Cancer Cell, Volume 32

## **Supplemental Information**

## **Integrated Molecular Meta-Analysis**

## of 1,000 Pediatric High-Grade

## and Diffuse Intrinsic Pontine Glioma

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Figure S1 (related to Figure 1) – Clinicopathological and molecular subgroups of pHGG/DIPG. (A) Venn diagram showing overlapping molecular data for the cohort. (B) Anatomical location of all cases separated by age at diagnosis <1 year (n=40). Radius of circle is proportional to the number of cases. Left – sagittal section showing internal structures; right – external view highlighting cerebral lobes. Orange = infant. Lighter shaded circles represent a non-specific designation of hemispheric, midline or brainstem. (C) Kaplan-Meier plot of overall survival of cases separated by age at diagnosis <1 year, between 1-3 years, and >3 years of age, p value calculated by the log-rank test (n=818). (D-F) Kaplan-Meier plot of overall survival of cases separated by histone mutation in (D) hemispheric (n=220), (E) midline (n=131), (F) brainstem locations (n=242), p value calculated by the log-rank test. (G) Anatomical location of all cases separated by BRAF V600E mutation (n=32, wild-types not shown). Radius of circle is proportional to the number of cases. Left - sagittal section showing internal structures; right external view highlighting cerebral lobes. Gold = BRAF V600E. Lighter shaded circles represent a non-specific designation of hemispheric, midline or brainstem. (H) Boxplot showing age at diagnosis of included cases, separated by BRAF V600E mutation (n=553). The lower and upper limits of the boxes represent the first and third quartiles, and the whiskers 1.5x the interquartile range. (I) Kaplan-Meier plot of overall survival of cases separated by BRAF V600E, p value calculated by the log-rank test (n=416). (J) Anatomical location of all cases separated by IDH1 R132 mutations (n=40, wild-types not shown). Radius of circle is proportional to the number of cases. Left - sagittal section showing internal structures; right external view highlighting cerebral lobes. Red = IDH R132. Lighter shaded circles represent a non-specific designation of hemispheric, midline or brainstem. (K) Boxplot showing age at diagnosis of included cases, separated by IDH1 R132 mutations (n=672). The thick line within the box is the median, the lower and upper limits of the boxes represent the first and third quartiles, and the whiskers 1.5x the interquartile range. \*\*\* p<0.0001, t-test. (L) Kaplan-Meier plot of overall survival of cases separated by IDH1 R132 mutations, p value calculated by the log-rank test (n=551).









В

**Figure S2** (related to Figure 2) – Methylation-based subclassification of pHGG/DIPG. (A) Boxplot of Heidelberg brain tumor subclassifier scores for pediatric HGG in the present study (solid circles, n=441) assigned to one of six HGG subgroups defined by a reference set (open circles). The thick line within the box is the median, the lower and upper limits of the boxes represent the first and third quartiles, and the whiskers 1.5x the interquartile range. H3 G34R/V – blue; H3 K27M – green; IDH1 – red; PXA-like – gold; LGG-like – tan; HGG WT – grey. (B) Barplots of number of cases with methylated *MGMT* promoter, subdivided by methylation subgroup (n=366). (C) Boxplots of mean methylation beta-values for cases subdivided by methylation subgroup (n=441). The thick line within the box is the median, the lower and upper limits of the boxes represent the first and third quartiles, and the whiskers 1.5x the interquartile range. \*\*\* p<0.001, \*\* p<0.01, \*p<0.05 *versus* rest, t-test.



























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а





Figure S3 (related to Figure 3) – DNA copy number aberrations in pHGG/DIPG. (A) DNA copy neutral cases. (a) Anatomical location (n=147). Radius of circle is proportional to the number of cases. Left - sagittal section showing internal structures; right - external view highlighting cerebral lobes. Pale grey = copy neutral cases. Lighter shaded circles represent a non-specific designation of hemispheric, midline or brainstem. (b) Boxplot showing age at diagnosis of included cases, separated by DNA copy neutral cases (n=798). The thick line within the box is the median, the lower and upper limits of the boxes represent the first and third quartiles, and the whiskers 1.5x the interquartile range. \*\*\* p<0.0001, t-test. (c) Kaplan-Meier plot of overall survival of cases separated by DNA copy neutral cases, p value calculated by the logrank test (n=638). (d) Venn diagrams showing platform distribution of copy neutral (left, n=147) and other cases (right, n=687). (B-G) Recurrent copy number changes. (a) Segmented exonlevel DNA copy number heatmaps for (B) 17p loss (n=156), (C) 9q gain (n=108), (D) 2p24 amplification (n=42), (E) 4q12 amplification (n=77), (F) chromosome 7 amplifications (n=59), (G) 9p21 deletion (n=102). Dark red, amplification; red, gain; dark blue, deletion; blue, loss. Chromosomal ideograms are provided indicating enlarged genome browser view and genes within common regions targeted across samples (grey). Clinicopathological and molecular annotations are provided as bars according to the included key. (B-F) (b) Kaplan-Meier plot of overall survival of subgroups of cases separated by indicated DNA copy number change, p value calculated by the log-rank test. (G) 9p loss. (b) Anatomical location (n=115). Radius of circle is proportional to the number of cases. Left - sagittal section showing internal structures; right – external view highlighting cerebral lobes. Dark blue = 9p loss cases. Lighter shaded circles represent a non-specific designation of hemispheric, midline or brainstem. (c) Boxplot showing age at diagnosis of included cases, separated by 9p loss cases (n=798). The thick line within the box is the median, the lower and upper limits of the boxes represent the first and third quartiles, and the whiskers 1.5x the interquartile range. \*\*\* p<0.0001, t-test. (d,e) Kaplan-Meier plot of overall survival of cases separated by 9p loss cases, p value calculated by the log-rank test (n=659).

