

*Supplementary Material*

**miR-148a-3p Mediates Notch Signaling to Promote the Differentiation and M1 Activation of Macrophages**

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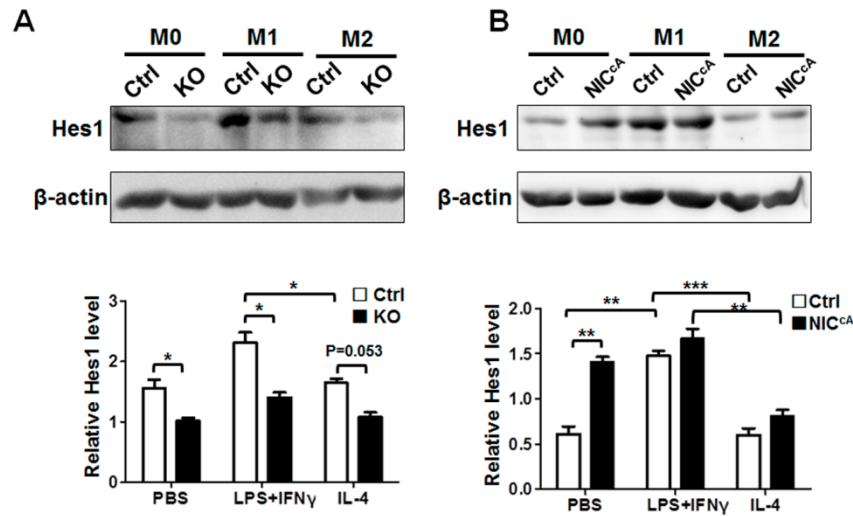
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**Table S1.** Primers and oligonucleotides used for PCR in this study.

