Supporting Information

Controlling the Excited-State Dynamics of Low Band Gap, NIR Absorbers via Proquinoidal Unit Electronic Structural Modulation

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1. Materials and Instrumentation

Materials. All manipulations were carried out under nitrogen or argon previously passed through an O₂ scrubbing tower (Schweitzerhall R3-11 catalyst) and a drying tower (Linde 3-Å molecular sieves) unless otherwise stated. Air sensitive solids were handled in a Braun 150-M glove box. Standard Schlenk techniques were employed to manipulate air-sensitive solutions. All solvents utilized in this work were obtained from Fisher Scientific (HPLC Grade). CH₂Cl₂ and tetrahydrofuran (THF) were distilled from CaH₂ and K/4-benzoylbiphenyl, respectively, under argon. Triethyleamine (TEA) was dried over KOH pellets and distilled under vacuum. All NMR solvents were used as received. The catalysts Pd(PPh₃)₄ and tris(dibenzylideneacetone)dipalladium(0) (Pd₂dba₃), as well as triphenylarsine (AsPh₃), were purchased from Sigma-Aldrich. Various materials (Scheme S1), such 4.9-bromo-6.7starting as $(\mathbf{Br}-\mathbf{TDQ}-\mathbf{Br})^{1}$ dimethyl[1,2,5]thiadiazolo[3,4-g]quinoxaline 4.7diethynylbenzo[c][1,2,5]thiadiazole (**Br-BTD-Br**),¹⁻² [5-triisopropylsilylethynyl-10,20-bis(2',6'bis(3,3-dimethyl-1-butyloxy)phenyl)porphinato]zinc(II) (ArPZnETIPS), [5-ethynyl-10,20- $(ArPZnE)^{3}$ bis(2',6'-bis(3,3-dimethyl-1-butyloxy)phenyl)porphinato]zinc(II) [5triisopropylsilylethynyl-10,15,20-tri(heptafluoropropyl)porphinato]zinc(II) (Rf₃PZnETIPS), and [5-ethynyl-10,15,20-tri(heptafluoropropyl)porphinato]zinc(II) (**Rf₃PZnE**)⁴ were prepared according to the published procedures (Scheme S1). The synthesis of and NMR characterization of 4,9-bis[(10,20-bis[2',6'-bis(3,3-dimethyl-1-butyloxy)phenyl]porphinato)zinc(II)-5-ylethynyl]-6.7-dimethyl[1,2,5]thiadiazolo[3,4-g]quinoxaline (ArPZnE-TDO-EArPZn) and 4,7-bis[(10,20bis[2',6'-bis(3,3-dimethyl-1-butyloxy)phenyl]porphinato)zinc(II)-5-

ylethynyl]benzo[c][1,2,5]thiadiazole (**ArPZnE-BTD-EArPZn**) have been reported previously (Scheme S1).¹ Flash and size exclusion column chromatography were performed on the bench top, using respectively silica gel (EM Science, 230–400 mesh) and Bio-Beads SX-1 as media.

Instrumentation. Electronic spectra were recorded on a Varian 5000 UV/vis/NIR spectrophotometry system. NMR spectra were recorded on a 400 MHz AC-Brucker instrument. Chemical shifts for ¹H NMR spectra are reported relative to residual protium in the deuterated solvents (CDCl₃ = 7.26 ppm, THF- d_8 = 1.72, 3.58 ppm). All J values are reported in Hertz. MALDI-TOF mass spectroscopic data were obtained with a Perspective Voyager DE Instrument (Department of Chemistry, Duke University). Samples were prepared as micromolar solutions in acetone, and 2-(4'-hydroxybenzeneazo)benzoic acid (Sigma-Aldrich) was utilized as the matrix. Microwave assisted reactions were performed with Emrys Personal Chemistry System (Biotage).



Scheme S1. Chemical structures of previously reported compounds, Br-TDQ-Br, Br-BTD-Br, ArPZnETIPS, ArPZnE, Rf₃PZnETIPS, Rf₃PZnE, ArPZnE-TDQ-EArPZn, and ArPZnE-BTD-EArPZn.

2. Synthetic Procedures and Characterization of New Chromophores



Scheme S2. Synthesis of **ArPPtE** from **ArPZnETIPS.** (a) trifluoroacetic acid, r.t., CH₂Cl₂, 30 min; (b) Pt(acac)₂, benzonitrile, 200 °C, 2h, microwave reaction; (c) TBAF, THF, 0 °C, 10 min, under argon.

