

Supplemental data for:

Unleashing natural competence in *Lactococcus lactis* by induction of the competence regulator ComX.

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Table S1: Comparison of competence protein identities and length of *L. lactis* KF147 to the competent strain *S. thermophilus* LMD-9.

	<i>comC</i>	<i>comEA</i>	<i>comEC</i>	<i>comFA</i>	<i>comFC</i>	<i>comGA</i>	<i>comGB</i>	<i>comGC</i>	<i>comGD</i>	<i>comGE</i>	<i>comGF</i>	<i>comGG</i>	<i>comX</i>	<i>coiA</i>	<i>ssbA</i>	<i>ssbB</i>	<i>recA</i>	<i>DprA</i>
Protein identity (%)	40	45	44	47	44	51	45	57	36	33	47	24	28	37	60	73	78	59
Positive AA score (%)	58	64	60	63	62	69	67	77	52	64	65	48	51	61	77	81	87	78
Alignment length (%)	96	67	97	87	100	99	97	97	96	98	97	55	90	89	98	100	98	99
Total length protein KF147 (AA)	221	215	736	440	216	312	291	127	143	98	141	94	163	329	129	166	387	282
Total length protein LMD-9(AA)	215	231	746	439	220	313	296	108	120	93	145	105	165	347	130	172	379	279

Fig. S1: Genetic organization of the canonical competence genes in *L. lactis* KF147 (A). The *cin*-box sequences indicated in Wydau *et al.*, 2006 were used to extract *cin*-box sequences in *L. lactis* KF147 (20). These sequences are depicted underneath the black arrow which reflects the promotor region. Numbers surrounding the gene clusters correspond with start and stop codons of the first and last gene of the competence gene/operon. Colors of the gene-representing arrows correspond with the protein-icons in the graphical representation of the core competence machinery (analogous to Johnston *et al.*, 2014, Muschiol *et al.*, 2015 and Mann *et al.*, 2013 (1-3)). Black gene-representing arrows indicate genes present within the *com*-associated operons that are predicted not to have a direct structural role in the core competence machinery.

Upon competence development, the pilus comprising ComGA, ComGB and ComGC entraps the exogenous dsDNA which subsequently connects with ComEA. One strand of the dsDNA is degraded by an unknown domain or protein in *L. lactis*, and the other strand translocates into the cell by transfer through the pore protein ComEC and is further processed and internalized by ComFA and ComFC. In the cytosol, the ssDNA is protected by single stranded binding proteins (SsbA and SsbB) and DprA, which interacts with RecA in order to facilitate DNA integration into the genome. CoiA is thought to fulfill a role in DNA processing and ComC and the minor pilins ComGD-GG are proposed to fulfill a role in priming of pilus assembly (8-10).

Figure S1:

