

**Figure S1**. (A) Proteins purified for this study. Pol  $\varepsilon$  (0.84 µg) and four-subunits complex of Pol  $\zeta$  (0.39 µg of Rev3) were separated by SDS-PAGE and stained by coomassie brilliant blue R250. (B) NGS substrates. Type A to Type F templates share the Ion-P1 adaptor (green box) and primer-binding site (underlined), but have different sequences between them (from #21 to #73 in the Figure). The variable regions in type A –D templates were excluding repeated nucleotides, and type E and F contain homodimers (like AA). (C) Control oligonucleotides that have two adaptors. TSO560 has an abasic site at the position indicated by "X".







**Figure S3.** Rates of single nucleotide insertions and deletions on six undamaged templates (type A-F) by Pol $\eta$ , Pol $\zeta$ , and Pol $\delta$ . For insertions, positions of colored bars indicate the positions after which insertions were observed.

**Table S1. Synthetic DNA oligos.** Type-A to F templates have same sequences except for the internal 53-nt region (variable region; Figure 1A and Figure S1B). Ion Torrent primers have same sequences except for the barcode (BC).

Name	Sequence (5'-3')	Use	Source
	CCTCTCTATGGGCAGTCGGTGATGACTAGATATCGACGT	Template (Type-A)	
TSO525	GATCAGACTCTAGATAGATGCTAGAGAGCTCTATCGATC	3'-Biotin	
	AGTGCGTGTGGCCTGTCAATC/3Bio/	Undamaged	IDT
	CCTCTCTATGGGCAGTCGGTGATGACTAGATATCGACGT	Template (Type-A)	
TSO526	GATCAGACTCTAGATAGATGCTAGAGAGCXCTATCGATC	3'-Biotin	
	AGTGCGTGTGGCCTGTCAATC/3Bio/	28AP (indicated as "X")	IDT
	CCTCTCTATGGGCAGTCGGTGATGACTAGATATCGACGT	Template (Type-A)	
TSO589	GATCAGACTCTAGAXAGATGCTAGAGAGCTCTATCGATC	3'-Biotin	
	AGTGCGTGTGGCCTGTCAATC/3Bio/	43AP (indicated as "X")	IDT
	CCTCTCTATCCCCCACTCCCCTCATGACCGTCGTCGACTCCCATA	Template (Type-B)	
TSO614	CATAGCTACACTGAGTATACGATCGCTAGCATCGTGATC	3'-Rintin	
	AGTGCGTGTGGCCTGTCAATC/3Bio/	Undamaged	IDT
'		Template (Type-()	
TS0615	AGACCCCCCTCTCTCGATCTGTGTGTGTGTGTGTCTCGATCTGGTCTCGATCTGGTCTCGATCTGGTCTCGATCTGGTGTGTGT	2'-Riotin	
100010	AGTGCGTGTGGCCTGTCAATC/3Bio/	5 -Diuliii Undamaged	דחו
		Tomplato (Type-D)	יטו
<b>TGO616</b>	CCTCTCTATGGGCAGTCGGTGATACGCACACAGTACATG		
120010	AGTGCGTGTGGCCTGTCAATC/3Bio/	3 -Blotin	
		Undamaged	וטו
<b>760705</b>		lemplate (Type-E)	
<b>TSO785</b>	TAGGACCAATTAAGTTTGCCTGGAACCGGTCTTCACGTC	3'-Biotin	
	AGIGCGIGIGGCCIGICAAIC[BUIIG]	Undamaged	Sigma
	CCTCTCTATGGGCAGTCGGTGATAACCGGATTCCCTTAA	Template (Type-F)	
TS0786	GGGTTCAACCAGGCCTGGAAAGGCAATTTGGTTACCGTC	3'-Biotin	
	AGTGCGTGTGGGCCTGTCAATC[Btnrg]	Undamaged	Sigma
т50590	CCTAACGTAACGATTGACAGGCCACACGCACTGA	Primer	
100000		for Gel assay	Fisher
TS0686	CCATCTCATCCCTGCGTGTCTCCGACTCAGTAAGGAGAA	Primer	
150000	CGATTGACAGGCCGCACGCACTGA	For IonTorrent (BC2)	Fisher
<b>™</b> ⊈∩687	CCATCTCATCCCTGCGTGTCTCCGACTCAGAAGAGGATT	Primer	
150007	CGATTGACAGGCCGCACGCACTGA	For IonTorrent (BC3)	Fisher
<b>TCO688</b>	CCATCTCATCCCTGCGTGTCTCCGACTCAGTACCAAGAT	Primer	
120000	CGATTGACAGGCCGCACGCACTGA	For IonTorrent (BC4)	Fisher
<b>TCO689</b>	CCATCTCATCCCTGCGTGTCTCCGACTCAGCAGAAGGAA	Primer	
150005	CGATTGACAGGCCGCACGCACTGA	For IonTorrent (BC5)	Fisher
700600	CCATCTCATCCCTGCGTGTCTCCGACTCAGCTGCAAGTT	Primer	
T50090	CGATTGACAGGCCGCACGCACTGA	For IonTorrent (BC6)	Fisher
70001	CCATCTCATCCCTGCGTGTCTCCGACTCAGTTCGTGATT	Primer	
L200AT	CGATTGACAGGCCGCACGCACTGA	For IonTorrent (BC7)	Fisher
	CCATCTCATCCCTGCGTGTCTCCGACTCAGTTCCGATAA	Primer	
TSO692	CGATTGACAGGCCGCACGCACTGA	For IonTorrent (BC8)	Fisher
	CCATCTCATCCCTGCGTGTCTCCGACTCAGTGAGCGGAA	Primer	
TSO693	CGATTGACAGGCCGCACGCACTGA	For IonTorrent (BC9)	Fisher
<u> </u> '	CCATCTCATCCCTCCGTCTCCCGACTCAGCTGACCGAA	Primer	
TSO694	CGATTGACAGGCCGCACGCACTGA	For IonTorrent (BC10)	Fisher

