## distribution in the Staphylococcus genus 2 3 Michal Bukowski<sup>1</sup>, Karolina Hyz<sup>1</sup>, Monika Janczak<sup>1</sup>, Marcin Hydzik<sup>1</sup>, Grzegorz Dubin<sup>2,3</sup>, 4 Benedykt Wladyka<sup>1,#</sup> 5 6 <sup>1</sup>Department of Analytical Biochemistry, Faculty of Biochemistry, 7 Biophysics and Biotechnology, Jagiellonian University, Krakow, Poland 8 9 <sup>2</sup>Malopolska Centre of Biotechnology, Jagiellonian University, Krakow, Poland 10 <sup>3</sup>Department of Microbiology, Faculty of Biochemistry, Biophysics and Biotechnology, 11 Jagiellonian University, Krakow, Poland 12 13 #Corresponding author: 14 Benedykt Wladyka 15 Department of Analytical Biochemistry 16 Faculty of Biochemistry, Biophysics and Biotechnology 17 Jagiellonian University 18 7 Gronostajowa St, 30-387 Krakow, Poland 19 phone: +48 12 664 65 11 20 fax: +48 12 664 69 15 21 e-mail: benedykt.wladyka@uj.edu.pl 22 23 Keywords: mazEF, mRNA interferase, pemIK, staphylococci, Staphylococcus, Staphylococcus aureus, TA 24 system, toxin-antitoxin system 25 26 27 SUPPLEMENTARY DATA 28 29 Supplementary note on construction of phylogenetic trees...... 2 30 31 32 Supplementary figure 3...... 5 33 Supplementary figure 4...... 6 34 Supplementary figure 5...... 7 35 Supplementary figure 6...... 8 Bibliography......9 36 37

Identification of novel mazEF/pemIK family toxin-antitoxin loci and their

Supplementary note on construction of phylogenetic trees. The following parameters were used for multiple sequence alignments of DNA sequences; gap open cost: 20.0; gap extension cost: 1.0; end gap cost: free; alignment mode: very accurate; and the following ones for protein sequences; gap open cost: 10.0; gap extension cost: 1.0; end gap cost: free; alignment mode: very accurate. For DNA sequences the model choice and parameters were as following; construction method: neighbour joining [1]; nucleotide substitution model: general time reversible, GTR [2]; transition/transversion ratio: 2.0; include rate variation: yes; number of substitution rate categories: 8; gamma distribution parameter: 1.0; estimate substitution rate parameters, topology and gamma distribution parameter: yes; bootstrap analysis with 100 replicates; and for protein sequences as following; construction method: neighbour joining [1]; protein substitution model: WAG [3]; transition/transversion ratio: 2.0; include rate variation: yes; number of substitution rate categories: 8; gamma distribution parameter: 1.0; estimate substitution rate parameters, topology and gamma distribution parameter: yes; bootstrap analysis with 100 replicates. Prototypical protein sequences of all MazF/PemK toxins were aligned with the following parameters: gap open cost: 20.0; gap extension cost: 1.0; end gap cost: as any other; alignment mode: very accurate; and visualised using CLC Main Workbench.