5-Triisopropylsilylethynyl-10,20-bis(2',6'-bis(3,3-dimethyl-1-butyloxy)phenyl)porphyrin (ArFbETIPS). In a 100 mL round bottom flask equipped with a magnetic stirring bar, ArPZnETIPS (300 mg, 0.270 mmol) was dissolved by dry methylene chloride (~50 mL). Trifluoroacetic acid (5 mL) was added to the round bottom flask and the react was stirred at room temperature under argon for 0.5 h and then quenched by saturated NaHCO₃. The organic layer was collected, washed with saturated NH₄Cl, and then dried with Na₂SO₄. After evaporation of the solvent, the residue was purified by a short column chromatography using 1:1 hexanes: methylene chloride as the eluent. Yield = 275 mg (98 %, based on 300 mg of the ArPZnETIPS starting material). ¹H NMR (400MHz, CDCl₃ as 7.26 ppm): δ 10.02 (s, 1H), 9.67 (d, 2H, J = 4.8), 9.16 (d, 2H, J = 4.4 Hz), 8.87 (d, 2H, J = 4.8 Hz), 8.85 (d, 2H, J = 4.4 Hz), 7.71 (t, 2H, J = 8.4 Hz), 7.01 (d, 4H, J = 8.4 Hz), 3.91 (t, 8H, J = 7.2 Hz), 1.49-1.43 (m, 21H), 0.85 (t,

8H, J = 7.2 Hz), 0.32 (s, 36H), -2.56 (s, 2H). MALDI-TOF: m/z = 1045.65 (calculated for $C_{67}H_{91}N_4O_4Si (M+H)^+ 1045.57$).

[5-Triisopropylsilylethynyl-10,20-bis(2',6'-bis(3,3-dimethyl-1-

butyloxy)phenyl)porphinato]platinum(II) (**ArPPtETIPS**). **ArFbETIPS** (70 mg, 0.067 mmol), platinum acetylacetonate (132 mg, 0.335 mmol) and a magnetic stirring bar were brought together into a 10 mL microwave reaction vial that was then sealed and charged with argon. These reagents were dissolved in ~6 mL of benzonitrile solvent that was previously purged with argon for 1 h. The reaction vial was stirred at 200°C for 2 h in a microwave irradiation cavity. After the reaction without removing the benzonitrile solvent, the reaction mixture was directly purified through a silica column chromatography using 65:35 hexanes: methylene chloride eluent, and the first greenish band was collected as the desired product. Yield = 174 mg (70%, products from 3 reactions in total, based on 210 mg of the **ArFbETIPS** starting material). ¹H NMR (400MHz, CDCl₃ as 7.26 ppm): δ 9.92 (s, 1H), 9.58 (d, 2H, J = 4.8), 9.03 (d, 2H, J = 4.8 Hz), 8.80 (d, 2H, J = 4.8 Hz), 8.78 (d, 2H, J = 4.8 Hz), 7.69 (t, 2H, J = 8.4 Hz), 6.98 (d, 4H, J = 8.4 Hz), 3.90 (t, 8H, J = 7.2 Hz), 1.47-1.41 (m, 21H), 0.88 (t, 8H, J = 7.6 Hz), 0.24 (s, 36H). MALDI-TOF: m/z = 1235.36 (calculated for C₆₇H₉₁N₄O₄PtSi (M+H)⁺ 1237.63).

[5-Ethynyl-10,20-bis(2',6'-bis(3,3-dimethyl-1-butyloxy)phenyl)porphinato]platinum(II) (ArPPtE). Tetrabutylammonium fluoride (1M in THF, 126 μ L, 0.126 mmol) was added to a solution of ArPPtETIPS (120 mg, 0.097 mmol) in THF (50 ml) under argon at 0°C. The reaction mixture was stirred for 10 min at 0 °C, quenched with water, extracted with CH₂Cl₂, dried over Na₂SO₄, and evaporated. The residue was chromatographed on silica gel using 85:15 hexanes:THF as the eluent. Yield = 100 mg (95%, based on 120 mg of ArPPtETIPS). ¹H NMR (400MHz, CDCl₃ as 7.26 ppm):): δ 9.93 (s, 1H), 9.59 (d, 2H, J = 4.8), 9.01 (d, 2H, J = 4.8 Hz), 8.82 (d, 2H, J = 4.8 Hz), 8.80 (d, 2H, J = 4.8 Hz), 7.66 (t, 2H, J = 8.4 Hz), 6.96 (d, 4H, J = 8.4 Hz), 4.13 (s, 1H), 3.90 (t, 8H, J = 7.2 Hz), 0.88 (t, 8H, J = 7.6 Hz), 0.24 (s, 36H). MALDI-TOF: m/z = 1082.12 (calculated for C₅₈H₇₀N₄O₄Pt (M+H)⁺ 1081.29).