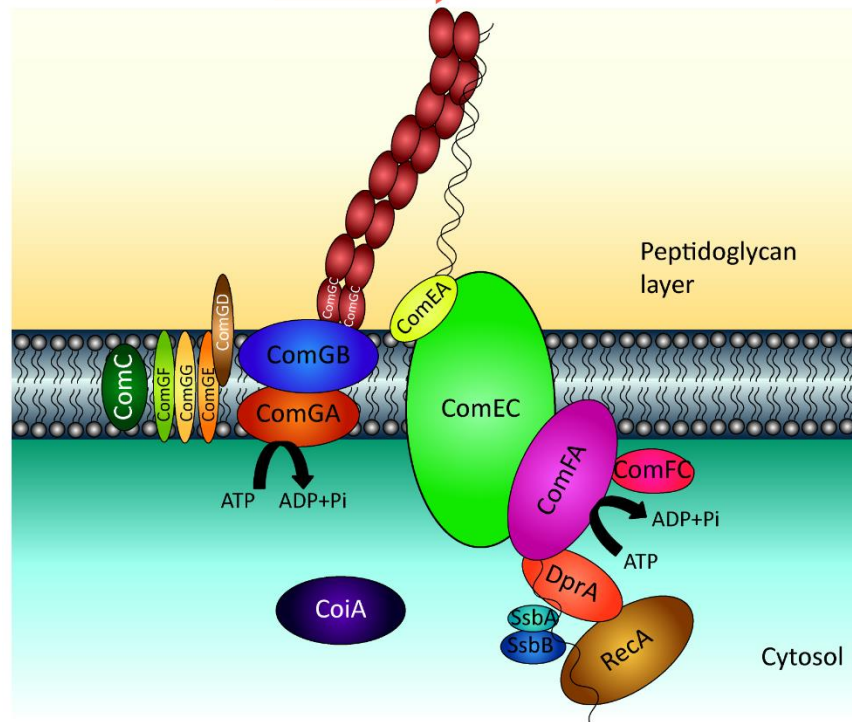
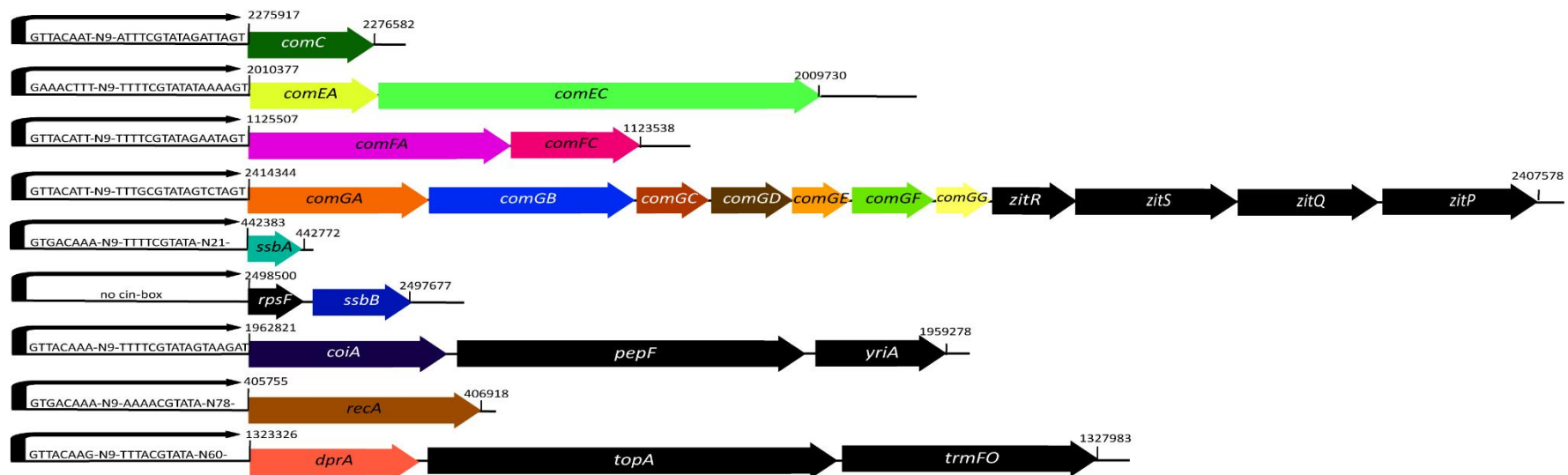


Fig. S2: Genomic analysis of 43 *L. lactis* strains to assess genetic capacity to develop natural competence. Analogous to Figure 1 of the main paper, except that full-length protein identity-scores (%) for the selected subset of late competence associated proteins are displayed in comparison to their homologues in strain *L. lactis* KW2 that was used as a reference. All other analyses, all labels used and references for strains are analogous to those presented in Figure 1. Genetic events leading to competence gene decay (black cells in the figure) are specified as premature stopcodon within the first 90% of the gene (a), transposon insertion (b), prophage insertion (c) absence of gene, mutated/alternative start or lengthened/fused protein; at least more than 25% of its total length (d) , followed by the position within the protein sequence where the event is detected relative to its N-terminus. The right-hand column represents the source of isolation of the strains in which P= plant, D= dairy, S= soil, W= water, H= human body and F= fruit (4).

