TRIM23 mediates virus-induced autophagy via activation of TBK1

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Supplementary Figures 1-9



Supplementary Figure 1. TRIM proteins induce autophagy. a, cDNA screen of 61 TRIM proteins to test their ability to induce GFP-LC3B puncta formation in HeLa cells transiently transfected with GFP-LC3B and the indicated TRIM proteins. At 48 h post-transfection, cells were treated with 10 µg/mL E64D/pepstatin A for 3 h. GFP-LC3B puncta were analyzed by confocal microscopy, and for each sample, the GFP-LC3B puncta of 30 randomly selected cells were quantified. Transfection of empty vector as well as rapamycin treatment (for 2 h) served as negative and positive control, respectively. Results represent total GFP-LC3B puncta of 30 cells ± SD (n=5). Numbers indicate individual TRIM proteins. b, Conversion of endogenous LC3B-I to LC3B-II in HEK293T cells transiently transfected with empty vector or the seven TRIM proteins (all V5-tagged) that showed the highest autophagy-inducing activity in the cDNA screen (a). At 48 h posttransfection, cells were treated with 10 µg/mL E64D/pepstatin A for 2 h, and WCLs were analyzed by immunoblot (IB) with anti-LC3B, anti-V5, and anti-actin antibodies. Stimulation of cells with rapamycin served as positive control. c, Exemplary image analysis for the TRIM siRNA screen shown in Figures 1c-f. Example images show results from Figure 1e for uninfected (Mock) or EMCV-infected HeLa cells stably expressing GFP-LC3B (GFP-LC3B-HeLa) that had been transfected with non-targeting control siRNA (si.NT) or ATG5-specific siRNA (si.ATG5). Confocal microscopy images of GFP-LC3B puncta were automatically extracted using an ImageJ Macro and converted into binary images. The pixel counts of the area of GFP-LC3B puncta in a 500 x 500 pixels section were quantified. Scale bar, 20 µm. d, GFP-LC3B puncta formation in GFP-LC3B-HeLa cells that were transiently transfected with specific siRNAs targeting the indicated TRIM proteins and subsequently treated with 3 µM rapamycin and 10 µg/mL E64D/pepstatin A for 4 h. Cells transfected with non-targeting control siRNA (NT) served as control. GFP-LC3B puncta areas were quantified using ImageJ software as described above. Box and whisker plots show the distribution of the area of GFP-LC3B for 3 individual images each containing ~30 cells. Numbers indicate the individual TRIM proteins that were silenced using siRNA. *p<0.05, ***p<0.0005 (ANOVA test). e, Representative knockdown efficiency of the individual TRIM proteins in GFP-LC3B-HeLa cells for the screens shown in (d) and Figures 1c-f, determined by gPCR analysis. Relative knockdown efficiency is shown as compared to values for si.NT-transfected control cells, which were set to 1. Results represent the mean ± SD (n=3). f, Viral infection efficiency in GFP-LC3B-HeLa cells for the screens shown in Figures 1cf, assessed by immunostaining of the indicated viral proteins (red) and immunofluorescence (IF) analysis. Nuclei were stained with DAPI (blue) to visualize individual cells. Results represent relative infection in % (n=75). Scale bar, 20 µm. ICP8, Infected cell protein 8. HA, Hemagglutinin. 3D, 3D polymerase. Data are representative of one screen (a), or at least two (b,d,f) or three (c,e) independent experiments.