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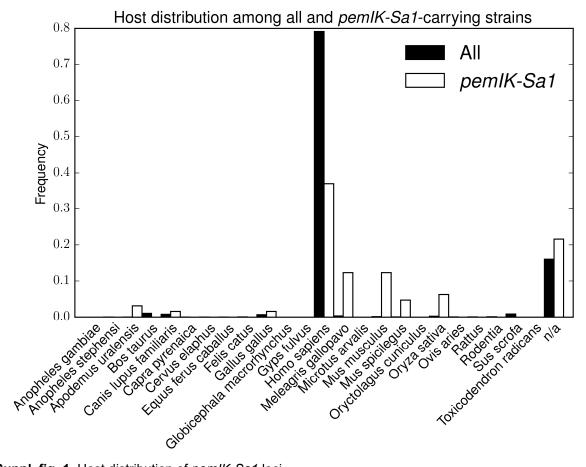
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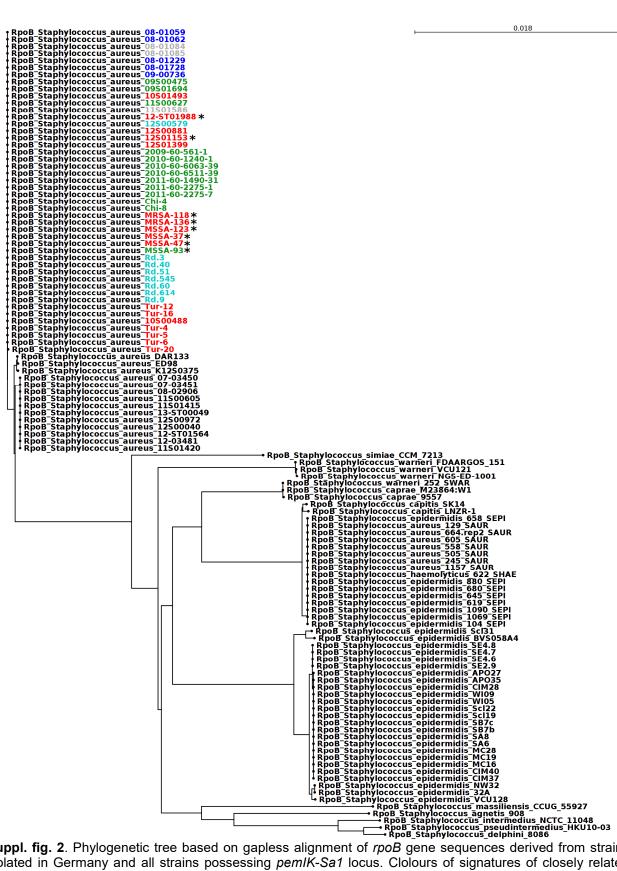
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Suppl. fig. 1. Host distribution of pemIK-Sa1 loci.

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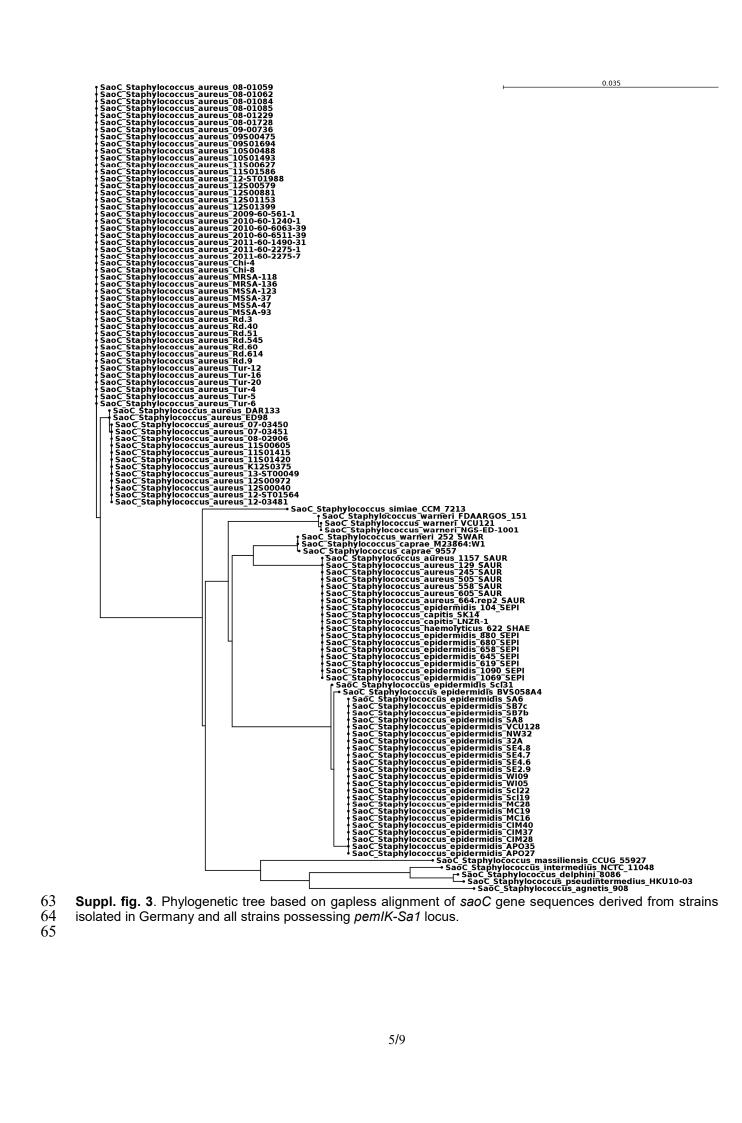


