

Figure S1

Hyaluronidase production is unaffected by *rocA* allele status.

Parental isolate MGAS2221, *rocA* deletion mutant derivative 2221Δ*rocA*, and complemented mutant strain 2221Δ*rocA*::*rocA*^{M1} were compared in a plate assay. Shown are the averages (± standard deviation) of duplicate samples ran in duplicate.

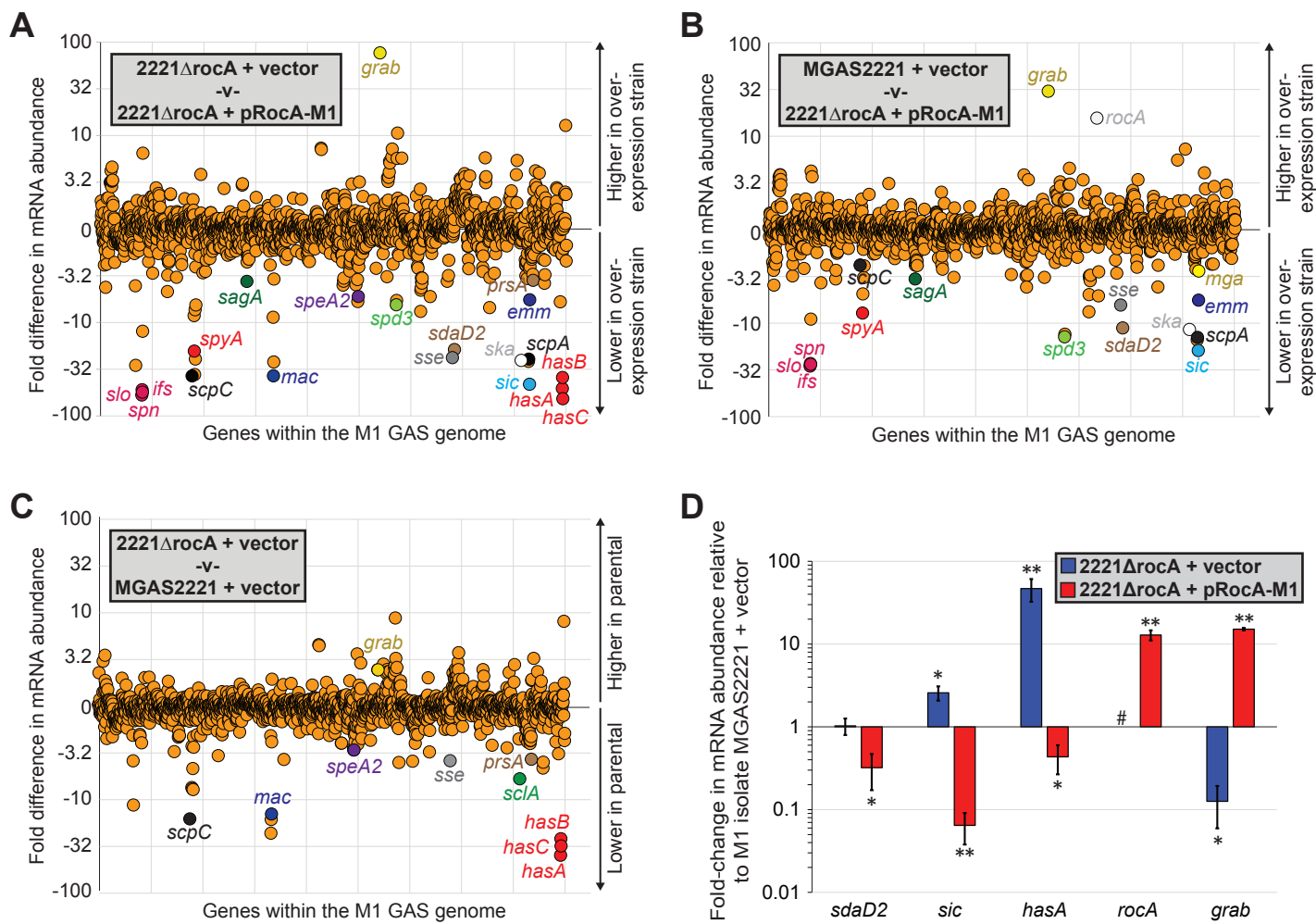


Figure S2

Roca overexpression selectively modifies transcript abundance.