Figure S2:

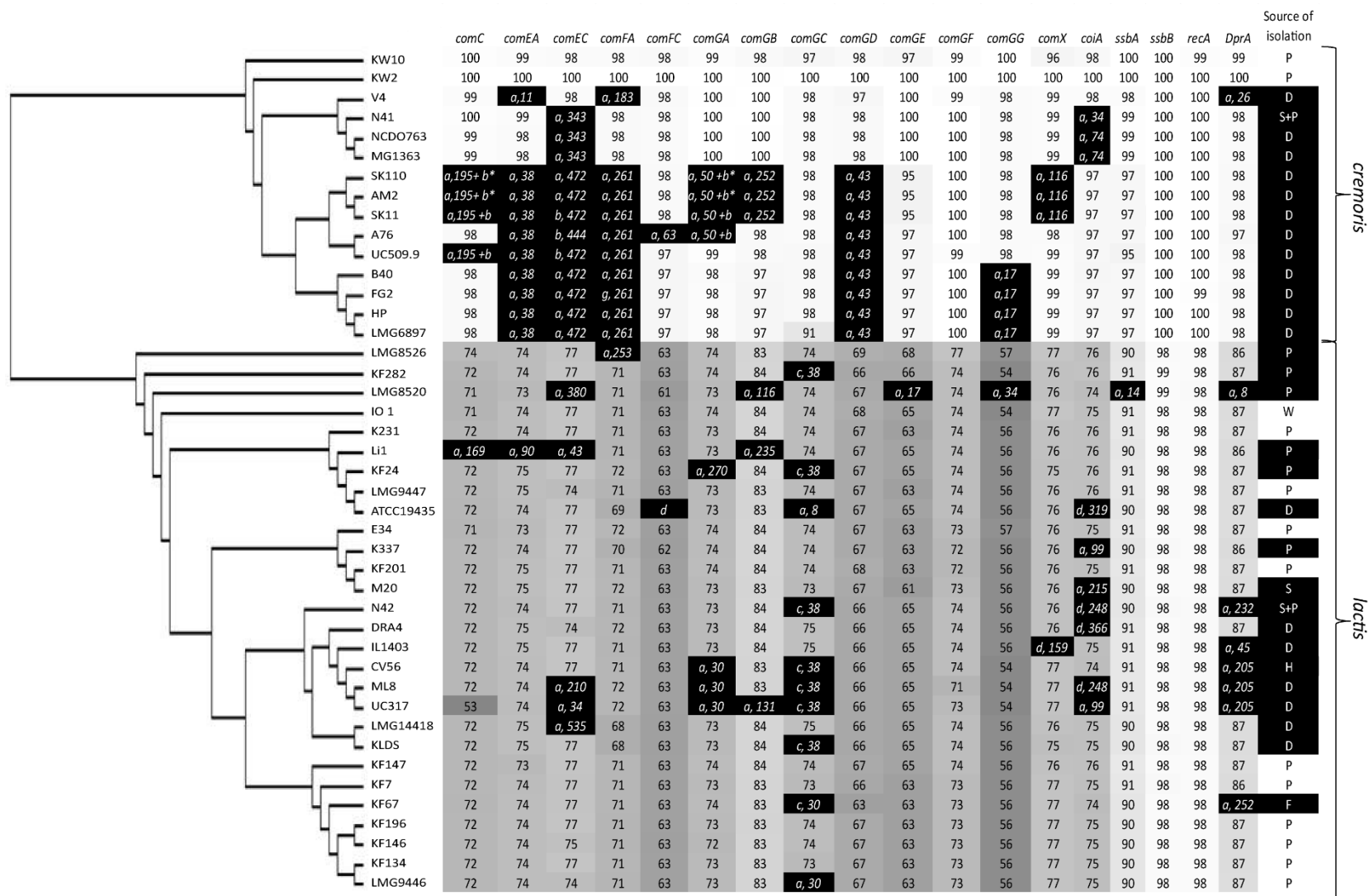


Fig S3: Premature stop-codons and transposon interruption within *comEC* among *L. lactis* subsp. *cremoris* strains SK11, A76 and UC509.9.

