

**Table S8**

Details of bacterial strains, plasmids and oligo's used during this study

Strain / Plasmid	Relevant Characteristics	Reference / source
<b><i>E. coli</i></b>		
CC118 $\lambda$ -pir	$\Delta(ara, leu)7697 araD139 \Delta lacX74 galE galK phoA20 thi-1 rpsE rpoB(RfR) argE(am) recA1 \lambda pir^+$	(1)
DH5 $\alpha$	$F^- \phi 80 dlacZ \Delta M15 \Delta(lacZYA-argF)U169 recA1 hsdRI7 (r_k m^+_k) supE44 \lambda^- thi-1 relA1 gyrA96$	(2)
SM10 $\lambda$ -pir	$thi thr leu tonA lacY supE recA::RP4-2-Tc::Mu Km \lambda pir$	(3)
<b><i>B. cenocepacia</i></b>		
<i>B. cenocepacia</i> H111	Clinical isolate from CF patient. Hanover Germany	(4)
<i>B. cenocepacia</i> J2315	Clinical isolate from CF patient. Edinburgh United Kingdom	(5)
<b>Plasmid</b>		
pSC200	<i>PrhaB</i> (rhamnose-inducible), Tp <sup>R</sup>	(6)
pRK2013	ori <sub>colE1</sub> , RK2 derivative, mob <sup>+</sup> tra <sup>+</sup> , Km <sup>R</sup>	(7)
pLG99	Transposon T23 (IS <sub>lacZ</sub> -P <sub>rhaB</sub> out-/FRT) Tp <sup>R</sup>	(8)
pSC200-BCAM0545	pSC200 derivative to create BCAM0545 conditional mutant	This study
pSC200-BCAM0632	pSC200 derivative to create BCAM0632 conditional mutant	This study
pSC200-BCAM0746	pSC200 derivative to create BCAM0746 conditional mutant	This study
pSC200-BCAM0855	pSC200 derivative to create BCAM0855 conditional mutant	This study
pSC200-BCAM0911	pSC200 derivative to create BCAM0911 conditional mutant	This study
pSC200-BCAM0913	pSC200 derivative to create BCAM0913 conditional mutant	This study
pSC200-BCAM0918	pSC200 derivative to create BCAM0918 conditional mutant	This study
pSC200-BCAM0963	pSC200 derivative to create BCAM0963 conditional mutant	This study
pSC200-BCAM0965	pSC200 derivative to create BCAM0965 conditional mutant	This study
pSC200-BCAM0967	pSC200 derivative to create BCAM0967 conditional mutant	This study
pSC200-BCAM0972	pSC200 derivative to create BCAM0972 conditional mutant	This study
pSC200-BCAM0986	pSC200 derivative to create BCAM0986 conditional mutant	This study
pSC200-BCAM0994	pSC200 derivative to create BCAM0994 conditional mutant	This study
pSC200-BCAM0995	pSC200 derivative to create BCAM0995 conditional mutant	This study
pSC200-BCAM0998	pSC200 derivative to create BCAM0998 conditional mutant	This study

pSC200-BCAM1362	pSC200 derivative to create BCAM1362 conditional mutant	This study
pSC200-BCAM1812	pSC200 derivative to create BCAM1812 conditional mutant	This study
pSC200-BCAM1931	pSC200 derivative to create BCAM1931 conditional mutant	This study
pSC200-BCAM2321	pSC200 derivative to create BCAM2321 conditional mutant	This study
pSC200-BCAM2358A	pSC200 derivative to create BCAM2358A conditional mutant	This study
pSC200-BCAM2650	pSC200 derivative to create BCAM2650 conditional mutant	This study
pSC200-BCAM2688	pSC200 derivative to create BCAM2688 conditional mutant	This study
pSC200-BCAM2833	pSC200 derivative to create BCAM2833 conditional mutant	This study

**Table S9**

Details of oligo's and the sequence of each oligo used during this study.

Oligonucleotide	Sequence
<b>Illumina Tn-Seq adapters</b>	
PAIR_ADAPT_6BC##_HI	ACACTCTTTCCCTACACGACGCTCTTCCGATCTBBBBBBT
PAIR_ADAPT_6BC##_LO_5PH	<5' phos>BBBBBBAGATCGGAAGAGCGGTTCAGCAGGAATGCCGAG
<b>Tn-Seq circle method collector probe</b>	
T23_PAIR_COLLECT_1	CTTCGGCGCGCCCTAGGGGATCCTCGGCATTCTGCTGAACCGCTCTTCCGATCT
<b>Tn-Seq amplification primers</b>	
T23_SLXA_PAIR_AmpF_3	AATGATACGGCGACCACCGAGATCTACACTAGAGAATAGGAACTTCGGAATAGGAACTTCTTAGATGTGTATAAGAG
SLXA_PAIR_REV_AMP	CAAGCAGAAGACGGCATAACGAGATCGGTCTCGGCATTCTGCTGAACCGCTCTTCCGATCT
<b>Tn-Seq Sequencing primers</b>	
T23_SEQ_G	AATAGGAACTTCGGAATAGGAACTTCTTAGATGTGTATAAGAGACAG
T23_INDEX_1	CTAGAGAATAGGAACTTCGGAATAGGAACTTCTTAGATGTGTATAAG
PE_READ2_SEQ	CGGTCTCGGCATTCTGCTGAACCGCTCTTCCGATCT
<b>pSC200 conditional mutant primers</b>	
BCAM0545 FW (NdeI)	TAT <u>CATATG</u> CAAACGCATCAGGCCGCC
BCAM0545 Rv (XbaI)	TAT <u>TCTAGA</u> TCAAACAGTGCGTTGCATGAG
BCAM0746 FW (NdeI)	TAT <u>CATATG</u> AGCACGATTCTCGAAAGC
BCAM0746 Rv (XbaI)	TAT <u>TCTAGA</u> CGGCCTGGTTCATGAATT

