Novel mechanism of regulation of the

5-lipoxygenase/leukotriene B₄ pathway by high-density lipoprotein in macrophages

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Supplemental Method:

Preparation of mice bone marrow derived macrophages (BMDM).

Bone marrow cells were sterilely obtained from femoral bones of C57BL6/J mice and differentiated to macrophages in 10% FBS containing RPMI with supplementation of M-CSF (10 ng/mL) for seven days.

PCR analysis.

Macrophage total RNA was extracted with TRIzol reagent (Invitrogen). cDNA was prepared from 1 µg total RNA by using PrimeScript RT reagent (RR047, Takara). To confirm the expression of BLT1 in RAW macrophages, PCR amplification for mouse BLT1 was performed with and without reverse transcription (RT). Primers for mouse BLT1 were Forward: CCTGGCACTAAGACAGATTCAAGGA, Reverse: CCTGGCACTAAGACAGATTCAAGGA, product size was 152bp. PCR product was separated by electrophoresis with 2% agarose gel.

Isolation of exosomes from plasma.

Exosomes were isolated from plasma with ExoTrap[™] (SHI-EXO-K010, COSMO BIO). Exosomederived proteins were eluted by SDS sample buffer, denatured by heating and applied for western blot analysis with anti-CD9 antibody (12A12, COSMO BIO) and anti-ApoA-I antibody (23030485, CHMEICON INTERNATIONAL)

Supplemental Tables:

Supplementary Table 1. Subject characteristics

| | Healthy group | CAD group |
|---------------------------|-----------------|---------------------|
| Sex | 4M | 3M/1F |
| Age (years) | 32 - 42 | 52 - 69 |
| Serum MPO mass (ng/ml) | 7.2 ± 6.4 | 275.7 ± 17.8* |
| Serum PON1 activity (U/L) | 253.0 ± 40.1 | 120.7 ± 62.7* |
| Serum MPO/PON1 | 0.03 ± 0.02 | $2.89 \pm 1.78^{*}$ |
| EPA/AA in HDL | 0.42 ± 0.02 | 0.25 ± 0.10 |
| DHA/AA in HDL | 0.58 ± 0.02 | 0.56 ± 0.19 |
| DPA/AA in HDL | 0.10 ± 0.02 | 0.12 ± 0.02 |

Values were represented as mean \pm SEM. **P* < 0.05 between two groups.

| | HDL _{Healthy} (pg/0.01mg protein/2h) | HDL _{CAD} (pg/0.01mg protein/2h) |
|------------------------------|--|--|
| | (pg) 0101111g proteini, 211 | (pg/0.01116 protein/211/ |
| AA Bioactive Metabolome | | |
| Lipoxin A ₄ | - | - |
| Lipoxin B ₄ | - | - |
| Prostaglandin D ₂ | 3.5 ± 1.1 | 23.5 ± 8.6* |
| Prostaglandin E ₂ | 2.0 ± 0.2 | 4.4 ± 1.9 |
| Prostaglandin $F_{2\alpha}$ | - | - |
| Thromboxane B ₂ | - | - |
| Leukotriene B ₄ | 0.4 ± 0.1 | 20.0 ± 21.2* |
| AA pathway markers | | |
| 5 HETE | 0.7 ± 0.2 | 7.2 ± 8.8* |
| 12 HETE | 5.9 ± 1.2 | 52.9 ± 33.0 |
| 15 HETE | 4.1 ± 0.7 | 7.8 ± 7.2 |
| EPA pathway markers | | |
| 5 HEPE | 2.6 ± 0.7 | 11.3 ± 0.9* |
| 12 HEPE | 20.5 ± 2.2 | 11.8 ± 4.9 |
| 15 HEPE | 2.3 ± 0.3 | 5.5 ± 3.6 |
| 18 HEPE | 2.3 ± 0.2 | 10.2 ± 4.6 |
| DHA pathway markers | | |
| 4 HDHA | 1.3 ± 0.4 | 5.5 ± 2.0* |
| 7 HDHA | 0.7 ± 0.1 | $2.8 \pm 0.6^{*}$ |
| 14 HDHA | 6.7 ± 1.1 | 8.0 ± 1.9 |
| 17 HDHA | 1.3 ± 0.7 | 5.4 ± 2.1 |

Supplementary Table 2. Lipid mediator biosynthesized from $HDL_{Healthy}$ or HDL_{CAD}

- = below detection limits.

Values are represented as mean \pm SEM. **P* < 0.05 between two groups.



Supplemental Figure 1. HDL_{CAD} enhanced phosphorylation of cPLA2 in macrophages.

