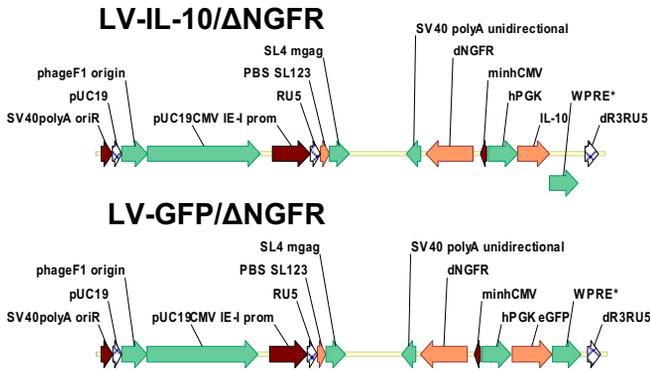
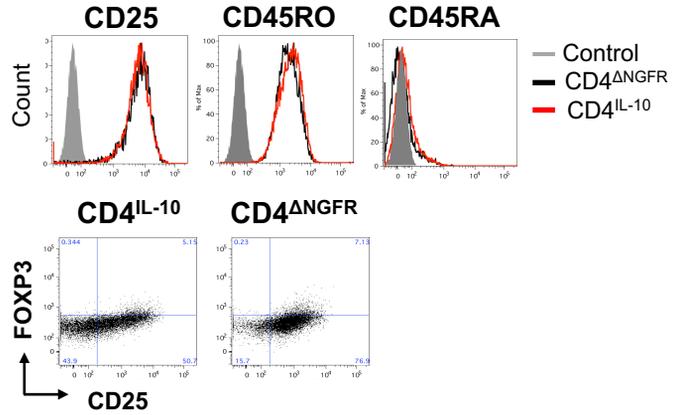
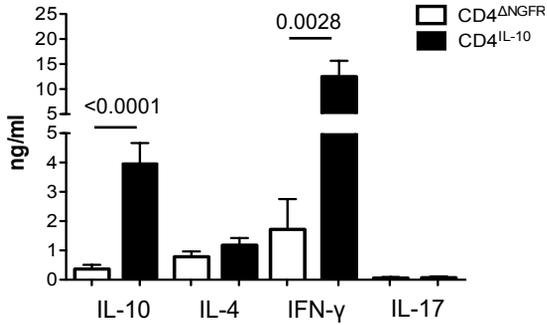
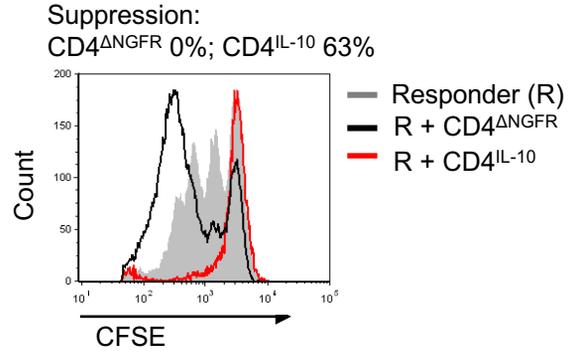


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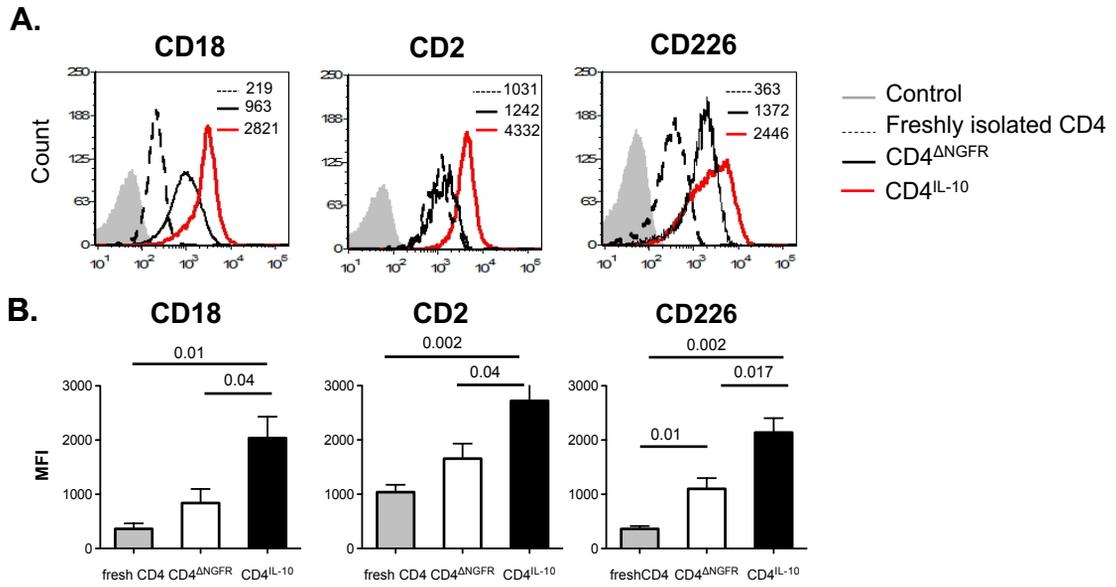
IL-10-Engineered Human CD4⁺ Tr1 Cells Eliminate Myeloid Leukemia in an HLA Class I-Dependent Mechanism