Scheme S3. Synthesis of **Rf₃PZnArPPtE** from **ArPZnETIPS** and **Rf₃PZnE**. (a) NBS, 0 °C, CH₂Cl₂:pyridine 20:1 mixture, 1 h, under argon; (b) trifluoroacetic acid, r.t., CH₂Cl₂, 30 min; (c) Pt(acac)₂, benzonitrile, 200 °C, 2h, microwave reaction; (d) Pd₂(dba)₃, AsPh₃, THF/*i*-Pr₂NH 10:1 mixture, 60 °C, 16 h, under argon; (e) TBAF, THF, 0 °C, 10 min, under argon.

[5-Bromo-15-triisopropylsilylethynyl-10,20-bis(2',6'-bis(3,3-dimethyl-1-

butyloxy)phenyl)porphinato]zinc(II) (**BrArPZnETIPS**). In a 250 mL round bottom flask, **ArPZnETIPS** (180 mg, 0.163mmol) was dissolved in methylene chloride (50 mL) and pyridine (5 mL). The reaction mixture was cooled to 0 °C and a solution of NBS (32 mg, 0.179 mmol) in methylene chloride (50 mL) was added via cannula. The reaction mixture was stirred at 0 °C under argon for 1 h and quenched by addition of H₂O (100 mL). The organic layer was separated and dried with anhydrous Na₂SO₄. After evaporation of the solvent, the residue was purified by column chromatography using 6:4 hexanes: methylene chloride as the eluent. Yield = 155 mg (80 %, based on 180 mg of the **ArPZnETIPS** starting material). ¹H NMR (400MHz, CDCl₃ as 7.26 ppm): δ 9.72 (d, 2H, J = 4.4), 9.66 (d, 2H, J = 4.4), 8.93 (d, 2H, J = 4.4 Hz), 8.90 (d, 2H, J = 4.8 Hz), 7.74 (t, 2H, J = 8.4 Hz), 6.98 (d, 4H, J = 8.4 Hz), 3.96 (t, 8H, J = 7.2 Hz), 1.53-1.47 (m, 21H), 0.91 (t, 8H, J = 7.2 Hz), 0.32 (s, 36H). MALDI-TOF: m/z = 1184.57 (calculated for C₆₇H₈₈BrN₄O₄SiZn (M+H)⁺ 1186.83).

5-Bromo-15-triisopropylsilylethynyl-10,20-bis(2',6'-bis(3,3-dimethyl-1-

butyloxy)phenyl)porphyrin (BrArFbETIPS). In a 100 mL round bottom flask equipped with a magnetic stirring bar, **BrArPZnETIPS** (120 mg, 0.101 mmol) was dissolved by dry methylene chloride (~50 mL). Trifluoroacetic acid (2 mL) was added to the round bottom flask and the react was stirred at room temperature under argon for 0.5 h and then quenched by saturated

NaHCO₃. The organic layer was collected, washed with saturated NH₄Cl, and then dried with Na₂SO₄. After evaporation of the solvent, the residue was purified by a short column chromatography using 1:1 hexanes: methylene chloride as the eluent. Yield = 111 mg (98 %, based on 120 mg of the **BrArPZnETIPS** starting material). ¹H NMR (400MHz, CDCl₃ as 7.26 ppm): δ 9.61 (d, 2H, J = 4.4), 9.54 (d, 2H, J = 4.4), 8.82 (d, 2H, J = 4.4 Hz), 8.78 (d, 2H, J = 4.8 Hz), 7.68 (t, 2H, J = 8.4 Hz), 6.91 (d, 4H, J = 8.4 Hz), 3.92 (t, 8H, J = 7.2 Hz), 1.53-1.47 (m, 21H), 0.89 (t, 8H, J = 7.2 Hz), 0.31 (s, 36H), -2.48 (s, 2H). MALDI-TOF: m/z = 1123.92 (calculated for C₆₇H₉₀BrN₄O₄Si (M+H)⁺ 1123.46).

[5-Bromo-15-triisopropylsilylethynyl-10,20-bis(2',6'-bis(3,3-dimethyl-1-

butyloxy)phenyl)porphinato]platinum(II) (**BrArPPtETIPS**). **BrArFbETIPS** (70 mg, 0.062 mmol), platinum acetylacetonate (123 mg, 0.312 mmol) and a magnetic stirring bar were brought together into a 10 mL microwave reaction vial that was then sealed and charged with argon. These reagents were dissolved in ~6 mL of benzonitrile solvent that was previously purged with argon for 1 h. The reaction vial was stirred at 200°C for 2 h in a microwave irradiation cavity. After the reaction without removing the benzonitrile solvent, the reaction mixture was directly purified through a silica column chromatography using 65:35 hexanes: methylene chloride eluent, and the first greenish band was collected as the desired product. Yield = 55 mg (68%, products from 3 reactions in total, based on 70 mg of the **BrArFbETIPS** starting material). ¹H NMR (400MHz, CDCl₃ as 7.26 ppm): δ 9.56 (d, 2H, J = 4.8), 9.51 (d, 2H, J = 4.8), 8.77 (d, 2H, J = 4.8 Hz), 8.75 (d, 2H, J = 4.8 Hz), 7.72 (t, 2H, J = 8.4 Hz), 7.01 (d, 4H, J = 8.4 Hz), 3.95 (t, 8H, J = 7.2 Hz), 1.51-1.44 (m, 21H), 0.95 (t, 8H, J = 7.2 Hz), 0.31 (s, 36H). MALDI-TOF: m/z = 1316.71 (calculated for C₆₇H₈₈BrN₄O₄PtSi (M+H)⁺ 1316.53).

