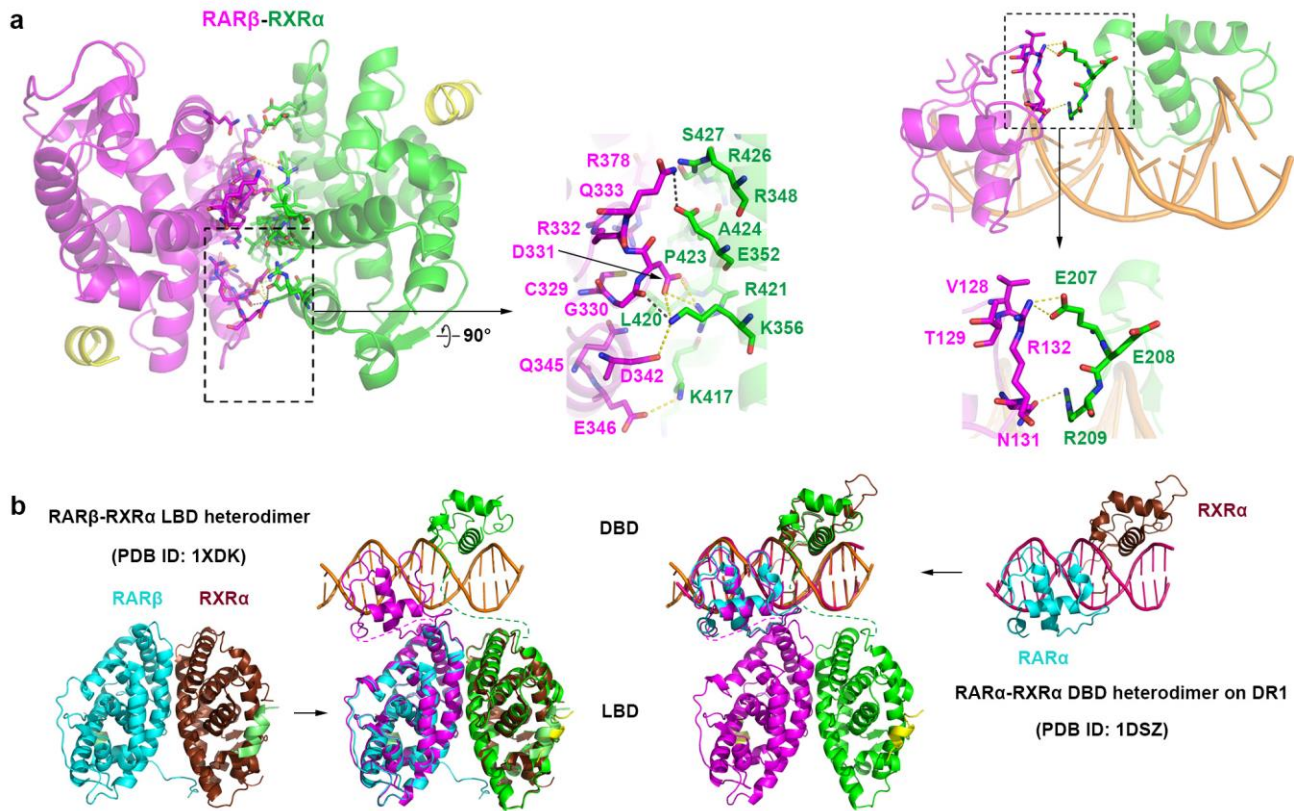
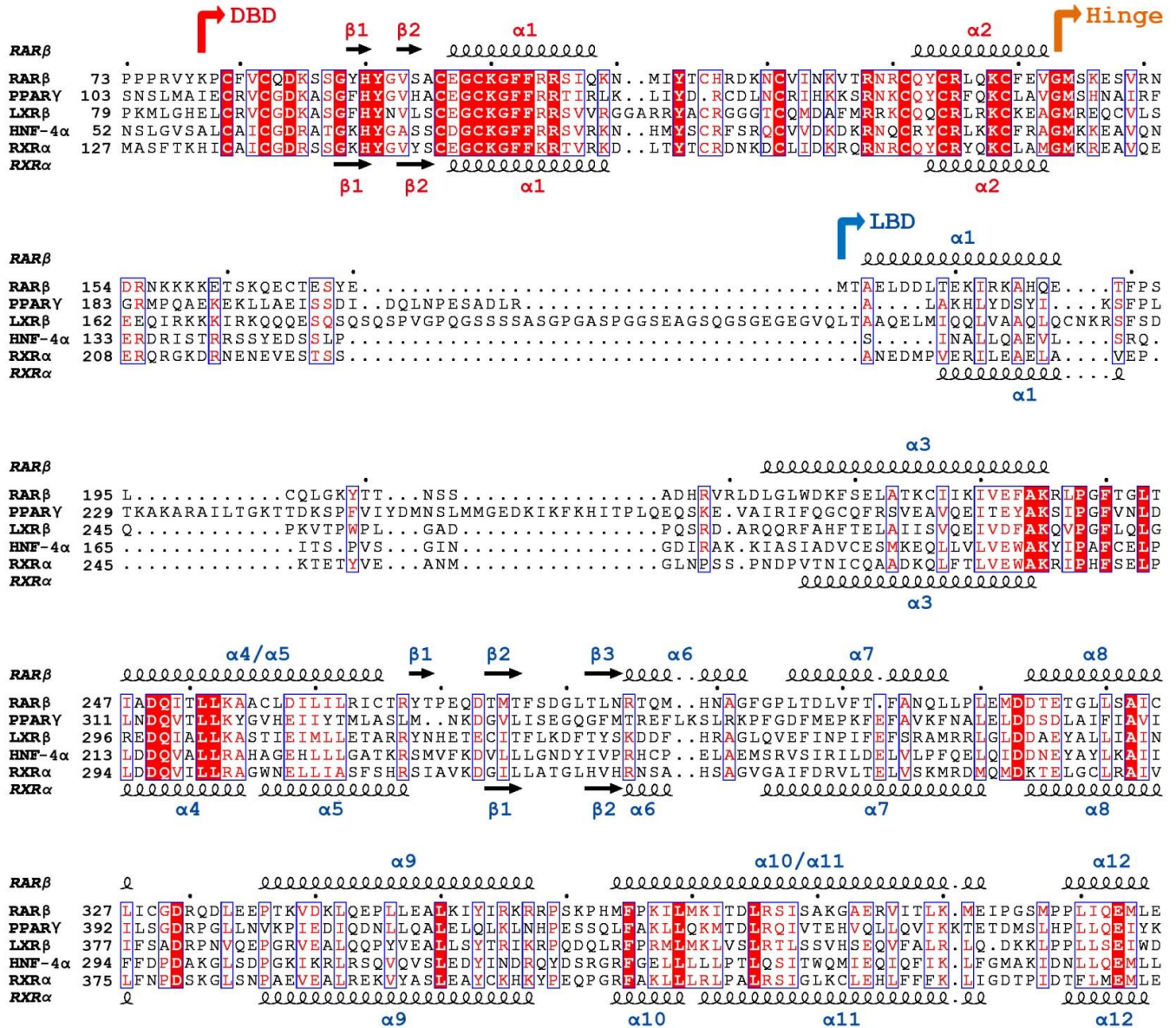


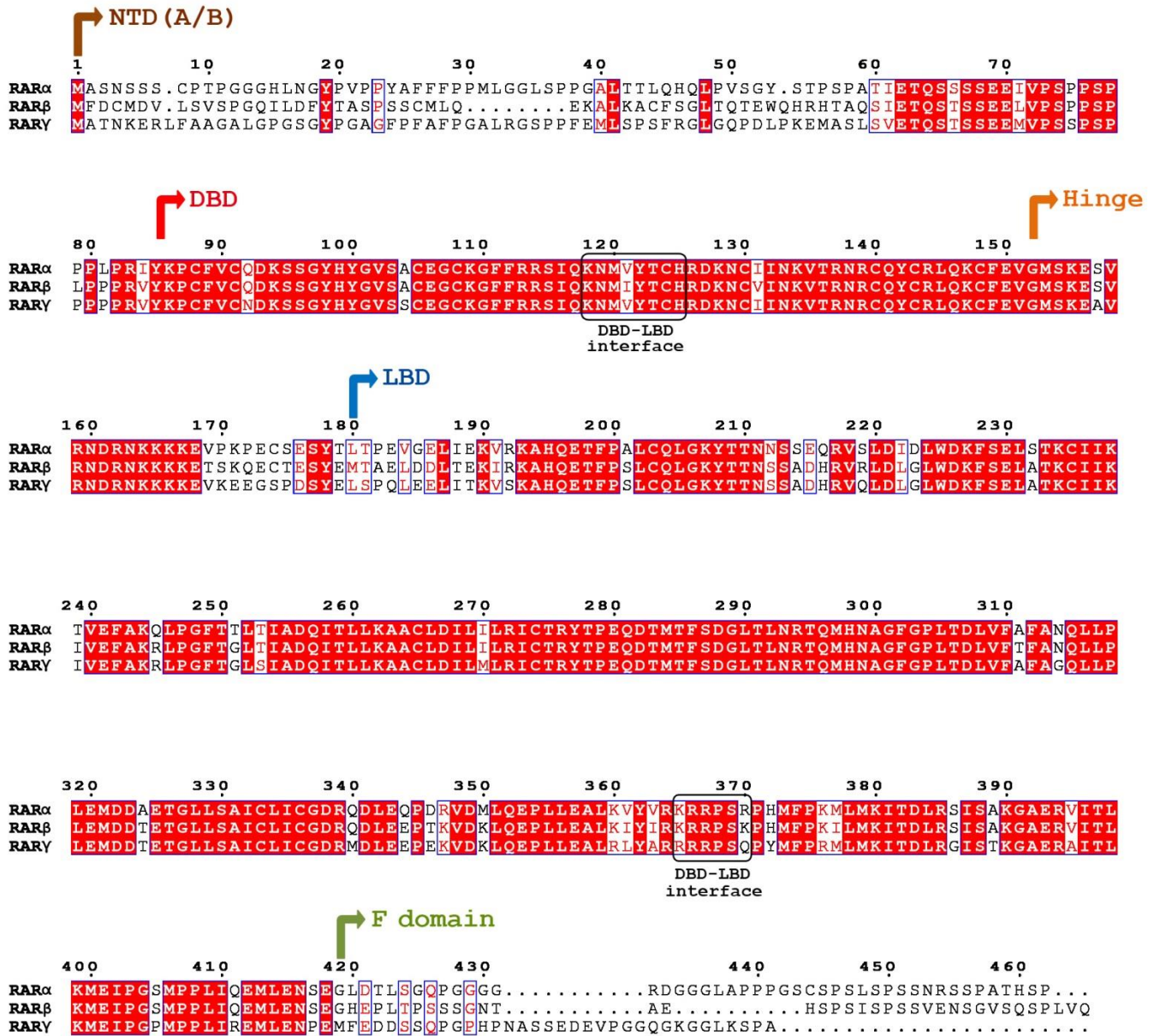
Supplementary Figure 1 Electron density maps of the RAR β -RXR α -DNA complex. (a) $2F_o - F_c$ map contoured at 1.0σ covers the overall RAR β -RXR α -DNA complex in two views. The colors used for RAR β , RXR α , DNA, NCOA2 peptides, REA, 9CR and density are magenta, green, orange, yellow, cyan, blue