Supporting Information

Redox-responsivePolysulfide-basedBiodegradableOrganosilicaNanoparticles for Delivery of Bioactive Agents

Seyyed Pouya Hadipour Moghaddam^{a, b}, Jiban Saikia^b, Mostafa Yazdimamaghani^{a, b}, and Hamidreza Ghandehari^{a, b, c, *}

^a Department of Pharmaceutics and Pharmaceutical Chemistry, University of Utah, Salt Lake City, UT 84112, USA

^b Utah Center for Nanomedicine, Nano Institute of Utah, University of Utah, Salt Lake City, UT 84112, USA

^c Department of Bioengineering, University of Utah, Salt Lake City, UT 84112, USA

* Corresponding Author. Utah Center for Nanomedicine, Nano Institute of Utah, 5205 SMBB, 36 S. Wasatch Dr, Salt Lake City, UT 84112, USA Email Address: hamid.ghandehari@utah.edu (H. Ghandehari)



Figure S1. STEM images for 60 Mesoporous D, 110 Mesoporous T, and 330 Nonporous D nanoparticles.



Figure S2. STEM spectra of (A) 60 Mesoporous D; (B) 110 Mesoporous T; and (C) 330 Nonporous D nanoparticles.



Figure S3. XRD graphs for (A) 60 Mesoporous D; (B) 110 Mesoporous T; (C) 120 Nonporous D; (D) 330 Nonporous D; and (E) regular non-degradable mesoporous SiNP. The degradable mesoporous nanoparticles did not exhibit the typical Bragg peaks like regular non-degradable mesoporous SiNP.



Figure S4. TGA analyses for the synthesized nanoparticles. Stöber 100 SiNP was used as a control.











Figure S5. XPS survey spectra of (A) 60 Mesoporous D; (B) 110 Mesoporous D; (C) 110 Mesoporous T; (D) 120 Nonporous D; and (E) 330 Nonporous D nanoparticles.



Figure S6. FTIR spectra of 110 Mesoporous D and 120 Nonporous D nanoparticles before and after washing. CTAB and BTESPD (disulfide-based precursor) were also analyzed for better comparison. Typical CTAB peaks around 3000 cm⁻¹ were not seen after washing the nanoparticles. This confirms complete removal of the surfactant from the pores.



Figure S7. Degradation images of mesoporous nanoparticles after 15 days in the presence of 8 μ M GSH; (A) 60 Mesoporous D; (B) 110 Mesoporous D; and (C) 110 Mesoporous T nanoparticles.



Figure S8. Degradation images of nonporous nanoparticles after 15 days in the presence of 8 μ M GSH; (A) 120 Nonporous D and (B) 330 Nonporous D nanoparticles.



Figure S9. Agglomeration of 60 Mesoporous D nanoparticles in solution containing 8 mM GSH.



Figure S10. Degradation of 110 Mesoporous D nanoparticles after 48 h in the presence of 8 mM GSH.



Figure S11. Normalized degradation percentages of all synthesized nanoparticles over the 15-day period of study.









Figure S12. ESI-MS for 3 precursors: (A) TEOS; (B) BTESPD; and (C) BTESPT. ESI-MS for the degradation products: (D) 60 Mesoporous D; (E) 110 Mesoporous D; (F) 110 Mesoporous T; (G) 120 Nonporous D; and (H) 330 Nonporous D nanoparticles.



Figure S13. Nanoparticle uptake comparison *via* flow cytometry in RAW 264.7 macrophages after 24 h incubation with the concentration of 80 μ g ml⁻¹. Data are mean ± SD (n = 3).



Figure S14. Dot plots from flow cytometry analyses of RAW 264.7 macrophages after 24 h incubation with $80 \,\mu g \, \text{ml}^{-1}$ nanoparticles.

Nanoparticle	0	С	S	Si
60 Mesoporous D	30.69	44.67	6.72	17.92
110 Mesoporous D	26.91	54.88	4.68	13.53
110 Mesoporous T	24.6	62	2.20	11.20
120 Nonporous D	19.07	56.73	11.39	12.81
330 Nonporous D	17.66	67.12	6.73	8.49
100 Stöber	31.01	56.14	0	12.84

Table S1. Atomic (top) and mass (bottom) percentages of the synthesized nanoparticles measured by XPS

Nanoparticle	0	С	S	Si
60 Mesoporous D	28.11	30.72	12.37	28.80
110 Mesoporous D	26.58	40.70	9.26	23.47
110 Mesoporous T	25.86	48.89	4.62	20.60
120 Nonporous D	17.82	39.81	21.34	21.03
330 Nonporous D	18.31	52.26	13.98	15.45
100 Stöber	32.40	44.04	0	23.56