(A-C) Summary of RNAseq data. Shown are pair-wise comparisons of the transcriptomes between the *rocA* deletion mutant strain containing either empty vector (pDCBB) or *rocA*-expressing plasmid (pRocA-M1) (A), between vector-containing MGAS2221 and 2221ΔrocA containing pRocA-M1 (B), and between vector-containing derivatives of MGAS2221 and 2221ΔrocA (C). The relative expression levels of all genes are graphed, with each represented by a circle. Select mRNAs of interest have been colored and labeled. (D) Taqman-based quantitative RT-PCR data confirming that Roca regulates the abundance of virulence factor-encoding mRNAs in GAS. The abundance of *sdaD2*, *sic*, *hasA*, *rocA*, and *grab* mRNAs were determined from triplicate exponential phase GAS cultures, with the mean (\pm standard deviation) shown. The hashtag highlights the lack of *rocA* transcript in the *rocA* deletion mutant strain. The asterisks highlight statistical significance relative to the parental M1 isolate (*t*-test, * $P < 0.05$, ** $P < 0.01$).

Feature ID	Fold change in transcript level in MGAS2221 relative to 2221ΔrocA
<i>M5005_Spy1541</i>	7.33
<i>scrK</i>	6.75
<i>spd3</i>	6.49
<i>M5005_Spy1746</i>	6.33
<i>endoS</i>	5.88
<i>M5005_Spy0859</i>	5.78
<i>M5005_Spy1170</i>	5.41
<i>scrA</i>	5.22
<i>M5005_Spy0028</i>	5.16
<i>nagB</i>	4.96
<i>xpt</i>	4.49
<i>M5005_Spy1668</i>	4.39
<i>salB</i>	4.30
<i>M5005_Spy1152</i>	4.22
<i>M5005_Spy1745</i>	4.19
<i>purD</i>	4.05
<i>M5005_Spy0169</i>	3.96
<i>M5005_Spy0023</i>	3.85
<i>M5005_Spy1750</i>	3.71
<i>purE</i>	3.61
<i>purM</i>	3.40
<i>purK</i>	3.40
<i>M5005_Spy0022</i>	3.30
<i>purF</i>	3.15
<i>sagD</i>	3.07
<i>smf</i>	3.00
<i>M5005_Spy0357</i>	-3.00
<i>M5005_Spy1018</i>	-3.17
<i>M5005_Spy1556</i>	-3.35
<i>M5005_Spy1015</i>	-3.63
<i>M5005_Spy0665</i>	-4.41
<i>speA2</i>	-4.81
<i>spyCEP</i>	-6.39
<i>M5005_Spy0355</i>	-6.41
<i>sclA</i>	-6.56
<i>M5005_Spy0354</i>	-7.23
<i>M5005_Spy0669</i>	-7.59
<i>M5005_Spy0115</i>	-11.27
<i>hasC</i>	-12.49
<i>M5005_Spy0352</i>	-13.16
<i>M5005_Spy0143</i>	-13.47
<i>hasA</i>	-13.80
<i>hasB</i>	-14.11
<i>M5005_Spy0666</i>	-18.93
<i>mac</i>	-23.12
<i>M5005_Spy0667</i>	-27.10
<i>M5005_Spy0144</i>	-58.73

Table S1

Transcripts that are differentially regulated between strains MGAS2221 and 2221ΔrocA.