Homo_sapie	1MATLVVN KLGAGVDS-GRQGS-RGTAV VKVLECGVCEDVFSLQGD-K VPR-LLLCGHTVCHDCLTRL PLHGRAIF	RCPF
Macaca mul	1MATLVVN KLGAGVDS-GRQGS-RGTAV VKVLECGVCEDVFSLQGD-K VPR-LLLCGHTVCHDCLTRL PLHGRAIF	RCPF
Mus muscul	1MAALAVN KPGAGVDS-GROGS-RGTAV VKVLECGVCEDVFSLOGD-K VPR-LLLCGHTVCHDCLTRL PLHGRAIF	RCPF
Danio reri	1MAAAVGV NKOASMESCVRHGRGTAANA VKVLECGVCEDVESLOGD-K VPR-LLLCGHTVCHDCLTRL PLHGRAVE	CPF
Xenopus la	1 MRMRNEAFRANSPECCAHAO REROGLESLDEKTK-GDILL TKVLECGVCEDIESLOGD-K VPR-LLLCGHTVCHDCLTRL PLHGRAIE	RCPF
C elecane	1	CDF
C. eregans		+++
01		TNOO
91 FDRQV1	SGVWGLKKNFALLELLERLQ NGPIGQYGAAEESIGI SGESIIRCDEDEAHLASVYC TVCATHLCSECSQVTHSTAT LAAHRRVPLADKPHEAT	IMCS
91 FDRQVI	SGVWGLKKNFALLELLERLQ NGPIGQYGAAEESIGI SGESIIRCDEDEAHLASVYC TVCATHLCSECSQVTHSTKT LAKHRRVPLADKPHEKI	IMCS
91 FDRQVI	SGVWGLKKNFALLELLERLQ NGHIGQYGAAEEAIGT SGESIIRCDEDEAHVASVYC TVCATHLCSDCSQVTHSTKT LAKHRRVPLADKPHEKI	IMCC
91 FDRQVI	SGVWGLKKNFALLELLERLQ NGATNQSGMSEDALRE MGECIIRCDEDETHTASMYC TVCATHLCAECSQLTHSTRT LAKHRRVPLADKPHEKN	1LCP
91 FDRQAI	SGVWGLKKNFALLELLERLQ NGASSQCSTVEEAIGG SGESIVRCDEDEAHMASVYC TVCATHLCADCSLHTHSTKT LAKHRRVPLADKPYEKI	FLCS
91 FDRTAT	GDLQNLKKNFALLELLEKIA DGGGLLEKSGEVVKFDRYSK ERLLNLECDEDSEHVAVIYC TVCDSNLCERCSESTHSTNV LSKHRRIPLTEKPPIN	/HCR
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201 QHQVHA	CLEEGCQT-S PLMCCVCKEYGKHQGHKHSV LEPEANQIRASILDMAHCIR TFTEEISDYSRKLVGIVQHI EGGEQIVEDGIGMAHTEHVP GTAENAF	RSCI
201 OHOVHA	CLEEGCOT-S PLMCCVCKEYGKHOGHKHSV LEPEANOIRASILDMAHCIR TFTEEISDYSRKLVGIVOHI EGGEOIVEDGIGMAHTEHVP GTAENAF	RSCI
201 OHOVHA	CLEEGCOT-S PLMCCVCKEYGKHOGHKHSV LEPEANOIRASILDMAHCIR TFTEEISDYSRKLVGIVOHI EGGEOIVEDGIGMAHTEHVP GTAENAF	RSCV
201 OHOVHZ	CLEDGCOP-G PLMCCVCKEYGKHOGHKHAV LEAEANOTRASILDMAHCIR TETEEVSEYSRKLVGIVOOT EGGEOIVEDSMGIAHTEHVP GTAESAF	RSCV
201 OHOVHZ	CLEDGCOT-S DIMCCVCREVGRHOGINHAU LETEASOLDATIONALCIR TETESSEVSRITCIVOLI EGEOIVEDGIGMAHTEHVS GTAENAL	RCV
201 10000	CUEICONTEC DIMONMODIVOBUCUCUUI IEVENDENTOFICITORIE VOETIONALUCIALI BOORGAUDOTIENDO OCIEFET	
201 103314		* ·
011 DAVDVD		
311 RAYFYL	LCROEEMALSVVDAHVERKL IWLRQQOEDMTILLSEVSAA CLHCEKTLQQDDCRVVLARQ EITRLETLQKQQQQFTEVA DHIQLDASIPVTFTKDD	1RVH
311 RAYFYL	LCRQEEMALSVVDAHVREKL IWLRQQQEDMTILLSEVSAA CLHCERTLQQDDCRVVLARQ EITRLLETLQRQQQQFTEVA DHIQLDASIPVTFTRDN	1RAH
311 RAYFSI	LCRQEEMALSVVDAHVREKL IWLRQQQEDMTILLSQVSTA CLHCEKTLQQDDCRVVLAKQ EITRLLETLQKQQQQFTEVA DHIQLDASIPVTFTKDN	IRVH