and gray, respectively. (b and c) The $2F_o - F_c$ maps at 1.0σ (b) and polder OMIT maps at 3.0σ (c) of two ligands REA and 9CR are shown in gray and green colors, respectively.



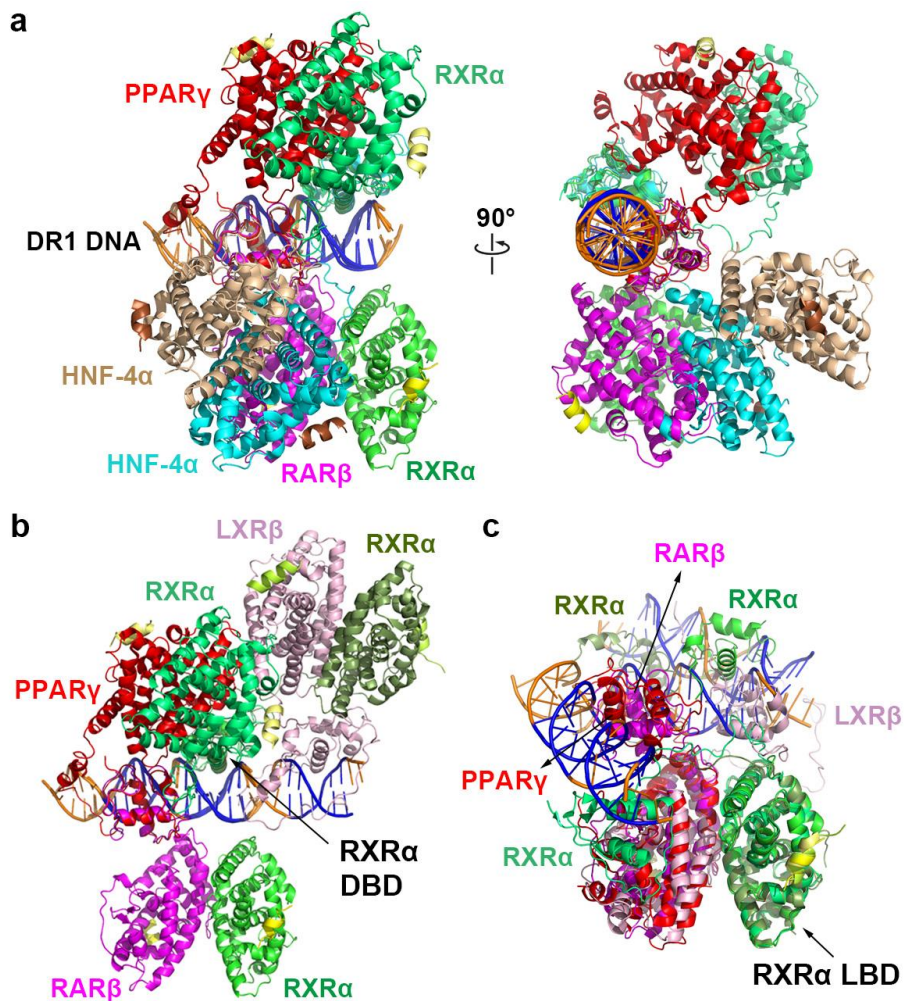
Supplementary Figure 2 Dimerization of the multi-domain RAR β -RXR α complex. (a) The dimerization interfaces between two LBDs (left) and two DBDs (right), with enlarged views