Feature ID	Fold change in transcript level in MGAS2221 relative to 2221 Δ rocA::rocA ^{M1}
<i>M5005_Spy0143</i>	3.07

Table S2

Transcripts that are differentially regulated between strains MGAS2221 and 2221 Δ rocA::rocA^{M1}.

Feature ID	Fold change in transcript level in 2221ΔrocA::rocA ^{M1} relative to 2221ΔrocA
<i>spd3</i>	5.79
<i>M5005_Spy1541</i>	5.50
<i>M5005_Spy1170</i>	5.46
<i>M5005_Spy1152</i>	5.09
<i>M5005_Spy0859</i>	4.95
<i>scrK</i>	4.84
<i>M5005_Spy0028</i>	4.72
<i>nagB</i>	4.59
<i>M5005_Spy1746</i>	4.42
<i>xpt</i>	4.41
<i>endoS</i>	4.01
<i>M5005_Spy1668</i>	3.80
<i>scrA</i>	3.43
<i>purD</i>	3.26
<i>grab</i>	3.23
<i>metB</i>	3.21
<i>purE</i>	3.04
<i>M5005_Spy0023</i>	3.00
<i>sic1.01</i>	-3.19
<i>M5005_Spy0665</i>	-3.38
<i>speJ</i>	-3.52
<i>M5005_Spy1407</i>	-3.57
<i>M5005_Spy1556</i>	-3.70
<i>M5005_Spy0113</i>	-3.88
<i>M5005_Spy0357</i>	-4.36
<i>M5005_Spy0114</i>	-5.14
<i>speA2</i>	-5.38
<i>M5005_Spy0354</i>	-9.54
<i>M5005_Spy0669</i>	-9.72
<i>M5005_Spy0355</i>	-12.28
<i>spyCEP</i>	-13.61
<i>sclA</i>	-17.98
<i>M5005_Spy0115</i>	-20.67
<i>M5005_Spy0666</i>	-20.75
<i>M5005_Spy0352</i>	-24.01
<i>hasC</i>	-24.76
<i>hasA</i>	-27.20
<i>hasB</i>	-27.36
<i>M5005_Spy0667</i>	-38.63
<i>M5005_Spy0143</i>	-41.30
<i>mac</i>	-41.79
<i>M5005_Spy0144</i>	-58.73

Table S3

Transcripts that are differentially regulated between strains 2221ΔrocA and 2221ΔrocA::rocA^{M1}.

Feature ID	Fold change in transcript level in MGAS2221 + vector relative to 2221ΔrocA + vector
<i>M5005_Spy_1178</i>	9.07
<i>M5005_Spy_T0067</i>	8.35
<i>xpt</i>	4.70
<i>M5005_Spy_0859</i>	4.58
<i>M5005_Spy_1440</i>	3.86
<i>M5005_Spy_1025</i>	3.73
<i>M5005_Spy_T0020</i>	3.62
<i>M5005_Spy_1189</i>	3.35
<i>M5005_Spy_1744</i>	3.35
<i>M5005_Spy_T0005</i>	3.12
<i>M5005_Spy_1437</i>	3.04
<i>M5005_Spy_1843</i>	-3.13
<i>M5005_Spy_0459</i>	-3.28
<i>prsA_2</i>	-3.58
<i>M5005_Spy_T0048</i>	-3.60
<i>sse</i>	-3.71
<i>M5005_Spy_0114</i>	-3.73
<i>M5005_Spy_1263</i>	-3.76
<i>M5005_Spy_1784</i>	-3.86
<i>M5005_Spy_1206</i>	-3.88
<i>dexS</i>	-4.45
<i>M5005_Spy_0354</i>	-4.51
<i>sclA</i>	-5.80
<i>M5005_Spy_0350</i>	-7.20
<i>M5005_Spy_0352</i>	-7.42
<i>M5005_Spy_0355</i>	-7.57
<i>M5005_Spy_0115</i>	-11.08
<i>mac</i>	-13.80
<i>scpC</i>	-15.57
<i>M5005_Spy_0667</i>	-15.81
<i>M5005_Spy_0666</i>	-22.13
<i>hasB</i>	-27.00
<i>hasC</i>	-32.28
<i>hasA</i>	-40.54

Table S4

Transcripts that are differentially regulated between strains MGAS2221 (+ vector) and 2221ΔrocA (+ vector).