TSO695	CCATCTCATCCCTGCGTGTCTCCGACTCAGTCCTCGAAT	Primer			
	CGATTGACAGGCCGCACGCACTGA	For IonTorrent (BC11)	Fisher		
TSO696	CCATCTCATCCCTGCGTGTCTCCGACTCAGTAGGTGGTT	Primer			
	CGATTGACAGGCCGCACGCACTGA	For IonTorrent (BC12)	Fisher		
TSO697	CCATCTCATCCCTGCGTGTCTCCGACTCAGTCTAACGGA	Primer			
	CGATTGACAGGCCGCACGCACTGA	For IonTorrent (BC13)	Fisher		
TSO698	CCATCTCATCCCTGCGTGTCTCCGACTCAGTTGGAGTGT	Primer			
	CGATTGACAGGCCGCACGCACTGA	For IonTorrent (BC14)	Fisher		
TSO699	CCATCTCATCCCTGCGTGTCTCCGACTCAGTCTAGAGGT	Primer			
	CGATTGACAGGCCGCACGCACTGA	For IonTorrent (BC15)	Fisher		
TSO700	CCATCTCATCCCTGCGTGTCTCCGACTCAGTCTGGATGA	Primer			
	CGATTGACAGGCCGCACGCACTGA	For IonTorrent (BC16)	Fisher		
TS0701	CCATCTCATCCCTGCGTGTCTCCGACTCAGTCTATTCGT	Primer			
120701	CGATTGACAGGCCGCACGCACTGA	For IonTorrent (BC17)	Fisher		
TS0702	CCATCTCATCCCTGCGTGTCTCCGACTCAGAGGCAATTG	Primer			
100/02	CGATTGACAGGCCGCACGCACTGA	For IonTorrent (BC18)	Fisher		
TS0703	CCATCTCATCCCTGCGTGTCTCCGACTCAGTTAGTCGGA	Primer			
150705	CGATTGACAGGCCGCACGCACTGA	For IonTorrent (BC19)	Fisher		
<b>TGO704</b>	CCATCTCATCCCTGCGTGTCTCCGACTCAGCAGATCCAT	Primer			
190104	CGATTGACAGGCCGCACGCACTGA	For IonTorrent (BC20)	Fisher		
<b>TGO705</b>	CCATCTCATCCCTGCGTGTCTCCGACTCAGTCGCAATTA	Primer			
150705	CGATTGACAGGCCGCACGCACTGA	For IonTorrent (BC21)	Fisher		
<b>TGO706</b>	CCATCTCATCCCTGCGTGTCTCCGACTCAGTTCGAGACG	Primer			
150706	CGATTGACAGGCCGCACGCACTGA	For IonTorrent (BC22)	Fisher		
TSO707	CCATCTCATCCCTGCGTGTCTCCGACTCAGTGCCACGAA	Primer			
	CGATTGACAGGCCGCACGCACTGA	For IonTorrent (BC23)	Fisher		
TC0708	CCATCTCATCCCTGCGTGTCTCCGACTCAGAACCTCATT	Primer			
150700	CGATTGACAGGCCGCACGCACTGA	For IonTorrent (BC24)	Fisher		
<b>TGO709</b>	CCATCTCATCCCTGCGTGTCTCCGACTCAGCCTGAGATA	Primer			
150709	CGATTGACAGGCCGCACGCACTGA	For IonTorrent (BC25)	Fisher		
<b>TGO710</b>	CCATCTCATCCCTGCGTGTCTCCGACTCAGTTACAACCT	Primer			
150/10	CGATTGACAGGCCGCACGCACTGA	For IonTorrent (BC26)	Fisher		
<b>Ψ</b> ⊆∩711	CCATCTCATCCCTGCGTGTCTCCGACTCAGAACCATCCG	Primer			
190/11	CGATTGACAGGCCGCACGCACTGA	For IonTorrent (BC27)	Fisher		
₩90712	CCATCTCATCCCTGCGTGTCTCCGACTCAGATCCGGAAT	Primer			
150/12	CGATTGACAGGCCGCACGCACTGA	For IonTorrent (BC28)	Fisher		
TC0712	CCATCTCATCCCTGCGTGTCTCCGACTCAGTCGACCACT	Primer			
120112	CGATTGACAGGCCGCACGCACTGA	For IonTorrent (BC29)	Fisher		
<b>TC</b> ∩714	CCATCTCATCCCTGCGTGTCTCCGACTCAGCGAGGTTAT	Primer			
190114	CGATTGACAGGCCGCACGCACTGA	For IonTorrent (BC30)	Fisher		
<b>₩</b> 9071 5	CCATCTCATCCCTGCGTGTCTCCGACTCAGTCCAAGCTG	Primer			
120/12	CGATTGACAGGCCGCACGCACTGA	For IonTorrent (BC31)	Fisher		
TC0716	CCATCTCATCCCTGCGTGTCTCCGACTCAGTCTTACACA	Primer			
120/10	CGATTGACAGGCCGCACGCACTGA	For IonTorrent (BC32)	Fisher		
₩60717	CCATCTCATCCCTGCGTGTCTCCGACTCAGTTCTCATTG	Primer			
120/1/	AACGATTGACAGGCCGCACGCACTGA	For IonTorrent (BC33)	Fisher		
TC0710	CCATCTCATCCCTGCGTGTCTCCGACTCAGTCGCATCGT	Primer			
190/18	TCGATTGACAGGCCGCACGCACTGA	For IonTorrent (BC34)	Fisher		
TC0710	CCATCTCATCCCTGCGTGTCTCCGACTCAGTAAGCCATT	Primer			
120/13	GTCGATTGACAGGCCGCACGCACTGA	For IonTorrent (BC35)	Fisher		