Α



0 10 20 30 40 50 40 30 20 10 0

% Frequency

p<0.0001

0 10 20 30 40 50 40 30 20 10 0

% Frequency



RUSCI ASHIL HON3 GBA FDPS

Histone

Wild-type

Amp Gain NC Loss Del

HDGF

Methylation subgroup

Location (Mb)

200

220

240

Location

Hemispheric Midline



PXA-like



Figure S4 (related to Figure 4) – Subgroup-specific copy number changes in pHGG/DIPG. (A) GISTIC and frequency barplots for whole chromosomal arm changes for (a) histone wildtype cases and (b) H3.3K27M cases subdivided by anatomical location. Log<sub>10</sub> values for GISTIC are plotted across the genome for both amplifications (dark red) and deletions (dark blue), with significantly enriched events labelled by likely driver genes. Subgroup-specific genes are highlighted by the appropriate color, as are significantly enriched whole arm alterations (p<0.0001, Fishers exact test). (B) Histone wild-type cases. (b) GISTIC and frequency barplots for whole chromosomal arm changes subdivided by simplified methylation subclass. Log<sub>10</sub> values for GISTIC are plotted across the genome for both amplifications (dark red) and deletions (dark blue), with significantly enriched events labelled by likely driver genes. Subgroup-specific genes are highlighted by the appropriate color, as are significantly enriched whole arm alterations (p<0.0001, Fishers exact test). (b) Segmented exon-level DNA copy number heatmaps for 1q loss in PXA-like cases (dark red, amplification; red, gain; dark blue, deletion; blue, loss; n=5). Chromosomal ideograms are provided indicating enlarged genome browser view and genes within common regions targeted across samples (grey). Clinicopathological and molecular annotations are provided as bars according to the included key. (c) Kaplan-Meier plot of overall survival of subgroups of cases separated by indicated DNA copy number change in PXA-like cases (n=35), p value calculated by the log-rank test. (C) Copy number losses in H3.3G34R/V cases. (a) 3q loss (n=29). (b) 5q loss (n=18). (c) 18q loss (n=20). Segmented exon-level DNA copy number heatmaps for losses in H3.3G34R/V cases (dark red, amplification; red, gain; dark blue, deletion; blue, loss). Chromosomal ideograms are provided indicating enlarged genome browser view and genes within common regions targeted across samples (grey). Clinicopathological and molecular annotations are provided as bars according to the included key.









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**Figure S5** (related to Figure 5) – Alterations targeting *FBXW7* in H3.3G34R/V pHGG and *TOP3A* in H3.3K27M DIPG. (A) Gene expression heatmap for all genes on chromosome 4q across three independent platforms, separated by those with 4q loss (blue) and those with normal copy number (grey). Agilent, left (n=67); Affymetrix, middle (n=102); RNAseq, right (n=82). (B) CIRCOS plots for 6 cases (5x H3.3K27M DIPG, 1x midline histone wild-type) with complex rearrangements leading to *TOP3A* amplification. In each case, plots provide DNA copy number changes (dark red, amplification; red, gain; dark blue, deletion; blue, loss) and loss of heterozygosity (yellow) on the inner rings, and intra- (orange) and inter- (blue) chromosomal translocations inside the circle.









С



Alteration





Coverage

Figure S6 (related to Figure 6) – Somatic mutations in pHGG/DIPG. (A) Mutation rate boxplots across non-hypermutator pHGG/DIPG separated according to location (n=318), histone mutation(n=326), methylation subgroup (n=129), pre/post-treatment sampling (n=326) and age <1 year (n=326), and then compared to 11 hypermutator cases (plotted on a  $log_{10}$  scale). The thick line within the box is the median, the lower and upper limits of the boxes represent the first and third quartiles, and the whiskers 1.5x the interquartile range. \*\*\* adjusted p<0.0001, \*\*<0.01, t-test. (B) Mutation signatures for the 11 hypermutator cases. (Top) Simple stacked barplot representation of the proportion of mutation types observed in individual hypermutator cases and the remaining accumulated dataset. Base changes given in the key. (Bottom) Mutation context given for each of the 96 mutated trinucleotides, represented by heatmap. The base located 5' to each mutated base is shown on the vertical axis, and the 3' base is on the horizontal axis. (C) Oncoprint representation of an integrated annotation of somatic mutations and DNA copy number changes in IDH1 mutant cases (n=14). Clinicopathological and molecular annotations are provided as bars according to the included key. (D) Mutual exclusivity analysis. Pairwise Fishers exact tests are calculated for each of the 30 most commonly altered genes in pHGG/DIPG. Log<sub>2</sub>-transformed odds ratios are plotted in a purple (negative correlation) to cyan (positive correlation) color scheme. White boxes represent no co-occuring events. \*\*\* adjusted p<0.001, \*\* adjusted p<0.01, \*adjusted p<0.05. (E) Coverage and variant allele frequency (VAF) statistics for rare variants. (a) Coverage and (b) VAF are provided as a density plot for those variants detected in 1-3 cases (blue lines) overlaid with the remaining variants (shaded grey). (c) Co-plot of VAF (blue points) and coverage (grey bars) for variants detected in a single case, ordered by VAF.