showing representative details of the interactions. These interfaces are mainly mediated by salt bridges (yellow dotted lines), hydrogen bonds (gray dotted lines) and hydrophobic interactions. (b) Comparison between multi-domain RAR β -RXR α complex and isolated DBD or LBD heterodimers. Structure superimpositions of RAR β -RXR α -DNA complex with previously published RAR β -RXR α LBD heterodimer (left) and RAR α -RXR α DBD heterodimer on DR1 DNA (right).



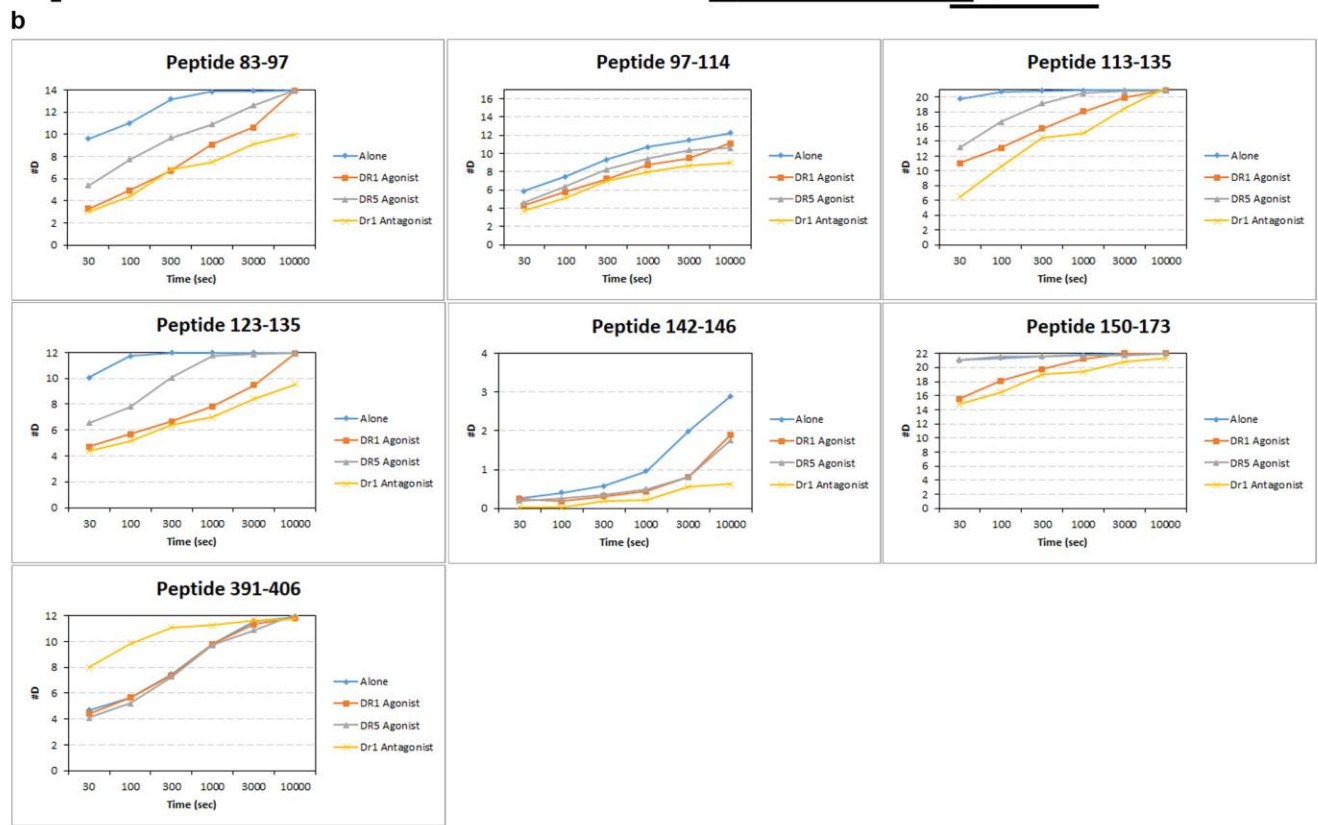
Supplementary Figure 3 Structure-based sequence alignment of human RARβ, PPARγ, LXRβ, HNF-4α and RXRα proteins. The starting positions of DBD, hinge and LBD segments are labelled in red, orange and blue, respectively. The secondary structure components of RARβ are shown above the alignment, while those of RXRα shown below.