	ССАТСТСАТСССТСССТСТСТСССАСТСАСААССААТСС	Primer	
TS0720	TCGATTGACAGGCCGCACGCACTGA	For IonTorrent (BC36)	Fisher
	ССАТСТСАТСССТСССТСТСТСССАСТСАССТТСАСА	Primer	
TS0721	GTCGATTGACAGGCCGCACGCACTGA	For IonTorrent (BC37)	Fisher
	CC2TCTC2TCCCTCCCTCTCTCCC2CTC2CCCCCCCCCC	Primer	
TS0722	GACGATTGACAGGCCGCACGCACTGA	For IonTorrent (BC38)	Fisher
		Primer	
TS0723	GCGATTGACAGGCCGCCCGCCCCGCACTCAGTAACAATCG	For IonTorrent (BC39)	Fisher
		Primer	
TS0724	TCGATTGACAGGCCGCACGCACTGA	For IonTorrent (BC40)	Fisher
		Primer	1 ioner
TS0725	CCATCTCATCCCTGCGTGTCTCCGACTCAGTTCCACTTC GCGATTGACAGGCCGCACGCACTGA	For IonTorrent (BC41)	Fisher
		Primer	TISHCI
TS0726	CGATTGACAGGCCGCACGCACTGA	For IonTorrent (BC42)	Fisher
			TISHCI
TS0727	CCATCTCATCCCTGCGTGTCTCCGACTCAGCTTGACACC	For IonTorrent (BC/13)	Fisher
		Drimor	1131101
TS0728		For IonTorrent (BC44)	Fishor
		Drimor	1131101
TS0729	CCATCTCATCCCTGCGTGTCTCCCGACTCAGTGGAGCTTC CTCGATTGACAGCCGCACGCACTGA	For IonTorrent (BC/15)	Fishor
TS0730		Primor	TISHEL
		Filler	Fichor
TS0731	ACGATIGACAGGCGCACGCACIGA	Drimor	FISHEI
		Primer	Fichor
	CACGATIGACAGGCCGCACGCACIGA	Drimor	FISHEI
TS0732		Filler	Fichor
		Poi foiffoiffeift (BC46)	1131161
TSO733		For IonTorrent (BC/19)	Fisher
		Primer	TISTICI
TS0734	CCATCTCATCCCTGCGTGTCTCCCGACTCAGCGGACAATG	For IonTorrent (BC50)	Fisher
		Primer	TISHCI
TS0584		For IonTorrent (BC60)	Fisher
		Primer	1 131101
TS0585		For IonTorrent (BC61)	Fisher
		Primer	
TSO586		For IonTorrent (BC62)	Fisher
		Primer	
TSO587	TCGATTGACAGGCCGCACGCACTGA	For IonTorrent (BC63)	Fisher
	ССЪ ТСТСЪ ТСССТССССТСТСТСССА СТСА ССТСА СТТСА С	Primer	
TSO588	GACGATTGACAGGCCGCACGCACTGA	For IonTorrent (BC64)	Fisher
		Control Oligo (with AP:	
TS0560		indicated as "X")	
	CTGAGTCGGAGACACGCAGGGATGAGATGG	For IonTorrent (BC51)	IDT
		Control Oligo (w/o	•
	$ \cdots $		
120201	ACTATCGATCAGTGCGATCGTTCCATGCGGCTGA	damage)	