311 RAYFAI	LCRQEEMALSVVDAHVRERL IWLRQQQEDMTILLSQVSTA CLHCEKTLQQDDCRVVLAKQ EINRLLETLQKQQQQFTELA DHIQLDAGIPVTFTKDN	IRVH
311 RAYFSI	LCRQEEMALSVVDAHVREQL MWLRQQQENMTILLSEVSTA CLHCEKALQQDDCRVVLAKQ EITRLLETLQKQQQQFTSLA DHIQMDSSIPVTFTKDN	JRVH
311 RNHFRF	LDRDEEDAVETVDRYARNRV ESLQTQKERLEAISSKIGNT CTTLQKALIMERGKILDRKD DLLALAESTAAEPTAVL DQSQLSTRIAFSFLNDF	KLH
* :*	* *:** *:** :.*::: *: *:*: : *::: * : *:*:* : ::: *::: * * : *: *	.::*
421 IGPKME	TLGLDGAGKT TILFKLKQDEFMQPIPT IGFNVETVEYKNLKFTIWDV GGKHKLRPLWKHYYLNTQAV VFVVDSSHRDRISEAHSELA KLLTEKE	ELRD
421 IGPKME	TLGLDGAGKT TILFKLKQDEFMQPIPT IGFNVETVEYKNLKFTIWDV GGKHKLRPLWKHYYLNTQAV VFVVDSSHRDRISEAHSELA KLLTEKE	ELRD
421 IGPKME	TLGLDGAGKT TILFKLKQDEFMQPIPT IGFNVETVEYKNLKFTIWDV GGKHKLRPLWKHYYLNTQAV VFVVDSSHRDRISEAHSELA KLLTEKE	ELRD
421 IGPKME	TLGLDGAGKT TILFKLKODEFMOPIPT IGFNVETVEYKNLKFTIWDV GGKHKLRPLWKHYYLNTOAV VFVIDSCHRDRLMESHSELA KLLTEKE	ELRD
421 IGPKME	TIGLDGAGKT TILFKLKODEFMOPIPT IGFNVETVEYKNLKFTIWDV GGKHKLRPLWKHYYLNTOAV VFVIDSSHRERVAEAHSELA KLLTEKE	ELRD
421 TGDETE	LIGLDGAGKT STURRIKKVONDTVMAPHPT IGENIETTHYKNYRLNEWDV GGLPKLRHLWKHYYSNAOAT FYVIDGYAVERESEAIKELN RVMSDPI	UVGT
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521 ATTTT		•
E21 ATTTT		
501 ALLLI	DWAGALSVEETIELESLAND COCKSWIIGCDAASGMGLI EGUDWISKOLVA-AGVEDVA 600	
531 ALLLI	DYAGALSYEBITELISINKI CCKSWIIGCDARSGMGLI EGIDWISKQIVA-AGVLDVA 600	
531 ALLLIE	DVPGAVSVEEMTELLSLHKL CCGRSWHIQGCDARSGMGLH EGLDWLSRQLVA-AGVLDVA 600	
531 ALLLIE	DVTGSLSVEEMTELLSLHKL CCGRSWYIQGCDARSGMGLY DGLDWLSRQLVA-AGVLDVA 600	
531 CPVIVA	DGYALNGHMDALLSQLE- ALPFQHHFHCCDAATGSGID QIIDQITVCLSRLNGTCPV- 600	
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Supplementary Figure 2. TRIM23 expression in human tissues and cells. a, Alignment of TRIM23 amino acid sequences from the indicated species using Clustal Omega. Numbers indicate amino acids. Asterisks (*) indicate positions which have a single, fully conserved residue. Colons (:) indicate conservation between groups of strongly similar properties, and periods (.) indicate conservation between groups of weakly similar properties. b, Absolute copy number of TRIM23 transcripts in the indicated human tissues, determined by qRT-PCR analysis. Data represent the mean ± SD (n=3). c, Relative transcript levels of

