Fossils, molecules, divergence times, and the origin of Salamandroidea. *Proc Natl Acad Sci USA* 109(15):5557–5558.
- S81. Rohlf FJ (2010) TpsDig, v. 2.16 Department of Ecology and Evolution, State University of New York at Stony Brook. (<http://life.bio.sunysb.edu/morph/>)

Voucher Numbers (See Table S9; Supplemental Reference S36)

Specimens use in the vertebral analyses were from the following museums: Biodiversity Research and Teaching Collection at Texas A&M University (BRTC), Florida Museum of Natural History (FLMNH), Museum of Natural Science at Louisiana State University (LSUMNS), Museum of Vertebrate Zoology at the University of California Berkeley (MVZ), Texas Natural History Collection (Texas Memorial Museum) at the University of Texas, Austin (TNHC), Tulane Museum of Natural History, New Orleans LA (TU), and University of Michigan Museum of Zoology (UMMZ). Abbreviations from field series: RMB (Ronald M. Bonett) and MAS (Michael A. Steffen).

Amphiumidae: *Amphiuma means*: FLMNH 18176; FLMNH Lot: 2707 (Specimen: 2-14); FLMNH 124145. *Amphiuma pholeter*: FLMNH 77797-77800. *Amphiuma tridactylum*: LSU 55707; RMB 5410-5411. Ambystomatidae: *Ambystoma annulatum*: RMB 5425-5429. Cryptobranchidae: *Cryptobranchus alleganiensis*: LSU 54897. Dicamptodontidae: *Dicamptodon copei*: UMMZ 171168; UMMZ 171171; UMMZ 171173-171174; UMMZ 171177-171178; UMMZ 171193; UMMZ 171196; UMMZ 171201-171202; UMMZ 171205. *Dicamptodon ensatus*: UMMZ 170933; UMMZ 170935; UMMZ 170949; UMMZ 171392-171395; UMMZ 171397; UMMZ 170928; UMMZ 171402. Plethodontidae: *Aneides ferreus*: UMMZ 170814-170822. *Aneides flavipunctatus*: UMMZ 151153. *Aneides hardii*: UMMZ-150094; UMMZ 151154. *Bolitoglossa morio*: UMMZ 151168. *Bolitoglossa rostrate*: UMMZ 151169-51170; UMMZ 151176-151177. *Bolitoglossa salvinii*: UMMZ 151166. *Bolitoglossa subpalmata*: UMMZ 183398. *Bolitoglossa adspersa*: UMMZ 128541. *Chiropterotriton multidentatus*: UMMZ 151194. *Chiropterotriton chiropterus*: UMMZ 151162; UMMZ 151196. *Desmognathus auriculatus*: UMMZ 183519; UMMZ 183520-183521. *Desmognathus marmoratus*: UMMZ 151385. *Desmognathus monticola*: UMMZ 151148; UMMZ 151158. *Desmognathus quadramaculatus*: UMMZ 154395. *Ensatina eschscholtzii*: UMMZ 170799-70800; UMMZ 170802-170808; UMMZ 170811-170812. *Eurycea chisholmensis*: TNHC 51141; TNHC 51142; TNHC 52770. *Eurycea cirrigera*: UMMZ 154705-154708; UMMZ 154711; UMMZ 154714-154728; UMMZ 54731-154732; UMMZ 154735-154736; UMMZ 154738; UMMZ 154740-154741; UMMZ 154743-154744; UMMZ 154746; UMMZ 154748; UMMZ 154750-154757; UMMZ 154761; UMMZ 154763; LSUMNS 54944-54948; MVZ 184745-184747; MVZ 184750-184751; MVZ 184753-184754; MVZ 184758; MVZ 184760-184766. *Eurycea guttolineata*: UMMZ 154631-154639; UMMZ 154648; UMMZ 154650-154651; UMMZ 154660-154662; LSUMNS 54951-54953; LSUMNS 54955-54963; MVZ184508. *Eurycea latitans*: BRTC 78086; TNHC 59933-59934; TNHC 54536. *Eurycea longicauda*: UMMZ 142533; UMMZ 151271-151276; MVZ 184509-184510. *Eurycea longicauda melanopleura*: LSUMNS 54970-54973. *Eurycea lucifuga*: UMMZ 154663-154675; LSUMNS 54975; MAS 0148; RMB 4156-4163. *Eurycea nana*: BRTC 78116-78120; BRTC 78128. *Eurycea naufragia*: TNHC 51008-51009; TNHC 51026; TNHC 57752. *Eurycea neotenes*: MVZ 120924-120925; MVZ 120927; MVZ 120929; MVZ 120931; MVZ 120933-120937; RMB 4091-4101; RMB 4129-4155. *Eurycea pterophila*: BRTC 78101-78105; TNHC 52764-52765; TNHC 52772-52773. *Eurycea quadridigitata*: UMMZ 135615; UMMZ 154764-154765; UMMZ 154767; UMMZ 183240-183241; UMMZ 183236; UMMZ 183509-183511; UMMZ 185926; LSUMNS 54978-54980; LSUMNS 54983; TU Lot: 11088 (Specimens: 5, 12, 15, 16, 19, 21, 23, 24, 27). *Eurycea rathbuni*: RMB 4300; RMB 4302-4306; RMB 4310-4312; RMB 4314; UMMZ 173560. *Eurycea sosorum*: TNHC 50915; TNHC 50921; TNHC 50923; TNHC 51178-51179. *Eurycea spelaea*: LSU 55084-55085; RMB 2245; RMB 2287; RMB 4217-4228; TU-Lot: 17999 (Specimens: 1-15). *Eurycea tonkawae*: BRTC 78106-78107. *Eurycea tridentifera*: BRTC 78132-78136; TNHC 31522; TNHC 31526; TNHC 53856. *Eurycea troglodytes*: BRTC 78091-78098; BRTC 78108-78115. *Eurycea tynerensis* Metamorphic: RMB 3801-3805; 3825-3829. *Eurycea tynerensis* Paedomorphic: RMB 3291-3318. *Eurycea wallacei*: TNHC 53821; TU 21063-21067. *Eurycea wilderae*: RMB 5412-5418; UMMZ 154710; UMMZ 154713; UMMZ 154724; UMMZ 154729; UMMZ 154745. *Gyrinophilus porphyriticus*: UMMZ 128706; UMMZ 128800; UMMZ 135751; UMMZ 151310; UMMZ 151313; UMMZ 151314; UMMZ 151330; UMMZ 151334; UMMZ 151338; UMMZ 151341; UMMZ 170276; UMMZ 181906; MVZ 184094; MVZ 184111; MVZ 184714-184717. *Hemidactylium scutatum*: UMMZ 141219; UMMZ 141230; UMMZ 141726; UMMZ 151380-151383; UMMZ 178702. *Hydromates brunus*: UMM 152690-152695. *Oedopina uniformis*: UMMZ 171461-171462. *Plethodon dunnii*: UMMZ 171209-171223; UMMZ 171225-171229. *Plethodon elongatus*: UMMZ