Supplementary Figure 4 Sequence alignment of human full-length RAR α , RAR β and RAR γ proteins. The starting positions of NTD (A/B domain), DBD, hinge, LBD and F domain segments are labelled in brown, red, orange, blue and green, respectively. The regions involved in DBD-LBD interface are also indicated by black boxes.



Supplementary Figure 5 Superimpositions of multi-domain NR crystal structures. (a) Superposed RAR β -RXR α heterodimer, PPAR γ -RXR α heterodimer and HNF-4 α homodimer structures by aligning the DR1 DNA sequences in two views. **(b and c)** Superposition of RAR β -RXR α , PPAR γ -RXR α and LXR β -RXR α structures, by aligning the common partner RXR α 's DBD **(b)** or LBD **(c)**, respectively.



Supplementary Figure 6 Peptide coverage map for RAR β in the HDXMS studies (a) and individual uptake curves for selected regions (b).

Supplementary Table 1 Sequences of the primers used in this study.

RXR α cloning primers	Forward	AAAAAACATATGAACCCCGTCAGCAGC
	Reverse	AAAAAACTCGAGCTAAGTCATTTGGTGCGG
RAR β cloning primers	Forward	AAAAAAGCTAGCATGCCTCCCCCTCGAGTGTAC
	Reverse	AAAAAAGCGGCCGCTCATTCATGTCCTTCAGAATTCTC
RAR β E99A mutation primers	Forward	GGGGTCAGCGCCTGTGCTGGATGTAAGGGCTTTTTC
	Reverse	GAAAAAGCCCTTACATCCAGCACAGGCGCTGACCCC
RAR β R106A mutation primers	Forward	TGTAAGGGCTTTTTCGCTAGAAGTATTCAGAAG
	Reverse	CTTCTGAATACTTCTAGCGAAAAAGCCCTTACA
RAR β M113E mutation primers	Forward	AGAAGTATTCAGAAGAATGAAATTTACACTTGTCCCGA
	Reverse	TCGGTGACAAGGTGAAATTTCACTTCTGAATACTTCT
RAR β T116V mutation primers	Forward	CAGAAGAATATGATTTACGTCTGTCACCGAGATAAG
	Reverse	CTTATCTCGGTGACAGACGTAAATCATATTCTTCTG
RAR β K358E mutation primers	Forward	CTAAAAATTTATATCAGAGAAAGACGACCCAGCAAGCCT
	Reverse	AGGCTTGCTGGGTCGTCTTTCTCTGATATAAATTTTAG
RAR β R359E mutation primers	Forward	AAAATTTATATCAGAAAAGAACGACCCAGCAAGCCTCAC
	Reverse	GTGAGGCTTGCTGGGTCGTTCTTTTCTGATATAAATTTT
RAR β R360E mutation primers	Forward	ATTTATATCAGAAAAGAGAACCAGCAAGCCTCACATG
	Reverse	CATGTGAGGCTTGCTGGGTTCTCTTTTCTGATATAAAT
RAR β K358E/R359E/R360E mutation primers	Forward	CTAAAAATTTATATCAGAGAAGAAGAACCAGCAAGCCTCACATG
	Reverse	CATGTGAGGCTTGCTGGGTTCTCTTCTCTGATATAAATTTTAG
RAR β S363E/K363E mutation primers	Forward	AGAAAAAGACGACCCGAAGAACCTCACATGTTTCAAAG
	Reverse	CTTTGAAACATGTGAGGTTCTTCGGGTCGTCTTTTCT