Feature ID	Fold change in transcript level in 2221ΔrocA + pRocA relative to MGAS2221 + vector
<i>grab</i>	30.28
<i>rocA</i>	15.64
<i>M5005_Spy_1668</i>	7.28
<i>salX</i>	5.56
<i>M5005_Spy_1206</i>	4.54
<i>metB</i>	4.23
<i>M5005_Spy_1746</i>	4.07
<i>M5005_Spy_0028</i>	3.87
<i>M5005_Spy_0995</i>	3.85
<i>purD</i>	3.82
<i>M5005_Spy_0152</i>	3.68
<i>purF</i>	3.67
<i>purM</i>	3.51
<i>purE</i>	3.44
<i>M5005_Spy_1029</i>	3.43
<i>M5005_Spy_1477</i>	3.42
<i>M5005_Spy_1263</i>	3.41
<i>M5005_Spy_1152</i>	3.24
<i>M5005_Spy_0023</i>	3.19
<i>M5005_Spy_0459</i>	3.12
<i>purH</i>	3.12
<i>M5005_Spy_1210</i>	3.07
<i>M5005_Spy_1204</i>	3.04
<i>purK</i>	3.02
<i>coaA</i>	-3.02
<i>M5005_Spy_1023</i>	-3.06
<i>M5005_Spy_1291</i>	-3.08
<i>M5005_Spy_1287</i>	-3.11
<i>M5005_Spy_T0034</i>	-3.12
<i>M5005_Spy_1008</i>	-3.14
<i>M5005_Spy_T0047</i>	-3.24
<i>M5005_Spy_1091</i>	-3.24
<i>M5005_Spy_0355</i>	-3.26
<i>M5005_Spy_1288</i>	-3.27
<i>sagA</i>	-3.37
<i>M5005_Spy_0247</i>	-3.37
<i>M5005_Spy_1289</i>	-3.38
<i>M5005_Spy_1290</i>	-3.52
<i>M5005_Spy_0189</i>	-3.70
<i>M5005_Spy_T0008</i>	-3.92
<i>M5005_Spy_1556</i>	-4.10
<i>M5005_Spy_1018</i>	-4.12
<i>M5005_Spy_0352</i>	-4.86
<i>M5005_Spy_1286</i>	-4.95

<i>M5005_Spy_T0053</i>	-5.10
<i>emm1.0</i>	-5.68
<i>Sse</i>	-6.41
<i>M5005_Spy_1285</i>	-7.56
<i>spyA</i>	-7.81
<i>M5005_Spy_0142</i>	-9.06
<i>sdaD2</i>	-11.26
<i>ska</i>	-11.70
<i>M5005_Spy_1170</i>	-13.14
<i>spd3</i>	-14.02
<i>scpA_2</i>	-14.23
<i>M5005_Spy_1714</i>	-15.12
<i>sic1.01</i>	-19.63
<i>ifs</i>	-27.07
<i>slo</i>	-27.61
<i>nga</i>	-28.80

Table S5

Transcripts that are differentially regulated between strains MGAS2221 (+ vector) and 2221 Δ rocA (+ pRocA).

