

Supporting Information

for

Chemical Synthesis of GPI Glycan-Peptide Conjugates by Traceless Staudinger Ligation

Sanyong Zhu and Zhongwu Guo*

Department of Chemistry, University of Florida, 214 Leigh Hall, Gainesville, Florida 32611,
United States

*Corresponding Author E-mail: zguo@chem.ufl.edu

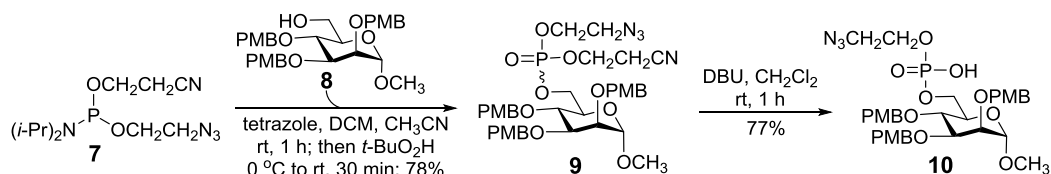
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I. Experimental

General methods. Commercial chemicals and materials were used as received without additional purification unless noted otherwise. Molecular sieves 4Å were flame-dried under vacuum and then cooled to rt under an N₂ atmosphere immediately before use. TLC was carried out on Silica Gel 60Å F254 plates with detection by a UV detector and/or by charring with 10% H₂SO₄ in EtOH (v/v). Mass spectrometry (MS) was recorded either on a high resolution ESI-TOF machine or on a normal resolution MALD-TOF machine. NMR spectra were recorded on a 400, 500 or 600 MHz machine with chemical shifts reported in ppm (δ) downfield from internal tetramethylsilane (TMS) or DHO references and with signals described as s (singlet), d (doublet), t (triplet) or m (multiplet) and coupling constants reported in Hz.

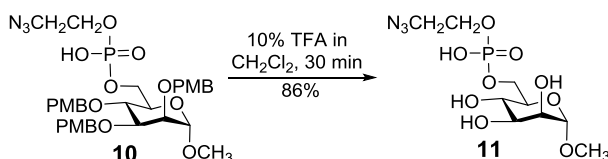
Methyl 6-*O*-[(2-azidoethoxy)-phosphono]-2,3,4-tri-*O*-(*p*-methoxybenzyl)- α -D-mannopyranoside (**10**).



To a solution of **8** (560 mg, 1.01 mmol) and freshly prepared **7** (870 mg, 3.03 mmol) in CH₂Cl₂/CH₃CN (v/v, 2/1, 10 mL) was added slowly an acetonitrile solution of tetrazole (~0.45 M, 11.22 mL, 5.05 mmol) at rt under an N₂ atmosphere. After the mixture was stirred at rt for 1 h, it was cooled to 0 °C and mixed with *tert*-butyl hydroperoxide (5.5 M solution in decane, 0.92 mL, 5.05 mmol). The mixture was stirred at rt for another 30 min and quench with aqueous Na₂S₂O₃. The aqueous layer was extracted with CH₂Cl₂ three times. The combined organic layer was washed with aqueous NaHCO₃ and brine, dried, and concentrated under vacuum. The residue was purified by column chromatography (EtOAc/hexane 2:1) to give **9** (595 mg, 78% yield) as a diastereomeric mixture (R/S 1:1). It was dissolved in CH₂Cl₂ (10 mL). After adding DBU (2 drops) and stirring at rt for 1 h, the solvent was evaporated, and the residue was purified by flash silica gel column chromatography (CH₃OH/CH₂Cl₂ 1:10) to give **10** (426.6 mg, 77%) as colorless syrup. R_f = 0.45 (CH₃OH/CH₂Cl₂ 1:4); ¹H NMR (500 MHz, CDCl₃) δ : 7.17 – 7.13 (m, 6H, Ph), 6.78 – 6.74 (m, 6H, Ph), 4.72 (d, J = 10.5 Hz, 1H, Ph-CH₂), 4.55 (d, J = 12 Hz, 1H, Ph-CH₂), 4.53 (d, J = 1.5 Hz, 1H, H-1), 4.50 (d, J = 10.5 Hz, 1H, Ph-CH₂), 4.48 (d, J = 10.0 Hz, 1H, Ph-CH₂), 4.44 (d, J = 12.0 Hz, 1H, Ph-CH₂), 4.41 (d, J = 11.5 Hz, 1H, Ph-CH₂), 4.08 (dd, J =

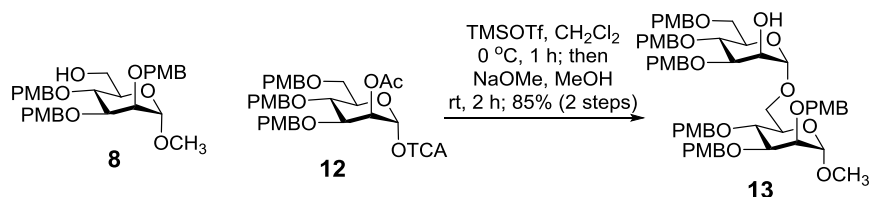
12.0, 5.0 Hz, 1H, H-6a), 4.02 (dd, $J = 11.0, 5.0$ Hz, 1H, H-6b), 3.92 – 3.88 (m, 2H, $-OCH_2CH_2-$), 3.72 (s, 3H, Ph- OCH_3), 3.71 (s, 3H, Ph- OCH_3), 3.70 (s, 3H, Ph- OCH_3), 3.66 – 3.64 (m, 3H, H-2, H-3, H-4), 3.58 – 3.56 (m, 1H, H-5), 3.26 – 3.24 (m, 2H, $-CH_2CH_2-N_3$), 3.19 (s, 3H, $-OCH_3$). ^{13}C NMR (125 MHz, $CDCl_3$) δ : 159.25, 159.13, 159.08, 130.53, 130.48, 130.06, 129.62, 129.50, 129.24, 113.71, 99.13(C-1), 79.67, 74.62, 74.27, 74.14, 72.49, 71.78, 71.28, 64.80, 64.17, 55.20, 54.62, 51.29. ^{31}P NMR (160 MHz, $CDCl_3$) δ : -2.49. HR ESI-TOF MS (m/z): calcd for $C_{33}H_{41}N_3O_{12}P$ [M - H] $^-$, 702.2433; found, 702.2446.

Methyl 6-*O*-[(2-azidoethoxy)-phosphono]- α -D-mannopyranoside (**11**).



To a solution of **10** (330 mg, 0.47 mmol) in CH_2Cl_2 (4.5 mL) was added TFA (0.5 mL). After stirring at rt for 30 min, the reaction mixture was co-evaporated with toluene three times. The residue was subjected to silica gel column chromatography ($CH_3OH/EtOAc$ 1:1) to produce **11** (138 mg, 86% yield) as colorless syrup. $R_f = 0.42$ ($CH_3OH/EtOAc$ 1:1); 1H NMR (600 MHz, CD_3OD) δ : 4.61 (d, $J = 1.8$ Hz, 1H, H-1), 4.14 – 4.06 (m, 2H, H-6a, H-6b), 4.03 – 4.00 (m, 2H, $-OCH_2CH_2-$), 3.76 (dd, $J = 3.6, 1.8$ Hz, 1H, H-2), 3.70 (dd, $J = 9.6, 5.4$ Hz, 1H, H-4), 3.66 (dd, $J = 9.6, 3.6$ Hz, 1H, H-3), 3.57 – 3.55 (m, 1H, H-5), 3.46 (t, $J = 5.4$ Hz, 2H, $-CH_2CH_2N_3$), 3.35 (s, 3H, $-OCH_3$). ^{13}C NMR (150 MHz, CD_3OD) δ : 102.79 (C-1), 73.44 (C-5), 72.47 (C-3), 72.05 (C-2), 68.17 (C-4), 66.08 (C-6), 65.42 ($-OCH_2CH_2-$), 55.26 ($-OCH_3$), 52.60 ($-CH_2CH_2N_3$). ^{31}P NMR (160 MHz, CD_3OD_3) δ : 0.61. HR ESI-TOF MS (m/z): calcd for $C_9H_{17}N_3O_9P$ [M - H] $^-$, 342.0708; found, 342.0717.

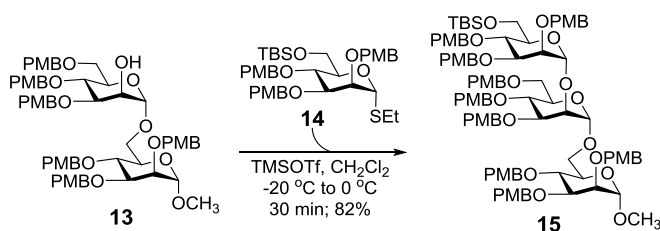
Methyl 3,4,6-tri-*O*-(*p*-methoxybenzyl)- α -D-mannopyranosyl-(1 \rightarrow 6)-2,3,4-tri-*O*-(*p*-methoxybenzyl)- α -D-mannopyranoside (**13**).



To a stirred mixture of **8** (118 mg, 0.213 mmol), **12** (196 mg, 0.319 mmol) and molecular sieves 4\AA in anhydrous CH_2Cl_2 (5 mL) was added slowly TMSOTf (5.8 μ L, 0.032 mmol) under an N_2 atmosphere at 0 $^\circ C$. After the mixture was stirred at 0 $^\circ C$ for 1 h, it was neutralized with Et_3N ,

filtered, and concentrated under vacuum. The residue was purified by silica gel column chromatography (EtOAc/hexane 1:2) to give the disaccharide as colorless syrup. This product was then dissolved in CH₃OH (5 mL) and mixed with CH₃ONa (11.5 mg, 0.213 mmol) at rt. After the mixture was stirred at rt for 2 h, it was concentrated under vacuum. The residue was purified by silica gel column chromatography (EtOAc/hexane 3:1) to afford **13** (195 mg, 85% for two steps) as colorless syrup. $R_f = 0.30$ (EtOAc/hexane 3:1); ¹H NMR (600 MHz, CDCl₃) δ : 7.30 – 7.24 (m, 8H, Ph), 7.16 (d, $J = 8.4$ Hz, 2H, Ph), 7.05 (d, $J = 8.8$ Hz, 2H, Ph), 6.87 – 6.78 (m, 12H, Ph), 5.05 (d, $J = 1.2$ Hz, 1H, H-1^{Man-II}), 4.82 (d, $J = 10.8$ Hz, 1H, Ph-CH₂), 4.72 (d, $J = 10.2$ Hz, 1H, Ph-CH₂), 4.67 (d, $J = 12.0$ Hz, 1H, Ph-CH₂), 4.66 (d, $J = 1.2$ Hz, 1H, H-1^{Man-I}), 4.63 – 4.48 (m, 6H, Ph-CH₂), 4.42 (d, $J = 10.2$ Hz, 1H, Ph-CH₂), 4.41 (d, $J = 12.0$ Hz, 1H, Ph-CH₂), 4.37 (d, $J = 10.2$ Hz, 1H, Ph-CH₂), 4.08 (dd, $J = 3.0, 4.8$ Hz, 1H), 3.88 – 3.70 (m, 8H), 3.81 (s, 3H, Ph-OCH₃), 3.79 (s, 3H, Ph-OCH₃), 3.77 (s, 3H, Ph-OCH₃), 3.76 (s, 3H, Ph-OCH₃), 3.75 (s, 3H, Ph-OCH₃), 3.72 (s, 3H, Ph-OCH₃), 3.67 (dd, $J = 10.8, 4.2$ Hz, 1H), 3.62 (dd, $J = 8.4, 4.8$ Hz, 1H), 3.59 (dd, $J = 10.8, 1.8$ Hz, 1H), 3.25 (s, 3H, -OCH₃), 2.35 (d, $J = 3.0$ Hz, 1H, -OH). ¹³C NMR (150 MHz, CDCl₃) δ : 159.43 (Ph-OCH₃), 159.33 (Ph-OCH₃), 159.26 (Ph-OCH₃), 159.23 (3C, Ph-OCH₃), 130.86, 130.82, 130.53, 130.11, 129.78, 129.70, 129.65, 129.53, 129.40, 113.99, 113.88, 113.86, 113.81, 113.76, 99.78(C-1^{Man-II}), 98.88(C-1^{Man-I}), 80.07, 79.18, 74.84, 74.77, 74.49, 74.45, 74.03, 73.08, 72.44, 71.83, 71.50, 71.14, 68.56, 68.16, 66.48, 55.41 (Ph-OCH₃), 55.39 (Ph-OCH₃), 55.37 (2C, Ph-OCH₃), 55.34 (Ph-OCH₃), 55.31 (Ph-OCH₃), 54.75 (-OCH₃). HR ESI-TOF MS (m/z): calcd for C₆₁H₇₂O₁₇Na [M + Na]⁺, 1099.4667; found, 1099.4663.

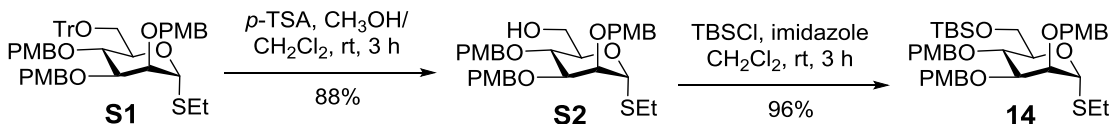
Methyl 6-*O*-*tert*-butyldimethylsilyl-2,3,4-tri-*O*-(*p*-methoxybenzyl)- α -D-mannopyranosyl-(1 \rightarrow 2)-3,4,6-tri-*O*-(*p*-methoxybenzyl)- α -D-mannopyranosyl-(1 \rightarrow 6)-2,3,4-tri-*O*-(*p*-methoxybenzyl)- α -D-mannopyranoside (15**).**



After the mixture of **13** (688 mg, 0.64 mmol), **14** (537 mg, 0.77 mmol), and molecular sieves 4Å in anhydrous diethyl ether (10 mL) was stirred at rt for 10 min, it was cooled to -20 °C, which was followed by the addition of NIS (346 mg, 1.54 mmol) and AgOTf (19.8 mg, 0.077 mmol). The mixture was stirred at 0 °C for 30 min and then quenched with Et₃N, filtered, and

concentrated under vacuum. The residue was purified by silica gel column chromatography (EtOAc/hexane 1:2) to produce **15** (899mg, 82%) as colorless syrup. $R_f = 0.60$ (EtOAc/hexane 1:1); $^1\text{H NMR}$ (600 MHz, CDCl_3) δ : 7.30 (d, $J = 8.4$ Hz, 2H, Ph), 7.26 – 7.18 (m, 12H, Ph), 7.15 (d, $J = 8.4$ Hz, 2H, Ph), 7.11 (d, $J = 8.4$ Hz, 2H, Ph), 6.86 – 6.73 (m, 18H, Ph), 5.24 (s, 1H, H-1^{Man-II}), 4.86 (s, 1H, H-1^{Man-III}), 4.83 (d, $J = 10.8$ Hz, 2H, Ph- CH_2), 4.75 (d, $J = 10.8$ Hz, 1H, Ph- CH_2), 4.68 (s, 1H, H-1^{Man-I}), 4.67 (d, $J = 12.0$ Hz, 1H, Ph- CH_2), 4.63 (d, $J = 12.0$ Hz, 1H, Ph- CH_2), 4.60 (d, $J = 12.0$ Hz, 1H, Ph- CH_2), 4.56 (d, $J = 10.8$ Hz, 1H, Ph- CH_2), 4.51 – 4.45 (m, 4H, Ph- CH_2), 4.44 – 4.35 (m, 7H, Ph- CH_2), 4.15 (d, $J = 2.4$ Hz, 1H), 3.96 – 3.85 (m, 5H), 3.83 – 3.59 (m, 11H), 3.81 (s, 3H, Ph- OCH_3), 3.80 (s, 3H, Ph- OCH_3), 3.78 (s, 3H, Ph- OCH_3), 3.76 (s, 3H, Ph- OCH_3), 3.74 (s, 6H, Ph- $\text{OCH}_3 \times 2$), 3.73 (s, 3H, Ph- OCH_3), 3.68 (s, 3H, Ph- OCH_3), 3.66 (s, 3H, Ph- OCH_3), 3.57 (d, $J = 10.8$ Hz, 1H), 3.25 (s, 3H, - OCH_3), 0.90 (s, 9H, - $t\text{Bu}$), 0.08 (s, 3H, - SiCH_3), 0.07 (s, 3H, - SiCH_3). $^{13}\text{C NMR}$ (150 MHz, CDCl_3) δ : 159.30 (Ph- OCH_3), 159.23 (2C, Ph- OCH_3), 159.18 (Ph- OCH_3), 159.15 (Ph- OCH_3), 159.14 (Ph- OCH_3), 159.08 (Ph- OCH_3), 159.03 (2C, Ph- OCH_3), 131.42, 131.13, 131.04, 130.89, 130.78, 130.47, 129.76, 129.57, 129.43, 129.37, 129.22, 113.89, 113.85, 113.81, 113.79, 113.74, 113.73, 113.71, 113.59, 99.33(C-1^{Man-II}), 98.88(C-1^{Man-III}), 98.72(C-1^{Man-I}), 80.28, 79.59, 74.72, 74.60, 74.57, 74.50, 74.27, 73.67, 73.07, 72.96, 72.32, 72.05, 71.84, 71.79, 71.54, 71.42, 71.08, 69.07, 66.69, 62.87, 60.53, 55.39 (2C, Ph- OCH_3), 55.36 (Ph- OCH_3), 55.32 (3C, Ph- OCH_3), 55.29 (Ph- OCH_3), 55.25 (Ph- OCH_3), 55.18 (Ph- OCH_3), 54.64 (- OCH_3), 26.14, 18.51, -4.83, -5.06. HR ESI-TOF MS (m/z): calcd for $\text{C}_{97}\text{H}_{120}\text{O}_{25}\text{SiNa}$ [$\text{M} + \text{Na}$] $^+$, 1735.7786; found, 1735.7780.

