

Supplementary Figure 1: (A) Primary human airway epithelial cells or (B) MAECs were treated with 50 ng/ml IFN-v or medium alone and protein lysates were analyzed for changes in the expression of Bak by Western blot. (C, D) Primary colon epithelial cells were isolated from C57BL/6 mice. Cells were plated on 6-well plates and infected with 100 MOI Ad-Bik^{L61G} or Ad-Bik. (C) Protein lysates analyzed for Bak expression levels by Western blotting. (D) Cells were quantified 24 h later. MAECs isolated from (E) $stat^{+/+}$ or $stat^{-/-}$ or (F) $noxa^{+/+}$ or $noxa^{-/-}$ mice treated with 50 ng/ml murine recombinant IFN-y or left untreated as controls and protein lysates were analyzed for Bak expression 24 h after treatment by Western blot. (G) $bik^{+/+}$ and $bik^{-/-}$ MAECs were treated with 50 ng/ml IFN-y or medium alone and Bak mRNA levels were analyzed 24 h later by gRT-PCR. HAECs treated with IFN-v or medium alone as controls in the presence of 20 µM pan-caspase inhibitor Q-VD-OPh for 24 h (H) or 72 (I) were analyzed for active Bak (Ab-1)-specific fluorescence by FACS. (J) HAECs infected with 100 MOI Ad-Bik or Ad-Bik^{L61G} for 24 h, the ER fraction of the protein lysates were analyzed for active Bak by Western blot. (K) Representative photomicrographs of HAECs treated with or without 50 ng/ml IFN- γ for 48 h in the presence of 20 μ M pan-caspase inhibitor Q-VD-OPh, fixed with paraformaldehyde and immunostained for activated Bak (Ab1) and calnexin. (L) Representative photomicrographs of HAECs infected with 100 MOI Ad-Bik or Ad-Bik^{L61G} for 24

h, fixed with paraformaldehyde and immunostained for activated Bak (Ab1) and calnexin. **(M)** HAECs stably expressing shCtr or shBik were treated with 50 ng/ml IFN- γ for 48 h. Protein lysates were analyzed for knockdown of Bik by Western blot. $bak^{+/+}/bax^{+/+}$ or $bak^{-/-}bax^{-/-}$ HCT116 cells were treated with medium alone or 50 ng/ml human recombinant IFN- γ for 48 h **(N)** or infected with 100 MOI Ad-Bik^{L61G} or Ad-Bik for 24 h **(O)**. Cells were quantified and protein lysates were analyzed by Western blot. **(P)** MAECs from $bak^{+/+}$ or $bak^{-/-}$ were infected with 100 MOI Ad-Bik^{L61G} or Ad-Bik for 24 h **(O)**. Cells were quantified and protein lysates were analyzed by Western blot. **(P)** MAECs from $bak^{+/+}$ or $bak^{-/-}$ were infected with 100 MOI Ad-Bik for 4, 8, or 16 h and stained with (Ca²⁺)m indicator, Rhod-2 (Red), counterstained for nuclei with DAPI (blue) and the percentage of cells with Rhod-2 positivity were quantified. Percent localization for each experiment in K and L were calculated by counting 200 cells. Scale bar, 5 μ m. Differences between two groups were assessed for significance by Student's t test. ANOVA was used to perform pair-wise comparison of data from more than two groups followed by Fisher least significant difference test. Error bars indicate \pm SEM, *=P<0.05; **=P<0.01.



Supplementary Figure 2: (A) HAECs were treated with 50 ng/ml human recombinant IFN- γ for 72 h and analyzed for annexin V positivity by FACS. **(B)** HAECs were infected with Ad-Bik or Ad-Bik^{L61G} and 24 h later analyzed for Annexin V positivity by FACs. Differences between two groups were assessed for significance by Student's t test. Error bars indicate ± SEM, *=P<0.05; **=P<0.01.



