# Unusual metabolic diversity of hyperalkaliphilic microbial communities associated with subterranean serpentinization

### at The Cedars

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### SUPPLEMENTARY INFORMATION

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# Supplementary Figure 1 | Genome clustering from the assembled contigs of metagenomic sequences of The Cedars springs GPS1 and BS5 in 2012.

Samples collected from GPS1 and BS5 in 2012 are shown as GPS12 or BS5B12 respectively. Colored circles indicate clusters of individual bin-genomes. Bin-genomes were identified using the estimated taxonomic classification (color of dots), length (size of dots), GC content and mean coverage of contigs. Colors for the circles denote groundwater sources where the bin-genomes are associated, i.e. red circles show the deep members, while blue circles show the shallow member. Numbers/letters shown near the circles correspond to the bin-genome IDs in Fig. 3A. (A, D) Clustering based on the mean coverage vs. GC% plot. (B) Clustering for the members of deep groundwater based on coverage-differences between the two springs. (E) Clustering for the members of shallow groundwater based on the coveragedifferences of two years' metagenomes. (C, F) Relative frequencies of each strain within GPS12 (C) and BS5B12 (F) estimated from a relative abundance of 16S rRNA gene (16S analysis), percentage of mapped raw reads to each bin-genome (Raw reads), an average value of reads per kilobase per million mapped reads (RPKMs) of each bin-genome (Coverage), an average RPKM value of 12 single-copied housekeeping genes (Core genes), and a RPKM value of twelve individual single-copied housekeeping genes.



# Supplementary Figure 2 | Blastn similarity of bin-genomes of deep members recovered from different metagenomic dataset from different springs (GPS1 and BS5) and years (2011 and 2012).

Due to the low coverage, bin-genomes of unc8 and fir15 were not allowed to recover from metagenomic data set of BS5 (2012) or GPS1 (2012) respectively.



Supplementary Figure 3 | Maximum Likelyhood RPS3 phylogeney of OD1 genomes recovered from The Cedars. Tree topologies are supported by bootstrap values for 100 replicates and the values greater than 70 were indicated

Red font denotes bin-genome ID from The Cedars serpentinization springs



# Supplementary Figure 4 | Intron-encoding 16S rRNA gene from the Candidate phylum OD1 bin-genomes.

numbers). Insertion regions are shown by red bars, while ORFs encoded in the insertion regions were identified by BLASTX against nr Alignment of 16S rRNA encoding region corresponding to Escherichia coli K12 gene positions (position described above bars by blue (orange bars) and applied to pfam search for functional annotation (orange words).



# Supplementary Figure 5 | GC content versus genome size for the bin-genomes of The Cedars members with reference genome sequences in each taxonomic group.

Red, blue and black dots denote the bin-genomes from The Cedars organisms, bacterial endosymbionts/obligate parasite of eukaryotes, or genomes from free-living prokaryote respectively. Genome size and GC contents information for the references were obtained from all completed bacterial and archaeal genomes in the NCBI genome database. Total numbers of reference genomes analyzed here are 3170.



# Supplementary Figure 6 | Maximum likelihood phylogenetic tree constructed for the A-type ATPase subunit A (NtpA).

Red letters show the position of NtpA from The Cedars organisms. Blue colored areas and nodes indicate bacterial phyla while green colored areas and nodes indicate the archaeal phyla. Reference sequences were obtained from KEGG database (K02117).



### Supplementary Figure 7 | Phylogenetic analysis of NtpA.

Red letters show the position of NtpA from The Cedars organisms. Closest relatives in NCBI database (nr) were employed as the reference sequences.



Supplementary Figure 8 | iRep values were determined for whole community (wc) and OD1 in each metagenomic dataset.



### Supplementary Figure 9 | CARD-FISH detection of OD1 organisms (green fluorescent) in GPS1.

A) A whole view of the CARD-FISH detection under fluorescent microscopy. Arrows indicate OD1 organisms. Detected OD1 organisms were associated with mineral particles.

B) A close view of the detection of OD1 organisms. The avarage cell size is around 0.2-0.3 µm.