Ethyl 6-*O*-*tert*-butyldimethyl- silyl-2,3,4-tri-*O*-(*p*-methoxybenzyl)-1-thio- α -D-mannopyranoside (14**)**

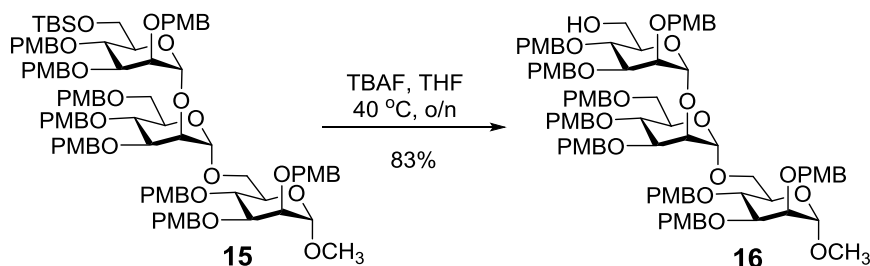


To a solution of **S1** (836 mg, 1.0 mmol) in $\text{CH}_3\text{OH}/\text{CH}_2\text{Cl}_2$ (1/1, v/v, 10 mL) was added *p*-TSA (19 mg, 0.1 mmol). After stirring at rt for 3 h, the reaction was quenched with Et_3N . The solvent was removed under vacuum, and the residue was purified by silica gel column chromatography (EtOAc/hexane 1:2) to give **S2** (513.9 mg, 88%) as colorless syrup. $R_f = 0.45$ (EtOAc/hexane 1:1); $^1\text{H NMR}$ (600 MHz, CDCl_3) δ : 7.28 – 7.22 (m, 6H, Ph), 6.86 – 6.84 (m, 6H, Ph), 5.24 (s, 1H, H-1), 4.85 (d, $J = 10.8$ Hz, 1H, Ph- CH_2), 4.64 (d, $J = 12.6$ Hz, 1H, Ph- CH_2), 4.61 (d, $J = 12.0$ Hz,

1H, Ph-CH₂), 4.55 – 4.48 (m, 3H, Ph-CH₂), 3.95 (dd, *J* = 9.6, 3.6 Hz, 1H, H-3), 3.92 (t, *J* = 9.6 Hz, 1H, H-4), 3.80 – 3.73 (m, 4H, H-2, H-5, H-6a, H-6b), 3.80 (s, 3H, Ph-OCH₃), 3.79 (s, 3H, Ph-OCH₃), 3.78 (s, 3H, Ph-OCH₃), 2.58 – 2.50 (m, 2H, -SCH₂CH₃), 1.94 (dd, *J* = 7.2, 6.0 Hz, 1H, -OH), 1.22 (t, *J* = 7.2 Hz, 1H, -SCH₂CH₃). ¹³C NMR (150 MHz, CDCl₃) δ: 159.41 (Ph-OCH₃), 159.37 (Ph-OCH₃), 159.32 (Ph-OCH₃), 130.74, 130.50, 130.24, 129.80, 129.65, 129.50, 113.94, 82.26(C-1), 80.14, 76.16, 74.96, 74.89, 72.48, 72.07, 71.87, 62.57, 55.40 (3C, Ph-OCH₃), 25.41 (-SCH₂CH₃), 15.01 (-SCH₂CH₃). HR ESI-TOF MS (*m/z*): calcd for C₃₂H₄₀O₈SNa [M + Na]⁺, 607.2342; found, 607.2332.

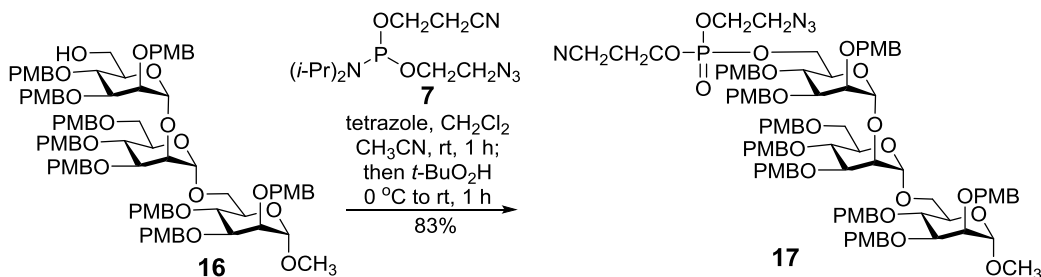
To a solution of **S2** (368 mg, 0.63 mmol) in CH₂Cl₂ (5 mL) was added imidazole (85.6 mg, 1.26 mmol). After stirring at rt for 10 min, TBSCl (142.7 mg, 0.95 mmol) was added, and the mixture was stirred at rt for another 3 h. The mixture was concentrated under vacuum, and the residue was purified by silica gel column chromatography (EtOAc/hexane 1:10) to give **14** (422.7 mg, 96%) as colorless syrup. *R_f* = 0.50 (EtOAc/hexane 1:5); ¹H NMR (600 MHz, CDCl₃) δ: 7.31 – 7.24 (m, 6H, Ph), 6.87 (d, *J* = 8.4 Hz, 6H, Ph), 5.31 (s, 1H, H-1), 4.85 (d, *J* = 10.8 Hz, 1H, Ph-CH₂), 4.63 (d, *J* = 12.6 Hz, 1H, Ph-CH₂), 4.61 (d, *J* = 12.0 Hz, 1H, Ph-CH₂), 4.55 – 4.49 (m, 3H, Ph-CH₂), 3.94 (dd, *J* = 9.6, 3.6 Hz, 1H, H-3), 3.88 (t, *J* = 9.6 Hz, 1H, H-4), 3.83 – 3.80 (m, 3H, H-5, H-6a, H-6b), 3.80 (s, 6H, Ph-OCH₃ x 2), 3.79 (s, 3H, Ph-OCH₃), 3.76 (dd, *J* = 3.0, 1.8 Hz, 1H, H-2), 2.64 – 2.50 (m, 2H, -SCH₂CH₃), 1.25 (t, *J* = 7.8 Hz, 1H, -SCH₂CH₃), 0.90 (s, 9H, -*t*Bu), 0.07 (s, 3H, -SiCH₃), 0.06 (s, 3H, -SiCH₃). ¹³C NMR (150 MHz, CDCl₃) δ: 159.23 (Ph-OCH₃), 131.06, 130.62, 130.36, 129.65, 129.49, 113.77, 81.36(C-1), 80.07, 76.15, 74.86, 74.78, 73.50, 71.75, 71.50, 62.86, 55.33 (3C, Ph-OCH₃), 26.00 (-*t*Bu), 25.02 (-SCH₂CH₃), 18.37 (-SiC(CH₃)₃), 14.98 (-SCH₂CH₃), -5.06 (-SiCH₃), -5.22 (-SiCH₃). HR ESI-TOF MS (*m/z*): calcd for C₃₈H₅₄O₈SSiNa [M + Na]⁺, 721.3206; found, 721.3231.

Methyl 2,3,4-tri-*O*-(*p*-methoxybenzyl)- α -D-mannopyranosyl-(1 \rightarrow 2)-3,4,6-tri-*O*-(*p*-methoxybenzyl)- α -D-mannopyranosyl-(1 \rightarrow 6)-2,3,4-tri-*O*-(*p*-methoxybenzyl)- α -D-mannopyranoside (16**).**



To a solution of **15** (678 mg, 0.395 mmol) in THF (4.0 mL) was added TBAF (0.60 mL, 1.0 M in THF, 0.60 mmol). The mixture was stirred at 40 °C overnight. After the solvent was removed under vacuum, the residue was purified by silica gel column chromatography (EtOAc/hexane 1:1) to afford **16** as colorless syrup (545 mg, 86%). $R_f = 0.50$ (EtOAc/hexane 2:1); $^1\text{H NMR}$ (600 MHz, CDCl_3) δ : 7.30 – 7.11 (m, 18H, Ph), 6.87 – 6.76 (m, 18H, Ph), 5.15 (d, $J = 1.2$ Hz, 1H, H-1^{Man-II}), 5.01 (d, $J = 1.8$ Hz, 1H, H-1^{Man-III}), 4.83 (d, $J = 10.8$ Hz, 1H, Ph- CH_2), 4.81 (d, $J = 10.8$ Hz, 1H, Ph- CH_2), 4.77 (d, $J = 10.2$ Hz, 1H, Ph- CH_2), 4.69 (d, $J = 1.2$ Hz, 1H, H-1^{Man-I}), 4.66 (d, $J = 13.2$ Hz, 2H, Ph- CH_2), 4.62 (d, $J = 12.0$ Hz, 1H, Ph- CH_2), 4.54 – 4.41 (m, 12H, Ph- CH_2), 4.12 (t, $J = 1.8$ Hz, 1H), 3.90 – 3.58 (m, 17H), 3.81 (s, 3H, Ph- OCH_3), 3.80 (s, 3H, Ph- OCH_3), 3.79 (s, 3H, Ph- OCH_3), 3.76 (s, 3H, Ph- OCH_3), 3.74 (s, 3H, Ph- OCH_3), 3.73 (s, 6H, Ph- $\text{OCH}_3 \times 2$), 3.67 (s, 6H, Ph- $\text{OCH}_3 \times 2$), 3.26 (s, 3H, - OCH_3). $^{13}\text{C NMR}$ (150 MHz, CDCl_3) δ : 159.30 (Ph- OCH_3), 159.21 (Ph- OCH_3), 159.17 (Ph- OCH_3), 159.11 (Ph- OCH_3), 159.03 (Ph- OCH_3), 130.89, 130.85, 130.83, 130.80, 130.77, 130.54, 130.39, 130.31, 129.63, 129.52, 129.34, 129.19, 113.91, 113.85, 113.82, 113.74, 113.72, 113.68, 99.71(C-1^{Man-II}), 99.61(C-1^{Man-III}), 99.00(C-1^{Man-I}), 80.06, 79.65, 78.84, 74.96, 74.77, 74.65, 74.56, 74.23, 74.18, 74.10, 72.97, 72.85, 72.30, 72.06, 71.94, 71.83, 71.60, 71.33, 68.99, 66.46, 62.78, 55.35 (Ph- OCH_3), 55.34 (2C, Ph- OCH_3), 55.30 (Ph- OCH_3), 55.28 (2C, Ph- OCH_3), 55.26 (Ph- OCH_3), 55.20 (Ph- OCH_3), 55.16 (Ph- OCH_3), 54.78 (- OCH_3). HR ESI-TOF MS (m/z): calcd for $\text{C}_{91}\text{H}_{106}\text{O}_{25}\text{Na}$ [$\text{M} + \text{Na}$]⁺, 1621.6921; found, 1621.6911.

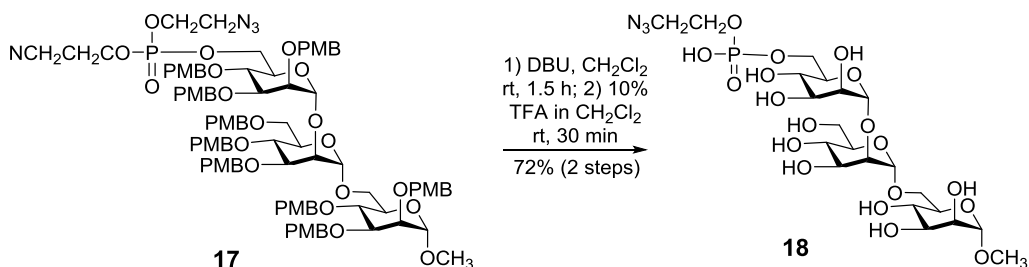
Methyl 6-*O*-[(2-azidoethoxy)-phosphono]-2,3,4-tri-*O*-(*p*-methoxybenzyl)- α -D-mannopyranosyl-(1 \rightarrow 2)-3,4,6-tri-*O*-(*p*-methoxybenzyl)- α -D-mannopyranosyl-(1 \rightarrow 6)-2,3,4-tri-*O*-(*p*-methoxybenzyl)- α -D-mannopyranoside (17**).**



To a solution of **16** (500 mg, 0.313 mmol) and freshly prepared **7** (269mg, 0.938 mmol) in anhydrous $\text{CH}_2\text{Cl}_2/\text{CH}_3\text{CN}$ (2:1, v/v, 8 mL) was added slowly an acetonitrile solution of tetrazole (~0.45 M, 6.96 mL, 3.13 mmol) under an N_2 atmosphere. After the mixture was stirred at rt for 1 h, it was cooled to 0 °C, and then *tert*-butyl hydroperoxide (5.5 M solution in decane, 0.57 mL,