Table S2. Summary of NGS analysis for abasic and undamaged type A templates.

	No-damage (Type A)				28AP (Type A	)	43AP (Type A)			
Polymerase	Barcode# Qualified N with 10G Read (N) (n)		Barcode#	Qualified Read (N)	N with 10G (n)	Barcode#	Qualified Read (N)	N with 10G (n)		
Polη	4	267,526	266,980	22	47,644	47,542	40	19,002	18,934	
ΡοΙη	10	116,744	116,366	28	48,137	48,046	46	5,964	5,946	
ΡοΙη	16	50,819	50,635	34	61,912	61,777	61	2,516	2,504	
ΡοΙζ	5	186,716	186,213	23	216,448	216,014	41	155,208	154,919	
Ροίζ	11	168,710	168,334	29	217,182	216,751	47	114,956	114,727	
ΡοΙζ	17	115,375	115,079	35	250,922	250,492	62	19,544	19,455	
Polη+ζ	6	123,736	123,458	24	78,384	78,248	42	100,393	100,176	
Polη+ζ	12	97,916	97,710	30	136,187	135,922	48	235,405	234,770	
Polη+ζ	18	123,360	123,126	36	130,582	130,362	63	30,027	29,893	
No Pol	7	2,594	2,578	25	2,258	2,245	43	1,224	1,210	
No Pol	13	1,318	1,310	31	2,775	2,757	49	2,618	2,600	
No Pol	19	2,262	2,242	37	1,823	1,816	64	555	550	
ΡοΙδ	2	235,223	234,352							
ΡοΙδ	8	83,650	82,970							
ΡοΙδ	14	145,990	145,535							
Pole	3	235,366	234,284							
Pole	9	167,361	166,884							
Pole	15	115,814	115,497							
	Control oli	go without c polymerase	lamage, no							
	Barcode#	Qualified Read (N)	N with 10G (n)							
No Pol	52	69,892	NA							
	Contro	ol oligo with polymerase	AP, no							
	Barcode#	Qualified Read (N)	N with 10G (n)							
No Pol	51	1,541	NA							