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TRIM23 and ISG15 (positive control) in HEK293T cells that were mock-treated, stimulated with 1,000 U/mL IFNa₂ for 18 h, or infected with 25 HA units per mL (HAU/mL) Sendai virus (SeV) for 18 h, determined by qRT-PCR analysis. Results are expressed as fold levels compared to mock-treated cells. Data represent the mean \pm SD (n=3). **d**, Protein abundance of endogenous TRIM23 as well as RIG-I and ISG15, which are both IFN-stimulated genes (ISGs) and therefore served as positive controls, in HEK293T cells that were mock-treated, stimulated with 1,000 U/mL IFNa₂ for 18 h, or infected with 25 HAU/mL SeV for 48 h, determined by IB with anti-TRIM23, anti-RIG-I, and anti-ISG15 antibodies. WCLs were further immunoblotted with anti-SeV and anti-actin antibodies. Data are representative of one experiment (**a**), or at least two independent experiments (**b-d**).



Supplementary Figure 3. TRIM23-mediated autophagy is dependent on ATG5. a, GFP-LC3B puncta formation in *ATG5*^{+/+} and *ATG5*^{-/-} MEFs transiently transfected with GFP-LC3B (green) together with empty vector or FLAG-tagged TRIM23 (red). At 48 h post-transfection, FLAG-TRIM23 was visualized by immunostaining with anti-FLAG antibody. Nuclei were stained with DAPI (blue). Scale bar, 20 μ m. **b**, Quantification of GFP-LC3B puncta for the experiment shown in (a). Results represent the mean GFP-LC3B puncta per cell ± SD (n=50). **c**, Endogenous LC3B-I to LC3B-II conversion in *ATG5*^{+/+} and *ATG5*^{-/-}

MEFs transfected with empty vector or FLAG-tagged TRIM23, determined by IB with anti-LC3B antibody at 50 h post-transfection. d, Laser scanning confocal microscopy images of GFP-LC3B puncta formation in HeLa cells transiently transfected with GFP-LC3B (green) and either TRIM23-targeting shRNA (sh.TRIM23) or non-targeting control shRNA (sh.NT) for 38 h and subsequently infected with mutHSV-1 (MOI 4) for 12 h, or left uninfected (Mock). Knockdown of endogenous TRIM23 was confirmed by immunostaining with anti-TRIM23 antibody (red). Nuclei were stained with DAPI (blue). Scale bar, 20 µm. e, Quantification of GFP-LC3B puncta for the experiment shown in (d). Results represent the mean GFP-LC3B puncta per cell ± SD (n=50). f, Quantification of GFP-LC3B puncta in HeLa cells that were transiently transfected with GFP-LC3B together with sh.TRIM23 or sh.NT for 38 h and subsequently infected with RSV (MOI 0.5) or IAV (MOI 1) for 20 h, or left uninfected (Mock). Results represent the mean GFP-LC3B puncta per cell ± SD (n=50). g, Representative laser scanning confocal microscopy images of GFP-LC3B puncta formation in HeLa cells that were transiently transfected with GFP-LC3B (green) and sh.TRIM23 or sh.NT for 46 h and subsequently treated with either DMSO or rapamycin for 2 h. Knockdown of endogenous TRIM23 was confirmed by immunostaining with anti-TRIM23 antibody (red). Nuclei were stained with DAPI (blue). Scale bar, 20 µm. h, Quantification of GFP-LC3B puncta for the experiment shown in (g). Results represent the mean GFP-LC3B puncta per cell ± SD (n=50). i, Endogenous LC3B-I to LC3B-II conversion in HEK293T cells that were transfected with sh.NT or sh.TRIM23 for 48 h and subsequently treated with DMSO or rapamycin for 2 h, assessed by IB with anti-LC3B antibody. Knockdown of endogenous TRIM23 was confirmed by IB with anti-TRIM23 antibody. ns, statistically not significant, ***p <0.001 (Student's *t*-test). Data are representative of at least two independent experiments (a-i).



Supplementary Figure 4. Virus- and rapamycin-induced autophagy is abrogated in TRIM23 -/- MEFs. a, Accumulation of endogenous LC3B (green) in TRIM23 +/+ and TRIM23 -/- MEFs that were mock-treated, stimulated with rapamycin for 2 h, or infected with mutHSV-1 (MOI 4) for 12 h, determined by immunostaining using anti-LC3B antibody and confocal microscopy analysis. Nuclei were stained with DAPI (blue). Scale bar, 20 µm. b, Representative laser scanning confocal microscopy images from the experiment shown in Figure 2d showing RFP-LC3B puncta formation in TRIM23 +/+ and TRIM23 -/- MEFs that were transiently transfected with RFP-LC3B (red) and subsequently infected with Ad-GFP (MOI 100) for 48 h. Nuclei were stained with DAPI (blue). Scale bar, 20 µm. c, Representative laser scanning confocal