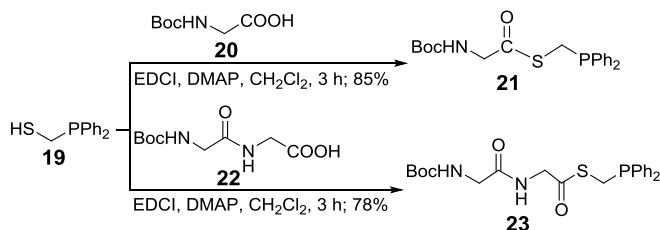
3.13 mmol) was added. The mixture was stirred at rt for 1 h and quenched with aqueous $\text{Na}_2\text{S}_2\text{O}_3$. The aqueous layer was extracted with CH_2Cl_2 three times. The combined organic layer was washed with aqueous NaHCO_3 and brine, dried, and concentrated under vacuum. The residue was purified by column chromatography (EtOAc/hexane 2:1) to give the phosphorylated product as a diastereomeric mixture (R/S \approx 1/1). This product was then dissolved in CH_2Cl_2 (8 mL) and mixed with DBU (2 drops). After stirring at rt for 1.5 h, the solution was concentrated, and the residue was subjected to silica gel column chromatography ($\text{CH}_3\text{OH}/\text{CH}_2\text{Cl}_2$ 1:10) to produce **17** (393.5 mg, 72% for 2 steps) as colorless syrup. $R_f = 0.55$ ($\text{CH}_3\text{OH}/\text{CH}_2\text{Cl}_2$ 1:5); ^1H NMR (600 MHz, CD_3OD) δ : 7.35 (d, $J = 8.4$ Hz, 2H, Ph), 7.25 – 7.11 (m, 12H, Ph), 7.06 (d, $J = 8.4$ Hz, 2H, Ph), 7.02 (d, $J = 8.4$ Hz, 2H, Ph), 6.86 – 6.75 (m, 14H, Ph), 6.71 (d, $J = 8.4$ Hz, 2H, Ph), 6.68 (d, $J = 8.4$ Hz, 2H, Ph), 5.09 (s, 1H, H-1^{Man-II}), 4.92 (s, 1H, H-1^{Man-III}), 4.82 (s, 1H, H-1^{Man-I}), 4.77 (d, $J = 10.8$ Hz, 1H, Ph- CH_2), 4.71 – 4.64 (m, 4H, Ph- CH_2), 4.61 (d, $J = 10.8$ Hz, 1H, Ph- CH_2), 4.51 (d, $J = 10.8$ Hz, 1H, Ph- CH_2), 4.45 – 4.32 (m, 10H, Ph- CH_2), 4.27 (d, $J = 10.8$ Hz, 1H, Ph- CH_2), 4.20 – 4.15 (m, 2H), 4.02 – 3.92 (m, 3H), 3.89 – 3.40 (m, 17H), 3.75 (s, 3H, Ph- OCH_3), 3.73 (s, 3H, Ph- OCH_3), 3.71 (s, 3H, Ph- OCH_3), 3.69 (s, 3H, Ph- OCH_3), 3.68 (s, 6H, Ph- $\text{OCH}_3 \times 2$), 3.62 (s, 3H, Ph- OCH_3), 3.61 (s, 3H, Ph- OCH_3), 3.57 (s, 3H, Ph- OCH_3), 3.32 (s, 3H, $-\text{OCH}_3$). ^{13}C NMR (150 MHz, CD_3OD) δ : 160.84 (Ph- OCH_3), 160.80 (Ph- OCH_3), 160.76 (Ph- OCH_3), 160.67 (Ph- OCH_3), 160.65 (Ph- OCH_3), 160.58 (Ph- OCH_3), 160.52 (Ph- OCH_3), 132.43, 132.08, 132.01, 131.97, 131.91, 131.75, 131.65, 131.38, 131.27, 130.97, 130.90, 130.81, 130.69, 130.66, 130.49, 130.43, 114.99, 114.81, 114.76, 114.71, 114.60, 114.57, 101.02(C-1^{Man-II}), 100.64(C-1^{Man-III}), 100.19(C-1^{Man-I}), 81.18, 80.32, 79.38, 76.05, 75.57, 75.34, 75.19, 75.12, 73.86, 73.37, 73.31, 73.16, 72.83, 72.72, 72.56, 72.46, 71.61, 69.86, 67.91, 65.47, 63.51, 55.70, 55.65, 55.58, 55.45, 53.58, 52.59. MALDI-TOF MS (m/z): calcd for $\text{C}_{93}\text{H}_{109}\text{N}_3\text{O}_{28}\text{PNa}$ [$\text{M} + 2\text{Na} - \text{H}$]⁺, 1792.673; found, 1792.610.

Methyl 6-O-[(2-azidoethoxy)-phosphono]- α -D-mannopyranosyl-(1 \rightarrow 2)- α -D-mannopyranosyl-(1 \rightarrow 6)- α -D-mannopyranoside (18**).**



To a solution of **17** (286 mg, 0.164 mmol) in CH₂Cl₂ (3.6mL) was added TFA (0.4 mL). After stirring at rt for 30 min, the reaction mixture was co-evaporated with toluene three times. The residue was subjected to flash silica gel column chromatography (CH₃OH/EtOAc 1:1) to afford **18** (96.3 mg, 88% yield) as a white foamy solid. *R_f* = 0.45 (CH₃OH/EtOAc 2:1); ¹H NMR (600 MHz, D₂O) δ: 5.13 (d, *J* = 1.2 Hz, 1H, H-1^{Man-II}), 5.02 (d, *J* = 1.2 Hz, 1H, H-1^{Man-III}), 4.74 (d, *J* = 1.2 Hz, 1H, H-1^{Man-I}), 4.15 (ddd, *J* = 11.4, 5.4, 1.8 Hz, 1H), 4.09 (t, *J* = 5.4 Hz, 1H), 4.07 – 4.03 (m, 3H, H-2^{Man-III}, -OCH₂CH₂-), 3.99 (dd, *J* = 3.6, 1.2 Hz, 1H, H-2^{Man-II}), 3.96 – 3.94 (m, 2H), 3.92 (dd, *J* = 3.6, 1.2 Hz, 1H, H-2^{Man-I}), 3.82 (dd, *J* = 9.6, 1.2 Hz, 1H), 3.76 – 3.68 (m, 7H), 3.52 (t, *J* = 4.8 Hz, 2H, -CH₂CH₂N₃), 3.39 (s, 3H, -OCH₃). ¹³C NMR (150 MHz, D₂O) δ: 102.33 (C-1^{Man-II}), 100.87 (C-1^{Man-III}), 97.81 (C-1^{Man-I}), 78.87, 72.64, 71.93, 70.62, 70.11, 69.81, 66.76, 66.54, 66.21, 65.75, 64.63, 60.79, 54.69 (-OCH₃), 50.89 (-CH₂CH₂N₃). ³¹P NMR (160 MHz, D₂O) δ: 0.30. HR ESI-TOF MS (*m/z*): calcd for C₂₁H₃₇N₃O₁₉P [M - H]⁻, 666.1764; found, 666.1764.

General procedure for the preparation of peptide phosphinothioesters.



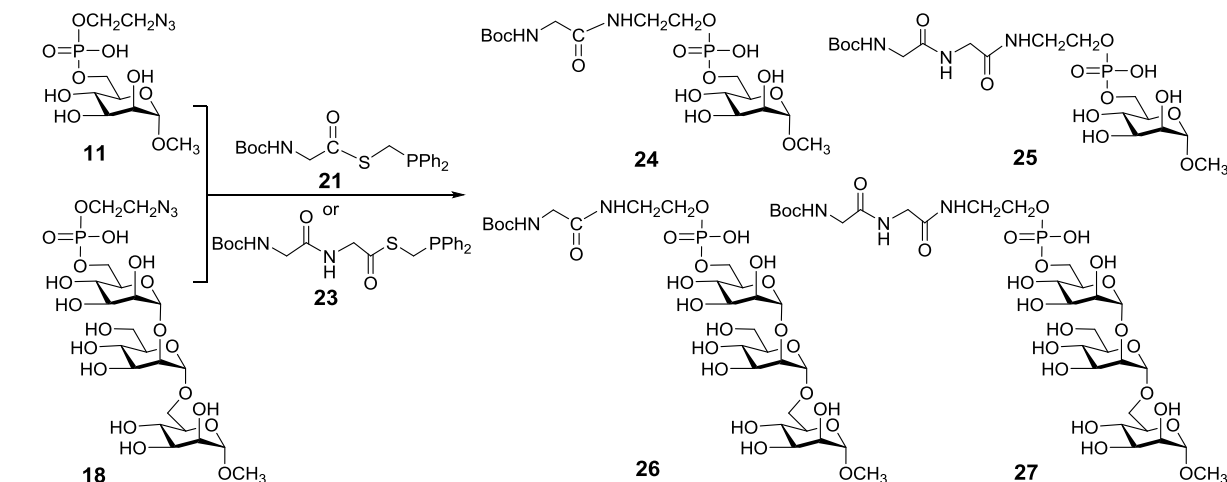
To a stirred solution of freshly prepared **19** (232.3 mg, 1.0 mmol) and amino acid **20** (175.2 mg, 1.0 mmol) or peptide **21** (232.2 mg, 1.0 mmol) in anhydrous CH₂Cl₂ (10 mL) were added EDCI (465.7 mg, 3.0 mmol) and DMAP (24.4 mg, 0.2 mmol). After stirring at rt for 3 h, the mixture was diluted with CH₂Cl₂, washed with water and brine, dried, and concentrated under vacuum. The residue was purified by silica gel column chromatography to produce **21** (350.2 mg, 85%) and **23** (348.7 mg, 78%).

21: a white solid. *R_f* = 0.65 (EtOAc/hexane 1:3); ¹H NMR (400 MHz, CDCl₃) δ: 7.45 – 7.34 (m, 10H, Ph), 5.16 (s, 1H, -NH-), 3.99 (d, *J* = 6.4 Hz, 2H, -NHCH₂CO-), 3.52 (d, *J* = 3.6 Hz, 2H, -SCH₂PPh₂), 1.43 (s, 9H, -*t*Bu). ¹³C NMR (100 MHz, CDCl₃) δ: 197.55 (-COS-), 155.52 (-OCONH-), 136.76, 132.69, 129.24, 128.60, 80.42 (-*Ot*Bu), 50.25 (-NHCH₂CO-), 28.36 (-*t*Bu), 25.23 (-SCH₂PPh₂). HR ESI-TOF MS (*m/z*): calcd for C₂₀H₂₄NO₃SPNa [M + Na]⁺, 412.1112; found, 412.1116.

23: a white solid. *R_f* = 0.35 (EtOAc/hexane 1:1); ¹H NMR (600 MHz, CDCl₃) δ: 7.42 – 7.35 (m, 10H, Ph), 6.86 (s, 1H, -NH-), 5.20 (s, 1H, -NH-), 4.17 (d, *J* = 6.0 Hz, 2H, -BocNHCH₂CO-), 3.83

(d, $J = 4.8$ Hz, 2H, $-\text{NHCH}_2\text{CO}-$), 3.52 (d, $J = 3.6$ Hz, 2H, $-\text{SCH}_2\text{PPh}_2$), 1.44 (s, 9H, $-t\text{Bu}$). ^{13}C NMR (150 MHz, CDCl_3) δ : 195.71 ($-\text{COS}-$), 169.80 ($-\text{OCONH}-$), 156.11 ($-\text{OCONHBoc}-$), 136.50, 132.76, 129.24, 128.58, 80.47 ($-\text{O}t\text{Bu}$), 48.78 ($-\text{BocNHCH}_2\text{CO}-$), 44.28 ($-\text{NHCH}_2\text{CO}-$), 28.28 ($-t\text{Bu}$), 25.47 ($-\text{SCH}_2\text{PPh}_2$). HR ESI-TOF MS (m/z): calcd for $\text{C}_{22}\text{H}_{28}\text{N}_2\text{O}_4\text{PS}$ [$\text{M} + \text{H}$] $^+$, 447.1507; found, 447.1503.

General procedure for peptide-GPI ligation.



entry	azide	thioester	temp ($^{\circ}\text{C}$)	solvent ^a	time (day)	product	yield ^b (%)
1	11	21	rt	DMF	2	24	55
2	11	21	40	DMF	1	24	96
3	11	21	40	DMF/ H_2O	1	24	93
4	11	23	40	DMF	1	25	93
5	11	23	40	DMF/ H_2O	1	25	92
6	18	21	40	DMF	1	26	93
7	18	21	40	DMF/ H_2O	1	26	91
8	18	23	40	DMF	1	27	92
9	18	23	40	DMF/ H_2O	1	27	89

An azide, **11** (5.15 mg, 0.015 mmol) or **18** (10.2 mg, 0.015 mmol) and a thioester, **21** (8.8 mg, 0.023 mmol) or **23** (10.1 mg, 0.023 mmol) were dissolved in DMF (0.5 mL) or DMF/ H_2O (1/1, v/v, 0.5 mL). After stirring at 40 $^{\circ}\text{C}$ for 1 day, the solution was concentrated under vacuum. The residue was purified by silica gel column chromatography to give **24**, **25**, **26**, and **27**, respectively (their yields are shown in Table 1)

24: a white foamy solid (6.83 mg, 96%, entry 2). $R_f = 0.50$ (EtOAc/ CH_3OH 1:1); ^1H NMR (600 MHz, D_2O) δ : 4.74 (s, 1H, H-1), 4.12 (dd, $J = 11.4, 5.4$ Hz, 1H, H-6a), 4.05 (dd, $J = 10.8, 5.4$ Hz, 1H, H-6b), 3.94 – 3.91 (m, 3H, $-\text{OCH}_2\text{CH}_2$, H-2), 3.77 – 3.69 (m, 5H, $-\text{CH}_2\text{NHBoc}$, H-3, H-4, H-5), 3.46 (s, 2H, $-\text{OCH}_2\text{CH}_2$), 3.39 (s, 3H, $-\text{OCH}_3$), 1.42 (s, 9H, $-t\text{Bu}$). ^{13}C NMR (150 MHz, D_2O) δ :

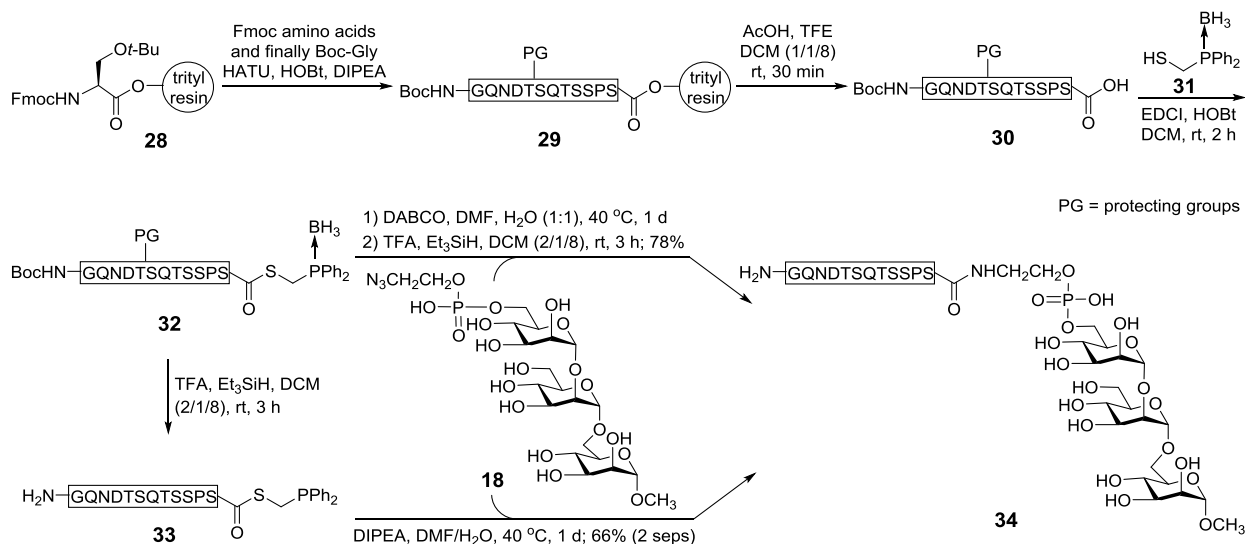
172.56 (-CONH), 158.00 (-*t*BuOCONH-), 100.83 (C-1), 81.56 (-*Ot*Bu), 71.31, 70.35, 69.74, 66.15, 64.39, 63.96, 54.68 (-OCH₃), 43.31 (-COCH₂NHBoc), 39.79 (-OCH₂CH₂NH), 27.49 (-*t*Bu). ³¹P NMR (160 MHz, D₂O) δ : 0.29. HR ESI-TOF MS (m/z): calcd for C₁₆H₃₀N₂O₁₂P [M - H]⁻, 473.1542; found, 473.1521.

25: a white foamy solid (7.33 mg, 92%, entry 5). *R_f* = 0.40 (EtOAc/CH₃OH 1:1); ¹H NMR (600 MHz, D₂O) δ : 4.74 (s, 1H, H-1), 4.12 (dd, *J* = 11.4, 5.4 Hz, 1H, H-6a), 4.05 (dd, *J* = 10.8, 5.4 Hz, 1H, H-6b), 3.93 – 3.91 (m, 5H, -OCH₂CH₂-COCH₂NH-, H-2), 3.81 (s, 2H, -CH₂NHBoc), 3.76 (dd, *J* = 9.0, 3.6 Hz, 1H, H-3), 3.70 – 3.69 (m, 2H, H-4, H-5), 3.46 (t, *J* = 5.4 Hz, 2H, -OCH₂CH₂), 3.39 (s, 3H, -OCH₃), 1.42 (s, 9H, -*t*Bu). ¹³C NMR (150 MHz, D₂O) δ : 173.18 (-CONH), 173.18 (-CONH), 156.94 (-*t*BuOCONH-), 100.81 (C-1), 81.65 (-*Ot*Bu), 71.29, 70.35, 69.74, 66.14, 64.37, 63.91, 54.66 (-OCH₃), 43.32 (-COCH₂NHBoc), 42.23 (-COCH₂NH), 39.88 (-OCH₂CH₂NH), 27.47 (-*t*Bu). HR ESI-TOF MS (m/z): calcd for C₁₈H₃₃N₃O₁₃P [M - H]⁻, 530.1756; found, 530.1728.