1-[(5-,10,15,20-Tri(heptafluoropropyl)porphinato)zinc(II)]- 2-[(5'-,15'-(triisopropylsilylethynyl)-10',20'-bis(2,6-bis(3,3-dimethyl-1-butyloxy)-

phenyl)porphinato)platinum(II)]-ethyne (**Rf₃PZnArPPtETIPS**). A 100 mL Schlenk flask equipped with a magnetic stirbar was charged with **Rf₃PZnE** (60 mg, 0.067 mmol), **BrArPPtETIPS** (55 mg, 0.042 mmol), $Pd_2(dba)_3$ (23 mg, 0.025 mmol) and AsPh₃ (38 mg, 0.126 mmol). A solvent mixture of THF (50 mL) and diisopropylamine (5ml) was degassed via five freeze-pump-thaw cycles, transferred to the reaction flask and stirred at 60 °C for 16 h. After the reaction endpoint was confirmed by TLC, the reaction mixture was evaporated to dry and the residue was sequentially purified by silica column chromatography using 1:1 hexanes: methylene chloride as the eluent, size exclusion column chromatography using THF as the eluent, and another silica column chromatography using 1:1 hexanes: methylene chloride as the eluent. Yield = 43 mg (48%, based on 55 mg of **BrArPPtETIPS**). ¹H NMR (400MHz, CDCl₃ as 7.26 ppm): δ 10.49 (d, 2H, J = 4.8), 10.12 (d, 2H, J = 4.8), 9.75 (s, 2H), 9.69 (s, 2H), 9.63 (s, 2H), 9.56 (d, 2H, J = 4.8), 8.99 (d, 2H, J = 4.8), 8.80 (d, 2H, J = 4.8), 7.73 (t, 2H, J = 8.4), 7.03 (d, 4H, J = 8.4), 3.99 (t, 8H, J = 7.2), 1.52-1.43 (m, 21H), 1.00 (t, 8H, J = 7.2), 0.33 (s, 36H). MALDI-TOF: m/z = 2138.19 (calculated for C₉₈H₉₆F₂₁N₈O₈PtSiZn (M+H)⁺ 2137.41).

1-[(5-,10,15,20-Tri(heptafluoropropyl)porphinato)zinc(II)]- 2-[(5'-,15'-(ethynyl)-10',20'bis(2,6-bis(3,3-dimethyl-1-butyloxy)- phenyl)porphinato)platinum(II)]-ethyne (Rf₃PZnArPPtE). Tetrabutylammonium fluoride (1M in THF, 26 μL, 0.026 mmol) was added to a solution of Rf₃PZnArPPtETIPS (43 mg, 0.020 mmol) in THF (20 ml) under argon at 0°C. The reaction mixture was stirred for 10 min at 0 °C, quenched with water, extracted with CH₂Cl₂, dried over Na₂SO₄, and evaporated. The residue was chromatographed on silica gel using 85:15 hexanes:THF as the eluent. Yield = 38 mg (95%, based on 43 mg of Rf₃PZnArPPtETIPS). ¹H NMR (400MHz, CDCl₃ as 7.26 ppm): δ 10.47 (d, 2H, J = 4.8), 10.11 (d, 2H, J = 4.8), 9.73 (s, 2H), 9.67 (s, 2H), 9.61 (s, 2H), 9.55 (d, 2H, J = 4.8), 8.97 (d, 2H, J = 4.8), 8.78 (d, 2H, J = 4.8), 7.72 (t, 2H, J = 8.4), 7.03 (d, 4H, J = 8.4), 4.12 (s, 1H), 3.99 (t, 8H, J = 7.2), 0.98 (t, 8H, J = 7.2), 0.32 (s, 36H). MALDI-TOF: m/z = 1979.36 (calculated for C₈₉H₇₆F₂₁N₈O₈PtZn (M+H)⁺ 1981.06).