Macrophages (1×10⁶) were incubated with vehicle or HDL (Healthy or CAD, 10 µg protein, 30 min at 37 °C). After incubation, 200 µl of lysis buffer (20 mM HEPES (pH 7.4), 150 mM NaCl, 1% NP40, 1% SDS) was added for western blot analysis. The expression levels of phosphorylation of cPLA2 and total cPLA2 were analyzed using an anti-phospho-cPLA2 antibody (2831, Cell Signaling Technology) and cPLA2 (5249, Cell Signaling Technology).



Supplemental Figure 2. HDL_{Healthy}, but not HDL_{CAD}, decreased LTB₄ production from mouse bone marrow-derived macrophages.

Mouse bone marrow-derived macrophages (1 × 10⁶) were activated by ZyA (30 min at 37 °C), then treated with HDL_{Healthy} or HDL_{CAD} (10 µg protein). LTB₄ production from macrophages was quantified by LC/MS/MS. The results are expressed as $pg/1 \times 10^6$ cells, mean ± SEM, N = 3 in each group.





Supplemental Figure 3. Macrophages showed proinflammatory cytokine profiles by HDL_{CAD}.

Macrophages (RAW 264.7 cell line, 1×10⁶) were activated by ZyA (30 min at 37°C), then treated with HDL_{Healthy} or HDL_{CAD} (10 µg protein, 30 min at 37°C). After incubation, total RNA was extracted, then M1 markers (**a**: IL-6, TNF- α , IL-1 β) and M2 markers (**b**: Arg1, Mrc1) were analyzed by real-time PCR. The results are shown as fold change compared to vehicle group; mean ± SEM, N = 3. **P* < 0.05 by post hoc analysis.



Supplemental Figure 4. ApoA-I expression levels in $HDL_{Healthy}$ and HDL_{CAD} .

Each type of HDL was purified by ultracentrifuge. Apo-AI (SLBN8688V, SIGMA-ALDRICH), HDL_{Healthy}, and HDL_{CAD} (10µg protein, respectively) were investigated by western blot analysis with anti-ApoA-I (#23030485, CHEMICON INTERNATIONAL).



LAMP1

Supplemental Figure 5. HDL partially colocalized with lysosomes in macrophages.

Macrophages were incubated with 10 μ g of HDL_{Healthy} at 37°C for 10, 60 and 120 min. The cells were fixed and immunostained with (**a**) anti-EEA1 (shown as red) and anti-human apoA-I (shown as green) antibodies or (**b**) anti-LAMP1 (shown as red) and anti-human apoA-I (shown as green) antibodies, followed by DAPI staining (shown as blue). Scale bars = 10 μ m. The arrows indicate apoA-I colocalization with lysosomes.



Supplemental Figure 6. HDL_{CAD} contained FLAP.

Each type of HDL was purified by ultracentrifuge. $HDL_{Healthy}$ and HDL_{CAD} (10 µg protein, respectively) were investigated by western blot analysis with anti-FLAP antibody (ab85227, abcam).



Supplemental Figure 7. HDL contained exosomes in circulating plasma.

Plasma_{Healthy} and Plasma_{CAD} were immunoprecipitated with anti-CD9 antibody (12A12, Cosmo Blo) using ExoTrap[™] Exosome Isolation Spin Column Kit (Cosmo Bio), then immunoblotted with anti-CD9 antibody antibody (12A12, Cosmo Bio) and anti-ApoA-I antibody (#23030485, CHEMICON INTERNATIONAL).



Supplemental Figure 8. RAW macrophages expressed BLT1.

Total RNA was extracted from zymosan stimulated RAW macrophages. PCR amplification for mice BLT1 was performed with or without reverse transcription (RT). PCR products were separated by electrophoresis with 2% agarose gel.

LTB_4



Supplemental Figure 9. LTB₄ selectively attenuated clathrin-mediated endocytosis.

Macrophages were pretreated for 30 min at 37°C with vehicle, LTB_4 (100 nM) with/without BLT1 antagonist U75302 (300 nM), or the clathrin-mediated endocytosis inhibitor Pitstop2 (20 μ M). Next, macrophages were co-incubated with 10 μ g/ml transferrin (shown as *green*) or dextran (shown as *red*) for 30 min at 37 °C, followed by DAPI staining (shown as blue) (scale bars = 10 μ m).



Supplemental Figure 10. LTB₄ treatment did not affect expression of ApoA-I receptors.

Macrophages were treated with or without 100 nM LTB₄ for 2hr at 37°C. Cell lysates were processed for western blot analysis with anti-ABCA1 antibody (NB400-105, Novus Biologicals), anti-ABCG1 antibody (NB400-132, Novus Biologicals), anti-SR-B1 antibody (NB400-104, Novus Biologicals), and anti-SR-A antibody (CD204, Trans Genic).

Supplemental Figure 11. Gels presented in the main figures.



Figure 4c (5-LO)





β-actin