2	Fig. S1. Bacterial growth, bacterial binding to H292 monolayers, and PMN transepithelial
3	migration are unaffected by pan-PLA ₂ or cPLA ₂ α inhibitors. (A) S. pneumoniae TIGR4 was
4	grown as described in <i>Materials and Methods</i> , in the presence or absence of pan-PLA ₂ inhibitors
5	ACA and ONO-RS-082 or of the cPLA ₂ α inhibitor. Growth was measured using a
6	spectrophotometer. Shown is a representative of two experiments. (B) H292 monolayers were
7	treated with pan-PLA ₂ inhibitors ACA (100 μ M) or ONO-RS-082 (20 μ M); or the cPLA ₂ α
8	inhibitor ($6\mu M$) (please see <i>Materials and Methods</i>). Monolayers were washed and infected with
9	S. pneumoniae TIGR4. The number of bacteria bound to monolayers was determined by viable
10	counts. Shown is a representative of two experiments. (C) H292 monolayers were treated with
11	pan-PLA ₂ inhibitors ACA (100 μ M) or ONO-RS-082 (20 μ M); or the DAG lipase inhibitor RHC-
12	80267 (labeled as RHC, 100 μ M); or the cPLA ₂ α inhibitor (6 μ M) (please see <i>Materials and</i>
13	Methods). The monolayers were washed and PMN transepithelial migration in response to fMLP
14	was quantified by myeloperoxidase (MPO) assay. Monolayers treated with DMSO and HBSS
15	+Ca/Mg were used as the vehicle control and negative control, respectively. Shown is a
16	representative of two experiments.
17	Figure S2. cPLA ₂ α promotes pulmonary inflammation in <i>S. pneumoniae</i> -infected mice. Mice
18	$(cPLA_2\alpha^{-2})$ or their WT littermates) were mock-infected with PBS or infected intratracheally with
19	S. pneumoniae TIGR4, as detailed in Materials and Methods. Mice were sacrificed and
20	bronchoalveolar lavage fluid (BALF) and lungs were collected. (A) Myeloperoxidase (MPO) in
21	the homogenized lung samples and BALF was determined biochemically. Figures are

representative of two independent experiments. **p < 0.005, ***p < 0.001, using one-way

ANOVA. (B) Serial dilutions of the lung homogenate and the BALF were plated on blood agar
plates and bacterial CFUs were counted, ns= not significant.







30 Table S1

	Cytotoxicity
HBSS, 2.5h	1.4 (0.4)
S. pneumoniae TIGR4, 2.5h	1.84 (0.4)
<i>S. pneumoniae</i> TIGR4, 5h	2.82 (0.8)
2% Triton X-100, 2.5h	68 (13.7)

31

Cytotoxicity in H292 monolayers in response to *S. pneumoniae* TIGR4 infection was assessed using the lactate dehydrogenase (LDH) assay [1]. Infection or treatment time is denoted in the table. Cytotoxicity was calculated as the amount of LDH in the supernatant as a percentage of total LDH (LDH inside the cell as well as released into the supernatant). Monolayers treated with Triton X-100 and HBSS (+Ca/Mg) were used as positive and negative controls, respectively. Shown is a representative of three experiments. Standard deviations are reported in parentheses.

Table S2

	HRP (ng/ml)
HBSS, 3.5h	7.4 (0.3)
<i>S. pneumoniae</i> TIGR4, 1.5h	11.67 (3)
S. pneumoniae TIGR4, 2.5h	8.216 (0.77)
S. pneumoniae TIGR4, 3.5h	12.73 (0.7)
Trypsin-EDTA, 2.5h	57.42 (2)

Effect of *S. pneumoniae* infection on the barrier integrity of H292 monolayers was assessed by
horseradish peroxidase (HRP) assay [1]. Barrier integrity was measured as the amount of HRP
(ng/ml) that could breach the H292 monolayer following infection. Monolayers treated with
trypsin-EDTA and HBSS (+Ca/Mg) were used as positive and negative controls, respectively.
Shown is a representative of two experiments. Standard deviations are reported in parentheses.

45 Table S3

	Cytotoxicity
HBSS	1.5 (0.4)
ACA (100µM)	2.7 (0.9)
ONO-RS-082 (20µM)	2.1 (0.8)
RHC-80267 (100µM)	1.74 (0.9)
$cPLA_2$ - α inhibitor (6 μ M)	2.7 (1.3)
2% Triton X-100	66.37 (13.5)

46

<sup>Cytotoxicity of H292 monolayers in response to pharmacological inhibitors used in this study
was assessed using LDH assay [1]. Inhibitor doses are denoted in the table. Cytotoxicity is
calculated as the amount of LDH in the supernatant as a percentage of total LDH (LDH inside
the cell as well as released into the supernatant). Monolayers treated with Triton X-100 and
HBSS (+Ca/Mg) were used as positive and negative controls, respectively. Shown is a
representative of two experiments. Standard deviations are reported in parentheses.</sup>

54 Table S4

	HRP (ng/ml)
HBSS	10.6 (2.5)
ACA (100µM)	13.07 (3.8)
ONO-RS-082(20µM)	10.48 (3.4)
RHC-80267 (100µM)	17.74 (6.2)
$cPLA_2$ - α inhibitor (6 μ M)	12.43 (4.5)
Trypsin-EDTA	59.8 (9.8)

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Effect of the pharmacological inhibitors on the barrier integrity of H292 monolayers was
assessed by horseradish peroxidase (HRP) assay [1]. Inhibitor doses are denoted in the table.
Barrier integrity was measured as the amount of HRP (ng/ml) that could breach the H292
monolayer in following the inhibitor treatment. Monolayers treated with trypsin-EDTA and
HBSS (+Ca/Mg) were used as positive and negative controls, respectively. Shown is a
representative of two experiments. Standard deviations are reported in parentheses.
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