

**Title: Improvement of a fermentation process for the production of two PfAMA1-DiCo-based malaria vaccine candidates in *Pichia pastoris***

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## Supplementary Text S1 - Mass spectrometry

Proteins were reduced, alkylated and digested in the polyacrylamide gel using trypsin (Promega, Mannheim, Germany) as previously described<sup>1</sup>. The resulting peptides were analyzed by nanoHPLC using an UltiMate 3000 HPLC system (LC PACKING, Dionex, Idstein, Germany) coupled to an amaZon ETD ion trap mass spectrometer (Bruker Daltonics, Bremen, Germany) with an ESI nanosprayer. The nanoHPLC system and the ion trap mass spectrometer were controlled using Bruker Compass HyStar v3.2 SR2 software. The liquid chromatography system comprised a reversed-phase pre-column (LC PACKINGS, Dionex) for sample desalting and a 15-cm PepMap 100 reversed-phase C18 column, 75  $\mu\text{m}$  inner diameter (LC PACKINGS, Dionex), for peptide fractionation. The peptides were separated using a 45-min linear gradient from 96% (v/v) solution A (2% (v/v) acetonitrile, 0.1% (v/v) formic acid in high-purity water) and 4% (v/v) solution B (98% (v/v) acetonitrile, 0.1% (v/v) formic acid in high-purity water) to 50% (v/v) solution A and 50% (v/v) solution B at a flow rate of 300  $\text{nL min}^{-1}$ . The electrospray was operated in positive ion mode with a spray voltage of  $-4000\text{ V}$  and 10 psi gas pressure. The end plate offset of the mass spectrometer was set to  $-500\text{ V}$  and Proteomics AutoMSMS Alternating Spectra CID-ETD Bruker trapControl v7.0 was used for data acquisition. Raw data files were evaluated using Compass DataAnalysis v4.0 SR5 software with embedded search engine Mascot Search v2.3.01 (Matrix Science Ltd, London, UK). The spectra were searched against the NCBI nr fungi database and the cRAP database (including VAMAX1 and VAMAX2 sequences) using the following parameters: enzyme = trypsin, up to one missed cleavage permitted, no fixed modifications, variable modifications allowed = carbamidomethyl (C), oxidation (M) and propionamide (C), mass tolerance for precursor ion  $\pm 0.3\text{ Da}$  and fragment ion  $\pm 0.3\text{ Da}$ .