Suppl. fig. 2. Phylogenetic tree based on gapless alignment of rpoB gene sequences derived from strains isolated in Germany and all strains possessing pemIK-Sa1 locus. Clolours of signatures of closely related strainsdenote the host:: unknown, light grey; Bos taurus, skyblue; Gallus gallus, green; Homo sapiens, blue; Meleagris gallopavo, red. The pemIK-Sa1-carrying strains are marked with asterisk.

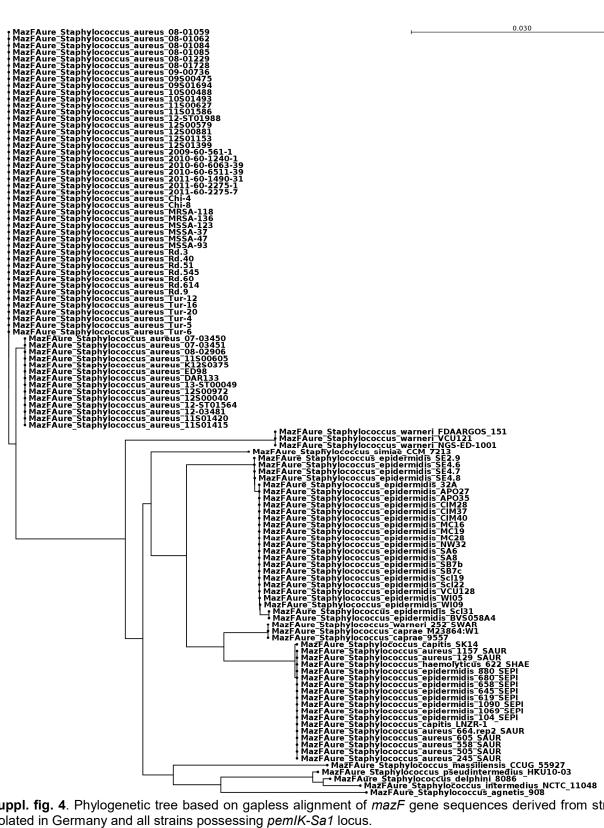
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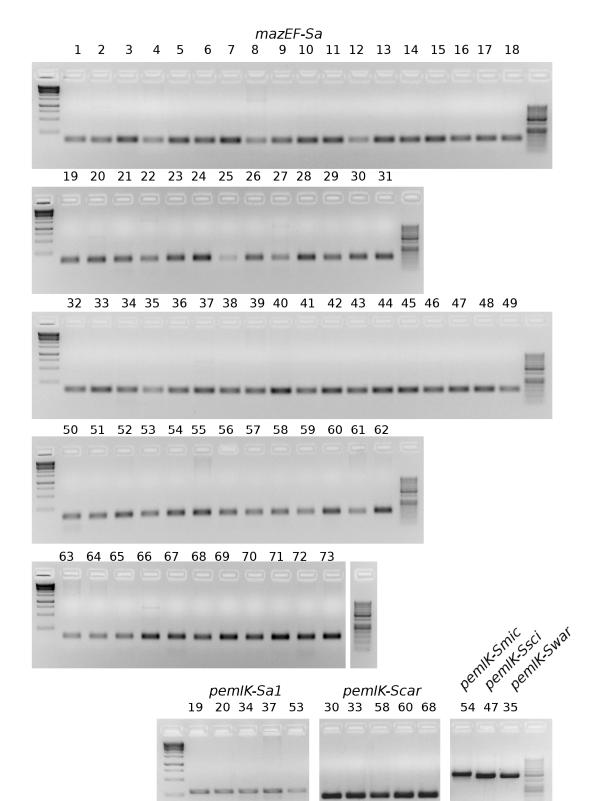
Suppl. fig. 4. Phylogenetic tree based on gapless alignment of mazF gene sequences derived from strains isolated in Germany and all strains possessing pemIK-Sa1 locus.