BCAM0855 FW (NdeI) TATCATATGAATCTGACTATCATCGGCAGC  
BCAM0855 Rv (XbaI) TATTCTAGAGCGCTTCGCGAGCTCCTC  
BCAM0911 FW (NdeI) TATCATATGTACGACTTGCTGAAAACCATC  
BCAM0911 Rv (XbaI) TATTCTAGAACATGTCGTTGTCGTTGAG  
BCAM0913 FW (NdeI) TATCATATGCTCGTTCTCGGCATCGAAA  
BCAM0913 Rv (XbaI) TATTCTAGAGAGCTTCGATACTTCCGGG  
BCAM0918 FW (NdeI) TATCATATGACCAAAAAGCTGAACGAAGTAT  
BCAM0918 Rv (XbaI) TATTCTAGAATCTCGCGCATGTACATC  
BCAM0963 FW (NdeI) TATCATATGAAGAAACTCCTGATCGCTACC  
BCAM0963 Rv (XbaI) TATTCTAGAGCTTCAGCTCGTTTCGAC  
BCAM0967 FW (NdeI) TATCATATGCCGCTGGCGCGCATATTGT  
BCAM0967 Rv (XbaI) TATTCTAGAGCGTGGTAGAAGAGGGAAAG  
BCAM0972 FW (NdeI) TATCATATGACTCCGTCTGATGTTAAAGCC  
BCAM0972 Rv (XbaI) TATTCTAGAACCTTGTACTCTTCGCACG  
BCAM0986 FW (NdeI) TATCATATGGTTCGGCAGCGTCCTGATG  
BCAM0986 Rv (XbaI) TATTCTAGACTCCTGCACCGAACCAT  
BCAM0994 FW (NdeI) TATCATATGAGCTGGCTCGACAACTGTT  
BCAM0994 Rv (XbaI) TATTCTAGACCATCTGCATCAGCGACA  
BCAM0995 FW (NdeI) TATCATATGAGCACTTTTCCCCTCTCGA  
BCAM0995 Rv (XbaI) TATTCTAGAAATTCGGTATGGTCGATGT  
BCAM1362 FW (NdeI) TATCATATGAGCGTAAAAAGAAGACCCAC  
BCAM1362 Rv (XbaI) TATTCTAGACCGCTTGAGCAGCACGAAC  
BCAM1812 FW (NdeI) TATCATATGACCGAACTTCTTCACGGCGA  
BCAM1812 Rv (XbaI) TATTCTAGACTGGCCGTAGCGCTCCG  
BCAM1931 FW (NdeI) TATCATATGAAACAAGACTCTGATCGTTGCA  
BCAM1931 Rv (XbaI) TATTCTAGACGCGACATCACCGTTCGTG  
BCAM2321 FW (NdeI) TATCATATGAATACGATCAAACGAATCGATC  
BCAM2321 Rv (XbaI) TATTCTAGACGCGTTGTCGGGCACGAA  
BCAM2358A FW (NdeI) TATCATATGTCGACCCACATCTGTTTCGC  
BCAM2358A Rv (XbaI) TATTCTAGACGCGCGCGGGCCGAGG  
BCAM2650 FW (NdeI) TATCATATGATCATCAAACCGCGCGT

BCAM2650 Rv (XbaI)

TATTCTAGAGTGCCGTCGATCTCTTCCT

BCAM2688 FW (NdeI)

TATCATATGGCCCGTTTCCCATTCCGT

BCAM2688 Rv (XbaI)

TATTCTAGAAGGATGTTTCGTCAGTTGCAG

## Strain references

1. Herrero M, de Lorenzo V, Timmis KN. 1990. Transposon vectors containing non-antibiotic resistance selection markers for cloning and stable chromosomal insertion of foreign genes in gram-negative bacteria. *J Bacteriol* 172:6557-67.
2. Hanahan D. 1983. Studies on transformation of *Escherichia coli* with plasmids. *J Mol Biol* 166:557-80.
3. Simon R, Priefer U, Puhler A. 1983. A Broad Host Range Mobilization System for In Vivo Genetic Engineering: Transposon Mutagenesis in Gram Negative Bacteria. *Nat Biotech* 1:784-791.
4. Romling U, Fiedler B, Bosshammer J, Grothues D, Greipel J, von der Hardt H, Tummler B. 1994. Epidemiology of chronic *Pseudomonas aeruginosa* infections in cystic fibrosis. *J Infect Dis* 170:1616-21.
5. Govan JR, Brown PH, Maddison J, Doherty CJ, Nelson JW, Dodd M, Greening AP, Webb AK. 1993. Evidence for transmission of *Pseudomonas cepacia* by social contact in cystic fibrosis. *Lancet* 342:15-9.
6. Ortega XP, Cardona ST, Brown AR, Loutet SA, Flannagan RS, Campopiano DJ, Govan JR, Valvano MA. 2007. A putative gene cluster for aminoarabinose biosynthesis is essential for *Burkholderia cenocepacia* viability. *J Bacteriol* 189:3639-44.
7. Figurski DH, Helinski DR. 1979. Replication of an origin-containing derivative of plasmid RK2 dependent on a plasmid function provided in trans. *Proc Natl Acad Sci U S A* 76:1648-52.
8. Gallagher LA, Ramage E, Patrapuvich R, Weiss E, Brittnacher M, Manoel C. 2013. Sequence-Defined transposon mutant library of *Burkholderia thailandensis*. *mBio* 4.