## Supplementary Figure S1

VAMAX1

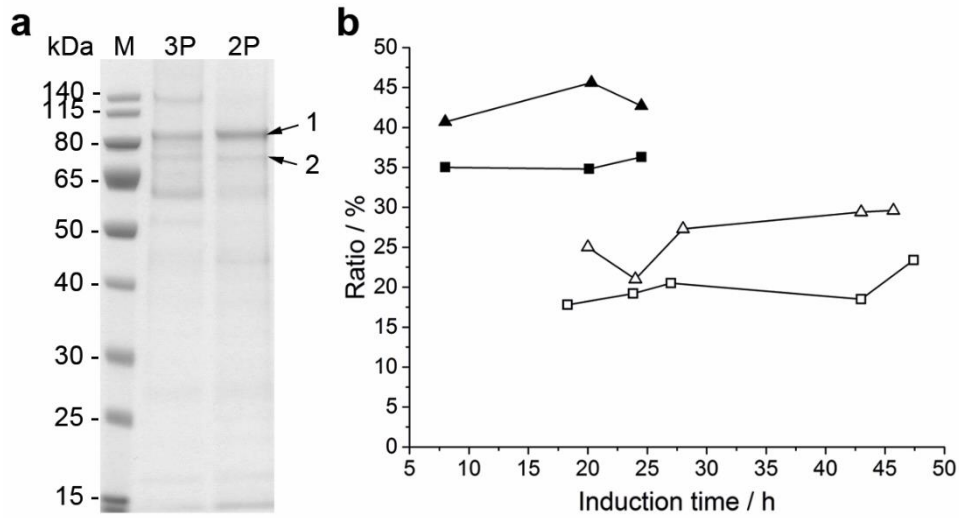


VAMAX2



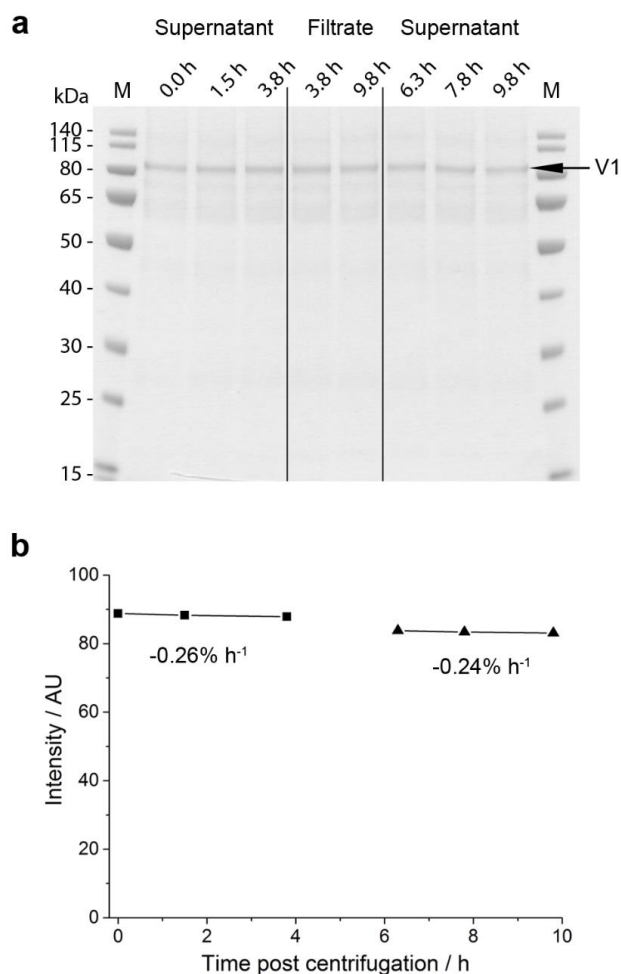
**Figure S1** Schematic representation of the VAMAX1 and VAMAX2 fusion antigens. DiCo1 and DiCo2 = diversity-covering variants of *PfAMA1* (dark gray); *Pfs25* = *Plasmodium falciparum* D7 sexual-stage surface antigen *Pfs25* (light gray); *PfCSP\_TSR* = thrombospondin-related region from *P. falciparum* 3D7 circumsporozoite protein *PfCSP* (hatched); *PfMSP1\_19* = C-terminal 19 kDa fragment of the merozoite surface protein 1 from *P. falciparum* strain FUP (cross-hatched).

## Supplementary Figure S2



**Figure S2** Investigation of potential proteolytic activity in the fermentation broth during the course of induction. **(a)** Samples of the supernatant during the induction phase of both strategies (three-phase = 3P, two-phase = 2P) were analyzed by LDS-PAGE and Coomassie-staining followed by the assessment of the background-corrected intensity of protein band 1 (VAMAX1) and protein band 2 (degradation product of VAMAX1) using AIDA Image Analyzer software (Raytest). Representative samples shown here from the end of the fermentation runs. M: marker. **(b)** To detect potential proteolytic activity in the fermentation broth the intensity ratio of band 1 to band 2 was plotted during the induction phase. Filled symbols: three-phase strategy. Open symbols: two-phase strategy. Squares: VAMAX1 fermentations. Triangles: VAMAX2 fermentations.