Supplementary Figure 3: (A) H1299 cells stably expressing Cyt C were infected with 100 MOI Ad-Bik or Ad-Bik^{L61G} or treated with 50 ng/ml IFN- γ or medium alone. Cells were analyzed for Cyt C release at 8 or 18 h post treatment by fluorescent microscopy. (B) HAECs stably expressing shCtr for Bik or shBik were infected with 100 MOI of Ad-Bik for 18 h. Protein lysates were fractionated into mitochondrial pellets and cytosolic supernatants and subjected to Western blotting. Cell lysates prepared from HAECs treated with 50 ng/ml human IFN-y or left NT (C) or infected with 100 MOI Ad-Bik or Ad-Bik^{L61G} for 24 h (D) were probed for cleaved caspase 3 by Western blotting. (E) Protein lysates from MTEC, MEF, Thymus, spleen and lung were analyzed for the expression of DAPk1 by Western blot. (F) Protein lysates from HAECs, H1975, Calu-6, H4006, H23, H2009, HT47D and MCF-7 cell lines were compared for the expression level of DAPk1 by Western blot. (G) Cells were infected with 100 MOI of Ad-Bik or Ad-Bik^{L61G} and analyzed for cell viability 24 h later. For all experiments n=3 independent experiments. (H) HAECs stably expressing shCtr or shDAPk were analyzed for Annexin Vpositivity by FACs 24 h after infection with100 MOI Ad-Bik. Differences between two groups were assessed for significance by Student's t test. ANOVA was used to perform pair-wise comparison of data from more than two groups followed by Fisher least significant difference test. Error bars indicate \pm SEM, n=5; *=p<0.05; **= p<0.01.



Supplementary Figure 4: (A) HAECs were infected with Ad-Bik or Ad-Bik^{L61G} for 6, 8, or 16 h. Cells were stained with Fluo-4 and ER-calcium release was analyzed using fluorimeter. (B) HAECs stably expressing shCTR or shBik were treated with 50ng/ml IFN-y for the indicated time point and stained with Ca²⁺-flux indicator, Fluo-4 (green) and counterstained for nuclei with DAPI (blue) and the percentage of cells with Fluo-4-positivity were quantified. (C) HAECs stably expressing shCtr or shBik were infected with Ad-Bik for the indicated time point and stained with Fluo-4 (green) and counterstained for nuclei with DAPI (blue) and the percentage of cells with Fluo-4-positivity were quantified. (D) HAECs stably expressing shBik or shCtr were infected with Ad-Bik for 6, 8, or 12 h and ER-calcium releases were analyzed using fluorimeter. (E) HAECs stably expressing shCtr or shBik were infected with Ad-Bik for 2, 4, and 8 h, stained with and [Ca²⁺]_m indicator Rhod-2 (red), counterstained for nuclei with DAPI (blue), and the percentage of cells with Rhod-2-positivity were quantified. Graphs show mean ± SEM of quantified cells with released Ca²⁺ (Rhod-2 positive cells (n=3, with >200 cells analyzed per condition. (F) Representative micrographs of shCtr- and shDAPk1-expressing HAECs treated with 50 ng/ml recombinant human IFN- γ or medium alone (NT) as control. Cells were stained with anti-CoxIV for mitochondria (green), with anti-calnexin for ER (red), and with DAPI for nuclei staining (blue). Confocal images of 3D reconstructions of ER and mitochondria were acquired. Scale bars, 5 µm. ER-mitochondria contacts were guantified by Manders' coefficient, means and SEM (n=5, at least 40 cells per experiment) of morphometric data. (n=5, with >200 cells analyzed per condition). For all experiments n=3 independent experiments. Differences between two groups were assessed for significance by Student's t test. ANOVA was used to

perform pair-wise comparison of data from more than two groups followed by Fisher least significant difference test. Error bars indicate \pm SEM, *=p<0.05; **=p<0.01.