Grazia Locafaro, Grazia Andolfi, Fabio Russo, Luca Cesana, Antonello Spinelli, Barbara Camisa, Fabio Ciceri, Angelo Lombardo, Attilio Bondanza, Maria Grazia Roncarolo, and Silvia Gregori

A.**B.****C.****D.**

Locafaro G. and Andolfi G. et al., Figure S1

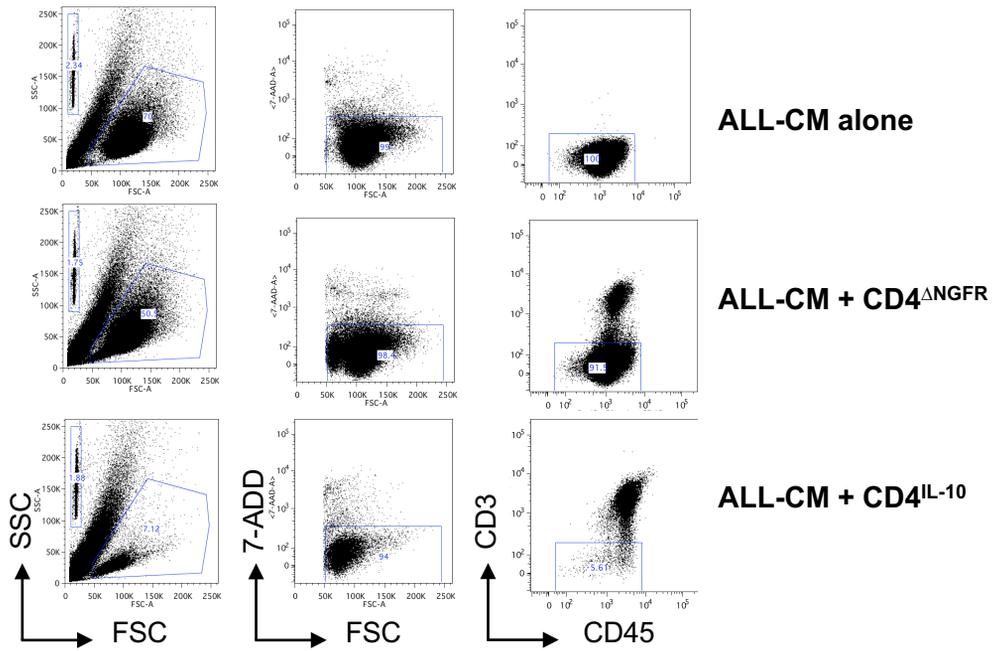
Figure S1. CD4^{IL-10} cells are phenotypically and functionally super-imposable to Tr1 cells. **A.** Scheme of the LV-IL-10/ Δ NGFR and LV-GFP/ Δ NGFR vectors. The presence of the bidirectional promoter (mCMV/PGK) allows the co-regulated expression of Δ NGFR and human IL-10 or GFP genes. Ψ , encapsidation signal including the 5' portion of GAG gene (GA); RRE, Rev-responsive element; cPPT, central poly-purine tract; polyA, poly-adenylation site from the Simian Virus 40; CTE, constitutive transport element; WPRE, woodchuck hepatitis virus post-transcription regulatory element. **B.** CD4^{IL-10} cells are CD25⁺, but not FOXP3⁺ memory T cells. CD4^{IL-10} and CD4 ^{Δ NGFR} cells were analyzed for the expression of CD25, CD45RO, CD45RA, and FOXP3 by FACS after gating on the CD4⁺ Δ NGFR⁺ population. One representative donor out of 4 tested is shown. **C.** Cytokine production profile of CD4^{IL-10} and CD4 ^{Δ NGFR} cells. CD4^{IL-10} and CD4 ^{Δ NGFR} cells were stimulated with immobilized anti-CD3 and soluble anti-CD28 mAbs for 48 hours and IL-10, IL-4, IFN- γ and IL-17 levels were measured by ELISA in culture supernatants. All samples were tested in duplicate-triplicate. Mean \pm SEM of n=15 for IL-10 and IL-4, n=12 for IFN- γ , and n=5 for IL-17 of CD4^{IL-10} cells and CD4 ^{Δ NGFR} cells are presented. *P*, Mann Whitney test. **D.** CD4^{IL-10} cells suppress T cell proliferation *in vitro*. Allogeneic PBMC cells were labeled with CFSE and stimulated with immobilized anti-CD3 and anti-CD28 mAbs alone or in the presence of CD4^{IL-10} cells or CD4 ^{Δ NGFR} cells at 1:1 ratio. After 4 days of culture, the percentage of proliferating PBMC was determined by CFSE/eFluor dilution of CD4⁺ Δ NGFR⁻ cells. One representative donor out of six tested is shown. The suppression mediated by CD4^{IL-10} cells or CD4 ^{Δ NGFR} cells was calculated as follows: ([proliferation responder-proliferation transduced]/proliferation responder] x 100).