## Supplementary Figure S3



**Figure S3** Stability of VAMAX1 (V1) in the supernatant after solid-liquid separation by centrifugation. At the end of the two-phase process, the cells were semi-continuously separated from the fermentation broth by centrifugation with a CARR Powerfuge P6 at  $5,500 \times g$  and a feed rate of  $0.75 \text{ kg min}^{-1}$ . The cell density was considerably reduced to an  $\text{OD}_{600}$  of  $2.48 \pm 0.01$  ( $n = 3$ ) in the supernatant. **(a)** The stability testing of VAMAX1 was performed during two consecutive filtration experiments starting directly (0.0 h) and 6.3 h after centrifugation, respectively, at room temperature. Samples of the supernatant were taken at the indicated times post-centrifugation and were analyzed by LDS-PAGE and Coomassie-staining. **(b)** The Coomassie-stained bands of VAMAX1 were evaluated by densitometric analysis using AIDA Image Analyzer software. The background-corrected integral of each band was plotted over time post-centrifugation. During the first filtration experiment, the apparent decrease in band intensity (representing product loss) was  $-0.26\%$  per hour, calculated using a linear regression function ( $R^2 = 0.97$ ). For the second filtration experiment, the decrease in band intensity was  $-0.24\%$  per hour ( $R^2 = 0.97$ ).

### Supplementary Table S1 – Mass spectrometry data

Candidate / Protein band	Accession	Protein	Score	Peptides	Sequence coverage
<b>VAMAX1</b>					
<b>Band 1</b>	VAMAX1_Dico1_pfs25FKO_CSP_TSR	VAMAX1	103.1	4	5.9%
	gi 328352741	Endochitinase [ <i>Komagataella phaffii</i> CBS 7435]	137.0	2	8.0%
<b>Band 2</b>	VAMAX1_Dico1_pfs25FKO_CSP_TSR	VAMAX1	396.1	8	12.9%
<b>VAMAX2</b>					
<b>Band 1</b>	VAMAX2_Dico2_pfs25FKO_MSP1_19	VAMAX2	313.2	8	12.4%
	gi 328352741	Endochitinase [ <i>K. phaffii</i> CBS 7435]	273.0	3	10.0%
	gi 254564921	Cell wall protein with similarity to glucanases [ <i>K. phaffii</i> GS115]	237.0	4	15.0%
	gi 254570078	Protein of the SUN family (Sim1p, Uth1p, Nca3p, Sun4p) that may participate in DNA replication [ <i>K. phaffii</i> GS115]	193.0	2	9.0%
<b>Band 2</b>	VAMAX2_Dico2_pfs25FKO_MSP1_19	VAMAX2	539.1	12	21.5%
	gi 254564921	Cell wall protein with similarity to glucanases [ <i>K. phaffii</i> GS115]	196.0	3	11.0%
	gi 254570078	Protein of the SUN family (Sim1p, Uth1p, Nca3p, Sun4p) that may participate in DNA replication [ <i>K. phaffii</i> GS115]	171.0	2	9.0%

## Supplementary Data S1

Sequences of the putative host cell proteins identified by mass spectrometry (Supplementary Table S1) were used as BLAST queries against the UniProtKB database with the following parameters:

- Threshold e-value: 0.001
- Matrix: auto
- Filtering: none
- Gapped: yes
- Hits: 100

Alignment with an endochitinase from *Saccharomyces cerevisiae* (UniProtKB entry P29029) found by using gi|328352741 (endochitinase [*Komagataella phaffii* CBS 7435]) as a BLAST query:

P29029		CHIT_YEAST - <b>Endochitinase</b> <i>Saccharomyces cerevisiae</i> (strain ATCC 204508 / S288c) (Baker's yeast)	
<b>E-value:</b> 6.8e-133			
<b>Score:</b> 1067			
<b>Ident.:</b> 48.0%			
<b>Positives:</b> 66.1%			
<b>Query Length:</b> 686			
<b>Match Length:</b> 562			
F2QUR1	F2QUR1_KOMPC	18	AFDNSAKNNVALYWGQNSAGSQRERLSYYCQSDSVDIVLLSFLYIFPANPLGLDFSNACGD 77
P29029	CHIT_YEAST	20	AFD SA N+A+YWGQNSAG+QE L+ YC+S DI LLSFL FP LGL+F+NAC D 77
F2QUR1	F2QUR1_KOMPC	78	QFPSGLLKCDTIAEDIQCQSLGKKVLLSLGGATGTYGFSSDSEAEFVAEVLWDTF-LGG 136
P29029	CHIT_YEAST	78	F GLL C IAEDI+TCQSLGKKVLLSLGGA+G+Y FS DS+AE FA+ LWDTF G 137
F2QUR1	F2QUR1_KOMPC	137	TFSDDLHCTQIAEDIETCQSLGKKVLLSLGGASGSYLFSDDSQAETFAQTLWDTFEGGT 196
P29029	CHIT_YEAST	138	STDERPFGDSILDGIDYDAENNNPTGYTALSAKLREFYASDPSRTYYIAAAPQCPYPDAS 196
F2QUR1	F2QUR1_KOMPC	197	ERPF +++DG D+D ENNN GY+AL+ KLR +A + ++ YY++AAPQCPYPDAS 256
P29029	CHIT_YEAST	197	GASERPFDASVVDGDFDFDIENNNEVGYSALATKLRFLFA-EGTKQYYLSAAPQCPYPDAS 255
F2QUR1	F2QUR1_KOMPC	257	VGDVLANADVDFVFIQFYNNYCALASTSFNWTWLDYAQNTSPNPNVKLYVGLPGGPTGA 315
P29029	CHIT_YEAST	256	VGD+L NAD+DF FIQFYNNY++ S FNW TWL YAQ SPN N+KL++GLPG + A 308
F2QUR1	F2QUR1_KOMPC	316	VGDLEENADIDFAFIQFYNNYCSV-SGQFNWDTWLTYAQTVSPNKNIKLFLGLPGSASAA 375
P29029	CHIT_YEAST	309	ASTSSSQAAATSQTTSTLATSISSTPGSSSTVSSSSSLSSSSSPSSSSSTTIWWTPTD 361
F2QUR1	F2QUR1_KOMPC	376	TS+SQ A T T +T TS +ST +S++ +S+S ++ S +S+ S + + +P 435
P29029	CHIT_YEAST	362	---TSASQTATT--TVATSKTSAASTSSASTSSASTSQKKTQSTTTSQSKSVTLSP-- 415
F2QUR1	F2QUR1_KOMPC	436	PQSSSSSVAAVSSETSSVLTSSVSTTQNSHGEVDSEGTLLTGTSTIWWTPSEAQSYETSS 494
P29029	CHIT_YEAST	416	+++SS++ ++T+ LTSS + T+S G +E T + T T +Q +S 474
F2QUR1	F2QUR1_KOMPC	495	--TASSAIKTSITQTTKTLTSS-TRTKSSLGTTTTSTLNSVAITSMKTTLSSQ---ITS 504
P29029	CHIT_YEAST	475	LSSVSSIPTGNKDVSSILVITDVTDSL-TKESSDSALTISTSLSSSPSLADSSRDGET 484
F2QUR1	F2QUR1_KOMPC		+ V+ T VSS + T +T +L+ +TK SS +L +T+ + SP+ +S G T
P29029	CHIT_YEAST		AALVTPQTITTSIVSSAPIQTAITSTLSPATKSSSVVSLQTATTSTLSPTTSTTS-SGST
F2QUR1	F2QUR1_KOMPC		STVVQVISST
P29029	CHIT_YEAST		S+ + ST
F2QUR1	F2QUR1_KOMPC		SSGSTSSDST
P29029	CHIT_YEAST		

Alignment with a putative family 17 glucosidase SCW10 from *S. cerevisiae* (UniProtKB entry Q04951) found by using gi|254564921 (cell wall protein with similarity to glucanases [*K. phaffii* GS115]) as a BLAST query:

Q04951	SCW10_YEAST - Probable family 17 glucosidase SCW10	Saccharomyces cerevisiae (strain ATCC 204508 / S288c) (Baker's yeast)		
<b>E-value:</b> 3.5e-124				
<b>Score:</b> 961				
<b>Ident.:</b> 55.5%				
<b>Positives :</b> 67.8%				
<b>Query Length:</b> 348				
<b>Match Length:</b> 389				
C4QVL7	C4QVL7_PICPG	26	HQHDKRGVVVVIK----TIVVDG-----STVEATAA	52
			H+H+KR VV T T+VV G ST+E I +	
Q04951	SCW10_YEAST	24	HKHEKRDVVTATVHAQVTVVVSGNSGETIIVPNENAVVAITTSSTAVASQATTSTLEPTTS	83
C4QVL7	C4QVL7_PICPG	53	AQV---QEHAETFAESTPSAVV--SSSSAPSSASSASAPASSGFSFA---GKGVITYSP	103
			A V Q+ T S ++ V S+SS+PSS+SS S+ ASS + S+ G KG+TYSP	
Q04951	SCW10_YEAST	84	ANVVTSQQQTSTLQSSAASTVGSSTSSPSSSSSTSSSASSSSASSISASGAKGITYSP	143
C4QVL7	C4QVL7_PICPG	104	YQAGGGCKTAEVASDLSQLTGYEIIIRLYGVDCNQVENVFKAKAPGQKFLFLGIFVDAIE	163
			Y G CK+ +VASDL QLTG++ IRLYGVDC+QVENV +AK QKFLFLGI++VD I+	
Q04951	SCW10_YEAST	144	YNDDGSCKSTAQVADLEQLTGFNIRLYGVDCSQVENVLQAKTSSQKFLFLGIYVVKRIQ	203
C4QVL7	C4QVL7_PICPG	164	SGVSAIASAVKSYGSWDDVHTVSVGNELVNGEATVVSQIGQYVSTAKSALRSAGFTGPVL	223
			V I SAV+SYGSWDD+ TVSVGNELVN G AT +Q+G+YVSTAKSAL SAG+IG V+	
Q04951	SCW10_YEAST	204	DAVDTIKSAVESYGSWDDITTVSVGNELVNGGSATTTQVGEYVSTAKSALTSAGYTGSSV	263
C4QVL7	C4QVL7_PICPG	224	SVDTFIAVINNPGLCDFADEYVAVNAHAFFDGGIAASGAGDWAAEQIQRVSSACGG-KDV	282
			SVDTFIAVINNP LC+++D Y+AVNAHA+FD AA AG W EQI+RV +ACGG KDV	
Q04951	SCW10_YEAST	264	SVDTFIAVINNPDLCLNYSD-YMAVNAHAYFDENTAAQDAGPWVLEQIERVYTACGGKKDV	322
C4QVL7	C4QVL7_PICPG	283	LIVESGWPSPKGDITNGAAVPSKSNQQAQVQSLGQKIGSSCIAFNAFNDYWKADGPFNAEKY	342
			+I E+GWPSKGDIT G AVPSK+NQ+AA+ S+ GSS F AFND WK DG + EKY	
Q04951	SCW10_YEAST	323	VITETGWPSKGDITYGEAVPSKANQEAAISSIKSSCGSSAYLFTAFNDLWKDDGQYGVVEKY	382
C4QVL7	C4QVL7_PICPG	343	WGILDS	348
			WGIL S	
Q04951	SCW10_YEAST	383	WGILSS	388



Alignment with a probable secreted  $\beta$ -glucosidase SUN4 from *S. cerevisiae* (UniProtKB entry P53616) found by using gi|254570078 (Protein of the SUN family [*K. phaffii* GS115]) as a BLAST query:

P53616		SUN4_YEAST - Probable secreted beta-glucosidase SUN4		Saccharomyces cerevisiae (strain ATCC 204508 / S288c) (Baker's yeast)	
<b>E-value:</b> 3.3e-163					
<b>Score:</b> 1231					
<b>Ident.:</b> 56.4%					
<b>Positives :</b> 69.9%					
<b>Query Length:</b> 431					
<b>Match Length:</b> 420					
C4R2Z5	C4R2Z5_PICPG	1	MKISALTACAVTLAGLA-IAAPAPKPED----CITTVQKRHQKRAVAYDYVYVTVTING	55	
P53616	SUN4_YEAST	1	MK+SA T A +L G + I + P D CITT HQHKRAVA YVY TVT++ MKLSATTLTAASLIGYSTIIVSALPYAADIDTGCTTTAGSHQHKRAVAVTVVYETVTVDK	60	
C4R2Z5	C4R2Z5_PICPG	56	QGETISPTVVAELETVSTPAT--SSSAVATSSAPATTSSIPETISAQVITSSVAESTSAQ	113	
P53616	SUN4_YEAST	61	G+I++PT TV++ T S S+V SS+ +SS E+I TTSS A++T NGQTVIPTSTEASSTVASTTILISESSVTKSSSKVASSS--ESTEQTATTSSSAQTTLIS	118	
C4R2Z5	C4R2Z5_PICPG	114	FTSSTEAEASSSVEQITQSSSSSSPSSSSSSSSSSSSSSSGSSSSGGIDGDLWSYSGPKNKFO	173	
P53616	SUN4_YEAST	119	+ST E+S + S+S S+S+ S++ S++G I GDL+ +SGP KF+ SETSTSESSVPI-----STSGSASTSSAASSATGSIYGDLDLDFSGPYEKFE	164	
C4R2Z5	C4R2Z5_PICPG	174	DGTIKCSEFPNGVVALDHLGFGGWSGLYHPSDTSTGGICEPDTYCSYACQSGMSKTQW	233	
P53616	SUN4_YEAST	165	DGTI C +FPNG GV+ + L GGWSG+ + +DSTGG C+ +YCSYACQ GMSKTQW DGTIPCGQFPNGQGVIPISWLDEGGWSGVEN-TDTSTGGCKEYCSYACQPGMSKTQW	223	
C4R2Z5	C4R2Z5_PICPG	234	PSEQPDNGVSVGGLYCDSGGLYRSKDDIDYLCEWGDVAVVSEIGKTAACRTDYPGT	293	
P53616	SUN4_YEAST	224	PS+QP +G S+GGL C DGYLYRS DTDYLCEWG+D AYVNSE+ AICRTDYPGT PSDQPSDGRSIGGLLC-KDGYLYRSNTDIDYLCEWGVDAAYVSELSNDVAICRTDYPGT	282	
C4R2Z5	C4R2Z5_PICPG	294	ENMVIPTVSPGDELPLTIVDQSSYYTWRGMKTSAQYYVNDAGVDWITGCTWGDYGTIDYG	353	
P53616	SUN4_YEAST	283	ENMVIPT V G LPLT VDQ +YYTW+G+KTSAQYYVN+AG+ C WG + G ENMVIPTVYVQAGDSLPLTIVDQDITYYTWQGLKTSAQYYVNNAGISVEDACVWGSSSSGVG	342	
C4R2Z5	C4R2Z5_PICPG	354	NWAPLNFAGYDNGISYLSLIPNPNKDSLNYNVKIVAYDSSVVIAGECKYENGSYNMG	413	
P53616	SUN4_YEAST	343	NWAPLNFAGAG +G++YLSLIPNPN ++LN+NVKIVA DDSS V GEC YENG++G G NWAPLNFAGSSDGVAYLSLIPNPNNGALNPNVNVKIVAAADSSIVNGECIYENGFSFG-G	401	
C4R2Z5	C4R2Z5_PICPG	414	SDGCTVAVNSGKAKFVLY	431	
P53616	SUN4_YEAST	402	SDGCTV+V +GKAKFVLY SDGCTVSVIAGKAKFVLY	419	

### Supplementary Reference:

1. Shevchenko, A., Tomas, H., Havlis, J., Olsen, J.V. & Mann, M. In-gel digestion for mass spectrometric characterization of proteins and proteomes. *Nat. Protoc.* **1**, 2856-60, doi:10.1038/nprot.2006.468 (2006).