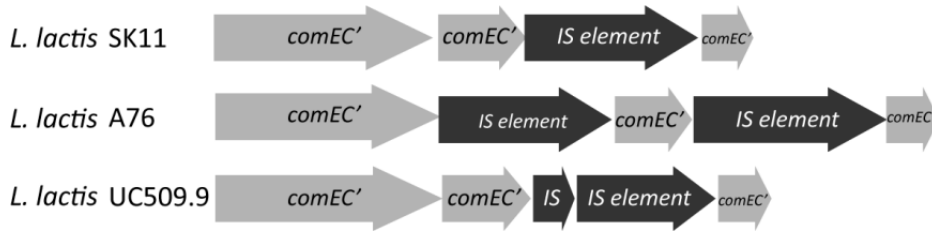


Fig. S4: Mutational events occurring in *L. lactis* subsp. *cremoris* strains that are focused on the late competence gene-sets used in this study. A mutational sequence of events was tentatively reconstructed for the late competence genes in different subspecies *cremoris* strains where the dot resembles the *cremoris* strain ancestor. The core-genome based SNP phylogenetic relatedness displays considerable topological congruency with the late-competence related evolutionary relatedness.

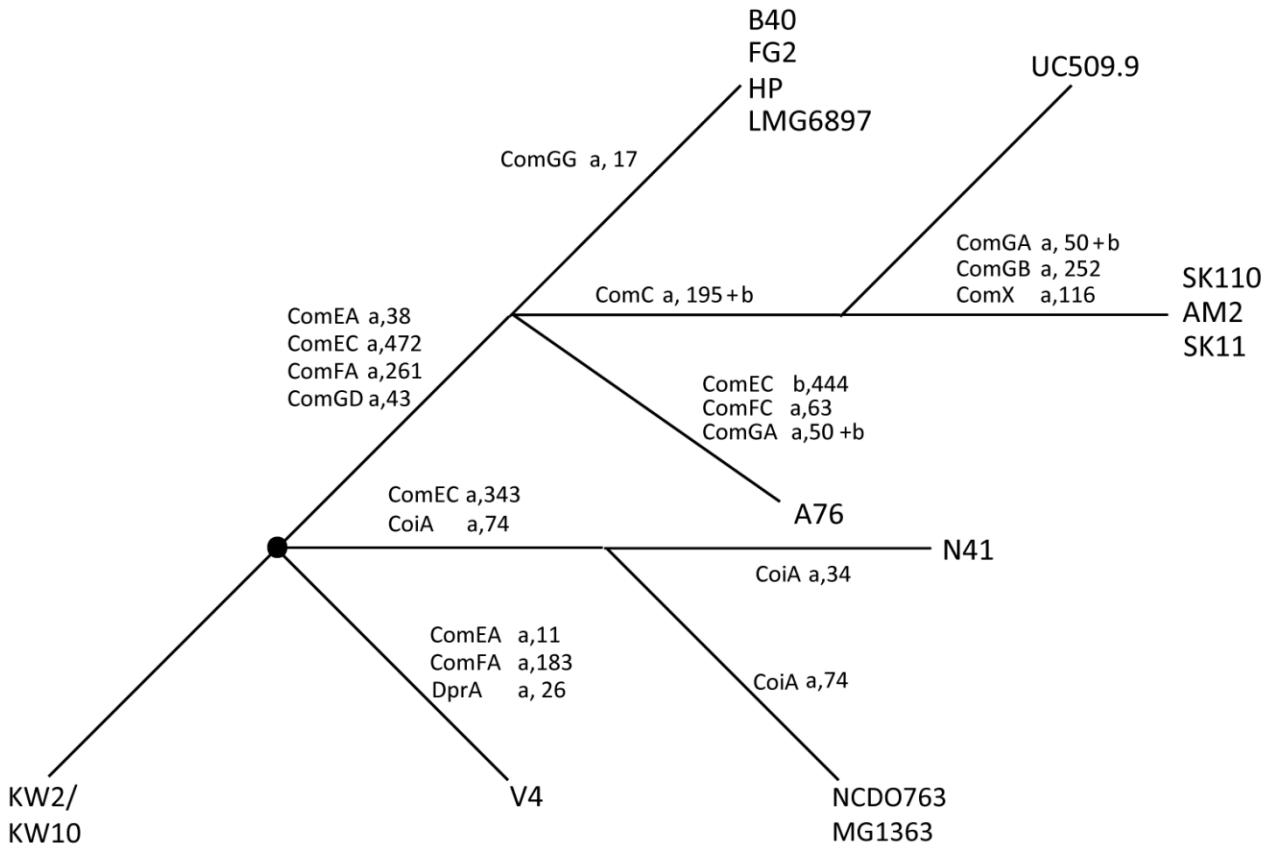


Fig. S5: Representative image of phage sequence insertions within *comGC* of *L. lactis* subsp. *lactis* strains KF282, KF24, N42, CV56, ML8, UC37, KLDS and KF67. Phage sequence arrows are scaled to the KF282 phage sequence arrow, whereas the *com* gene arrows are scaled to the KF282 *comGB* gene. On the right, the conserved insertion sequence within *comGC* in *L. lactis* subsp. *lactis* strains is depicted.