microscopy images from the experiment shown in Figure 2e showing GFP-LC3B puncta formation in TRIM23 +/+ and TRIM23 -/- MEFs that were transiently transfected with GFP-LC3B (green) and subsequently infected with SINV (MOI 5) for 24 h. Nuclei were stained with DAPI (blue). Scale bar, 20 µm. d, Representative confocal microcopy images of GFP-LC3B puncta formation in TRIM23 +/+ and TRIM23 -/- MEFs that were transiently transfected with GFP-LC3B (green) and incubated in starvation medium (Hank's buffered salt solution) for 2 h. Nuclei were stained with DAPI (blue). Scale bar, 20 µm. e, Quantification of GFP-LC3B puncta from the experiment shown in (d). Results represent the mean GFP-LC3B puncta per cell ± SD (n=50). f, Relative transcript levels of p62 in TRIM23 +/+ and TRIM23 -/- MEFs that were either left uninfected or infected with mutHSV-1 (MOI 4) for 12 h, determined by gRT-PCR. Analysis of IFNB1 transcripts served as control. Values were normalized to GAPDH mRNA levels, and are presented as fold induction relative to values for uninfected control samples, which were set to 1. g. Protein abundance of endogenous RIG-I, ISG15, LC3B-I and LC3B-II in HEK293T cells that were left uninfected (Mock), or infected with WT HSV-1 or mutHSV-1 (each MOI 4) for 16 h, or SeV (300 HAU/mL) for 48 h, determined by IB with the indicated antibodies. WCLs were further immunoblotted with anti-ICP8 (HSV-1) and anti-SeV antibodies to confirm efficient viral infection. ns, statistically not significant, **p <0.01, ***p <0.001 (Student's *t*-test). Data are representative of at least two independent experiments (**a-g**).



Supplementary Figure 5: Ubiquitination and autophagy-inducing abilities of TRIM23 WT and mutants. a, Ubiquitination of FLAG-TRIM23 in transiently transfected HEK293T cells that were either mock-treated or treated with rapamycin for 4 h, assessed by Immunoprecipitation (IP) with anti-FLAG and IB with antiubiquitin (Ub) at 52 h post-transfection. Asterisk indicates unspecific band. **b**, Ubiquitination of FLAGtagged TRIM23 WT or mutants in transiently transfected HEK293T cells, determined by IP with anti-FLAG and IB with anti-Ub at 48 h post-transfection. **c**, Schematic representation of TRIM23 domain structure as well as the ubiquitinated lysine residues in TRIM23 identified by mass spectrometry analysis of affinitypurified FLAG-TRIM23 from HEK293T cells. **d**, Ubiquitination of FLAG-tagged TRIM23 WT or the indicated mutants in transiently transfected HEK293T cells, determined as in (b). **e**, Representative laser scanning confocal microscopy images of GFP-LC3B puncta formation in HeLa cells that were transiently transfected with GFP-LCB (green) together with empty vector or FLAG-tagged TRIM23 WT or mutants (red). At 48 h post-transfection, cells were stained with anti-FLAG antibody, followed by confocal microscopy analysis. Nuclei were stained with DAPI (blue). Scale bar, 20 μm. Data are representative of one (**c**), or at least two (**a,b,d,e**) independent experiments.



Supplementary Figure 6. Characterization of GTPase- or K27-polyubiguitin-deficient mutants of TRIM23. a, Ubiquitination of FLAG-tagged TRIM23 WT or the mutant K458I in transiently transfected HEK293T cells, assessed by IP with anti-FLAG and IB with anti-Ub at 48 h post-transfection. Transfection of empty vector served as control. Asterisk indicates unspecific band. b, In vitro GTPase activity of purified FLAG-TRIM23 WT or the indicated mutants. Purified proteins were incubated *in vitro* with GTP for 2 h, followed by measuring the generation of free phosphate using a colorimetric reaction. Results are expressed as mean ± SD (n=2). c, Cladogram showing the relation of the TRIM23 ARF domain to other small GTPases: Sar1/Arf family: SAR1A, ARF1-6 and ARL1-5; Ras family: RalB, RalA; Rho/Rac family: RhoA; Rab family: Rab5α. d, Co-localization of myc-tagged TRIM23 WT or mutants (red) with FLAG-tagged ATG16L (green) in transiently transfected HeLa cells. At 48 h post-transfection, cells were immunostained with anti-myc and anti-ATG16L antibodies. DAPI, nuclei (blue). Scale bar, 20 µm. e, Color-coded intensity profiles of the merged confocal images from (d) (white lines). f, Co-localization analysis of FLAG-tagged TRIM23 WT or mutants (red) and endogenous LAMP1 (green) in transiently transfected HeLa cells. At 48 h post-transfection, cells were immunostained using anti-FLAG and anti-LAMP1 antibodies. DAPI, nuclei (blue). Scale bar, 20 µm. Data are representative of one (c), or at least two (a, b, d-f) independent experiments.