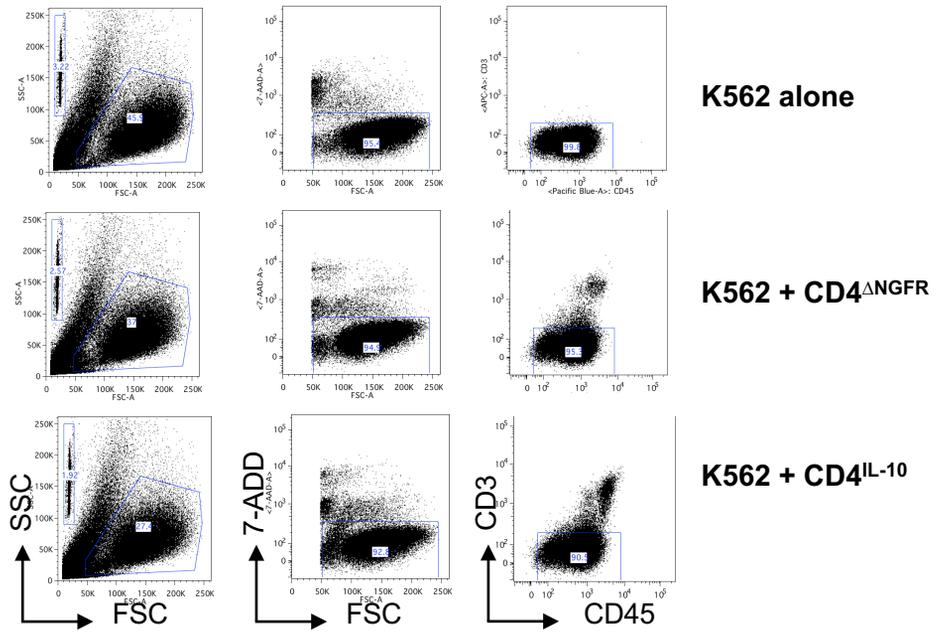
Locafaro G. and Andolfi G. et al., Figure S2

Figure S2. CD4^{IL-10} cells express CD2, CD226 and CD18. CD4^{IL-10}, CD4^{ΔNGFR}, freshly isolated CD4⁺ T cells were analyzed for the expression of CD18, CD2, and CD226 by FACS after gating on the CD4⁺ ΔNGFR⁺ population. **A.** One representative donor out of 6 tested is shown. Numbers indicate the MFI. **B.** Mean ± SEM of 9 donors tested is shown. *P*, Mann Whitney test for comparison between CD4^{IL-10} and CD4^{ΔNGFR} cells, unpaired T cells for comparison between CD4^{IL-10} or CD4^{ΔNGFR} and freshly isolated CD4⁺ T cells.

A.