С

30

b



100

80

60

4

20

0

Pons

Midline

Hemispheric

Histone subgroup (% cases)

\*\*\*

+++

H3.3 K27M

H3.3 G34RV



d





е

100

80

4

20

0

Location (% cases) 60





В

а



Frontal lobe

Thalamus

Pons

С

d

С







d



С

D



е

Ε

d



С





F

b

Hemispheric

Basal ganglia Parietal lobe Frontal Joh Pa 1.0 MAPK Frontal lot 40 × 0.8 <sup>30</sup> <sup>00</sup> <sup>01</sup> <sup>10</sup> ... Survival 0.4 0.6 q oital lobe Thalam Temporal lobe Pon Cerebellum Occipital lobe 10 0.2 Brainstem p=0.0197 0.0 0 Spinal cord 40 60 80 Time (months) None -Ó 20 100 120 MAPK -MAPK е 100 100 100 100 Methylation subclass (% cases) Histone subgroup (% cases) 20 40 60 80 Location (% cases) 40 60 80 Age <1 year (% cases) 8 \*\* 60 4 20 20 0 0 0 0 Wild-type Midline Pons

H3.3 G34RV H3.3 K27M H3.1 K27M С

d

HGG WT LGG-like PXA-like

H3 G34RV H3 K27M <1 year

Older



G

b



С

d







е

d

HGG WT LGG-like



С





Figure S7 (related to Figure 7) – Integrated pathway analysis of pHGG/DIPG. (A) TP53 / DNA repair pathway, (B) chromatin readers, writers, remodellers, (C) cell cycle pathway, (D) receptor tyrosine kinases, (E) PI3K/mTOR pathway, (F) MAPK pathway, (G) BMP pathway, (H) WNT pathway and (I) maintenance of genome integrity. (a) Oncoprint representation of an integrated annotation of somatic mutations and DNA copy number changes in cases targeting one or more members of a specific pathway. Clinicopathological and molecular annotations are provided as bars according to the included key. (b) Anatomical location of pathwaytargeted cases. Radius of circle is proportional to the number of cases. Left - sagittal section showing internal structures; right - external view highlighting cerebral lobes. Black = given pathway alteration. Lighter shaded circles represent a non-specific designation of hemispheric, midline or brainstem. (c) Boxplot showing age at diagnosis of included cases, separated by pathway-targeted cases. The thick line within the box is the median, the lower and upper limits of the boxes represent the first and third quartiles, and the whiskers 1.5x the interquartile range. (d) Kaplan-Meier plot of overall survival of cases separated by pathwaytargeted cases, p value calculated by the log-rank test. (e) Barplots showing distribution of pathway-targeted cases by anatomical location, histone subgroup, methylation subgroup and age. \*\*\* p<0.001, \*\* p<0.01, \*p<0.05, n.s., not significant, t-test.



Location
Histone
Methylation subgroup
Cluster
Platform

I Hemispheric
Midline
Pons
IH3.3 G34RV
IH3.1 K27M
Wild-type
IH3 G34RV
IH3 K27M
INId-type
IH3 G34RV
IH3 K27M
INId-type
IH3 G34RV
IH3 C34RV
IH3

**Figure S8** (related to Figure 8) – Integrated analysis of H3/IDH1 wild-type pHGG/DIPG. (A-C) Unsupervised hierarchical clustering of the most variable genes in samples profiled by (A) Agilent arrays (n=67), (B) Affymetrix arrays (n=102) and/or (C) RNA sequencing (n=82). (D) A combined dataset across all platforms (n=220) is subjected to differential expression analysis for H3.3G34R/V, H3.3K27M and H3.1K27M mutations, with significant genes plotted in the heatmap. (E) Gene set enrichment analysis for selected significant gene sets for each comparison. (F) Combined dataset for H3/IDH1 wild-type clusters WT-A, WT-B and WT-C (n=21), with significantly differentially expressed genes plotted in the heatmap. (G) Gene set enrichment analysis for selected significant gene sets for each comparison. Clinicopathological and molecular annotations are provided as bars according to the included key.