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Supplemental Information

**In Vivo Production of Monoclonal Antibodies
by Gene Transfer via Electroporation Protects
against Lethal Influenza and Ebola Infections**

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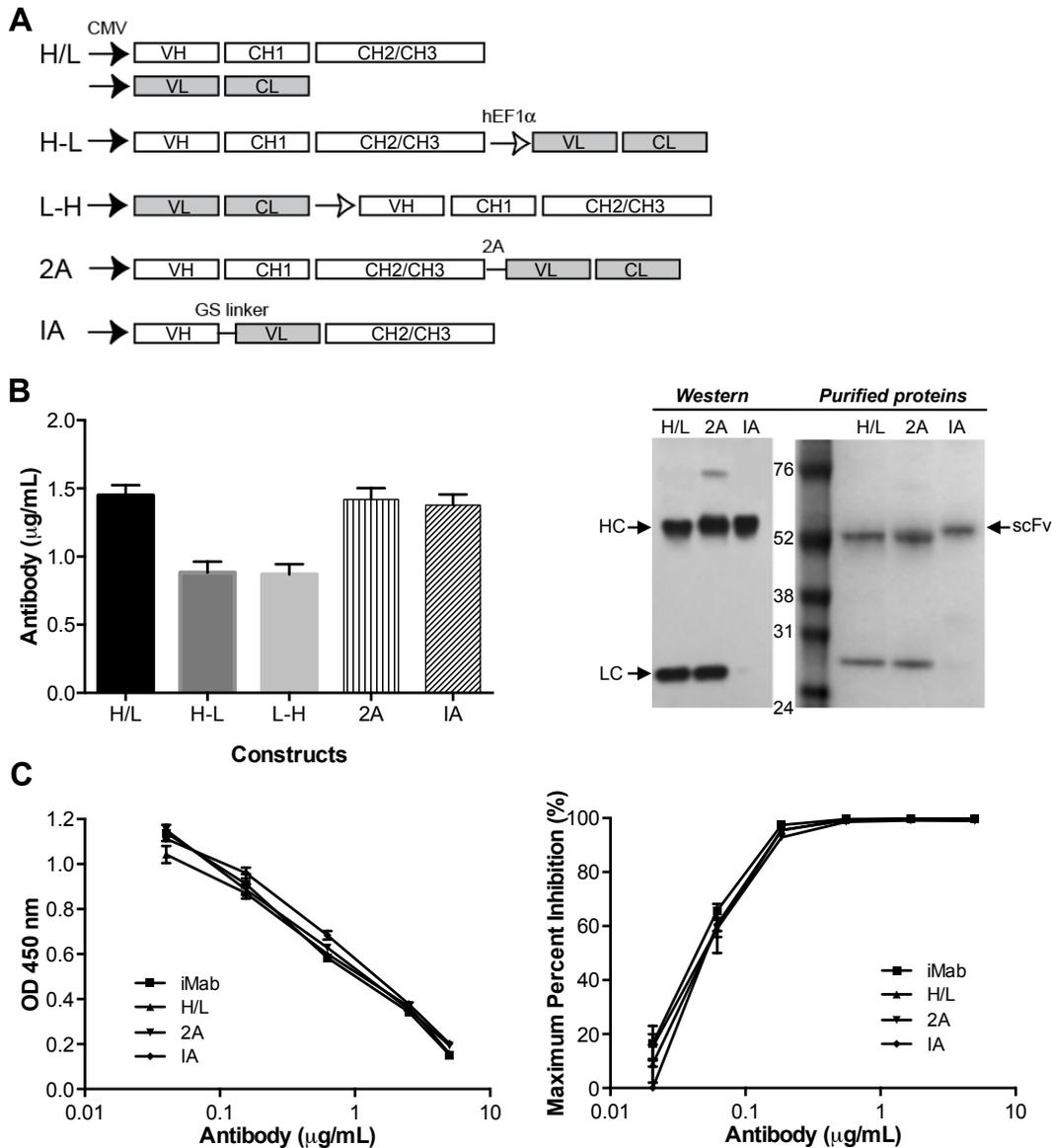


Figure S1

5A8 expression cassettes and *in vitro* characterizations.

