SIPIN1 regulates auxin efflux to affect flower abscission process

Zihang Shi^{1,2*}, Yun Jiang^{1,2*}, Xinqi Han^{1,2}, Xin Liu^{1,2}, Ruishu Cao^{1,2}, Mingfang Qi^{1,2}, Tao Xu^{1,2}& Tianlai Li^{1,2}

1 College of Horticulture, Shenyang Agricultural University, Shenyang 110866, Liaoning, People's Republic of China. 2 Key Laboratory of Protected Horticulture of Ministry of Education, No. 120 Dongling Road, Shenhe District 110866, People's Republic of China. *These authors contributed equally to this work. Correspondence and requests for materials should be addressed to T.X. (email: syauxutao@qq.com) and T.L. (email: tianlaili@126.com)

b а 7 100 6 80 5 Pedicel abscission (%) Relative Expression 60 4 3 40 2 20 1 0 \mathbb{A} 0 10 20 зо 40 50 60 70 5 15 ò Ó 10 20 25 Time after flower removal (h) Time after flower removal (h)

Supplementary Figure S1. Abscission process and *SIPIN1* expression profiles following the IAA treatments at different ends. The arrows indicate the opposite directions of incubation. The data are presented as the mean (\pm SD) values corresponding to three independent tests. For each test of abscission rate, 40 to 50 pedicels were investigated.

Supplementary information



Supplementary Figure S2. Full-length images in SIPIN1 western blotting experiments. a, SIPIN1 western blotting of the AZ protein samples as described in Fig. 3 a. A specific signal among the 62-70 KD (arrowhead) was detected. b, the Coomassie-staining of the AZ protein samples.



Supplementary Figure S3. *FEI2* expression profiles in the AZ during abscission. *FEI2* expression decreased at 8-24 h after flower removal, while 1-MCP treatment prevented the down-regulation of *FEI2* during abscission. The data are presented as the mean (\pm S D) values corresponding to three independent tests.

Supplementary Table S1. Primers used in this study

Primer name	Sequence (5' -3')		
Forward primer of <i>SIPIN1</i> for RT-PCR	CAGGCAGCTCTACCACAAGG		
Reverse primer of <i>SIPIN1</i> for RT-PCR	TGTAATCGGCAACGCAATC		
Forward primer of <i>FEI2</i> for RT-PCR	TCTGGTCTCCACTGCTCAAGA		
Reverse primer of FEI2 for RT-PCR	CCAAGGGAACCGTAGCACAG		
Forward primer for vector pSPT19 construction (Eco RI)	GG <u>GAATTC</u> ACTTATTGTTCTTGGGGTTCTTGCT		
Reverse primer for vector pSPT19 construction (Hind III)	GG <u>AAGCTT</u> TTTTCCATCTTCTTTTAACTCTGCT		
Forward primer of <i>SIPIN1</i> for VIGS construction (Eco RI)	CCG <i>GAATTC</i> TGGTACTGCTGGGAGAAACTCAAAC		
Reverse primer of <i>SIPIN1</i> for VIGS construction (BamHI)	CGC <u>GGATCC</u> GCTTTGTTGCCATTGTTACTAGGTG		
Forward primer of <i>SIPIN1</i> for bait construction of Y2H (Eco RI)	CCG <i>GAATTC</i> AGAGGTGCAAGACTGCTAATTTCT		
Reverse primer of <i>SIPIN1</i> for bait construction of Y2H (BamHI)	CGC <u>GGATCC</u> GGACTGTGCAATTATAGCAGGC		
Forward primer of <i>SIPIN1</i> for vector construction of BiFC (KpnI)	CGG <i>GGTACC</i> AGAGGTGCAAGACTGCTAATTTCT		
Reverse primer of <i>SIPIN1</i> for vector construction of BiFC (Spel)	CGG <u>ACTAGT</u> GGACTGTGCAATTATAGCAGGC		
Forward primer of <i>PP2C</i> for vector construction of BiFC (KpnI)	CGG <i>GGTACC</i> ATGACTGGAGGCAAAGAAATCATCG		
Reverse primer of <i>PP2C</i> for vector construction of BiFC (Eco RI)	CCG <i>GAATTC</i> TTGGAACCTTACAACTACGCAGG		

Supplementary Table 52. Prosphopeptides of SiPINI					
December lated particle and site (in lowercase)	pRS	Mascot	Relative quantity		
Phosphorylated peptide and site (in lowercase)	Score	Score	12h/0h	24h/0h	
DDFsFANR	108	35	0.582	0.378	
FNNYHGGAAAPQsNsNtHyPAPNPGMFsPsNNGNk	57	11	0.700	0.761	

Supplementary Table S2. Phosphopeptides of SIPIN1