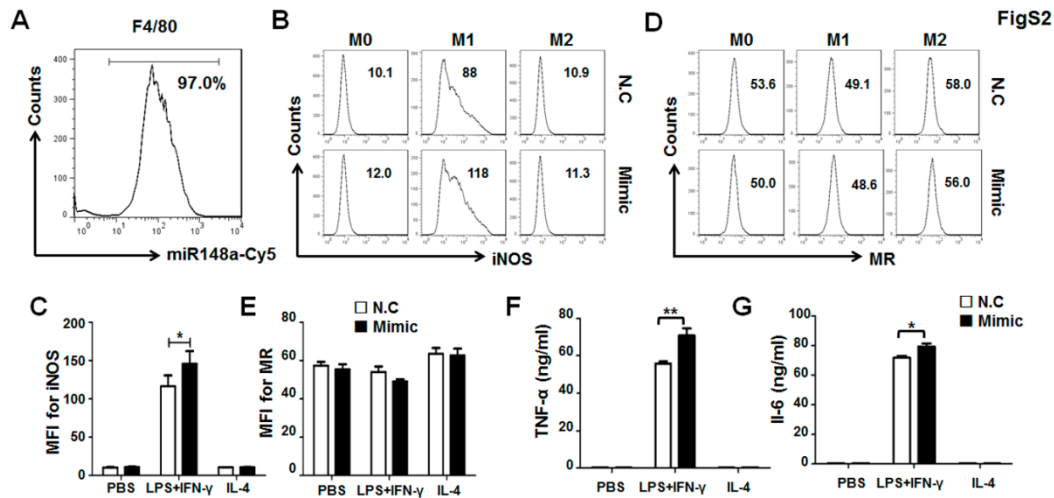
<b>Name</b>	<b>Purpose</b>	<b>Sequence</b>
Cre-F*	Genotyping	5'-CCGGTCGATGCAACGAGTGATGAGG
Cre-R*	Genotyping	5'-GCCTCCAGCTTGCATGATCTCCGG
RBP-J-F	Genotyping	5'-GTTCTTAACCTGTTGGTCGGAACC
RBP-J-WT-R	Genotyping	5'-GCTTGAGGCTTGATGTTCTGTATTGC
RBP-J-floxed-R	Genotyping	5'-ACCGGTGGATGTGGAATGTGT
NICD-F	Genotyping	5'-AAAGTCGCTCTGAGTTGTTAT
NICD-WT-R	Genotyping	5'-TAAGCCTGCCCAGAAGACTC
NICD-floxed-R	Genotyping	5'-GAAAGACCGCGAAGAGTTTG
PTEN-F	RT-PCR	5'-ATCAAGAGGATGGATTCCG
PTEN-R	RT-PCR	5'-GGCGGTGTCATAATGTCT
PTEN mut1-F	PCR	5'-CAATGGGCTGTCCGAGACTTAATA
PTEN mut1-R	PCR	5'-TATTAAGTCTCGGACAGCCCATTTG
PTEN-mut2-F	PCR	5'-AAATGCTACCGAGACAGGATACAC
PTEN-mut2-R	PCR	5'-GTGTATCCTGTCTCGGTAGCATTT
PTEN 3'UTR-F	PCR	5'-CGGAATTCGAAATCTGTACACCCCTTGTCTT
PTEN 3'UTR-R	PCR	5'-AACTGCAGCACCCACACAATGACAAGAATGAG
PTEN siRNA	siRNA	5'-TCTTCAAGGGCAATTTGCTCATTAA
iNOS-F	RT-PCR	5'-TCGACATCCGCAACGACTATC
iNOS-R	RT-PCR	5'-CCAGGGCGTAGTTGTAGAAGAG
IL-6-F	RT-PCR	5'-ATGGCCCATTACAAAGCCG
IL-6-R	RT-PCR	5'-TTTCTGGAGTAGCAGCTCCTAA
TNF- $\alpha$ -F	RT-PCR	5'-TCGACATCCGCAACGACTATC
TNF- $\alpha$ -R	RT-PCR	5'-CCAGGGCGTAGTTGTAGAAGAG
MR-F	RT-PCR	5'- AAACACAGACTGACCCTTCCC
MR-R	RT-PCR	5'- GTTAGTGTACCGCACCCCTCC
$\beta$ -actin	RT-PCR	5'-CATCCGTAAAGACCTCTATGCCAAC
$\beta$ -actin	RT-PCR	5'-ATGGAGCCACCGATCCACA
control mimics	nucleotides	5'-UUUGUACUACACAAAAGUACUG
control inhibitors	nucleotides	5'-CAGUACUUUUGUGUAGUACAAA
miR-148a-3p mimics	nucleotides	5'-UCAGUGCACUACAGAACUUUGU
miR-148a-3p inhibitors	nucleotides	5'-ACAAAGUUCUGUAGUGCACUGA

Note: F, Forward; R, Reverse.