General Procedure for the Preparation of ArPPtE-TDQ-EArPPt, ArPPtE-BTD-EArPPt, Rf₃PZnE-TDQ-ERf₃PZn, and Rf₃PZnArPPtE-TDQ-EArPPtRf₃PZn. *meso*-ethynyl functionalized (porphinato)metal(II) derivatives (ArPPtE, Rf₃PZnE, and Rf₃PZnArPPtE) and proquinoidal spacer moieties (Br-TDQ-Br, and Br-BTD-Br) were placed in a Schlenk flask equipped with a magnetic stir bar. Pd₂(dba)₃ (0.3 eq., based on the (porphinato)zinc synthon) and AsPh₃ (2 eq., based on the (porphinato)zinc synthon) were added under nitrogen atmosphere. A solvent mixture of 10:1 HPLC grade THF:triethylamine was degassed by a small stream of dry argon for approximately 2 h. Enough of this solvent mixture was added to the reaction vessel via cannula to completely dissolve all reactants, and the resulting solution was heated to 50-60 °C for 16 h. Consumption of all the starting materials was confirmed by thin layer chromatography (6:4 hexanes:CH₂Cl₂ as eluent for ArPPtE-TDQ-EArPPt and ArPPtE-BTD-EArPPt, 65:35 Rf₃PZnE-TDQ-ERf₃PZn and Rf₃PZnArPPtE-TDQhexanes:THF eluent for as EArPPtRf₃PZn). The solution was then cooled to room temperature and evaporated. The crude product was purified by column chromatography using 6:4 hexanes:CH₂Cl₂ as eluent for ArPPtE-TDQ-EArPPt and ArPPtE-BTD-EArPPt, 65:35 hexanes: THF as eluent (due to solubility reason) for Rf₃PZnE-TDQ-ERf₃PZn and Rf₃PZnArPPtE-TDQ-EArPPtRf₃PZn. The product fractions was concentrated to dryness, then further purified through size exclusion column chromatography (Bio-beads SX-1 medium) using THF as the eluent, followed by a short silica gel column chromatography using the corresponding solvent mixture as eluent (vide Additional column chromatography was performed if the level of purity was supra). unsatisfactory as assessed by NMR spectroscopy.



Scheme S4. Schematic summary for the synthesis of ArPME-TDQ-EArPM, ArPME-BTD-EArPM ($M = Zn^{II}$, or Pt^{II}), Rf_3PZnE -TDQ-ER f_3PZn , and Rf_3PZnA rPPtE-TDQ-EArPPtR f_3PZn .

4,9-Bis[(10,20-bis[2',6'-bis(3,3-dimethyl-1-butyloxy)phenyl]porphinato)platinum(II)-5ylethynyl]-6,7-dimethyl[1,2,5]thiadiazolo[3,4-g]quinoxaline (ArPPtE-TDQ-EArPPt). Reagents: ArPPtE (50 mg, 0.046 mmol), Br-TDQ-Br (9 mg, 0.023 mmol), Pd₂(dba)₃ (13 mg, 0.014 mmol), and AsPh₃ (28 mg, 0.092 mmol). Reaction solvent: 22 mL of 10:1 THF:triethylamine. Isolated yield = 30 mg of ArPPtE-TDQ-EArPPt (56%, based on 9 mg of Br-TDQ-Br). ¹H NMR (400MHz, CDCl₃ as 7.26 ppm): δ 10.39 (d, 4H, J = 4.8), 9.94 (s, 2H), 9.05 (d, 4H, J = 4.8), 8.98 (d, 4H, J = 4.8), 8.80 (d, 4H, J = 4.4), 7.76 (d, 4H, J = 8.4), 7.05 (d, 8H, J = 8.8), 3.97 (t, 16H, J = 7.2), 3.14 (s, 6H), 0.95 (t, 16H, J = 7.2), 0.25 (s, 72H). MALDI-TOF: m/z = 2371.35 (calculated for $C_{126}H_{141}N_{12}O_8Pt_2S$ (M+H)⁺ 2373.81).

4,7-Bis[(10,20-bis[2',6'-bis(3,3-dimethyl-1-butyloxy)phenyl]porphinato)platinum(II)-5ylethynyl]benzo[c][1,2,5]thiadiazole (**ArPPtE-BTD-EArPPt**). Reagents: **ArPPtE** (50 mg, 0.046 mmol), **Br-BTD-Br** (7 mg, 0.023 mmol), Pd₂(dba)₃ (13 mg, 0.014 mmol), and AsPh₃ (28 mg, 0.092 mmol). Reaction solvent: 22 mL of 10:1 THF:triethylamine. Isolated yield = 28 mg of **ArPPtE-BTD-EArPPt** (53%, based on 7 mg of **Br-BTD-Br**). ¹H NMR (400MHz, CDCl₃ as 7.26 ppm): δ 9.92 (d, 4H, J = 4.8), 9.80 (s, 2H), 8.94 (s, 4H), 8.92 (d, 4H, J = 4.8), 8.79 (d, 4H, J = 4.8), 7.98 (s, 2H), 7.75 (t, 4H, J = 8.4), 7.04 (d, 8H, J = 8.4), 3.97 (t, 16H, J = 7.2), 0.97 (t, 16H, J = 7.2), 0.26 (s, 72H). MALDI-TOF: m/z = 2294.81 (calculated for C₁₂₂H₁₃₇N₁₀O₈Pt₂S (M+H)⁺ 2293.72).