(A) Schematic representation of mAb transgene configurations (heavy chain genes shown in white and light chain shown in gray) in the pVAX1 vector. Cytomeglovirus (CMV) and human elongation factor-1 alpha (hEF1 α) promoters are indicated by closed black or open white arrows, respectively. The 2A cassette contains H- and L-chain genes separated by a furin cleavage site coupled with a P2A self-processing peptide (2A). The immunoadhesin (IA) contains a single-chain variable fragment (scFv) fused to the Fc region. (B) Left panel: *In vitro* 5A8 expression in supernatant 72 hours after pDNA transfection of 293A cells measured by CD4 binding ELISA (mean \pm SEM). Right panel: Western blot of cell culture supernatants detected by anti-mouse HRP and mAbs purified from supernatants were resolved by SDS-PAGE and visualized with Coomassie Blue. The faint ~80kDa band detected in the 2A samples by Western blot is unprocessed antibody. Molecular weight ladder represents kDa. (C) Left panel: Purified mAbs compete with iMab-HRP for sCD4 binding. Right panel: Purified mAbs neutralize HIV-1 pseudotyped virus tested in TZM-bl cells.

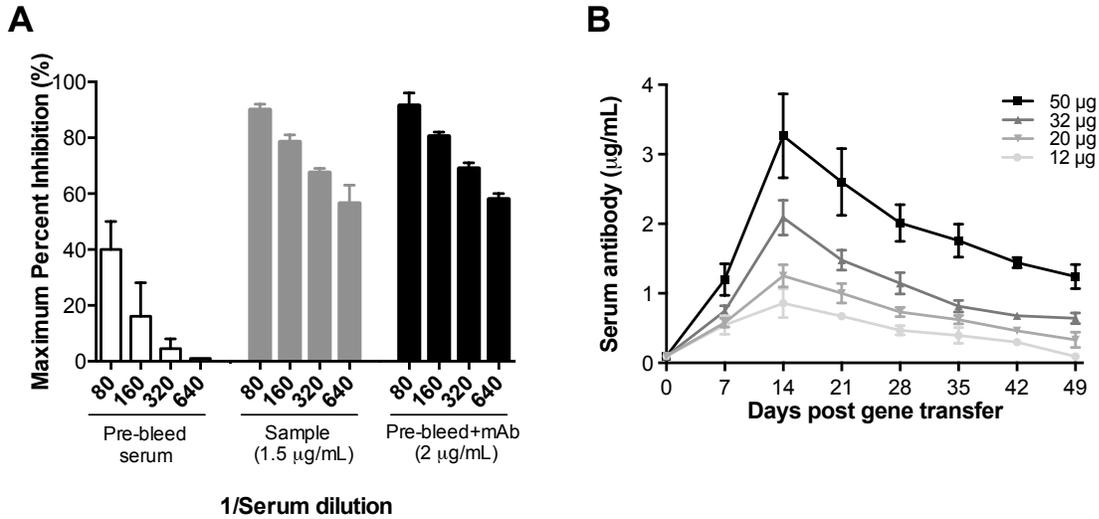


Figure S2

EP delivery of pDNA results in functional dose-dependent serum mAb in mice.

(A) Mice were administered 50 µg H/L 5A8 pDNA in the pVAX1 vector followed by EP. To verify functional activity of the expressed mAb, maximum percent inhibition (MPI) measured by pseudotyped HIV-1 neutralization in TZM-bl cells was compared between serially diluted pre-bleed serum, serum samples taken after DNA/EP, and pre-bleed serum mixed with *in vitro* produced mAb. Mouse serum prior to gene transfer (pre-bleed) was evaluated for antibody neutralization of HIV-1, showing non-specific activity at low serum dilutions. Endogenously expressed mAb retains functional activity comparable to *in vitro* produced mAb. (B) mAb expression is pDNA dose-dependent. BALB/c mice (n=6) were injected with 12, 20, 32 or 50 µg of H/L 5A8 pDNA in the pVAX1 vector followed by EP. Serum was collected weekly and mAb concentration was determined by sCD4 binding ELISA. All data are presented as mean ± SEM.