FigS1

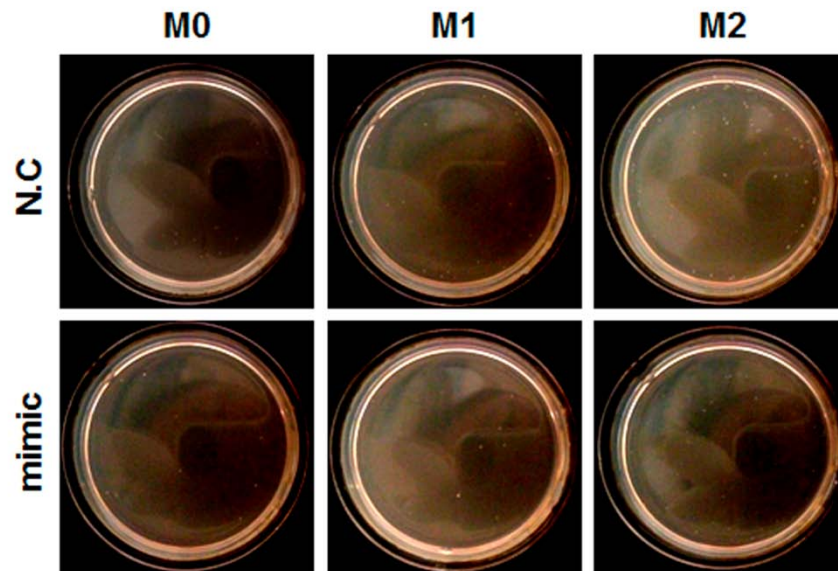


**Figure S1. The efficiency of Notch knockdown or activation in macrophages was detected. (A)** BMDMs from RBP-J<sup>CKO</sup> or control (Ctrl) mice were stimulated with PBS (M0), LPS+IFN- $\gamma$  (M1), or IL-4 (M2). The expression level of HES1, one downstream molecules of Notch signaling, was detected by Western blotting, using  $\beta$ -actin as a reference control and relative levels of HES1 compared quantitatively (n=3). **(B)** BMDMs were prepared from NIC<sup>cA</sup> or control (Ctrl) mice, and stimulated with PBS (M0), LPS+IFN- $\gamma$  (M1), or IL-4 (M2). The expression level of HES1 was detected by Western blotting, using  $\beta$ -actin as a reference control and relative levels of HES1 compared quantitatively (n=5). Bars, mean  $\pm$  SD; \* $P$  < 0.05; \*\* $P$  < 0.01; \*\*\* $P$  < 0.001; n.s, not significant.



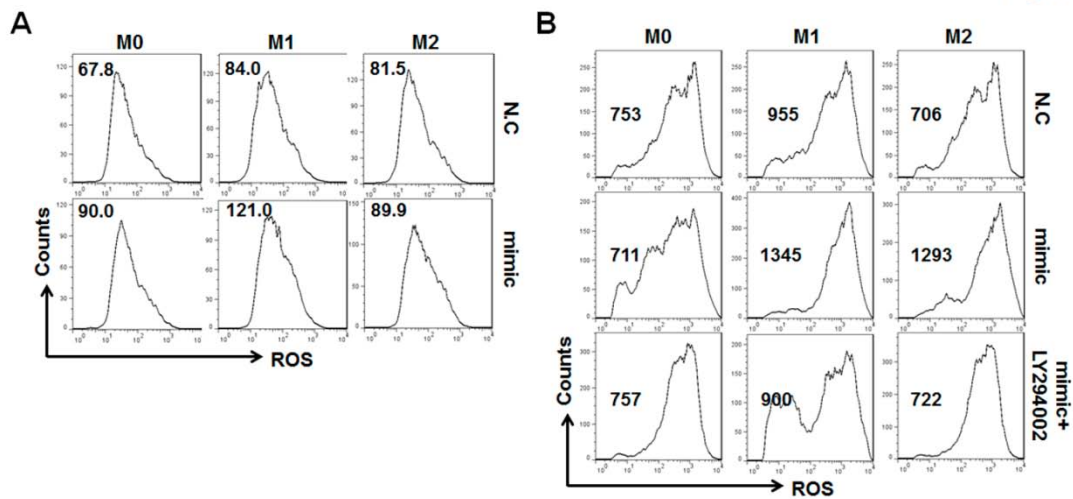
**Figure S2. miR-148a-3p can promote M1 macrophage polarization.** (A) The transfection efficiency of miR-148 in macrophages was detected by labeling miR-148a with Cy5 using FACS (n=3). (B, C) BMDMs were transfected with miR-148a-3p mimic or control (N.C). Cells were then polarized with PBS (M0), LPS+IFN-γ (M1), or IL-4 (M2) for 24 h. The expression level of M1 marker iNOS was analyzed using FACS intracellular staining (B), and then the mean fluorescent intensity (MFI) for iNOS was quantitatively compared (C)(n=3). (D, E) BMDMs were transfected with miR-148a-3p mimic or N.C. Cells were then polarized with PBS (M0), LPS+IFN-γ (M1), or IL-4 (M2) for 24 h. The expression level of M2 marker MR was analyzed using FACS intracellular staining (D), and then the mean fluorescent intensity (MFI) for MR was quantitatively compared (E)(n=3). (F, G) BMDMs were transfected with miR-148a-3p mimic or N.C. Cells were then polarized with PBS (M0), LPS+IFN-γ (M1), or IL-4 (M2) for 24 h. After that, the supernatant of cultured medium was collected and the expression level of M1 marker TNF-α (F) and IL-6 (G) was analyzed using ELISA (n=8). Bars, mean ± SD; \**P* < 0.05; \*\**P* < 0.01.