26: a white foamy solid (11.80 mg, 92%, entry 8). *R_f* = 0.50 (EtOAc/CH₃OH 1:2); ¹H NMR (600 MHz, D₂O) δ : 5.12 (s, 1H, H-1^{Man-III}), 5.02 (s, 1H, H-1^{Man-II}), 4.74 (s, 1H, H-1^{Man-I}), 4.11 – 4.02 (m, 3H), 3.98 – 3.92 (m, 6H), 3.86 – 3.84 (m, 3H), 3.82 (d, *J* = 11.4 Hz, 1H), 3.77 – 3.64 (m, 9H), 3.48 (t, *J* = 3.6 Hz, 2H, -OCH₂CH₂), 3.39 (s, 3H, -OCH₃), 1.43 (s, 9H, -*t*Bu). ¹³C NMR (150 MHz, D₂O) δ : 172.56 (-CONH), 157.99 (-*t*BuOCONH-), 102.32 (C-1^{Man-III}), 100.88 (C-1^{Man-I}), 97.80 (C-1^{Man-II}), 81.58 (-*Ot*Bu), 78.81, 72.65, 71.98, 70.65, 70.12, 69.81, 66.76, 66.54, 66.24, 65.75, 64.65, 64.01, 60.80, 60.03, 54.69 (-OCH₃), 43.34 (-COCH₂NHBoc), 39.88 (-OCH₂CH₂NH), 27.53 (-*t*Bu). ³¹P NMR (160 MHz, D₂O) δ : 0.50. HR ESI-TOF MS (m/z): calcd for C₂₈H₅₀N₂O₂₂P [M - H]⁻, 797.2598; found, 797.2605.

27: a white foamy solid (6.83 mg, 96%, entry 2). *R_f* = 0.45 (EtOAc/CH₃OH 1:2); ¹H NMR (600 MHz, D₂O) δ : 5.12 (s, 1H, H-1^{Man-III}), 5.02 (s, 1H, H-1^{Man-II}), 4.74 (s, 1H, H-1^{Man-I}), 4.11 – 4.03 (m, 3H), 3.99 – 3.92 (m, 7H), 3.87 – 3.79 (m, 6H), 3.77 – 3.63 (m, 8H), 3.48 (t, *J* = 5.4 Hz, 2H, -OCH₂CH₂), 3.39 (s, 3H, -OCH₃), 1.43 (s, 9H, -*t*Bu). ¹³C NMR (150 MHz, D₂O) δ : 173.16 (-CONH), 171.34 (-CONH), 159.88 (-*t*BuOCONH-), 102.31 (C-1^{Man-III}), 100.88 (C-1^{Man-I}), 97.79 (C-1^{Man-II}), 81.69 (-*Ot*Bu), 78.80, 72.64, 72.02, 70.64, 70.12, 69.81, 66.76, 66.54, 66.24, 65.73, 64.57, 63.91, 60.79, 60.03, 54.69 (-OCH₃), 43.36 (-COCH₂NHBoc), 42.28 (-COCH₂NH), 39.93 (-OCH₂CH₂NH), 27.52 (-*t*Bu). ³¹P NMR (160 MHz, D₂O) δ : 0.56. HR ESI-TOF MS (m/z): calcd for C₃₀H₅₃N₃O₂₃P [M - H]⁻, 854.2813; found, 854.2810.

Synthesis of CD52 peptide phosphinothioesters **32** and **33**.



CD52 peptide **30** was prepared by solid-phase synthesis on a peptide synthesizer using standard Fmoc chemistry. The synthesis started from commercial serine-modified 2-chlorotrytil resin **28**, and the completed peptide **30** was released from the resin with 10% AcOH in TFE and DCM (1/1/8 v/v/v). The product was purified by silica gel column chromatography (CH₃OH/CH₂Cl₂ 1:5) and characterized with MALDI-TOF MS (calcd for C₁₃₅H₁₇₉N₁₅O₂₆Na [M + Na]⁺, 2449.3; found, 2449.8). Peptide **30** (86 mg, 0.035 mmol) was then dissolved in CH₂Cl₂ (2 mL), and to the solution were added mercaptan **31** (13.08 mg, 0.053 mmol), EDCI (20.16 mg, 0.106 mmol), and HOBT (9.45 mg, 0.07 mmol) at rt. After stirring at rt for 2 h, the mixture was concentrated, and the residue was purified by silica gel column chromatography (CH₃OH/CH₂Cl₂ 1:10) to produce **32** (87.5 mg, 93%) as a white solid. MALDI-TOF MS: calcd for C₁₃₅H₁₇₉N₁₅O₂₆Na [M – BH₃ + Na]⁺, 2663.3; found, 2663.5.

To a solution of **32** (87.5 mg, 0.033 mmol) in CH₂Cl₂ (1.6 mL) were added TFA (0.4 mL) and Et₃SiH (0.2 mL). After stirring at rt for 3 h, the mixture was poured into cold ethyl ether (20 mL). The precipitated product **33** was filtered, dried under vacuum, and applied directly to the next step. MALDI-TOF MS: calcd for C₅₈H₈₃N₁₅O₂₃PS [M – H]⁻, 1420.5; found, 1420.7.

Staudinger ligation between **32 and **18**.** To a solution of **32** (2.3 mg, 0.87 nmol) and **18** (0.85 mg, 1.30 nmol) in DMF and H₂O (1:1, v/v, 300 μL) was added DABCO (0.3 mg, 2.60 nmol). After stirring at 40 °C for 1 day and MS indication of completed reaction, the mixture was concentrated under vacuum. The residue was dissolved in 20% TEA in Et₃SiH and DCM (2/1/8, v/v/v, 0.5 mL). After stirring at rt for 3 h, 5 mL of cold ether was added to the mixture to precipitate the

reaction product, which was finally purified by reversed-phase HPLC to afford **34** (1.24 mg, 78%) as a white solid. HPLC conditions: C18 column (5 μm , 250 x 4.6 mm), gradient elution with 10~60% CH_3CN in H_2O , 1.5 mL/min flow rate, UV detection at 220 nm, retention time 9.5 min. MALDI-TOF MS: calcd for $\text{C}_{66}\text{H}_{110}\text{N}_{16}\text{O}_{42}\text{P}$ $[\text{M} - \text{H}]^-$, 1829.7; found, 1829.9.

Staudinger ligation between 33 and 18. To a solution of **33** (1.5 mg, 1.05 nmol) and **18** (1.1 mg, 1.60 nmol) in DMF and H_2O (1:1, v/v, 0.5 mL) was added DIPEA (1.84 μL , 10.5 nmol). After stirring at 40 $^\circ\text{C}$ for 1 day, the mixture was concentrated and the product was purified by HPLC as described above to give **34** (1.24 mg, 66%) as a white solid (identical to the product obtained from the above procedure).

II. MS spectra and HPLC diagrams of CD52 peptide, its phosphinothioesters, and its GPI conjugate

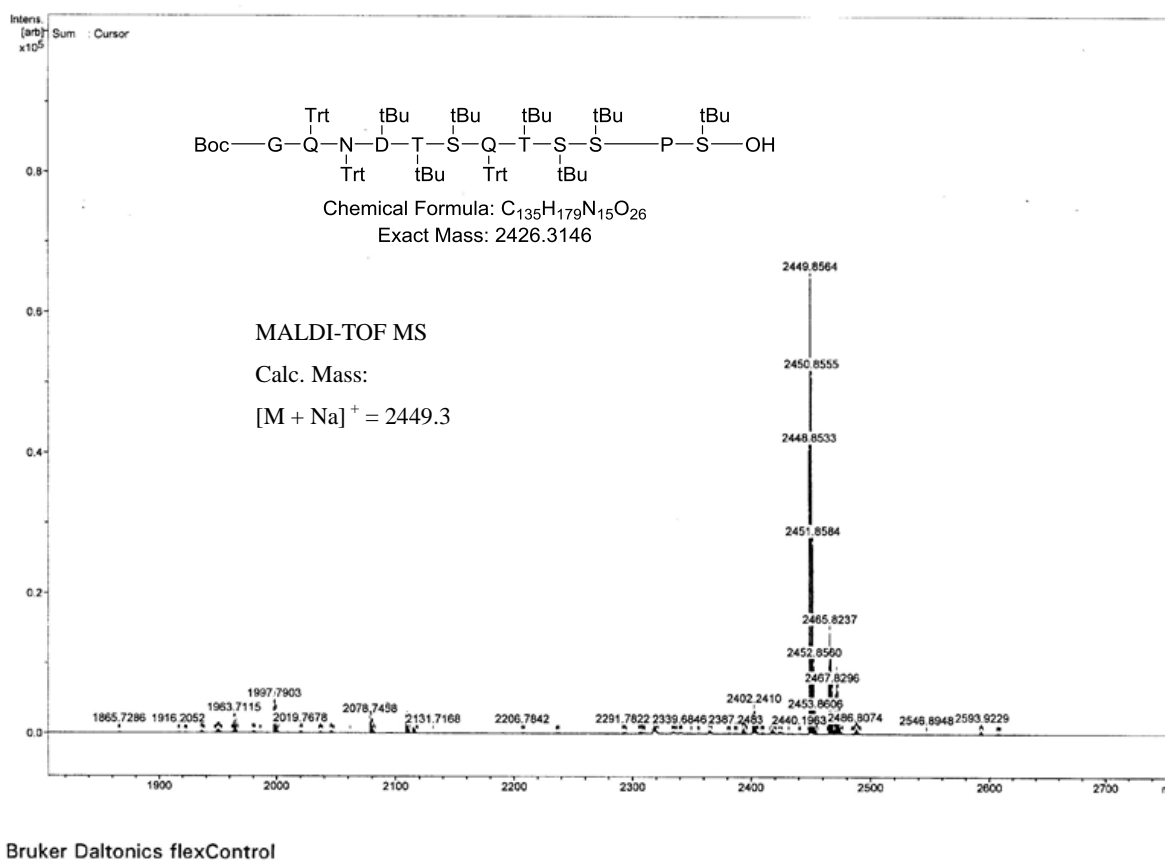


Figure S1. MALDI-TOF MS spectrum of the peptide **30** obtained from solid-phase synthesis