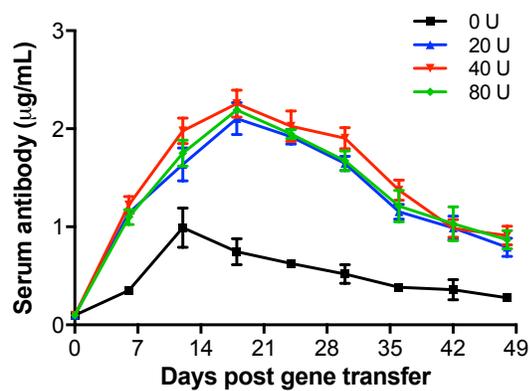


Figure S3

Hyaluronidase pretreatment enhances mAb expression.

BALB/c mice (n=4-5) were pretreated with 20, 40 or 80 U of hyaluronidase or saline (0 U) as a control 2 hours prior to administration of 20 µg of H/L 5A8 pDNA in the pVAX1 vector followed by EP. Serum was collected weekly and mAb concentration was determined by sCD4 binding ELISA. Antibody expression was evaluated by area-under-the-curve measurements during the 48-day period studied. All data are presented as mean ± SEM.

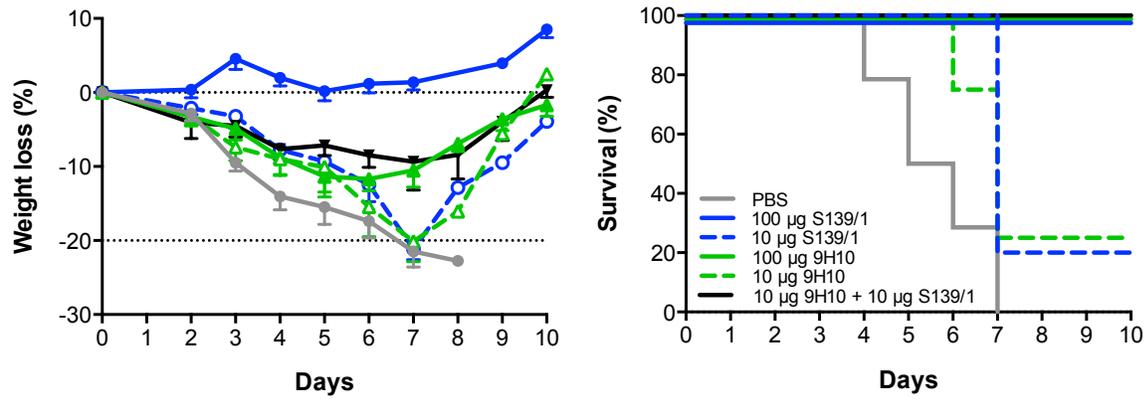


Figure S4

Passive administration of anti-influenza mAbs in mice defines the partially protective dose against influenza challenge.

BALB/c mice (n=3-5) were passively administered mAb via the intraperitoneal route one day prior to influenza challenge. Mice were challenged on day 0 with 13 MLD₅₀ of H3N2 (A/Aichi/2/68) delivered intranasally. Left panel: mean weight loss (%) ± SEM compared to day 0. Right panel: survival (%) as depicted in Kaplan-Meier plots following challenge.