**FigS3**



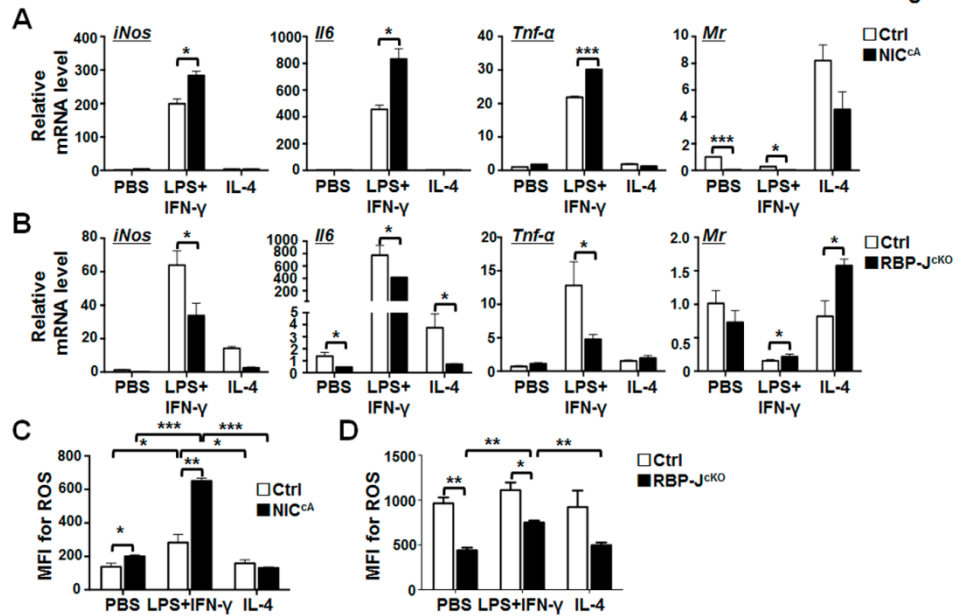
**Figure S3. miR-148a-3p enhances bactericidal activity of macrophages.** BMDMs derived from normal mice were transfected with miR-148a-3p mimic or control (N.C) and stimulated with PBS (M0), LPS+IFN- $\gamma$  (M1), or IL-4 (M2). Then macrophages ( $1 \times 10^6$ ) were co-cultured with *E. coli* ( $1 \times 10^7$  CFU) that had been transformed with an *EGFP*-expressing vector for 2 h. Macrophages that had engulfed *EGFP*<sup>+</sup> bacteria were incubated for a further 6 h. Cells were then lysed and plated on ampicillin-containing agar plates to observe the growth of bacterial colonies (n=3).

FigS4



**Figure S4. miR-148a-3p increases ROS level in macrophages in an AKT-dependent manner. (A)** BMDMs were transfected with miR-148a-3p mimic or control (N.C), and then polarized with PBS (M0), LPS+IFN- $\gamma$  (M1), or IL-4 (M2). ROS levels were determined by FACS (n=3). **(B)** BMDMs were transfected with miR-148a-3p mimic or control (N.C), and then polarized with PBS (M0), LPS+IFN- $\gamma$  (M1), or IL-4 (M2), in the presence of LY294002. ROS levels were determined by FACS.

FigS5



**Figure S5. Notch signaling promotes M1 and attenuates M2 polarization of macrophages. (A, B)**

BMDMs from mice with different genotypes were stimulated with PBS (M0), LPS+IFN- $\gamma$  (M1), or IL-4 (M2). mRNA levels of *iNos*, *Il6*, *Tnf- $\alpha$* , and *Mr* were determined by qRT-PCR, using  $\beta$ -actin as a reference control (n=3). (C, D) BMDMs from mice with different genotypes were stimulated with PBS (M0), LPS+IFN- $\gamma$  (M1), or IL-4 (M2). ROS levels were determined by FACS and quantified by MFI (n = 3). Bars, mean  $\pm$  SD; \* $P$  < 0.05; \*\* $P$  < 0.01; \*\*\* $P$  < 0.001.