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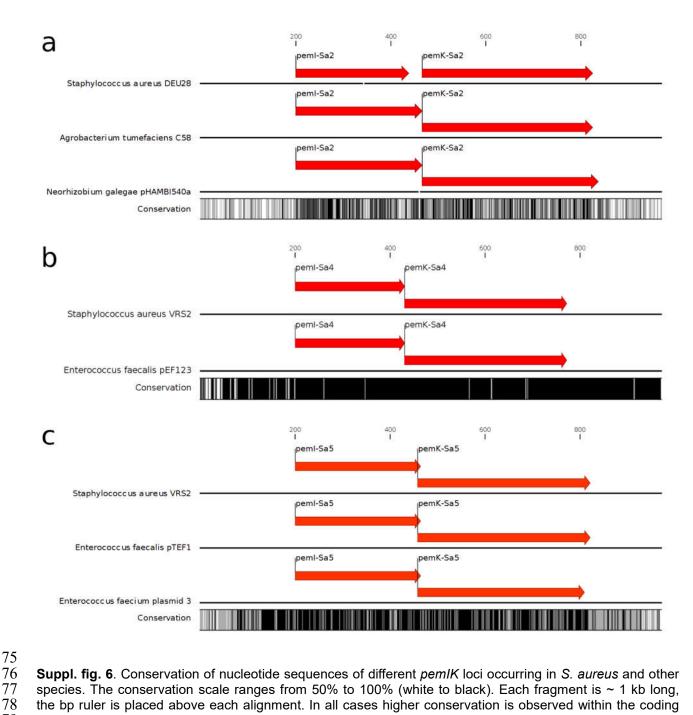
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**Suppl. fig. 5**. Whole-lane images used for preparation of fig. 10: PCR screen for *mazEF/pemIK* loci. The numbers correspond to the number in fig. 10. The first and the last lane: length standards, respectively Gene Ruler 1 kb and Gene Ruler 50 bp (Thermocientific). White gaps: irrelevant lanes were removed. All fragments in one row are fragments of the same gel image.



**Suppl. fig. 6**. Conservation of nucleotide sequences of different *pemIK* loci occurring in *S. aureus* and other species. The conservation scale ranges from 50% to 100% (white to black). Each fragment is ~ 1 kb long, the bp ruler is placed above each alignment. In all cases higher conservation is observed within the coding sequences, which when translated are more than 80% similar. The adjacent non-coding sequences are more diverse, likely due to variability in species-specific promoter and other regulatory sequences. (a) *pemIK-Sa2* loci are the most divergent, still much higher conservation of coding sequences is clearly visible. (b) On the other hand, although *pemIK-Sa4* sequences are highly conserved, there are clear differences in 5'-UTRs. (c) *pemIK-Sa5* loci are more diverse than co-occurring with them *pemIK-Sa4* ones and as in other cases higher variability in non-conding sequences is observed.

## Bibliography

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