Supplementary Figure 5: (A) Sequence composition of FAM-Bik BH3 and its L61G mutant peptides screened for their effect in causing cell death in different cell lines. "X" denotes the position of the non natural amino acid with (S)-2-(2'-pentenyl) Ala residues within each peptide. NHBEs (AALEB, N3, HBEC2) and EGFR mutant cancer cell line (H1975) were plated in 24-well plates and treated with 5 µM scramble, DHS-Bik^{L61G}, SHS-Bik2 or DHS-Bik^{WT} peptides. Cells were harvested and quantified 72 h later. (B) AALEB cells were seeded on 6-well plates and treated with 0, 1.0, 2.5, 5.0, or 10 µM DHS-Bik^{WT} BH3 helix and guantified 72 hours later. There was no significant difference in sensitivity of the cells between 2.5, 5.0, and 10 µM treatments. (C) Primary normal human bronchial epithelial cells (NHBEs) from five donors were plated in 24well plates and treated with 5 µM of single (SHS-Bik1^{WT}, SHS-Bik2^{WT}), double (DHS-Bik^{WT}, DHS-Bik^{L61G}), or scramble hydrocarbon stapled Bik BH3 helixes. Cells were harvested and quantified 72 hours later. DHS-Bik^{WT} was found to be the most effective in reducing cell number. (D) Cultured HAECs were treated with vehicle, FAM-Bik^{L61G} or FAM-Bik^{WT} BH3 peptides (1 µM) for 2 hours, followed by centrifugation, trypsinization, washing, lysate preparation, electrophoresis, and fluorescence detection. (E) HAECs treated with 5 µM of FAM-DHS-Bik^{WT} or FAM-DHS-Bik^{L61G} peptides for 3 h were analyzed for active Bak (Ab1-specific) fluorescence by FACS. (F, G) C57BL/6 mice were exposed to 250 mg/m³ CS 6 h/d, 5 d/wk for 3 weeks and instilled with 5 uM of FAM-DHS-Bik^{WT} or FAM-DHS-Bik^{L61G} peptides in 50ul of PBS on 2

consecutive days and lungs harvested the following day. The number of epithelial cells (**F**) Muc5ac-positive cells per mmBL and (**G**) were significantly reduced in the airways of mice instilled with FAM-DHS-Bik^{WT} compared with those instilled with FAM-DHS-Bik^{L61G} or PBS. (**H**) Differentiated HAECs were treated with 4000 ng/ml CSE for 48 h followed by treatments with 5 uM of FAM-Bik^{WT} or FAM-DHS-Bik^{L61G} peptides for 48 h before harvest. FAM-DHS-Bik^{WT} peptides reduce MUC5AC mRNA levels and percentage of MUC5AC-positive cells in CS-treated cells as analyzed by qRT-PCR and immunostaining, respectively. FAM-DHS-Bik^{WT} reduced MUC5AC positivity significantly compared to FAM-DHS-Bik^{L61G}. (n = 5 individuals/group, done in triplicate). Differences between two groups were assessed for significance by Student's t test. ANOVA was used to perform pair-wise comparison of data from more than two groups followed by Fisher least significant difference test. Error bars indicate ± SEM, Graphs and error bars show means±SEM, *=p<0.05; **= p<0.01.

Supplementary Figure 6. Uncropped images related to Figure 1



Figure 1A



Figure 1B

Figure 1D

Figure 1G

Figure 1K

Supplementary Figure 7. Uncropped images related to Figure 2

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Figure 2E

Supplementary Figure 8. Uncropped images related to Figure 3

Figure 3A

Figure 3C

Figure 3D

Figure 3F

Figure 3G

Figure 3I

Figure 3J

Figure 3L

Figure 3M

Supplementary Figure 9. Uncropped images related to Figure 6

Figure 6B

Figure 6C

Supplementary Figure 10. Uncropped images related to Supplementary Figure 1

Supplementary Figure 1a

Supplementary Figure 1b

Supplementary Figure 1e

Supplementary Figure 1f

Supplementary Figure 11: Uncropped images related to Supplementary Figure 3

Supplementary Figure 3b

Supplementary Figure 3d

Supplementary Figure 3f

SHS-Bik1^{wT}

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Supplementary Figure 12: High-performance liquid chromatography (HPLC) and Mass spectrometry (MS) data for single hydrocarbon stapled wild-type Bik1 (SHS-Bik1^{WT}) peptide.

SHS-Bik2^{WT}

Supplementary Figures 13: High-performance liquid chromatography (HPLC) and Mass spectrometry (MS) data for single hydrocarbon stapled wild-type Bik1 (SHS-Bik2^{WT}) peptide.

Scrambled Peptide

Supplementary Figures 14: High-performance liquid chromatography (HPLC) and Mass spectrometry (MS) data for single hydrocarbon stapled scramble peptide.

DHS-Bik1^{wT}

Supplementary Figures 15: High-performance liquid chromatography (HPLC) and Mass spectrometry (MS) data for single hydrocarbon stapled wild-type Bik1 (DHS-Bik1^{WT}) peptide.

DHS-Bik2^{L61G}

Supplementary Figures 16: High-performance liquid chromatography (HPLC) and Mass spectrometry (MS) data for single hydrocarbon stapled L61G mutant Bik2 (DHS-Bik2^{WT}) peptide.