## **Supporting Information**

## A biomimetic stress sensitive hydrogel controlled by DNA nanoswitches

Swapneel R. Deshpande, Roel Hammink, Frank H.T. Nelissen, Alan E. Rowan, and Hans A. Heus\*

Institute for Molecules and Materials, Radboud University, Heyendaalseweg 135, 6525 AJ Nijmegen, the Netherlands

**Corresponding Author** 

E-mail: h.heus@science.ru.nl



Samples	[DNA-DBCO] at 0 mins (Absorbance)	[DNA-DBCO] after 12 hours	
	(This of builde)	(Absorbance)	
DNA A	~100 µM (0.958)	~ 4 µM (0.108)	
DNA B	~100 µM (0.860)	~ 10 µM (0.112)	
Control	$\sim 5 \ \mu M \ (0.05)$	~ 5 µM (0.06)	
PIC, no DBCO			

**Figure S1.** Yields of DBCO-DNA conjugation to N<sub>3</sub>-functionalized PIC polymers. A standard curve was generated by measuring the absorbance of free concentrations of DBCO at 309 nm using a Nanodrop spectrophotometer (ND-1000, Thermo Scientific). The amount of DNA-DBCO clicked on DNA A and DNA B was determined by the absorbance of residual free DNA-DBCO after the click reaction. Comparing the amount of free DBCO before (0 min) and after incubation (12 hours) with the polymer reveals a high reaction efficiency of ~90 - 95 % for both DNA A and B oligonucleotides.



**Figure S2**. Frequency dependent rheology of PIC DNA hydrogels cross-linked with 0.9 equivalents DNA LC at pH 7.4 (a) or 5.2 (b). The hydrogels were stable and did not relax as demonstrated by measuring G' and G'' over a range of frequencies ( $10^{-3}$  to 10 Hz). All measurements were performed at 30 °C in PBS, 10 mM MgCl<sub>2</sub>, pH 7.4.



**Figure S3**. Temperature dependent rheological analysis of PIC-DNA hydrogel, covalently cross-linked with DNA AB (single-stranded) or DNA AB + C (double stranded). (1  $^{\circ}$ C min<sup>-1</sup> at (1 Hz) and strain (2%) PBS, 10 mM MgCl<sub>2</sub>.)



**Figure S4**. CD spectra to confirm formation of an i-motif in solution (25 °C, PBS.10 mM MgCl<sub>2</sub>). The spectrum of DNA LC shows a large positive peak at  $\sim$  260 nm at pH 7.4 and a larger positive peak, shifted to  $\sim$  285 nm at pH 5.2. Simultaneously, the small negative peak becomes smaller and shifts to higher wavelength. Both are indicators of a switch to an i-motif structure at low pH.



**Figure S5.** pH dependent rheological analysis of PIC-DNA cross-linked with 0.9 equivalents DNA LC to form the i-motif at 30 °C in PBS,  $10 \times 10^{-3}$  M MgCl<sub>2</sub>. a) Time evolution of storage modulus G' after addition of DNA cross-linker LC at pH 6.0 (grey) and pH 6.5 (green). b) *K*<sup>1</sup>/*G*<sub>0</sub> as function of  $\sigma$  at pH 6.0 (grey) and pH 6.5 (green).

DNA construct		Rheological properties		
Cross-linker Sequence	Contour Length (nm)	G <sub>o</sub> (Pa)	σ <sub>c</sub> (Pa)	σ <sub>max</sub> (Pa)
ss-DNA LC (pH 6.0)	~ 16 - 19	79 ± 9	4 ± 0.5	30
ss-DNA LC (pH 6.5)	~ 16 - 19	68.5 ± 8	4.5 ± 0.5	30

Table S1 Mechanical properties of hydrogel cross-linked with i-motif at pH 6.0 and 6.5



**Figure S6**. Rheological analysis of DNA triplex cross-linked PIC hydrogel at pH 7.4 and pH 5.2. Temperature dependent rheology was measured with ramping of 1 °C min<sup>-1</sup> at (1 Hz) and strain (2%) PBS, 10 mM MgCl<sub>2</sub>.



**Figure S7.** pH dependence of PIC DNA hydrogels covalently cross-linked with DNA AB + C at pH 7.4 (orange) and 5.2 (green).



**Figure S8** Rheology measurements of the PIC-DNA hydrogel cross-linked with DNA LC in the presence of 10  $\mu$ g/ml BSA. a) Time evolution of the storage modulus (*G'*). b) G' and G'' over a range of frequencies (10<sup>-3</sup> to 10 Hz). Measurements were performed at 30 °C in PBS, 10 mM MgCl<sub>2</sub>, pH 7.4.