4,9-Bis[(10,15,20-Tri(heptafluoropropyl)porphinato)zinc(II)-5-ylethynyl]-6,7-

dimethyl[1,2,5]thiadiazolo[3,4-g]quinoxaline (Rf₃PZnE-TDQ-ERf₃PZn). Reagents: Rf₃PZnE (42 mg, 0.046 mmol), Br-TDQ-Br (9 mg, 0.023 mmol), Pd₂(dba)₃ (13 mg, 0.014 mmol), and AsPh₃ (28 mg, 0.092 mmol). Reaction solvent: 22 mL of 10:1 THF:triethylamine. Isolated yield = 23 mg of Rf₃PZnE-TDQ-ERf₃PZn (50%, based on 9 mg of Br-TDQ-Br). ¹H NMR (400MHz, THF- d_8 as 1.72, 3.58 ppm): δ 10.35 (s, 4H), 9.68 (s, 4H), 9.60 (s, 4H), 9.32 (s, 4H), 3.28 (s, 6H). MALDI-TOF: m/z = 2014.76 (calculated for C₇₂H₂₃F₄₂N₁₂SZn₂ (M+H)⁺ 2016.80).

4,9-Bis[[15-[(10',15',20'-tri(heptafluoropropyl)porphinato)zinc(II)-5'-ylethynyl]-10,20bis(2',6'-bis(3,3-dimethyl-1-butyloxy)-phenyl)porphinato]platinum(II)-5-ylethynyl]-6,7dimethyl[1,2,5]thiadiazolo[3,4-g]quinoxaline (Rf₃PZnArPPtE-TDQ-EArPPtRf₃PZn). Reagents: Rf₃PZnE (42 mg, 0.046 mmol), Br-TDQ-Br (9 mg, 0.023 mmol), Pd₂(dba)₃ (13 mg, 0.014 mmol), and AsPh₃ (28 mg, 0.092 mmol). Reaction solvent: 22 mL of 10:1 THF:triethylamine. Isolated yield = 23 mg of Rf₃PZnArPPtE-TDQ-EArPPtRf₃PZn (50%, based on 9 mg of Br-TDQ-Br). ¹H NMR (400MHz, THF- d_8 as 1.72, 3.58 ppm): δ 11.02 (d, 4H, J = 4.8), 10.66 (d, 4H, J = 4.8), 10.29 (s, 4H), 10.23 (s, 4H), 10.17 (s, 4H), 10.10 (d, 4H, J = 4.8), 9.53 (d, 4H, J = 4.8), 9.34 (d, 4H, J = 4.8), 8.27 (t, 4H, J = 8.4), 7.57 (d, 8H, J = 8.4), 4.53 (t,

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16H, J = 7.2), 3.25 (s, 6H), 1.54 (t, 16H, J = 7.2), 0.89 (s, 72H). MALDI-TOF: m/z = 4174.16 (calculated for $C_{188}H_{155}F_{42}N_{20}O_8Pt_2SZn_2$ (M+H)⁺ 4173.35).

3. Steady-State Emission Spectra

The emission spectroscopy for all the **PM-Sp-PM** supermolecular species were investigated at ambient temperature under both deoxygenated and oxygenated condition. Deoxygenation was achieved through three cycles of freeze-pump-thaw, oxygenation was done by purging with air for 30 min. The optical density at all the different excitation wavelengths is ≤ 0.1 . For lowtemperature experiments, prior to freezing the sample by liquid nitrogen, three cycles of freezepump-thaw was applied for deoxygenation.



Figure S1. Comparative steady-state electronic absorption (solid lines) and emission spectra (dash lines) of **ArPZnE-BTD-EArPZn** in THF (blue lines) and toluene (orange lines). Stokes shift is computed based on the difference of the lowest-energy absorption and emission maxima.



Figure S2. Comparative steady-state electronic absorption (solid lines) and emission spectra (dash lines) of **ArPZnE-TDQ-EArPZn** in THF (black lines) and toluene (red lines). Stokes shift is computed based on the difference of the lowest-energy absorption and emission maxima.



Figure S3. Steady-state emission spectra of **ArPPtE-BTD-EArPPt** measured under deoxygenated (black line) and oxygenated (red line) condition. Experimental conditions: $\lambda_{exc} = 480$ nm, ambient temperature, solvent = THF.