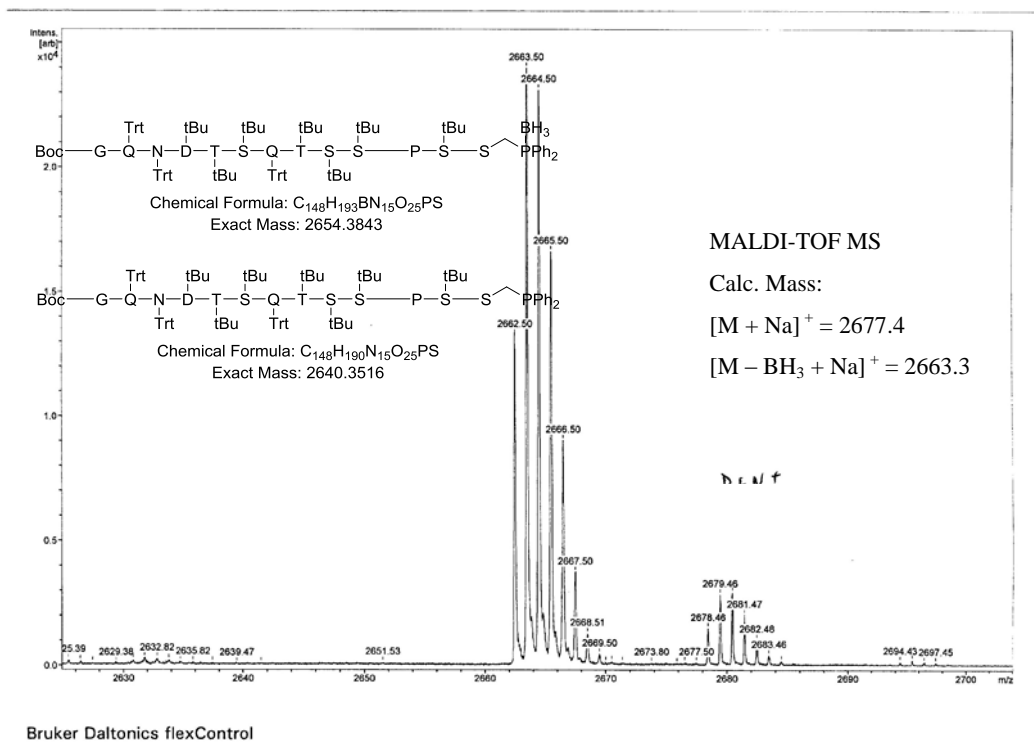


Figure S2. MALDI-TOF MS spectrum of peptide phosphinothioester **32**

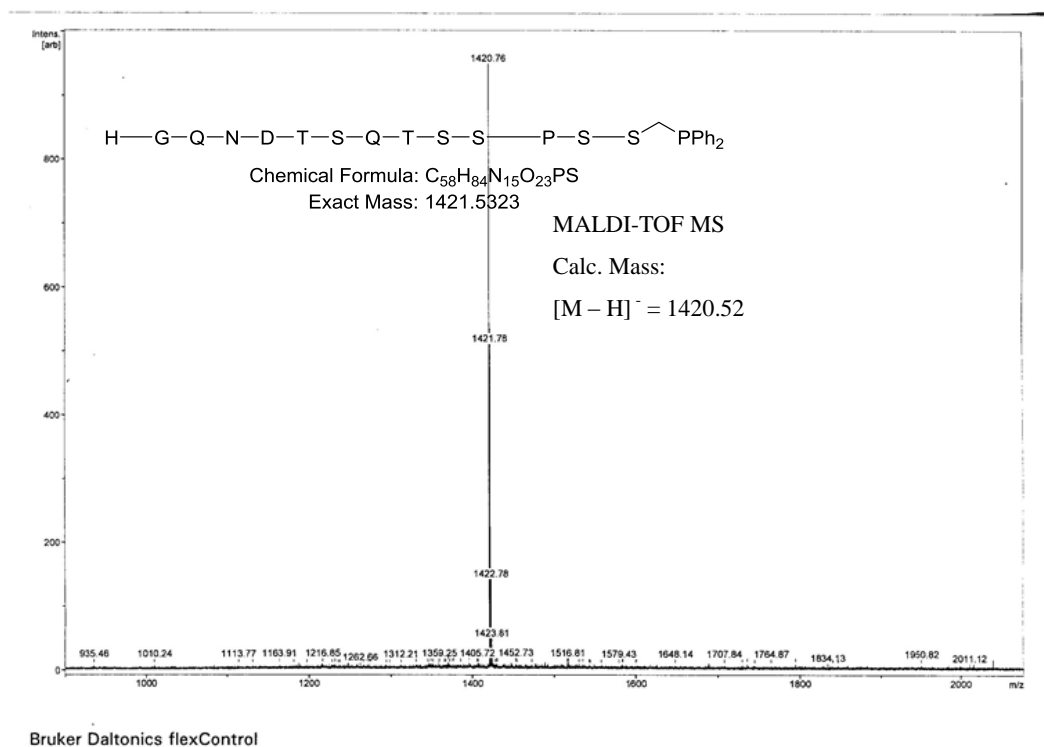


Figure S3. MALDI-TOF MS spectrum of peptide phosphinothioester **33**

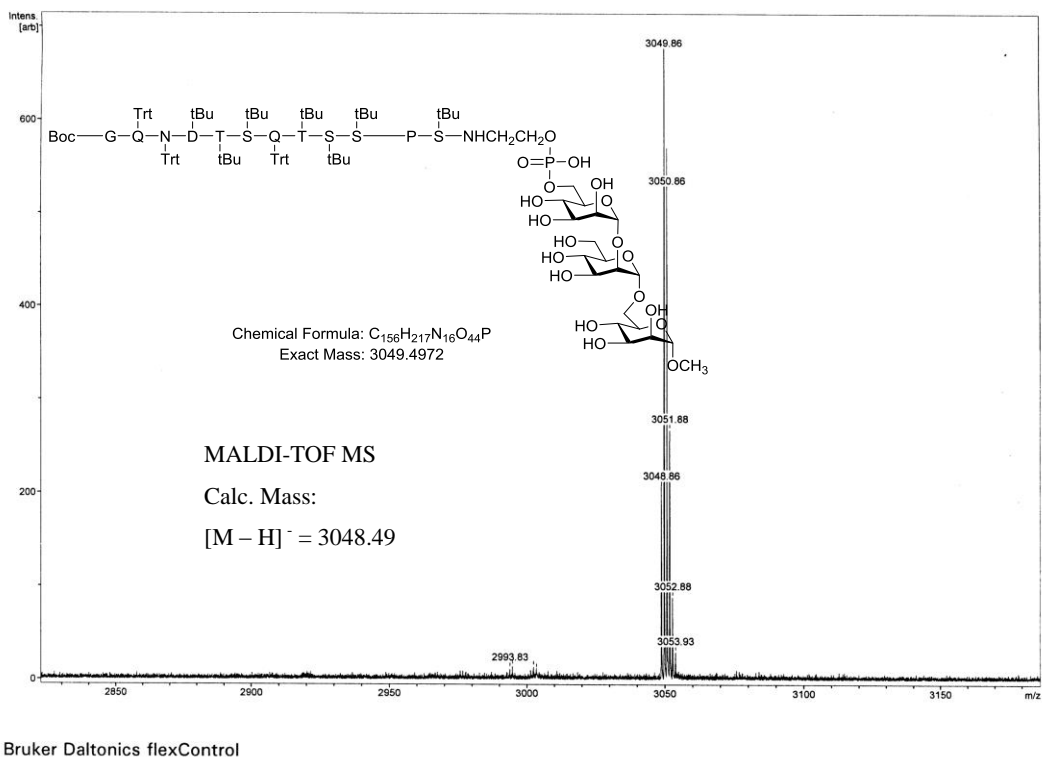


Figure S4. MALDI-TOF MS spectrum of the reaction product of **18** and **32** (i.e., protected **34**)

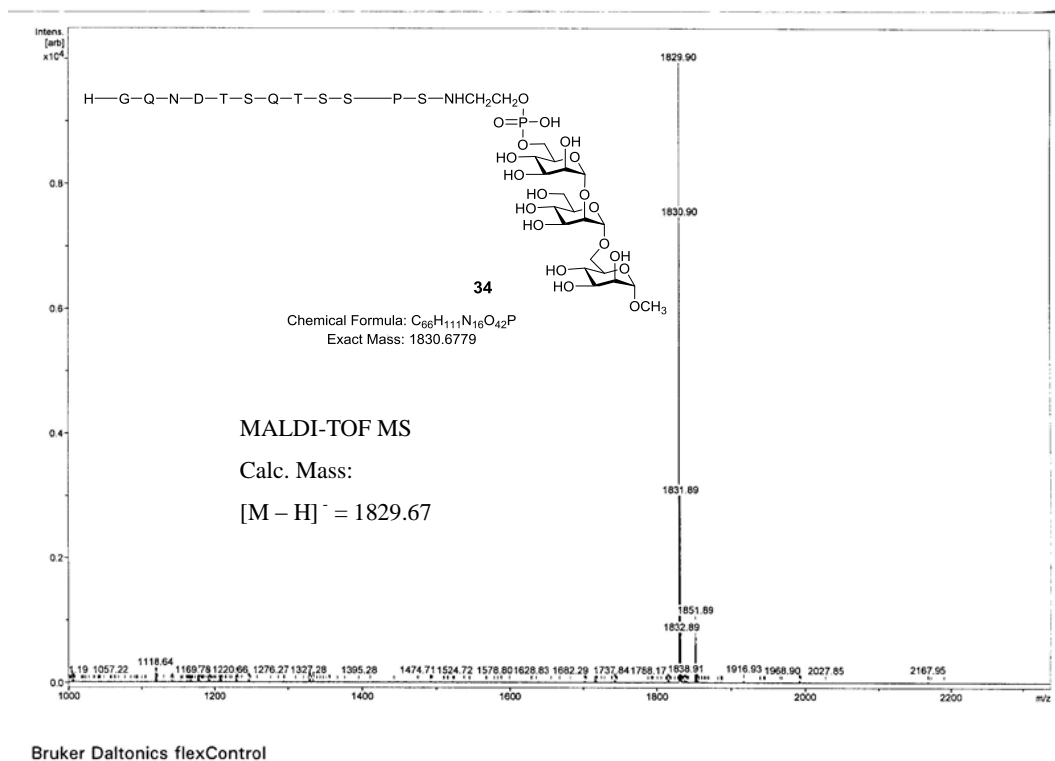


Figure S5. MALDI-TOF MS spectrum of CD52 analog **34**

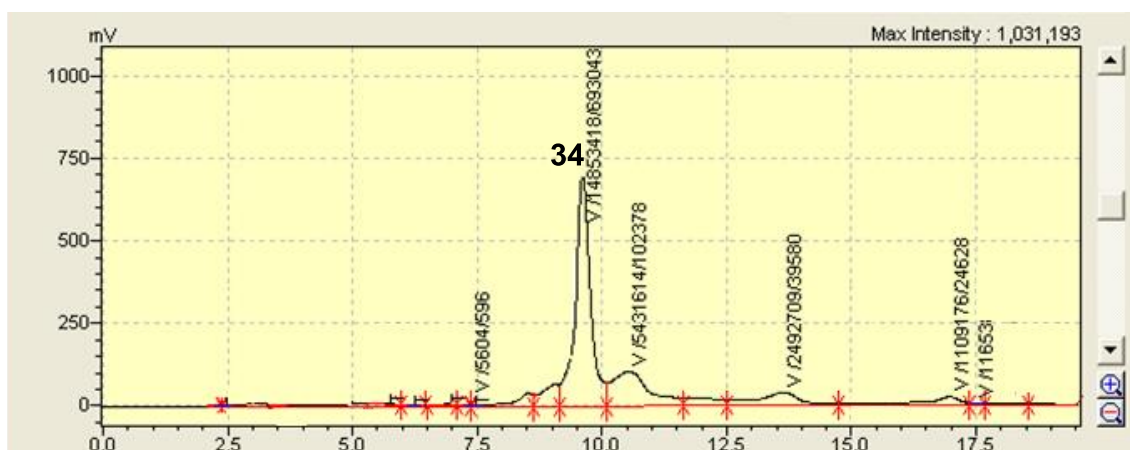


Figure S6. HPLC diagram of **34** derived from **18** and **32** (after the deprotection step). C18 column (5 μ m, 250 x 4.6 mm); gradient eluent: 10-60% CH₃CN in H₂O; flow rate: 1.5 mL/min, UV detection at 220 nm.

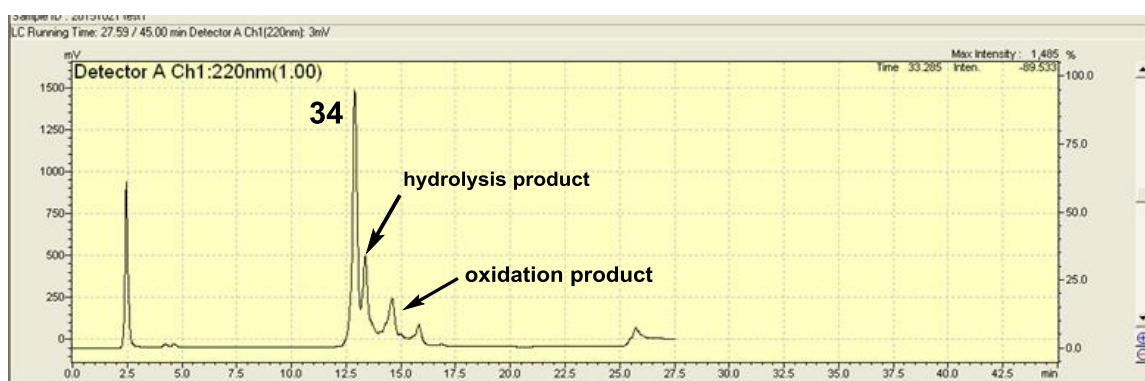


Figure S7. HPLC diagram of **34** derived from **18** and **33**. C18 column (5 μ m, 250 x 4.6 mm); gradient eluent: 5-60% CH₃CN in H₂O; flow rate: 1.5 mL/min, UV detection at 220 nm.