170863-170883. *Plethodon larselli*: UMMZ 171265-171269. *Plethodon neomexicanus*: UMMZ 151458; UMMZ 171465. *Plethodon netting*: UMMZ 154380. *Plethodon stormi*: UMMZ 170849-170858. *Plethodon vandykei*: UMMZ 170839-170848. *Plethodon vehiculum*: UMMZ 171206-171208; UMMZ 171224; UMMZ 171232-171233; UMMZ 171254. *Pseudoeurycea robertsi*: UMMZ 151478; UMMZ 151479. *Pseudotriton montanus*: UMMZ 170954. *Pseudotriton ruber*: UMMZ 130416; UMMZ 151485; UMMZ 171466; UMMZ 181908; UMMZ 183396-183397; MVZ 218640; MVZ 218642-218643; MVZ 218652-218654; MVZ 218658-218661. *Stereochilus marginatus*: UMMZ 151487, UMMZ 154379, UMMZ 171463-171464. *Thorius tryglodytes*: UMMZ 151495. Proteidae: *Necturus beyeri*: LSUMNS 54898. Rhyacotritonidae: *Rhyacotriton cascadae*: MVZ 173325-173326; MVZ 173328; MVZ 173335, MVZ 173341. *Rhyacotriton olympicus*: MVZ 173353-173356; MVZ 173358. *Rhyacotriton variegatus*: MVZ 173304; MVZ 173306; MVZ 173308; MVZ 173310; MVZ 173312. Sirenidae: *Pseudobranchius axanthus*: RMB 5419. *Pseudobranchius striatus*: LSUMNS 54892-54896. *Siren intermedia*: RMB 5420. *Siren lacertina*: RMB 5421-5424.

Acknowledgements for specimen access and loans

For access to specimens for vertebral column examination we are grateful to the following museums, collection managers, and curators: Florida Museum of Natural History (K. Krysko and M. Nickerson), Museum of Natural Science at Louisiana State University (E. Rittmeyer and C. Austin), Museum of Vertebrate Zoology at the University of California Berkeley (C. Spencer, D. Wake and J. McGuire), Texas Biodiversity Research and Teaching Collections (T. Hibbitts and L. Fitzgerald), Texas Natural History Collection (T. LaDuc and D. Cannatella), Tulane Museum of Natural History (H. Dundee), and University of Michigan Museum of Zoology (G. Schneider, A. Davis, and D. Rabosky).