Supplementary Figure 7. Ternary complex formation of TRIM23, TBK1 and p62. a, Amino acid sequence of unique peptides of TBK1 and p62 identified by mass spectrometry analysis of affinity-purified FLAG-TRIM23 from HEK293T cells that were infected with mutHSV-1 (MOI 1) for 12 h. b, Co-localization of FLAG-TBK1 (red), GFP-p62 (green), and TRIM23-V5 (magenta) in transiently transfected HeLa cells. At 48 h post-transfection, cells were stained with anti-FLAG and anti-V5 antibodies, followed by confocal

microscopy analysis. DAPI, nuclei (blue). Scale bar, 20 µm. **c**, Co-localization of endogenous TRIM23 (green), endogenous p62 (red), and endogenous TBK1 (blue) in HeLa cells that were either treated with DMSO (Mock) or rapamycin for 8 h, assessed by immunostaining with anti-TRIM23, anti-p62, and anti-TBK1 antibodies and confocal microscopy. DAPI-stained nuclei are depicted in white. Scale bar, 20 µm. **d**, Binding of endogenous p62 and TBK1 to endogenous TRIM23 in HEK293T cells that were either mock-treated, stimulated with rapamycin for 4 h, or infected with mutHSV-1 (MOI 1) for 24 h, determined by IP with anti-TRIM23 antibody, followed by IB with anti-p62 and anti-TBK1 antibodies. **e**, *In vitro* binding of recombinant His-tagged TBK1 and purified recombinant GST or GST-TRIM23, determined by pulldown (PD) assay using GST-agarose, followed by IB with anti-TBK1. Data are representative of one (**a**) or at least two (**b-e**) independent experiments.

b а α-FLAG Merge GFP-LC3B + DAPI + DAPI 3 Viral titer (log10 pfu) Vector 2 WT Jector W BUN DAY 1 ΔARF С ctor RIM23. M d :MG132 RIM23 :DMSO MB2 :TRIM23-V5 :HA-Ub :HA-Ub :GST-TBK1 + :GST-TBK1 + (kDa) (kDa) IP: IP: HA (Ub) HA(Ub) α -GST α -GST 150-100-GST 150-GST 100-75 V5 FLAG 75-250 150-HA (Ub) WCL HA(Ub) WCL 100-50-10-HA

actin

Supplementary Figure 8. Effect of TRIM23 WT and mutants on viral growth and TBK1 ubiquitination. a, GFP-LC3B puncta formation as well as localization of TRIM23 WT and Δ ARF mutant in *TRIM23* \rightarrow MEF cells transiently transfected with GFP-LC3B (green) together with empty vector, or FLAG-tagged TRIM23

(Ub monomer)

àctin

WT or ΔARF mutant. TRIM23 WT and mutant proteins were immunostained using anti-FLAG antibody (red). Nuclei were stained with DAPI (blue). Scale bar, 20 μm. **b**, Replication of mutHSV-1 in TRIM23 ^{-/-} MEFs that were transiently transfected with either empty vector, or TRIM23 WT or mutants, and subsequently infected with mutHSV1 (MOI 0.1) for 48 h. Viral titers in the cell supernatant were determined by plaque assay. Results are expressed as mean ± SD (n=2). **c**, Ubiquitination of GST-TBK1 in transiently transfected HEK293T cells that co-expressed HA-tagged Ub and either empty vector or FLAG-tagged TRIM23 WT or C34A, or MIB2 (positive control), determined by IP with anti-GST and IB with anti-HA at 48 h post-transfection. **d**, Ubiquitination of GST-TBK1 in transiently transfected HEK293T cells that co-expressed V5-tagged TRIM23 and HA-Ub. At 24 h post-transfection, cells were treated with MG132 or DMSO for 4 h, followed by IP with anti-GST and IB with anti-GST and IB are representative of at least two independent experiments (**a-d**).

















Supplementary Figure 3i:				
anti-LC3B	anti-TRIM23	anti-actin		
75- 50- 25- 15-	75– ––– –	75- 50- 37-		













Supplementary Figure 9. Complete western blot images of the indicated figures of the manuscript. Red boxes indicate the cropped areas shown in the indicated figures. Numbers indicate molecular weight markers (in kDa).