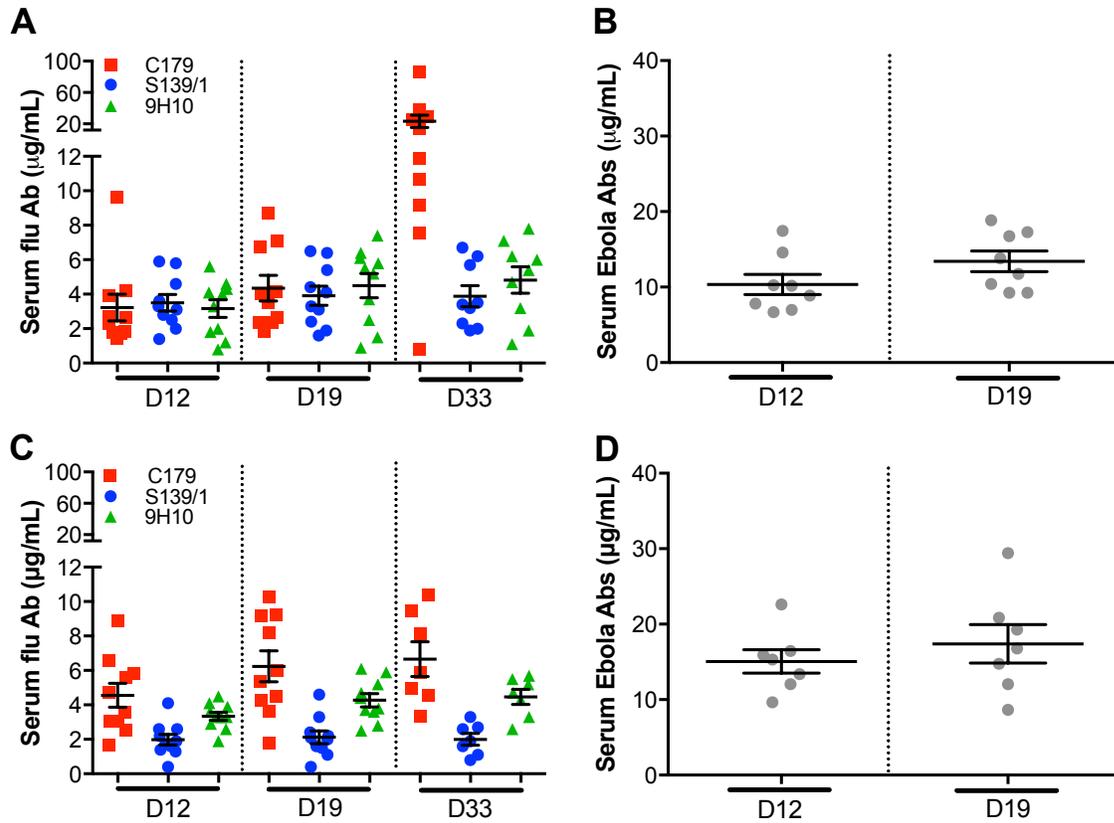


Figure S5

Serum mAb concentrations from mice challenged with group 1 or group 2 influenza viruses.

Mice were administered pDNA followed by EP and challenged as described in Fig. 3. mAb expression from mice challenged with (A-B) H1N1 and (C-D) H3N2. (A, C) Individual mAb concentrations for C179, S139/1 and 9H10. (B, D) Total anti-Ebola mAb concentrations. Each symbol represents an individual animal and error bars indicate mean \pm SEM.

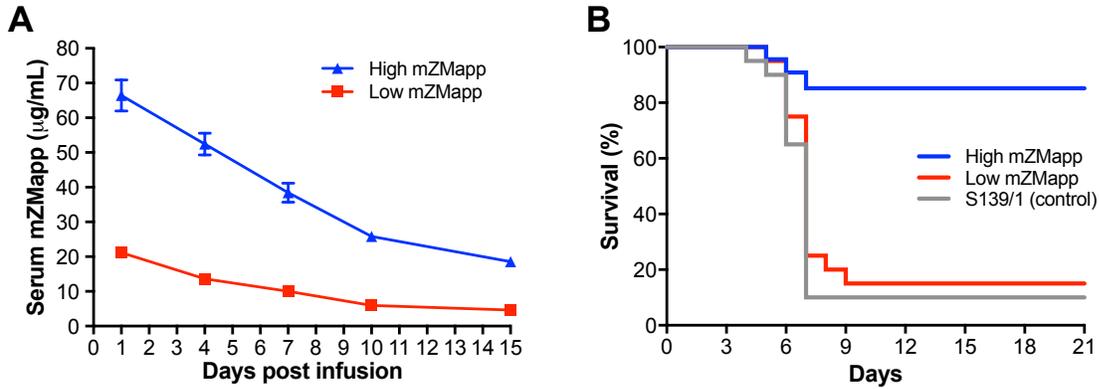


Figure S6

Passive infusion of high-dose mZMapp protects mice from Ebola challenge.