Figure S4. Steady-state emission spectra of **ArPPtE-TDQ-EArPPt** measured at deoxygenated (black line) and oxygenated (red line) condition. Experimental conditions: $\lambda_{exc} = 649$ nm, ambient temperature, solvent = THF.



Figure S5. Steady-state emission spectra of **ArPPtE-TDQ-EArPPt** measured at low temperature (77K, green line) and room temperature (298K, red line). Experimental conditions: $\lambda_{exc} = 649$ nm, solvent = Methyl-THF.



4. Picosecond Fluorescence Lifetime Results

Figure S6. Picosecond fluorescence decay of ArPZnE-TDQ-EArPZn in THF (A) and toluene (B), ArPZnE-BTD-EArPZn in THF (C) and toluene (D), ArPPtE-TDQ-EArPPt in THF (E), Rf₃PZnE-TDQ-ERf₃PZn in THF (F), and Rf₃PZnArPPtE-TDQ-EArPPtRf₃PZn in THF (G). Experimental conditions: $\lambda_{exc} = 405$ nm, instrument response function (IRF) ~ 150-160 ps, ambient temperature, magic angle polarization.

5. Nanosecond-to-Microsecond Transient Absorption and Triplet Excited-State Lifetime.

Nanosecond-to-microsecond transient absorption for all chromophores were measured with air-free spectro-cell, three cycles of freeze-pump-thaw was applied for deoxygenation prior to measurement. Oxygen quenching experiments were performed by purging the sample with air for \sim 30 min before measurement. Kinetic traces were fitted with single-exponential decay model to acquire the excited triplet state lifetime.



Figure S7. Nanosecond-to-microsecond transient absorption spectra recorded at selected time delays (under deoxygenated condition) and transient kinetic traces (under deoxygenated and oxygenated condition, respectively) for **ArPPtE-TDQ-EArPPt** (A) and (B), **ArPPtE-BTD-EArPPt** (C) and (D), and **Rf₃PZnArPPtE-TDQ-EArPPtRf₃PZn** (E) and (F). Experimental condition: $\lambda_{exc} = 480$ nm (**ArPPtE-TDQ-EArPPt**, **ArPPtE-BTD-EArPPt**), 500 nm (**Rf₃PZnArPPtE-TDQ-EArPPtRf₃PZn**), pump power = 1.5 mJ / pulse (**ArPPtE-TDQ-EArPPt**, **ArPPtE-BTD-EArPPt**), 1.3 mJ / pulse (**Rf₃PZnArPPtE-TDQ-EArPPtRf₃PZn**), solvent = THF, ambient temperature.

6. Other Supplementary Spectra.



6.1 Global Fitting and Decay Associated Difference Spectra

Figure S8. DADS of the NIR regime for ArPZnE-BTD-EArPZn (A), ArPZnE-TDQ-EArPZn (B), ArPPtE-BTD-EArPPt (C), and ArPPtE-TDQ-EArPPt (D), revealing solvent relaxation dynamic (~ 1 ps), structural (torsional) relaxation dynamics (~ 10-30 ps), and the intrinsic S₁ state decay. Offsets account for the transient absorption signal of the evolved excited triplet states, which persists beyond the delay limit of the transient optical system. Steady-state absorption and emission are plotted as the inverted dash lines.

6.2 Estimation of ISC Quantum Yields for ArPPtE-TDQ-EArPPt

The *ISC* quantum yield for **ArPPtE-TDQ-EArPPt** was determined from fs transient absorption spectral data via comparison of spectra acquired at t = 30 ps, which probes the excited singlet population without influence from solvent and structural relaxation dynamics (these processes are in the time domain of ~ 1 ps and 10-30 ps), and at a delay time (*e.g.*, t = 1.32 ns) at which only ground and excited triplet state populations of **ArPPtE-TDQ-EArPPt** exist. The ISC quantum yield can be estimated using equation (1).

$$\Phi_{ISC} = \frac{\Delta A (715 nm, t=1.32 ns)}{\Delta A (715 nm, t=30 ps)}$$
(1)