## Table S3. Summary of NGS analysis with type B to F templates.

Polymerase

No Pol

Polymerase

No Pol

Barcode#

52

Barcode#

51

Qualified N with 10G Read (N) (n)

polymerase Qualified N with 10G Read (N) (n)

NA

NA

10,493

Control oligo with AP, no

0

	No-damage (Type B)		No-damage (Type C)			No-damage (Type D)		No-damage (Type E)			No-damage (Type F)				
	Barcode#	Qualified Read (N)	N with 10G (n)	Barcode#	Qualified Read (N)	N with 10G (n)	Barcode#	Qualified Read (N)	N with 10G (n)	Barcode#	Qualified Read (N)	N with 10G (n)	Barcode#	Qualified Read (N)	N with 10G (n)
Ροίδ	48	216,737	210,080	49	153,759	149,863	50	66,491	64,946	60	24,145	24,046	61	43,070	42,724
ΡοΙδ										63	35,605	35,424	64	19,399	19,208
ΡοΙη	19	411,091	400,792	21	56,446	55,159	26	15,978	15,528	28	10,056	10,004	30	25,064	24,846
ΡοΙη	32	17,989	17,661	34	32,710	31,341	36	23,196	22,579	38	27,832	27,713	41	11,124	10,968
Polη										43	4,643	4,622	45	5,445	5,391
Ροίζ	20	56,948	56,126	25	225,590	220,875	27	79,807	77,450	29	47,933	47,702	31	67,407	66,561
Ροίζ	33	49,059	48,074	35	74,496	73,075	37	89,395	86,783	39	31,353	31,205	42	16,456	15,787
Ροίζ										44	10,430	10,379	47	22,311	22,123
No Pol										18	452	57			
	Control oligo without damage, no polymerase									_					

## Table S4. Base-substitution rates on undamaged templates.

	P	olη			P	οΙζ		
From	То	Avr (%)	STD	From	То	Avr (%)	STD	
А	С	0.072001	0.231915	А	С	0.018665	0.063343	
Α	G	0.584463	0.339584	А	G	0.043357	0.085708	
Α	Т	0.152433	0.152056	А	Т	0.019069	0.057238	
С	А	0.01706	0.017012	С	Α	0.072231	0.046962	
С	G	0.033472	0.044221	С	G	0.212069	0.080492	
С	Т	0.050209	0.032468	С	Т	0.035905	0.025726	
G	А	0.042319	0.026869	G	А	0.058707	0.034413	
G	С	0.014348	0.038731	G	С	0.011038	0.015567	
G	Т	0.004897	0.015081	G	Т	0.01668	0.012629	
Т	А	0.049124	0.029224	Т	А	0.090879	0.091182	
Т	С	0.141028	0.310944	Т	С	0.02506	0.021895	
Т	G	0.062356	0.050967	Т	G	0.079522	0.056363	
Total subs	stitutions	1.22371		Total sub:	stitutions	0.683184		
Misnair	ed templa	te hase		Misnai	red templa	te hase		
(% in to	tal substit	utions)		(% in total substitutions)				
A-templa	te base	20.63461		A-templa	ate base	28.6103		
C-templa	ate base	5.030889		C-templ	ate base	12.65033		
G-templa	ate base	8.232498		G-templ	ate base	46.86967		
T-templa	ite base	66.102		T-templa	ate base	11.8697		
Total N	Aisinserte	d hase		Total	Misinserte	d hase		
(% in to	tal substit	utions)		(% in to	ntal substit	utions)		
Δ-Mici	$\Lambda_{-}$ Misinsort			Δ-mic	insert	37 46873		
C-Mici	nsert	18 5800		C-mis	insort	2 015822		
G-Mici	nsert	55 59252		G-Mic	insert	49 02763		
T-Misi	nsert	16 95988		T-Mis	insert	10 4883		

Data obatined with type A to F templates were averaged after subtraction of background rates obtained in Pol-delta mediated reactions.