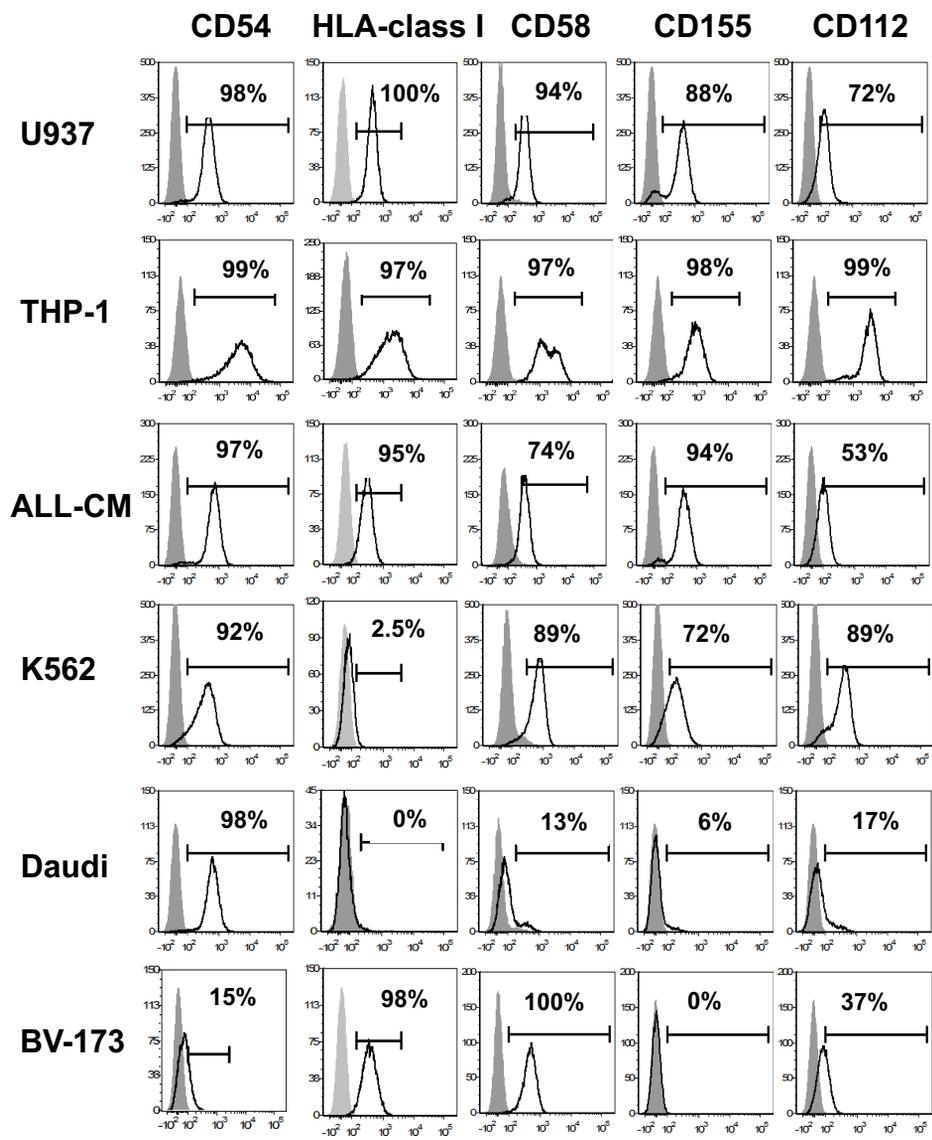


B.



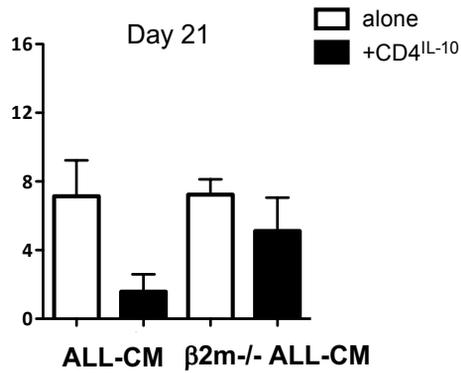
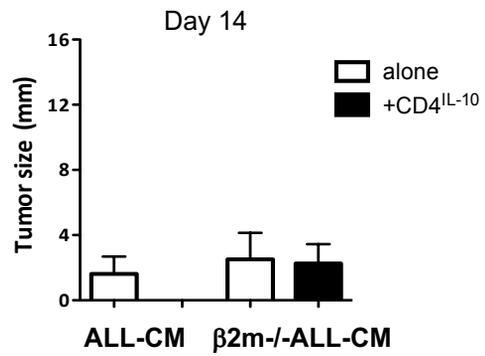
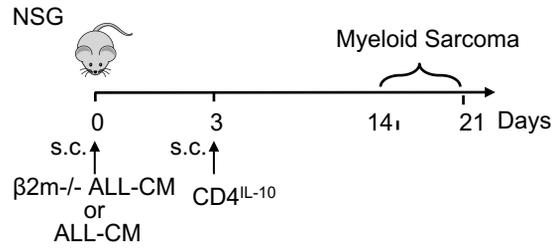
Locafaro G. and Andolfi G. et al., Figure S3

Figure S3. Gating strategy to define residual leukemic cell lines after co-culture experiments. CD4^{IL-10} and CD4^{ΔNGFR} cells were co-cultured with ALL-CM (**A**) or K562 cell lines (**B**) at 1:1 ratio for 3 days. Residual leukemic cell lines (CD45^{low}CD3⁻) were analyzed and counted by FACS according to the gating strategy depicted. One representative donor is shown.