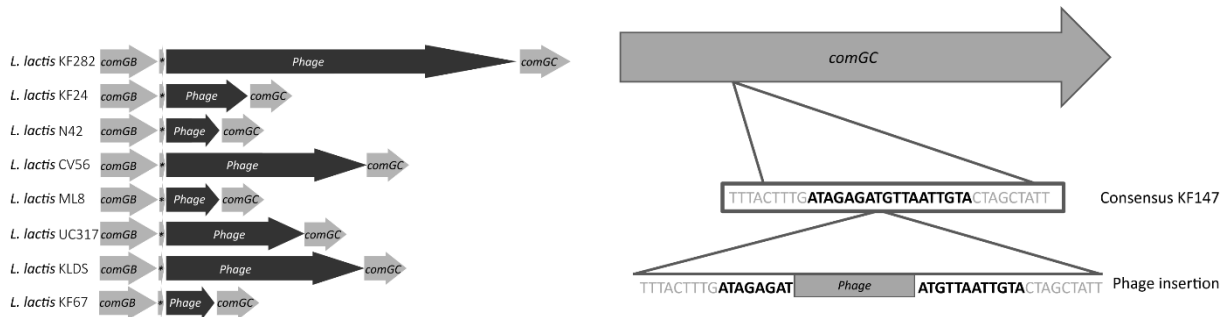


Fig. S6: Growth inhibition upon full induction with nisin is only observed in *L. lactis* KF147 harboring pNZ6200. Full induction with nisin in *L. lactis* KF147 harboring pNZ8150 (empty vector control) and *L. lactis* KF147 harboring pNZ8040 (*pepN* under nisin control) shows the same growth curve as uninduced conditions showing that nisin induction itself and overexpression of a protein does not lead to growth inhibition. In contrast, growth inhibition of *L. lactis* KF147 harboring pNZ6200 can be observed relative to uninduced conditions in this strain.