(A) The pharmacokinetic properties of mZMapp were initially evaluated in BALB/c mice. Mice (n=6/group) were infused via the intraperitoneal route with high dose mZMapp (40 µg/mAb; 120 µg total; ~6 mg/kg total) or low dose mZMapp (10 µg/mAb, 30 µg total; ~1.5 mg/kg total) on day 0. Blood was collected at the indicated time points and total mZMapp concentrations were evaluated by GP binding ELISA. Mean ± SEM are shown. (B) In a separate experiment, mice (n=20/group) were infused with a low or high dose of mZMapp or an anti-influenza mAb, S139/1 (30 µg total) as a control, one day prior to intraperitoneal challenge with 100 PFU mouse-adapted Ebola virus. Mice were monitored for lethality for 21 days. The Kaplan-Meier plot depicting survival is shown.

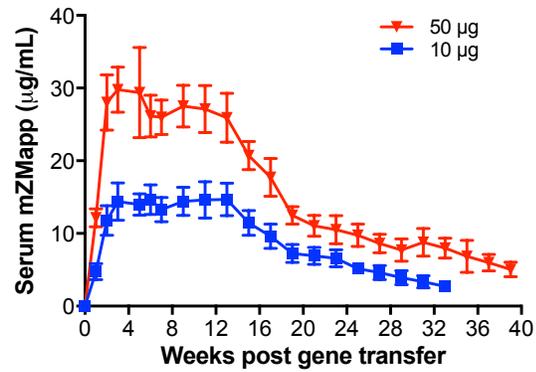


Figure S7

Duration of mZMapp expression following pDNA/EP administration.

BALB/c mice (n=6-9) were pretreated with 20 U of hyaluronidase and administered 10 or 50 µg H/L gWiz pDNA encoding each 13C6, 2G4 and 4G7 at distinct sites followed by EP. Total serum mAb concentrations were measured by GP-binding ELISAs. All data are represented as mean ± SEM.

Supplemental Methods

5A8 / 5A8-like Ab *in vitro* expression

5A8 constructs were evaluated for mAb expression by transient transfection in 293A cells using 5 µg/mL polyethylenimine and 1 µg pDNA. Transfected cells were maintained in Hybridoma-SFM medium for 72 hours. CD4 binding ELISA was used to quantify 5A8 in culture supernatants. Culture supernatants were collected, filtered, purified using Protein G Plus Agarose (Pierce). Eluted mAbs were concentrated by centrifugation (Millipore). mAbs were analyzed by SDS-PAGE visualized with Simply Blue and Western blot detected with anti-mouse HRP.

5A8 assessment for soluble CD4 binding in competition ELISA

In vitro produced 5A8 was assessed for its ability to compete with HRP-labeled iMab for soluble CD4 binding as previously described¹.

HIV-1 pseudovirus-based *in vitro* neutralization assays

Molecular cloned Env-pseudotyped virus G16 was prepared² and neutralization was assessed using TZM-bl cells (NIH AIDS Research and Reference Reagent Program, Division of AIDS, NIAID, NIH) with a luciferase-based assay as previously described¹.

Supplemental References

1. Seaman, MS, Janes, H, Hawkins, N, Grandpre, LE, Devoy, C, Giri, A, *et al.* (2010). Tiered categorization of a diverse panel of HIV-1 Env pseudoviruses for assessment of neutralizing antibodies. *Journal of virology* **84**: 1439-1452.
2. Sun, M, Pace, CS, Yao, X, Yu, F, Padte, NN, Huang, Y, *et al.* (2014). Rational design and characterization of the novel, broad and potent bispecific HIV-1 neutralizing antibody iMabm36. *J Acquir Immune Defic Syndr* **66**: 473-483.