	BS5	GPS1		
рН	11.6 ± 0.1	11.9 ± 0.1		
Temp. (°C)	$17.4 \pm 0.5$	17.1 ± 0.1		
E <sub>h</sub> (mV)	- 585±33	-656±9		
Cond. (µS/cm)	870 ± 70	3010 ± 10		
DO (mM)	<dl< td=""><td><dl< td=""></dl<></td></dl<>	<dl< td=""></dl<>		
$N_2$ (% by vol)	53.6	53.6 36.6±0.4		
H <sub>2</sub> (% by vol)	34	50.9±1.1		
$CH_4$ (% by vol)	5.3 ± 1.2	7.4±3.1		
Na⁺ (mM)	1.98±0.06	14.69±0.41		
K⁺ (mM)	0.03 ± 0.01	0.13 ± 0.01		
NH4 <sup>+</sup> (mM)	<dl< td=""><td><dl< td=""></dl<></td></dl<>	<dl< td=""></dl<>		
Ca <sup>2+</sup> (mM)	1.17 ± 0.11	0.94 ± 0.10		
Mg <sup>2+</sup> (mM)	$0.036 \pm 0.040$	$0.004 \pm 0.004$		
Cl⁻ (mM)	1.49 ± 0.11	8.73 ± 0.10		
SO4 <sup>2-</sup> (mM)	0.001± 0.000	<dl< td=""></dl<>		
PO4 <sup>3-</sup> (mM)	<dl< td=""><td><dl< td=""></dl<></td></dl<>	<dl< td=""></dl<>		
NO <sub>3</sub> <sup>-</sup> (mM)	<dl< td=""><td><dl< td=""></dl<></td></dl<>	<dl< td=""></dl<>		
TIC (mM)	0.07	0.035		
DOC (mM)	0.02	0.17		
f <sub>deep</sub> a	$0.14 \pm 0.02$	1		
Reference	Morrill et al 2013			

Supplementary Table 1 Water geochemistry of BS5 and GPS1

Average data from multiple mesurements is shown.

<sup>a</sup> Average fraction of deep groundwater

% Mapped back (%) <sup>a</sup>	91.14	91.91	86.58	91.14
Mapped raw reads count <sup>a</sup>	49,337,795	42,017,066	48,255,451	46,333,039
Original raw reads count	54,543,392	45,983,339	55,675,843	50,843,647
Max Contig length	114,683	100,250	179,628	428,608
Contigs over 1 kbp	12,612	8,521	16,255	19,135
N50 Length (bp)	2,490	2,475	3,969	3,753
Number of contig	52,742	33,758	66,400	67,210
Total bases (bp)	59,351,457	38,098,402	88,663,216	94,598,029
Sample	GPS1 (2011)	GPS1 (2012)	BS5B (2011)	BS5B (2012)

Supplementary Table 2. Stats of metagenome assembly.

<sup>a</sup> Numbers of reads to perform mapping raw reads back to the assembled contigs (with length cutoff 0.8 and similarity cutoff 0.8). <sup>b</sup> Numbers of reads to perform mapping raw reads back to the assembled contigs as paired (with length cutoff 0.8 and similarity cutoff 0.8).

## Lists of Additional Data

**Supplementary Data 1** | Single-copied housekeeping gene list for validation of bin-genome association.

**Supplementary Data 2** | Single copied gene list for validation of draft genome assembly within domain Archaea

**Supplementary Data 3** | Genomic features of all bin-genomes associated from metagenomes of GPS1 and BS5 springs in 2011 and 2012.

**Supplementary Data 4** | Analysis of ANIb and TETRA among the recovered bin-genomes affiliated with the phylum OD1.

**Supplementary Data 5** | Analysis of ANIb and TETRA among the recovered bin-genomes affiliated with the class Proteobacteria.

**Supplementary Data 6** | Analysis of ANIb and TETRA among the recovered bin-genomes affiliated with the Firmicutes.

**Supplementary Data 7** | Analysis of ANIb and TETRA among the recovered bin-genomes affiliated with the phylum Chloroflexi.

Supplementary Data 8 | Analysis of ANIb and TETRA among the recovered bin-genomes.

**Supplementary Data 9** | Marker genes related to cell activity in The Cedars microbial communities.

**Supplementary Data 10** | Metabolic marker genes related to respiration and fermentation in The Cedars microbial communities.

Supplementary Data 11 | MAPLE pathway analysis of dominant representative bin-genomes.

Supplementary Data 12 | MAPLE complex analysis of dominant representative bin-genomes.

Supplementary Data 13 | MAPLE functional analysis of dominant representative bin-genomes.

Supplementary Data 14 | Percentage of predicted genes per genome.