To ensure the accuracy of the calculated Φ_{ISC} , the ΔA utilized in equation (1) has to be a pure

signal from ground state bleach (GSB). In this regard, note that the O-band region (600-800 nm) GSB at t = 30 ps overlaps well with the ground-state absorption (Figure S9A), and due to the large Stokes shift, stimulated emission signal is greatly separated from the GSB signal, suggesting ΔA (715 nm, t = 30 ps) can be attributed only to the depletion of ground state populations (due to excited singlet populations). However, the *O*-band region GSB at t = 1.32 ns does not overlap with the ground-state absorption (Figure S9A), particular in the spectral domain of 700-800 nm that is due to the triplet excited state absorption. To eliminate the influence from the triplet excited state absorption on the *Q*-band region GSB at t = 1.32 ns, spectra deconvolution is required. The O-band ground-state electronic absorption can be deconvoluted using three Gaussian peaks (Figure S9B), therefore, a similar deconvolution pattern was utilized for *Q*-band region GSB at t = 1.32 ns (Figure S9C, Fit Peak 3-5). Another two Gaussian peaks that account for the triplet excited state absorption were required to fit transient absorption spectra (t = 1.32 ns) in the 575-1075 nm spectral domain (Figure S9C, Fit Peak 1-2). Thus, the pure signal from *Q*-band region GSB at t = 1.32 ns can be acquired by addition of the three negative Gaussian peaks in Figure S9C (Fit Peak 3-5), and ΔA (715 nm, t = 1.32 ns) obtained based on such deconvoluted spectra can be attributed only to the depletion of ground state populations (due excited triplet populations) (Figure S9D). Then, Φ_{ISC} (ArPPtE-TDQ-EArPPt) $= \Delta A (715 \text{ nm}, t = 1.32 \text{ ns}) / \Delta A (715 \text{ nm}, t = 30 \text{ ps}) = (-28.66 \text{ mOD}) / (-14.37 \text{ mOD}) = 0.5$. In the similar manner, the Φ_{ISC} for Rf₃PZnArPPtE-TDQ-EArPPtRf₃PZn can also be calculated, resulting in Φ_{ISC} (**Rf₃PZnArPPtE-TDQ-EArPPtRf₃PZn**) = 0.73.



Figure S9. Spectral deconvolution for transient absorption at t = 1.32 ns. Comparative spectra among ground-state electronic absorption (red dotted lines, rescaled), transient absorption at t = 30 ps (black solid line), and transient absorption at t = 1.32 ns (green solid line) (A). Spectral deconvolution of *Q*-band ground-state electronic absorption (B). Spectra deconvolution of transient absorption at t = 1.32 ns in the spectral domain of 575-1075 nm. *Q*-band region ground state bleach recovery calculations (D).

6.3 Dynamic Stokes Shift in Ultrafast Time Domain



Figure S10. (A) Femtosecond transient absorption spectra of *Q*-band bleach and stimulated emission region in the time domain of solvation dynamics, experimental condition: $\lambda_{exc} = 900$ nm, power = 260 nJ / pulse, solvent = THF, ambient temperature, magic angle polarization. (B) Schematic illustration of dynamics Stokes shift of **ArPZnE**-

TDQ-EArPZn in THF, where potential energy curve displacement (Δq) between S₀ and S₁ states along the solvation coordinate plays a key role.

7. TD-DFT Calculations and Population Analysis

All calculations were performed upon structures with aliphatic chains truncated to methyl groups (Figure S59, molecular structures on top). For these oligo(porphinato)metal(II) species, C_{2V} conformeric minima could be formulated. Ground-state structure optimizations were performed with Density Functional Theory (DFT) using Gaussian 09, Rev C.1.⁵ The M11⁶ functional was employed for all calculations. Optimizations were performed with minimal symmetry constraints using tight optimization criteria; initial optimizations used smaller basis sets but the final optimizations and TD-DFT calculations employed the 6-311g(d) basis set⁷ as implemented in Gaussian 09. TD-DFT result files were post-processed using the GaussSum package;⁶ this software partitions the wavefunction amplitudes onto molecule fragments using Mulliken population analysis.⁸



Figure S11. Comparison of TD-DFT predicted transitions and experimentally determined ground-state absorption spectra for ArPZnE-TDQ-EArPZn (A), ArPZnE-BTD-EArPZn (B), ArPPtE-TDQ-EArPPt (C), and ArPPtE-BTD-EArPPt (D).

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Figure S12. Frontier orbitals plotted as 0.02 isodensity surfaces for (left) **ArPZnE-TDQ-EArPZn** and (right) **ArPZnE-BTD-EArPZn**, along with a diagram comparing their calculated energies. Arrows depict the major oneelectron configurations with percentages representing each excitation's contribution to the lowest-energy transition. The coefficients of each molecular orbital contributed by porphyrin, spacers (**ETDQE** and **EBTDE**), and zinc atoms are listed, determined by population analysis.



Figure S13. Frontier orbitals plotted as 0.02 isodensity surfaces for (left) **ArPPtE-TDQ-EArPPt** and (right) **ArPPtE-BTD-EArPPt**, along with a diagram comparing their calculated energies. Arrows depict the major oneelectron configurations with percentages representing each excitation's contribution to the lowest-energy transition.

The coefficients of each molecular orbital contributed by porphyrin, spacers (ETDQE and EBTDE), and zinc atoms are listed, determined by population analysis.

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