Feature ID	Fold change in transcript level in 2221ΔrocA + pRocA relative to 2221ΔrocA + vector
<i>grab</i>	76.61
<i>M5005_Spy_T0067</i>	12.82
<i>M5005_Spy_1178</i>	10.63
<i>xpt</i>	7.34
<i>M5005_Spy_1153</i>	7.18
<i>M5005_Spy_0859</i>	7.09
<i>metB</i>	6.47
<i>M5005_Spy_1152</i>	5.91
<i>scrK</i>	5.78
<i>M5005_Spy_1210</i>	5.73
<i>M5005_Spy_1174</i>	5.69
<i>M5005_Spy_1668</i>	5.41
<i>M5005_Spy_1143</i>	5.30
<i>M5005_Spy_1144</i>	5.08
<i>purD</i>	4.60
<i>purM</i>	4.20
<i>M5005_Spy_1744</i>	4.19
<i>M5005_Spy_1142</i>	4.06
<i>M5005_Spy_1760</i>	3.97
<i>M5005_Spy_1457</i>	3.91
<i>M5005_Spy_0376</i>	3.84
<i>M5005_Spy_1664</i>	3.82
<i>M5005_Spy_1477</i>	3.71
<i>scrA</i>	3.62
<i>M5005_Spy_1459</i>	3.62
<i>nagB</i>	3.57
<i>M5005_Spy_1135</i>	3.54
<i>endoS</i>	3.53
<i>M5005_Spy_1437</i>	3.46
<i>M5005_Spy_1440</i>	3.36
<i>purK</i>	3.33
<i>purE</i>	3.30
<i>M5005_Spy_0028</i>	3.25
<i>M5005_Spy_1137</i>	3.22
<i>M5005_Spy_1025</i>	3.18
<i>M5005_Spy_0022</i>	3.18
<i>purN</i>	3.16
<i>purF</i>	3.15
<i>M5005_Spy_1541</i>	3.13
<i>M5005_Spy_0023</i>	3.06
<i>M5005_Spy_1463</i>	3.05
<i>agaS</i>	3.05
<i>cfa</i>	3.03
<i>M5005_Spy_1746</i>	3.01
<i>M5005_Spy_1731</i>	-3.06
<i>M5005_Spy_1784</i>	-3.09
<i>dacA2</i>	-3.12
<i>M5005_Spy_1391</i>	-3.15
<i>dppA</i>	-3.17
<i>prsA_1</i>	-3.24
<i>M5005_Spy_0281</i>	-3.29
<i>mutY</i>	-3.31
<i>M5005_Spy_1288</i>	-3.32
<i>M5005_Spy_1291</i>	-3.36

<i>ciaR</i>	-3.36
<i>M5005_Spy_1709</i>	-3.38
<i>M5005_Spy_0669</i>	-3.43
<i>M5005_Spy_1007</i>	-3.47
<i>prsA_2</i>	-3.55
<i>ciaH</i>	-3.62
<i>M5005_Spy_1386</i>	-3.63
<i>sagA</i>	-3.63
<i>M5005_Spy_1289</i>	-3.70
<i>M5005_Spy_1290</i>	-3.80
<i>pepO</i>	-3.81
<i>speA2</i>	-3.96
<i>coaA</i>	-3.98
<i>M5005_Spy_0114</i>	-4.05
<i>M5005_Spy_T0008</i>	-4.22
<i>M5005_Spy_1287</i>	-4.26
<i>M5005_Spy_1286</i>	-4.52
<i>M5005_Spy_0247</i>	-4.67
<i>M5005_Spy_1018</i>	-5.27
<i>M5005_Spy_T0053</i>	-5.28
<i>emm1.0</i>	-5.72
<i>M5005_Spy_1170</i>	-5.82
<i>M5005_Spy_1008</i>	-5.97
<i>M5005_Spy_1843</i>	-6.02
<i>spd3</i>	-6.46
<i>M5005_Spy_0357</i>	-6.50
<i>M5005_Spy_1556</i>	-6.93
<i>M5005_Spy_0354</i>	-7.32
<i>M5005_Spy_1285</i>	-7.62
<i>M5005_Spy_0350</i>	-8.44
<i>M5005_Spy_0666</i>	-8.79
<i>M5005_Spy_0142</i>	-12.43
<i>sdaD2</i>	-19.48
<i>spyA</i>	-20.27
<i>Sse</i>	-23.79
<i>M5005_Spy_0355</i>	-24.64
<i>scpA_2</i>	-24.69
<i>ska</i>	-25.47
<i>M5005_Spy_1714</i>	-26.11
<i>mac</i>	-26.45
<i>M5005_Spy_0115</i>	-28.82
<i>M5005_Spy_0352</i>	-36.11
<i>M5005_Spy_0667</i>	-37.29
<i>spyCEP</i>	-37.35
<i>hasB</i>	-38.83
<i>sic1.01</i>	-45.90
<i>hasA</i>	-50.89
<i>ifs</i>	-52.48
<i>slo</i>	-55.79
<i>nga</i>	-59.74
<i>hasC</i>	-65.58