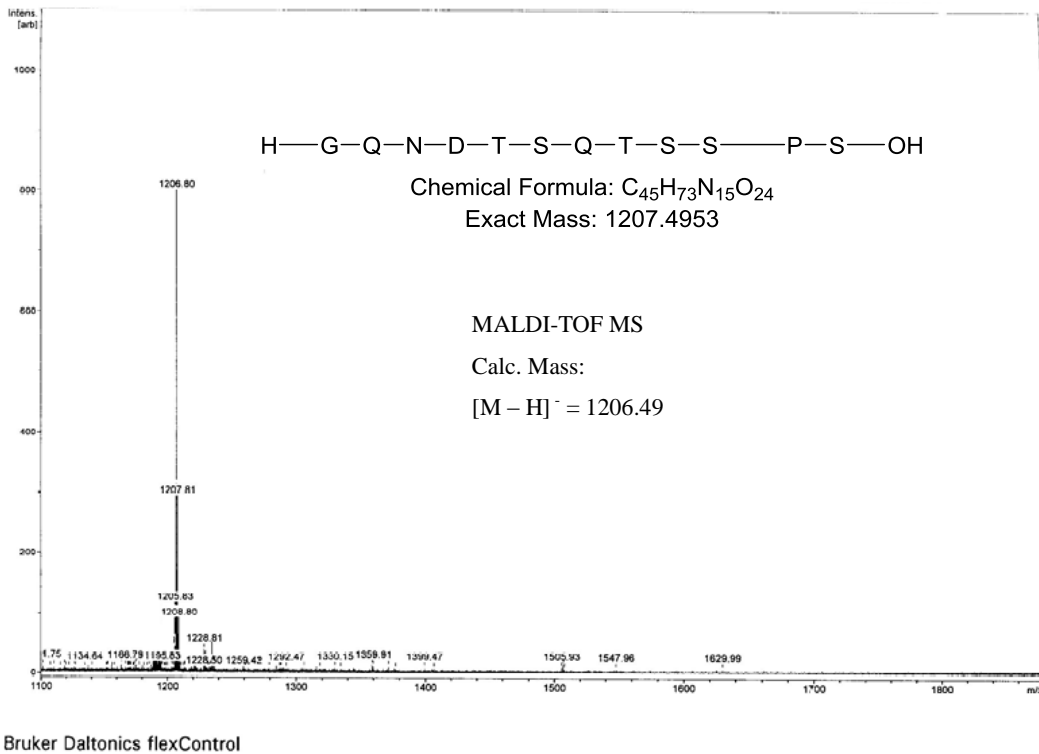


Figure S8. MALDI-TOF MS spectrum of the hydrolysis product of **33**

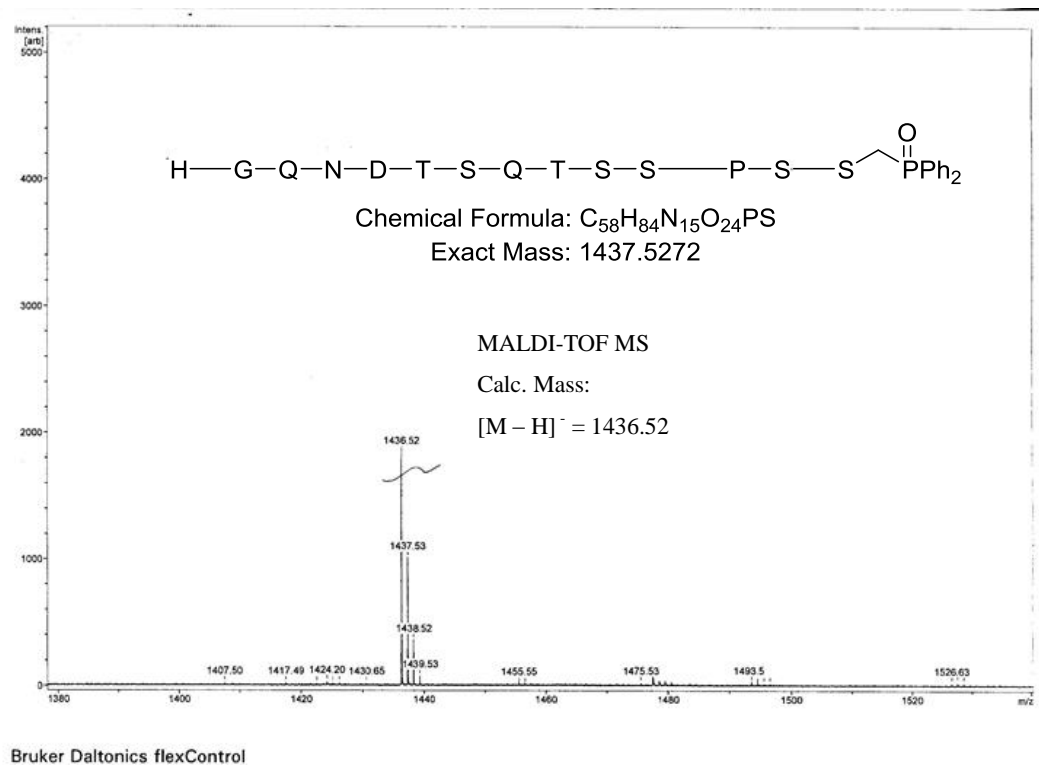
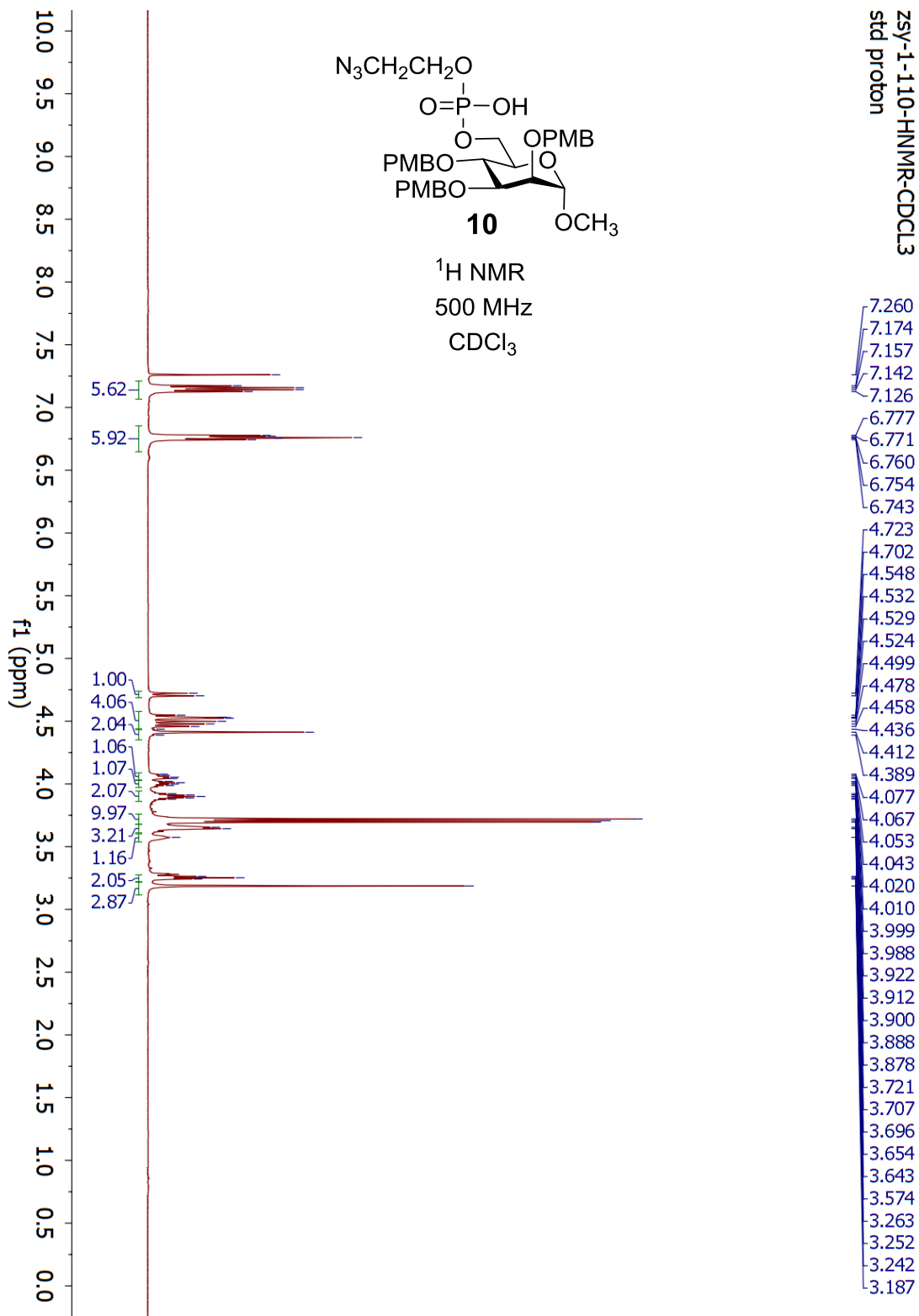
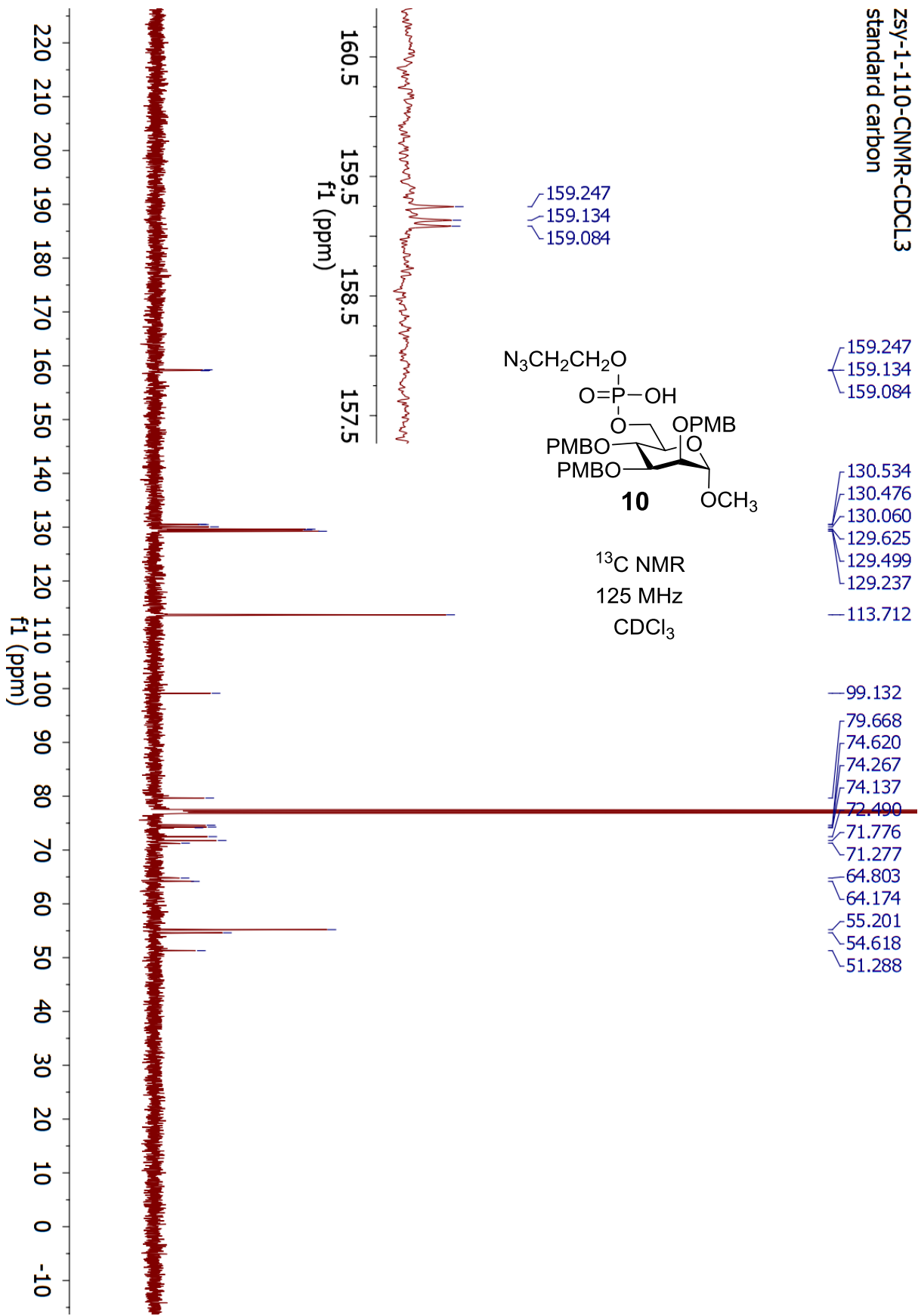


Figure S9. MALDI-TOF MS spectrum of the oxidation product of **33**

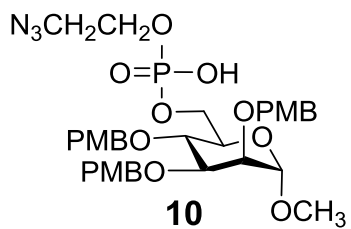
III. NMR and MS spectra of synthetic intermediates and other GPI-peptide conjugates



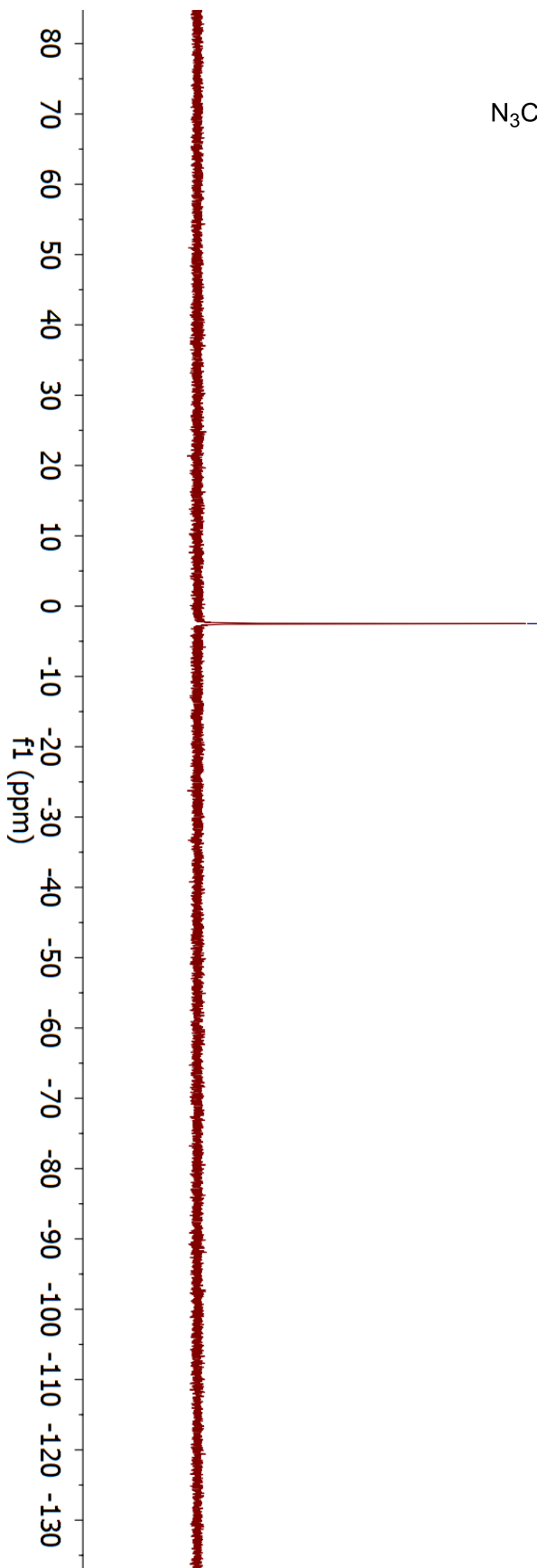
zsy-1-110-CNMR-CDCL3
standard carbon



zsy-1-110-PNMR-CDCl3
P31 spectrum

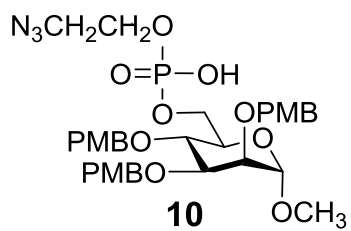


³¹P NMR
160 MHz
CDCl₃

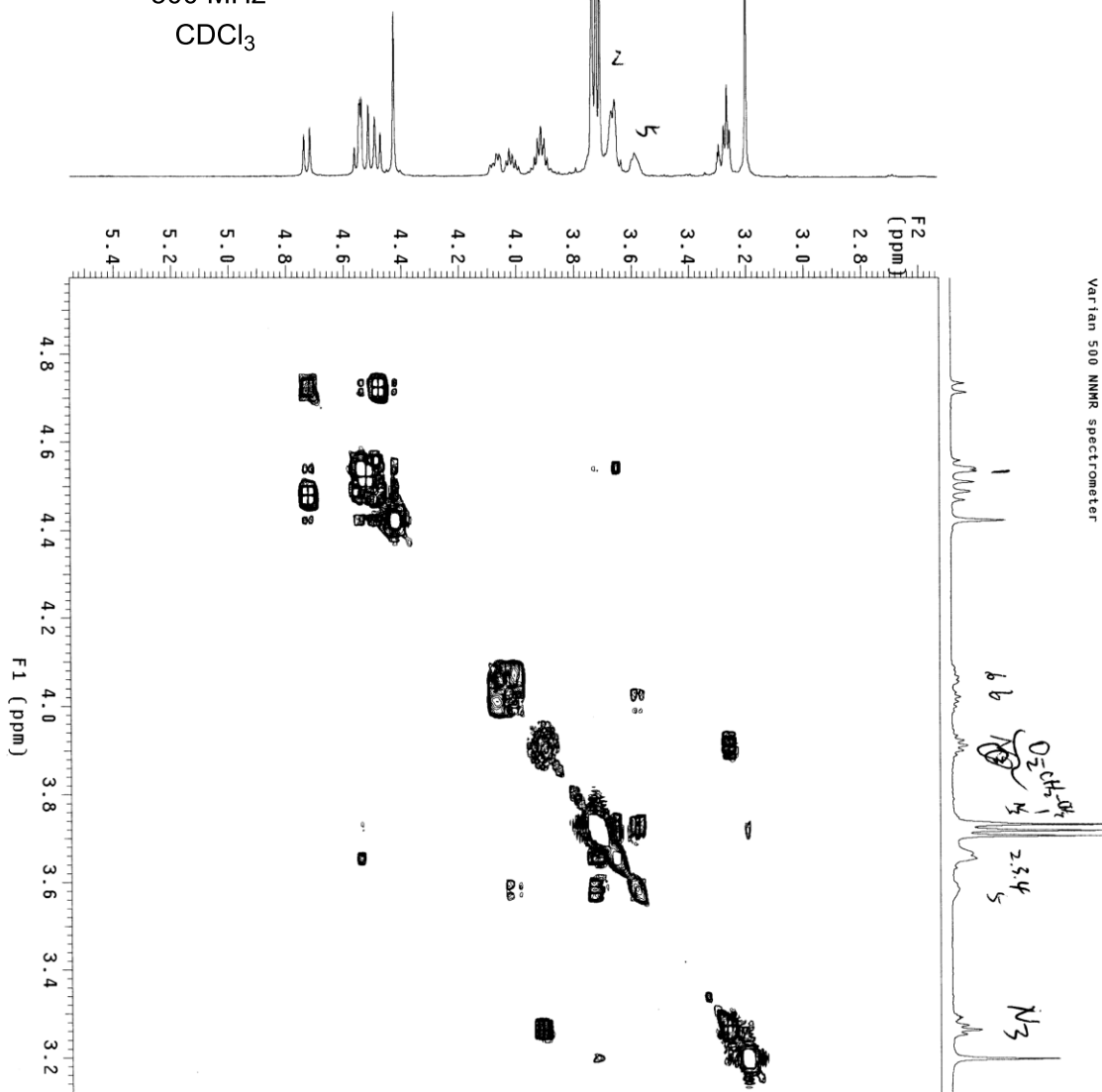


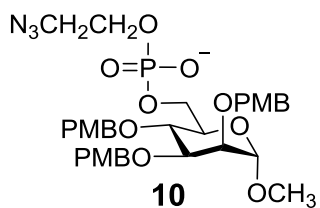
—2.491

ZSY-1-110-gcosy-cdcl3
File: xp
Pulse Sequence: gcosy



^1H - ^1H COSY
500 MHz
 CDCl_3





Exact Mass: 702.2433

HR ESI-TOF MS

Calc. Mass:

[M - H]⁻ = 702.2433

Elemental Composition Report

Single Mass Analysis

Tolerance = 5.0 PPM / DBE: min = -1.5, max = 100.0

Element prediction: Off

Number of isotope peaks used for i-FIT = 6

Monoisotopic Mass, Even Electron Ions

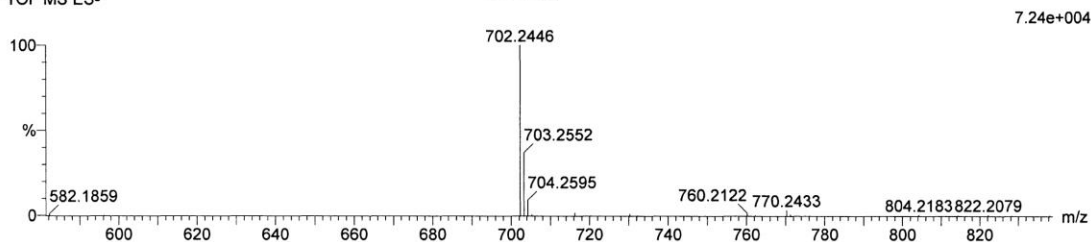
225 formula(e) evaluated with 1 results within limits (all results (up to 1000) for each mass)

Elements Used:

C: 33-33 H: 0-50 N: 0-5 O: 0-20 31P: 0-1

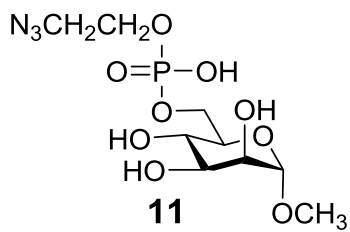
LCT Premier KD128
TOF MS ES-

S.Zhu ZSY-1-110 inMeOH Cone(V)50

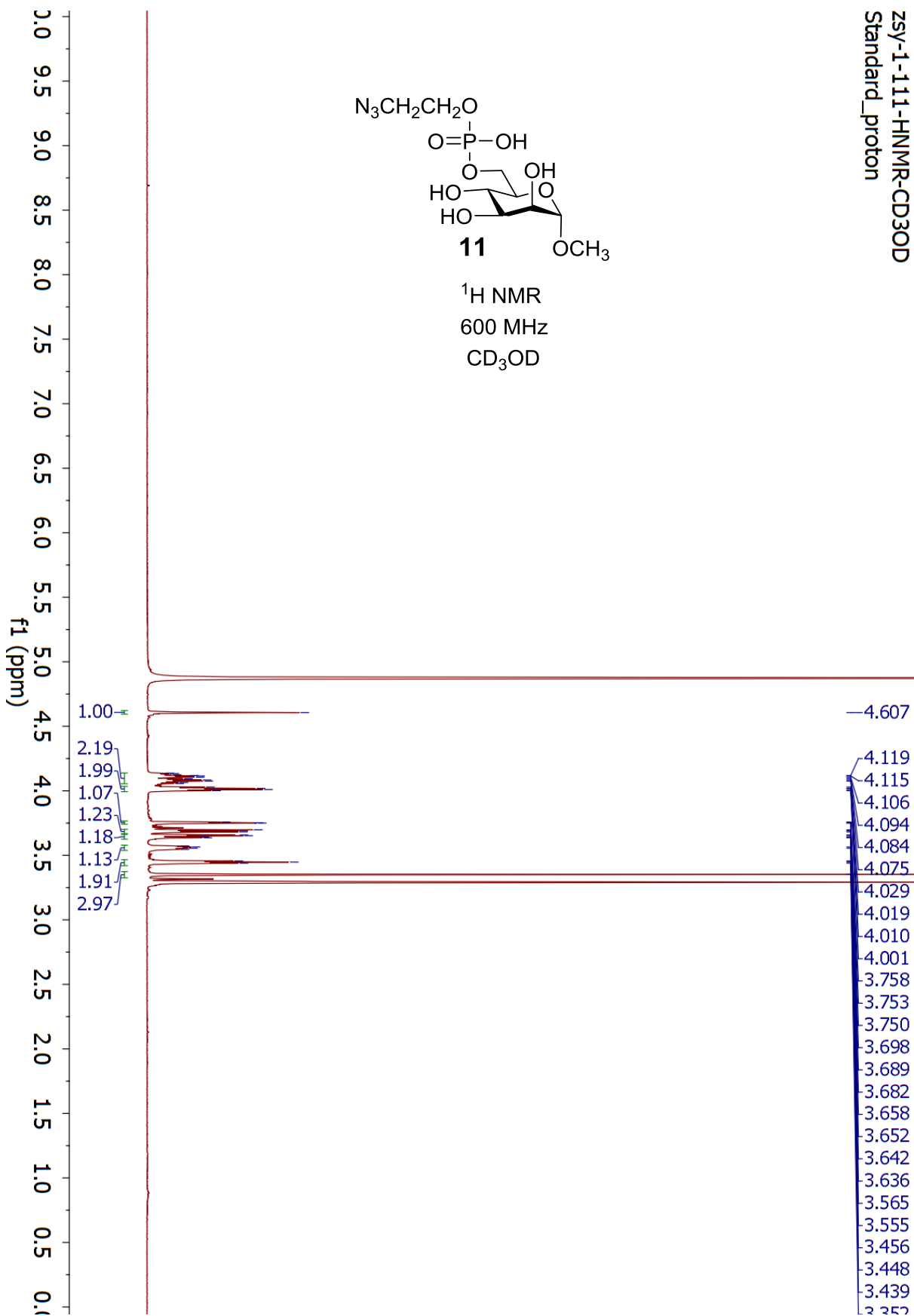


Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	i-FIT (Norm)	Formula
702.2446	702.2428	1.8	2.6	15.5	51.8	0.0	C33 H41 N3 O12 31P