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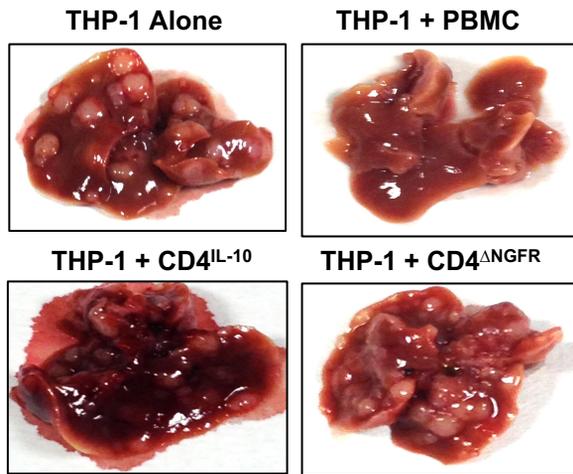
Figure S4. CD4^{IL-10} cells specifically kill *in vitro* leukemic cell lines expressing CD54, HLA-class I, CD58, CD155, and CD112. The expression of the CD54, HLA-class I, CD58, CD155, and CD112 on U937, THP-1, ALL-CM, K562, Daudi and BV-173 cell lines was analyzed by FACS. Numbers indicate the percentages of positive cells (black solid line) according to isotype control (filled light gray histogram). Data representative of at least three independent experiments are shown.



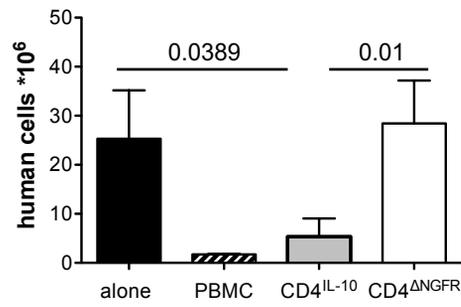
Locafaro G. and Andolfi G. et al., Figure S5

Figure S5. CD4^{IL-10} cells do not kill HLA-class I deficient cells *in vivo*. NSG mice were sub-cutaneously injected with $\beta 2m^{-/-}$ ALL-CM or ALL-CM (2×10^6 cells/mouse) and on day 3, mice received allogeneic CD4^{IL-10} cells (1×10^6 cells/mouse). Tumor growth was measured at the indicated time points. Data obtained from one experiment are presented.

A.



B.



Locafaro G. and Andolfi G. et al., Figure S6

Figure S6. CD4^{IL-10} cells mediate anti-tumor activity in a model of THP-1 extra-medullary tumor. NSG mice were intravenously injected with THP-1 leukemia cells (2×10^6 cells/mouse) and 14 days later mice received allogeneic PBMC (2×10^6 cells/mouse), CD4^{IL-10} or CD4^{ANGFR} cells (1×10^6 cells/mouse). A. Liver cistis in control untreated and PBMC-, CD4^{IL-10}- and CD4^{ANGFR}-treated mice. One representative liver of n=12 mice for THP-1, n=3 mice THP-1+PBMC, n=8 mice for THP-1+CD4^{IL-10} cells, and n=7 mice for THP-1+CD4^{ANGFR} cells. B. Absolute number of human THP-1 cells infiltrating the liver of mice. Mean \pm SEM of n=9 mice for THP-1, n=3 mice THP-1+PBMC, n=7 mice for THP-1+CD4^{IL-10} cells, and n=7 mice for THP-1+CD4^{ANGFR} cells. Data obtained from two independent experiments are presented; *P*; two-sided Mann Whitney test.

Table S1. Polyclonal CD4^{IL-10} cells recovery

	CD4^{ΔNGFR}	CD4^{IL-10}
% of transduction (n=23)	44.5±12.8	51.3±12.5
% of recovery after beads selection (n=8)	44.8±15	57±19
Fold increase after 1 feeder (n=6)	3.7±0.7	2.3±0.6
Fold increase after 2 feeders (n=3)	5.2±0.7	3.0±0.7

Mean±StD are presented

Table S2. Allo-CD4^{IL-10} cells recovery

	CD4^{ANG+K}	CD4^{IL-10}
% of transduction (n=24)	46±11	52.5±13.8
% of recovery after beads selection (n=10)	52.2±18	45±9.9
Fold increase after 1 feeder (n=6)	5±4.3	2.1±1
Fold increase after 2 feeders (n=6)	3.9±2	3.8±1.9

Mean±StD are presented