Table S6

Transcripts that are differentially regulated between strains 2221 Δ rocA (+ vector) and 2221 Δ rocA (+ pRocA).

Primer name	Sequence (5' - 3')	Use
HASATMF	ATGATCGATGTTTAACAAATTATGCTATTG	Taqman primer for <i>hasA</i>
HASATMR	TAAATAACTTTTTAATTGGAAAGGTACATCAG	Taqman primer for <i>hasA</i>
HASATMP	ACGCACTGTCTACCAATCAACAGCTAGATGTG	Taqman probe for <i>hasA</i>
PROSTMF	TACCACTGGCAAATCGTACC	Taqman primer for <i>proS</i>
PROSTM	CATTCAACAGCACCGATCT	Taqman primer for <i>proS</i>
PROSTMP	CACGCATGATGGTCTTGAATTTCTCA	Taqman probe for <i>proS</i>
SCPCTMF	AAGGAGCTTGGGACAAGGGATA	Taqman primer for <i>scpC</i>
SCPCTMR	TGATGGGCCGGATCGA	Taqman primer for <i>scpC</i>
SCPCTMP	CAATAACTGCGACAACCTTGCCTTGTCTT	Taqman probe for <i>scpC</i>
SLOTMF	GACCTTTAAAGAGTTGCAACGAAAA	Taqman primer for <i>slo</i>
SLOTMR	GACCATAAGCTACGTTACTCACAAAGA	Taqman primer for <i>slo</i>
SLOTMP	TGTCAGCAATGAAGCCCGCC	Taqman probe for <i>slo</i>
SPD3TMF	GGCGATCTTGACAATCTGCAA	Taqman primer for <i>spd3</i>
SPD3TMR	CAGCCAGGAGGATTGAATTTAAGT	Taqman primer for <i>spd3</i>
SPD3TMP	CCAACTTCGCACATCCAGCTAAAAG	Taqman probe for <i>spd3</i>
SKATMF	CGGCTACTTTGAGGTCATTGATT	Taqman primer for <i>ska</i>
SKATMR	CCGAACCATCTTTGTCAGCAA	Taqman primer for <i>ska</i>
SKATMP	CAAGCGATGCAACCATCTAGTATCGAAAC	Taqman probe for <i>ska</i>
ROCATMF	AGGGCTATAAGCGCAAAGAA	Taqman primer for <i>rocA</i>
ROCATMR	GGCTTTCTTCCAGACCAT	Taqman primer for <i>rocA</i>
ROCATMP	TGAGCCAACATCACAACATCAAGAATG	Taqman probe for <i>rocA</i>
GRABTMF	GCATCAGTATTAGTCGGTTCAACAGT	Taqman primer for <i>grab</i>
GRABTMR	GGTCCGCCATTTGGAATAA	Taqman primer for <i>grab</i>
GRABTMP	TGTTGACTCACCTATCGAACAGCCTCGA	Taqman probe for <i>grab</i>
SICTMF	CTGAGGACACCCCTCGTTTC	Taqman primer for <i>sic</i>
SICTMR	TCTGTGGATTTTTTGAGGAGTATG	Taqman primer for <i>sic</i>
SICTMP	TGAACCTCGTGTGACAGAAAACCGCA	Taqman probe for <i>sic</i>
SDAD2TMF	AAGAAAATGAAATCGAACTACTGATGAC	Taqman primer for <i>sdad2</i>
SDAD2TMR	GCCATTACTTGCCACATATACAG	Taqman primer for <i>sdad2</i>
SDAD2TMP	AAAACGAGCCTGACACGGATGCACTAAA	Taqman probe for <i>sdad2</i>