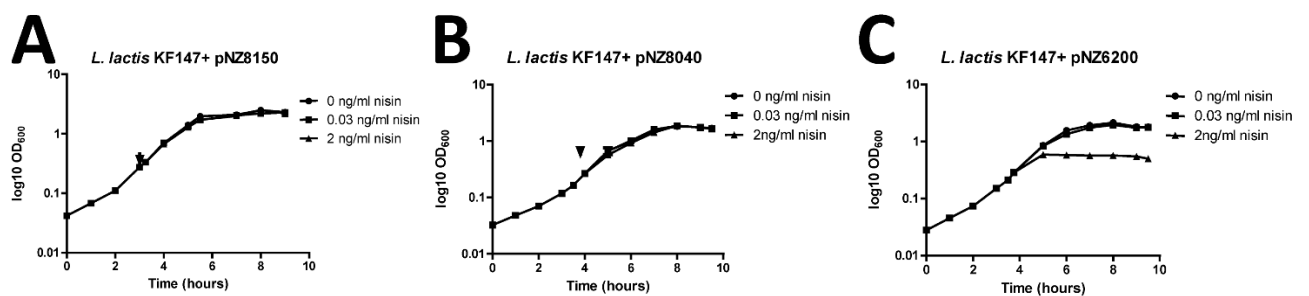
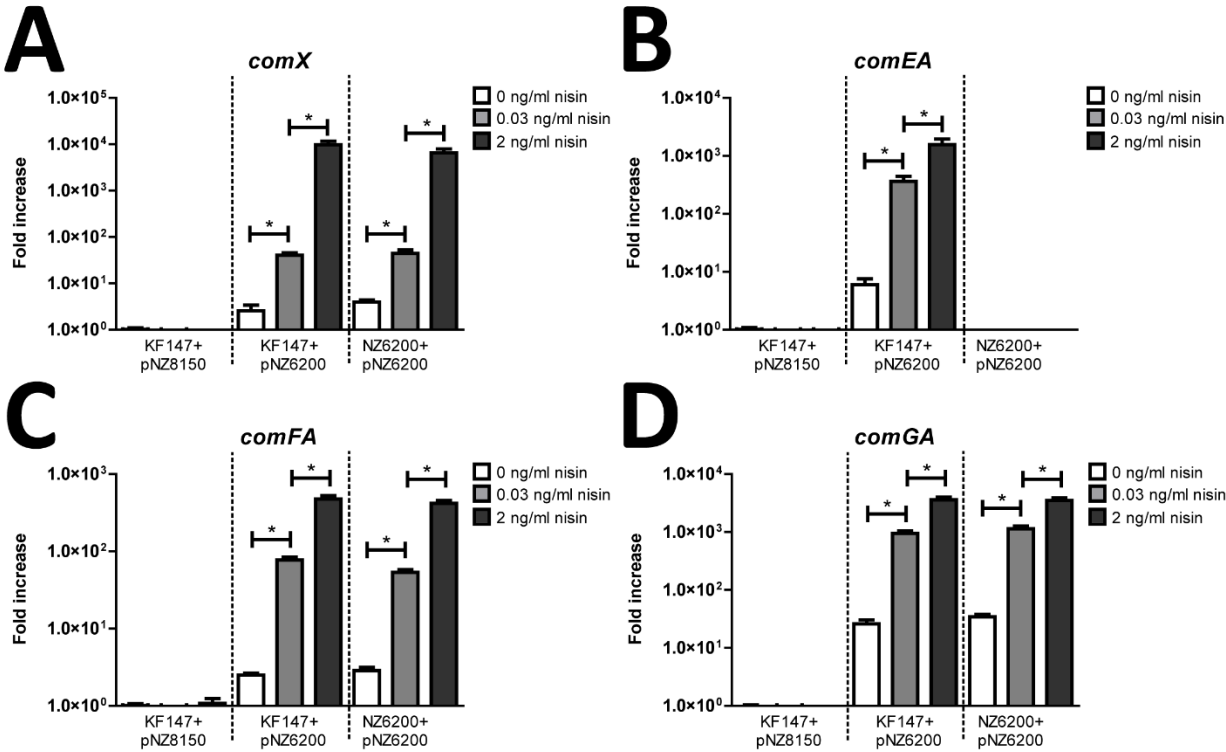


Fig. S7: Gene expression analysis of competence genes in KF147 and NZ6200. Transcript levels of *comX* (A), *comEA* (B), *comFA* (C) and *comGA* (D) in *L. lactis* KF147 harboring pNZ8150 or pNZ6200 and *L. lactis* NZ6200 harboring pNZ6200 in uninduced (0 ng/ml nisin), moderately induced (0.03 ng/ml nisin) and fully induced (2 ng/ml nisin) conditions. Values were obtained from triplicate measurements and statistical significance (P-value<0,05; indicated by *) is based on the non-parametric Mann-Whitney U-test (one-tailed).



References

1. **Mann JM, Carabetta VJ, Cristea IM, Dubnau D.** 2013. Complex formation and processing of the minor transformation pilins of *Bacillus subtilis*. *Mol Microbiol* **90**:1201-1215.
2. **Johnston C, Martin B, Fichant G, Polard P, Claverys JP.** 2014. Bacterial transformation: distribution, shared mechanisms and divergent control. *Nat Rev Microbiol* **12**:181-196.
3. **Muschiol S, Balaban M, Normark S, Henriques-Normark B.** 2015. Uptake of extracellular DNA: competence induced pili in natural transformation of *Streptococcus pneumoniae*. *Bioessays* **37**:426-435.
4. **Dijkstra AR, Setyawati MC, Bayjanov JR, Alkema W, van Hijum SA, Bron PA, Hugenholtz J.** 2014. Diversity in robustness of *Lactococcus lactis* strains during heat stress, oxidative stress, and spray drying stress. *Appl Environ Microbiol* **80**:603-611.