Supporting Information

Fluorinated molecular beacons as functional DNA nanomolecules for cellular imaging

Cheng Jin,^{a,†} Ting Fu,^{a,†} Ruowen Wang,^{a, b, c} Hui Liu,^a Jiamei Zou,^a Zilong Zhao,^a Mao Ye,^a Xiaobing Zhang^a and Weihong Tan^{a, b}

- a) Molecular Sciences and Biomedicine Laboratory, State KeyLaboratory for Chemo/Biosensing and Chemometrics, College of Chemistry and Chemical Engineering and College of Biology, Collaborative Innovation Center for Chemistry and Molecular Medicine, Hunan University, Changsha 410082, China
- b) Department of Chemistry and Department of Physiology andFunctional Genomics, Center for Research at the Bio/Nano Interface, Health Cancer Center, UF Genetics Institute and McKnight Brain Institute, University of Florida, Gainesville, Florida 32611-7200, United States
- c) Department of Biotechnology and Biomedicine, Yangtze DeltaRegion Institute of Tsinghua University, Zhejiang, 314006, China

[†]These authors contributed equally to this paper.

Correspondenceauthors: W.T. (<u>tan@chem.ufl.edu</u>) and R.W. (<u>wangwrw@yahoo.com</u>).



Scheme S1. Synthesis route of F base phosphoramidite

The synthesis procedures of fluorinated base phosphoramidite monomer refer to the protocol developed in our laboratory.¹ ¹HNMR, ¹⁹FNMR, ³¹PNMR and ESI-MS data of 3,5-bis(trifluoromethyl) benzoyl phosphoramidite monomer (3) are as follows:

¹H NMR (300 MHz, CDCl₃) δ : 8.43 (s, 1H), 8.41 (s, 1H), 8.23 (s, 1H), 8.17 (s, 1H), 7.51 (d, J=6.0 Hz, 2H), 7.37 (d, J=6.6 Hz, 4H), 7.26-7.30 (m, 2H), 7.187.21 (m, 1H), 6.83-6.87 (m, 4H),4.29-4.35 (m, 1H), 3.62-3.93 (m, 12H), 3.17-3.34 (m, 2H), 2.70-2.74 (m, 1H), 2.60-2.63 (m, 1H), 1.10-1.25 (m, 12H); ¹⁹F NMR (CDCl₃) δ : -64.23, -64.29; ³¹P NMR (CDCl₃) δ : 139.62, 140.99. MS (ESI+): m/z 833.3 (Calculated M+Na: 856.40).



Figure S1. Fluorescence spectroscopy of FMBs before (black line) and after (red line) hybridization with cDNA. (a) F2 MB, (b) F4 MB and (c) F6 MB.



Figure S2. Fluorescence spectroscopy kinetics of F6 MB (a) and N6 MB (b) treated with cDNA (black line) and synthetic one-base mismatch target (1m-cDNA) (red line).



Figure S3. Fluorescence spectroscopy of 1 μ M N6 MB (a) and F6 MB (c) treated with cDNA. b & d) The fitted linear relation of N6 MB (b) and F6 MB (d) function to the concentration of cDNA. In both cases, the lowest detectable concentration of target DNA is 50 nM.



Figure S4. S/B ratios of F6 MB (red column) and N6 MB (black column) hybridized with cDNA at 20, 30 and 37°C.



Figure S5. Melting curves of 200 nM FMBs and N6 MBs.



Figure S6. Melting curves of F6N6 MB and N6 MB. F6N6 MB represents FMBs with additional six natural base pairs in stem region.



Figure S7. Comparison between the S/B ratio of F6 MB (red column) and N6 MB (black column) with various incubation times to CCRF-CEM cell lysate.



Figure S8. (a) Flow cytometry analysis and (b) the relative fluorescence intensity of MCF-7 cells (with and without LPS treatment) after incubation with AS1411-linked F6 MnSOD MBs.



Figure S9. Cytotoxicity assay of MCF-7 cells treated with F6 DNA hairpin.

| Name | Sequence (from 5'to 3') |
|------------|--|
| F2 MB | FAM FF TCT AAA TCA CTA TGG TCG C FF Dabcyl |
| F4 MB | FAM FFFF TCT AAA TCA CTA TGG TCG C FFFF Dabcyl |
| F6 MB | FAM FFFFFF TCT AAA TCA CTA TGG TCG C FFFFFF Dabcyl |
| F8 MB | FAM FFFFFFF TCT AAA TCA CTA TGG TCG C FFFFFFFF Dabcyl |
| F6N6 MB | FAM FFFFFF CCG AGC TCT AAA TCA CTA TGG TCG C GCT CGG |
| | FFFFF Dabcyl |
| N6 MB | FAM CCG AGC TCT AAA TCA CTA TGG TCG C GCTCGG Dabcyl |
| cDNA | GCG ACC ATA GTG ATT TAG A |
| 1m-cDNA | GCG ACC ATA G A G ATT TAG A |
| Oligo 2 | CCA TAG TGA TTT AGA GCT CGG |
| Oligo3 | TAG TGA TTT AGA GCT CGG |
| Oligo4 | TGATTT AGA GCT CGG |
| AS1411- | Dabcyl FFFFFF AGT TAC ATT CTC CCA GTT GAT T FFFFFF FAM TTT |
| F6MnSOD MB | GGT GGT GGT GGT TGT GGT GGT GGT GGT TT |
| AS1411-F6 | Dabcyl FFFFFF TCT AAA TCG CTA TGG TCG C FFFFFF FAM TTT |
| control MB | GGT GGT GGT GGT TGT GGT GGT GGT GGT TT |

 Table S1. DNA sequences designed in this article.

Note: **F** refers to F base

Reference

 R. Wang, C. Wang, Y. Cao, Z. Zhu, C. Yang, J. Chen, F.-L. Qing and W. Tan, Chemical Science, 2014, 5, 4076-4081.