SCLATMF	TGCAGAAAAACCTGCTGACAA	Taqman primer for <i>sclA</i>
SCLATMR	GGTGTCTTAGCCATATTGCCATT	Taqman primer for <i>sclA</i>
SCLATMP	AGCTAACCAAACGCCAGAACGCCG	Taqman probe for <i>sclA</i>
ENDOSTMF	CGCAAGGACGAGGACAAAGC	Taqman primer for <i>endoS</i>
ENDOSTMR	CCCACCTGTCTTAGTTGCCA	Taqman primer for <i>endoS</i>
ENDOSTMP	TGCATACCGTTCGGCACGCGTTCCAGT	Taqman probe for <i>endoS</i>
SAGCTMF	CTGCTGAGTTGCTAGCTGAACAAC	Taqman primer for <i>sagC</i>
SAGCTMR	CTTTGGTAATCGCACAAAGGCATCTG	Taqman primer for <i>sagC</i>
SAGCTMP	CGGCGATGCTCAAAGCATCTAAACGCGAAC	Taqman probe for <i>sagC</i>
UNR24	AGTCATGCATAAAAAGAGTCTGGGACTATTGTCTCAG	To be used with UNR25 to amplify <i>rocA</i> from M1 GAS genome in order to make plasmid pRocA-M1 (or from M3/M18 genomes to make pRocA-M3/M18)
UNR25	AGTCAGATCTGAGATCAACAACCTGATAAATCTGAG	To be used with UNR24 to amplify <i>rocA</i> from M1 GAS genome in order to make plasmid pRocA-M1 (or from M3/M18 genomes to make pRocA-M3/M18)
UNR544	TGATGTTAAAGGTATGAATAATCACTCAAAATCAACTTAAGAGTCTGAGACAATAG	To be used with UNR547, UNR548, UNR608 and UNR610 for amplifying truncated <i>rocA</i> alleles for creation of plasmids pRocA-M1 ¹⁻³²⁸ , pRocA-M1 ¹⁻²⁶⁷ , pRocA-M1 ¹⁻²²¹ , and pRocA-M1 ¹⁻⁸⁹
UNR547	GATTTTGAGTGATTATTCATACCTTTAACATCACTTCTTATTTTTGCTCTCAATTAATTTAGCTAAACAATATG	To be used with UNR544 for amplifying a truncated <i>rocA</i> allele for the creation of plasmid pRocA-M1 ¹⁻³²⁸
UNR548	GATTTTGAGTGATTATTCATACCTTTAACATCAATCAATTTTGTTCGATTCCAATCTTAAGACGACTAGACACGGTAGTTAATAGC	To be used with UNR544 for amplifying a truncated <i>rocA</i> allele to create plasmid pRocA-M1 ¹⁻²⁷⁶
UNR608	GATTTTGAGTGATTATTCATACCTTTAACATCAACGCTCTAAATGAA TTAATAAAGTAAGAAAAC	To be used with UNR544 for amplifying a truncated <i>rocA</i> allele to create plasmid pRocA-M1 ¹⁻²²¹
UNR610	GATTTTGAGTGATTATTCATACCTTTAACATCAAAACTTAGCTTTTATTGATTTACCATAG	To be used with UNR544 for amplifying a truncated <i>rocA</i> allele to create plasmid pRocA-M1 ¹⁻⁸⁹

Table S7
Table of primers and probes used in this study.