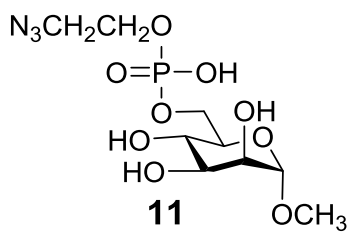
zsy-1-111-HNMR-CD3OD
Standard_proton



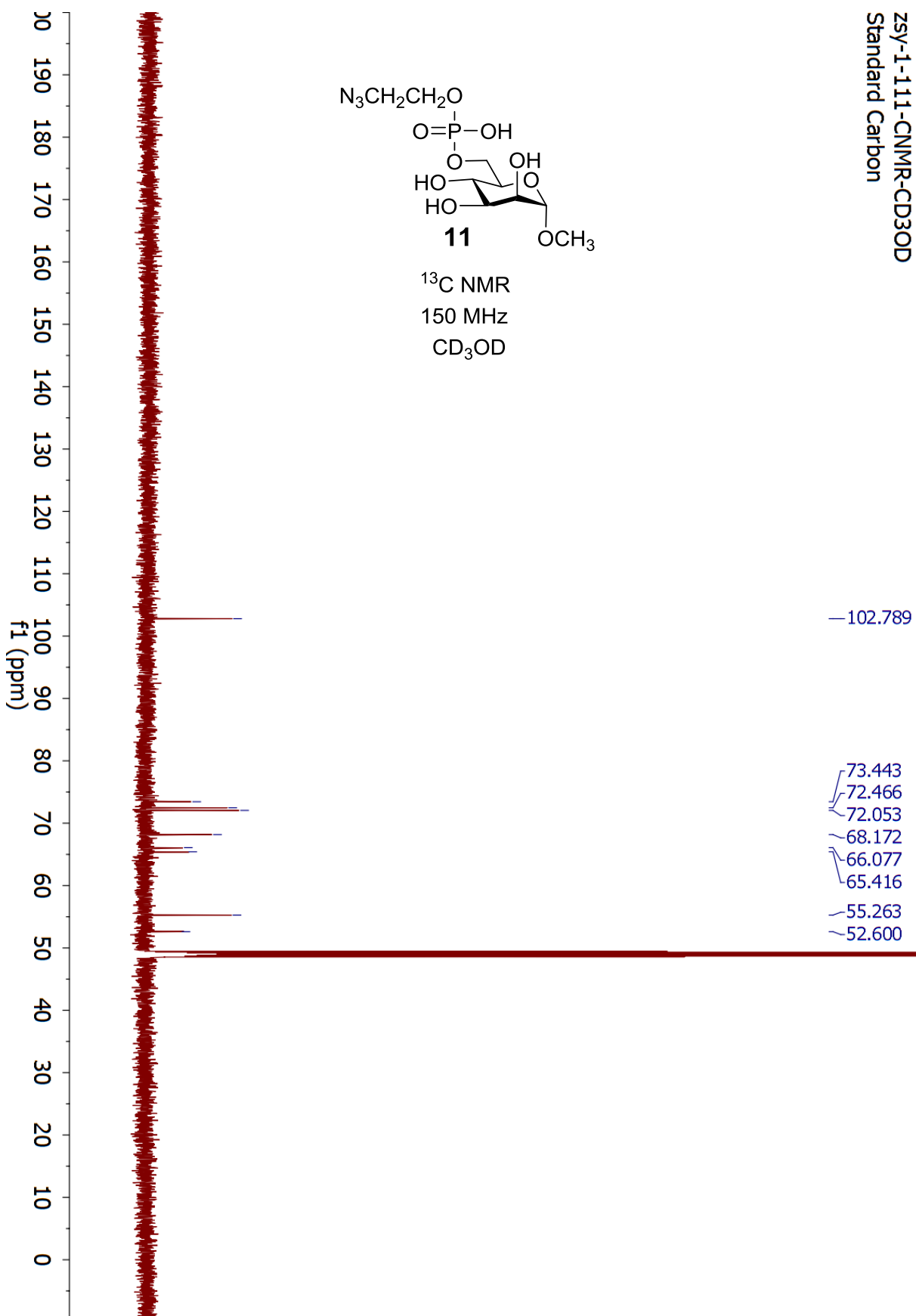
¹H NMR
600 MHz
CD₃OD



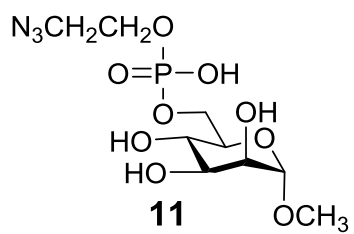
zsy-1-111-CNMR-CD3OD
Standard Carbon



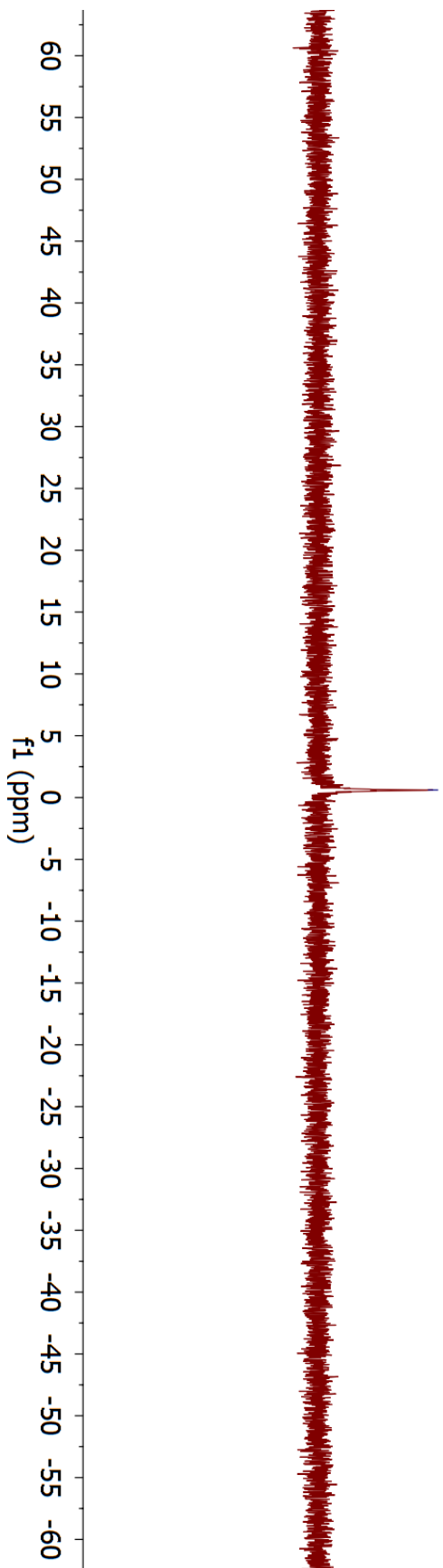
^{13}C NMR
150 MHz
 CD_3OD



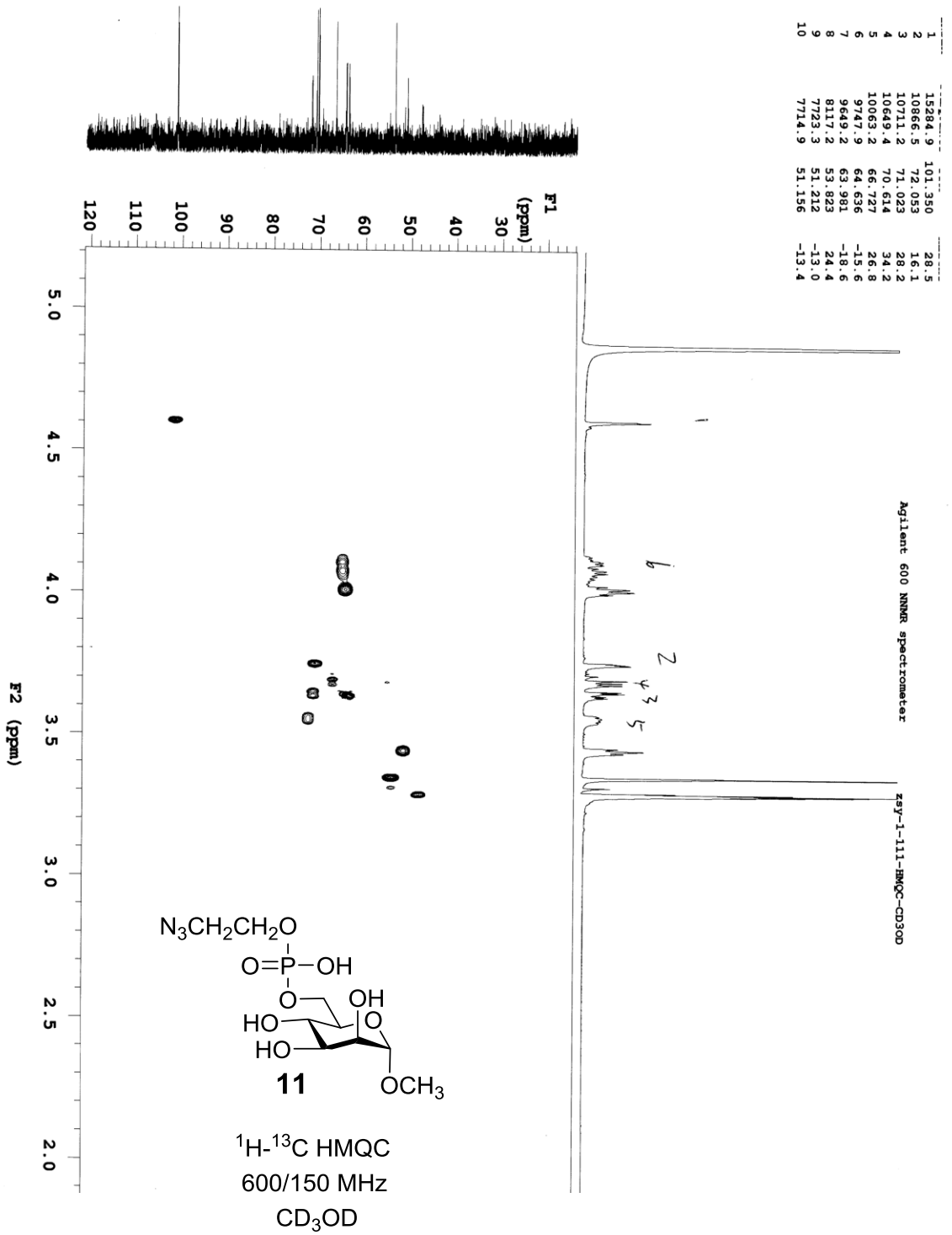
zsy-1-111-PNMR-CD3OD
P31 spectrum

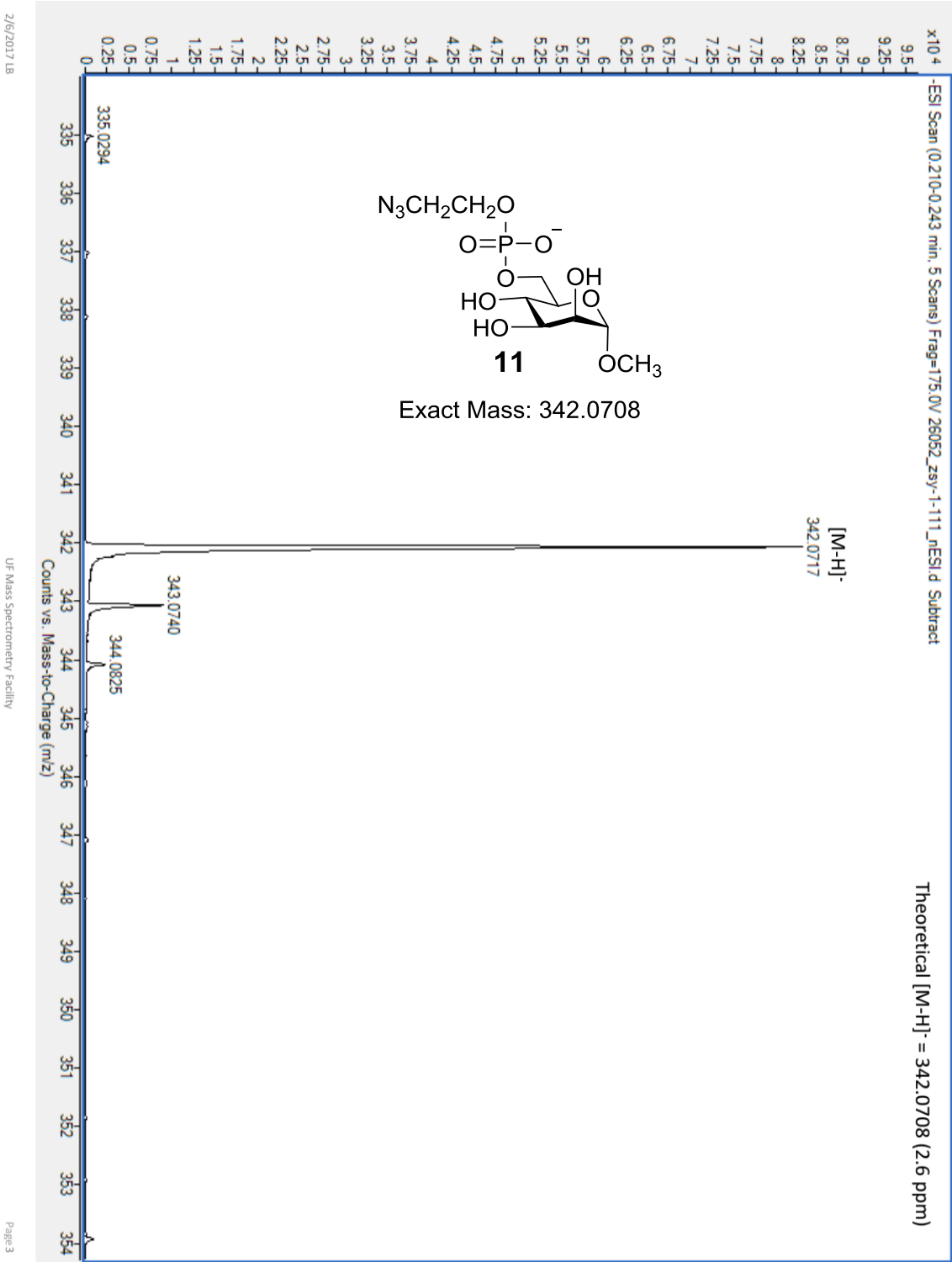


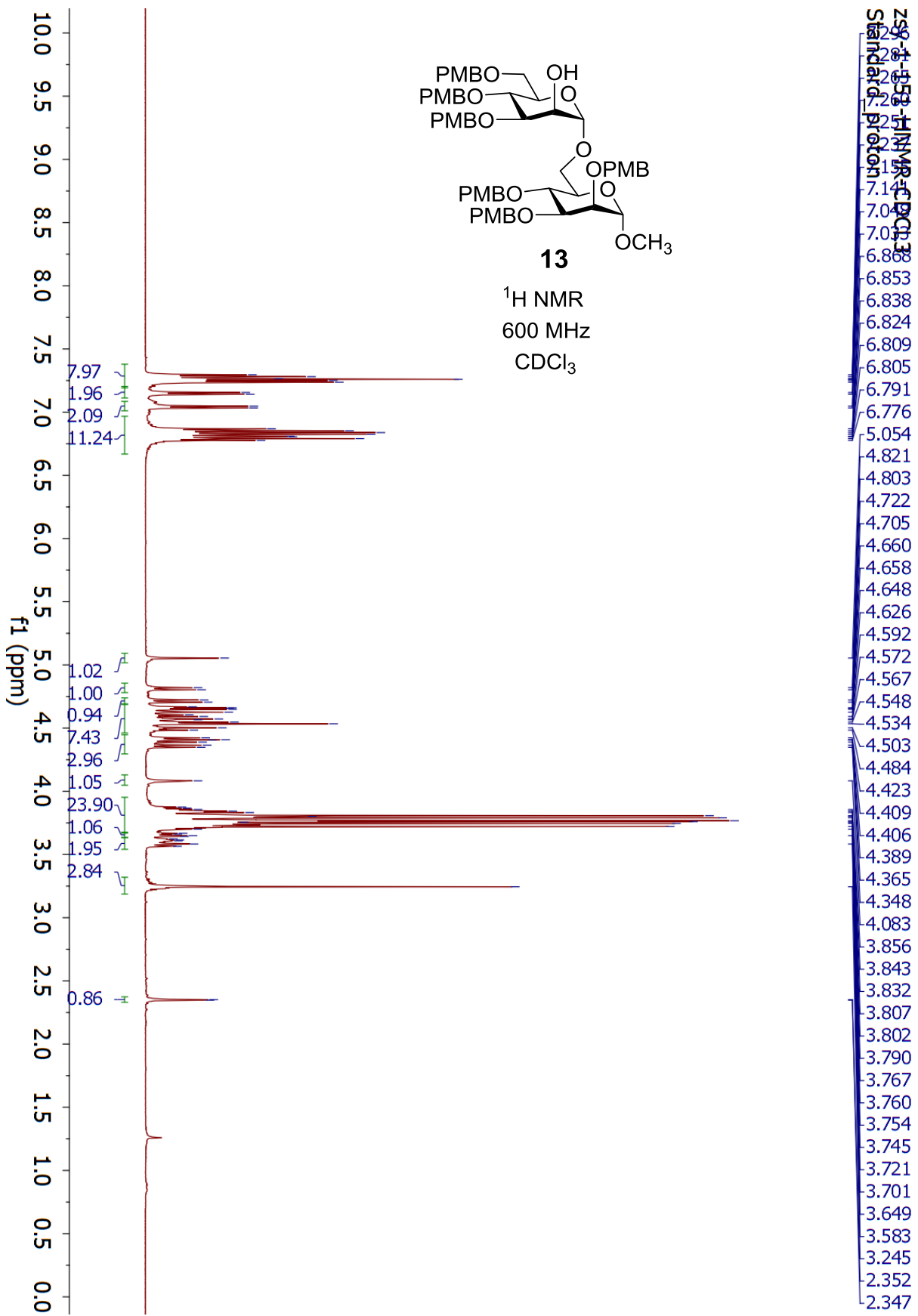
³¹P NMR
160 MHz
CD₃OD



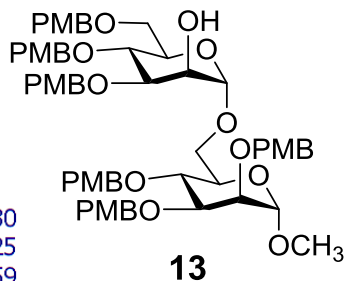
—0.609





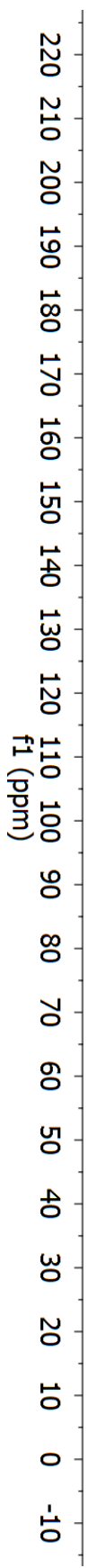
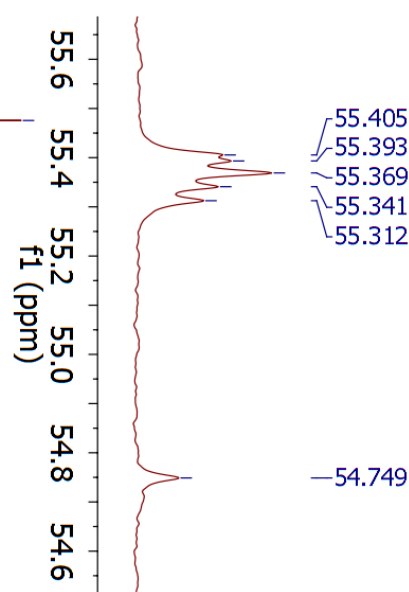
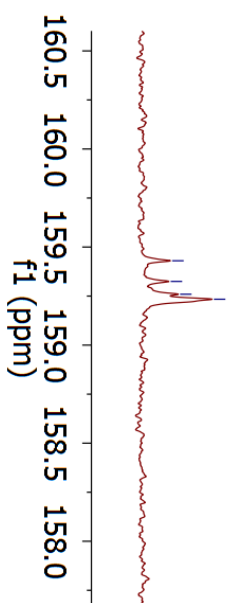


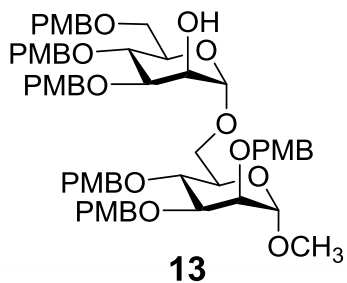
zsy-1-151-CNMR-CDCL3
Standard Carbon



¹³C NMR
150 MHz
CDCl₃

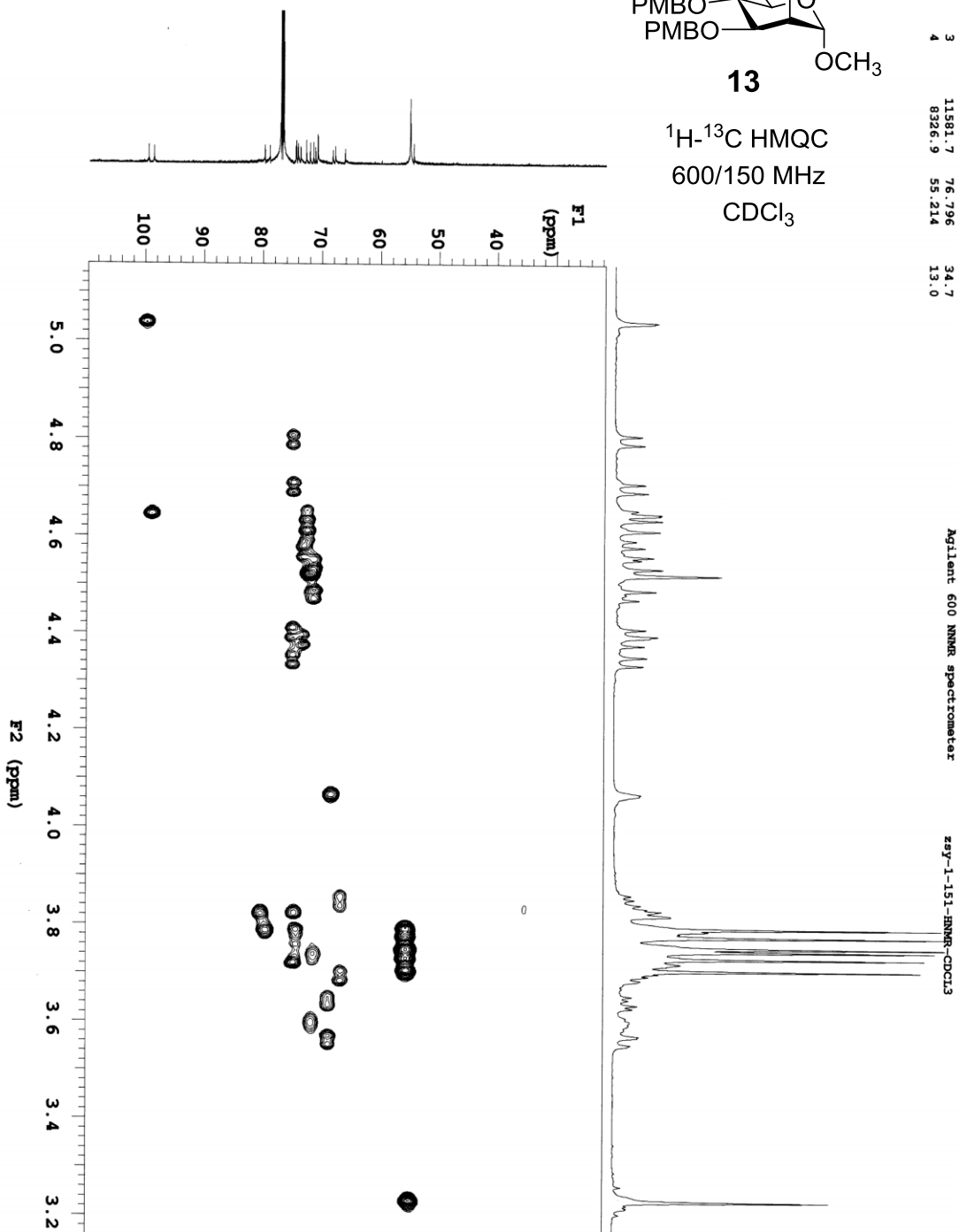
- 159.430
- 159.325
- 159.259
- 159.234
- 130.860
- 130.824
- 130.534
- 130.110
- 129.774
- 129.696
- 129.650
- 129.528
- 129.400
- 113.991
- 113.875
- 113.855
- 113.807
- 113.762
- 99.777
- 98.884
- 80.069
- 79.184
- 74.837
- 74.773
- 74.485
- 74.446
- 74.032
- 73.078
- 72.444
- 71.832
- 71.500
- 71.138
- 68.558
- 68.163
- 66.484
- 55.405
- 55.393
- 55.369
- 55.341
- 55.312
- 54.749

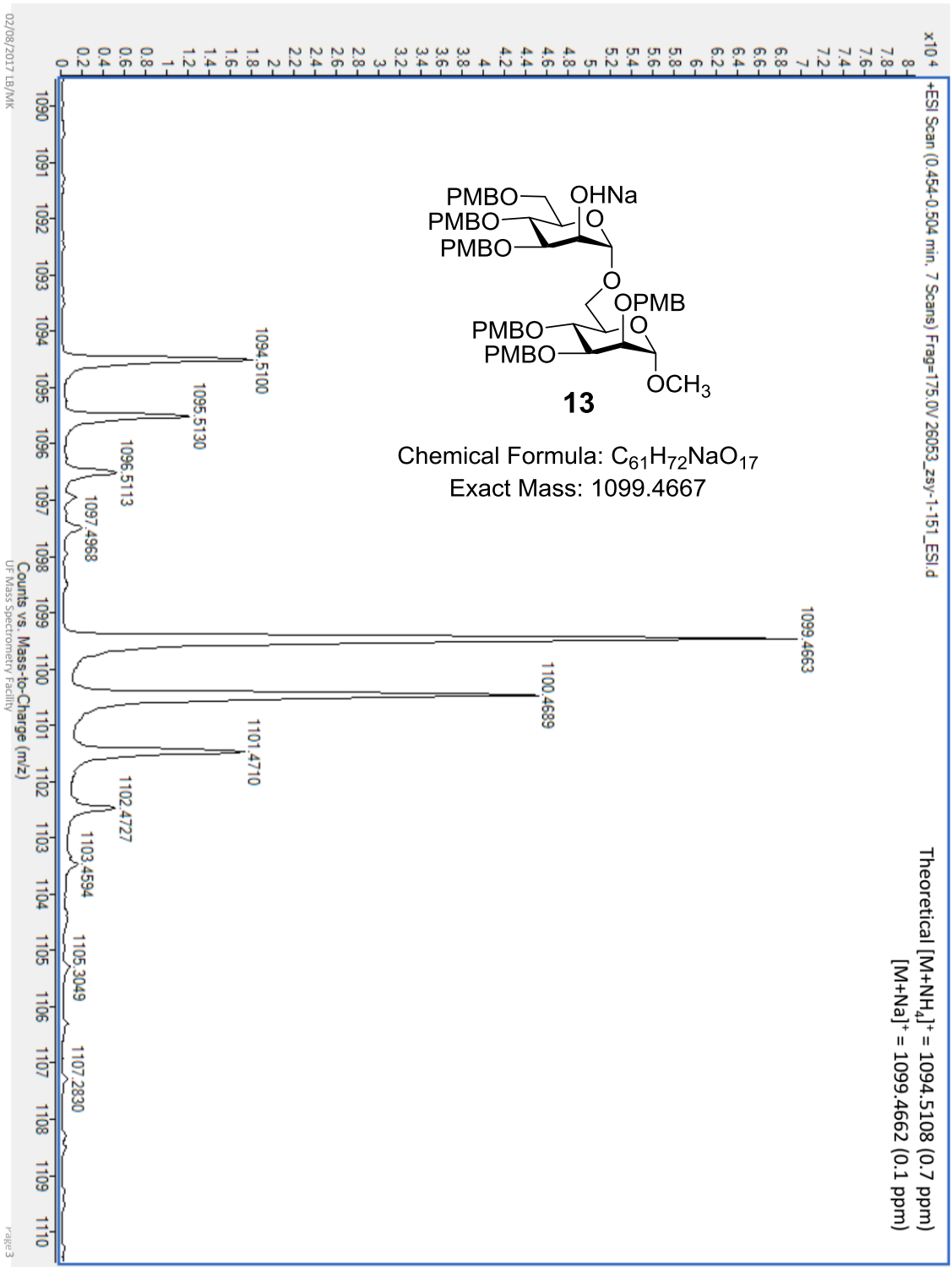


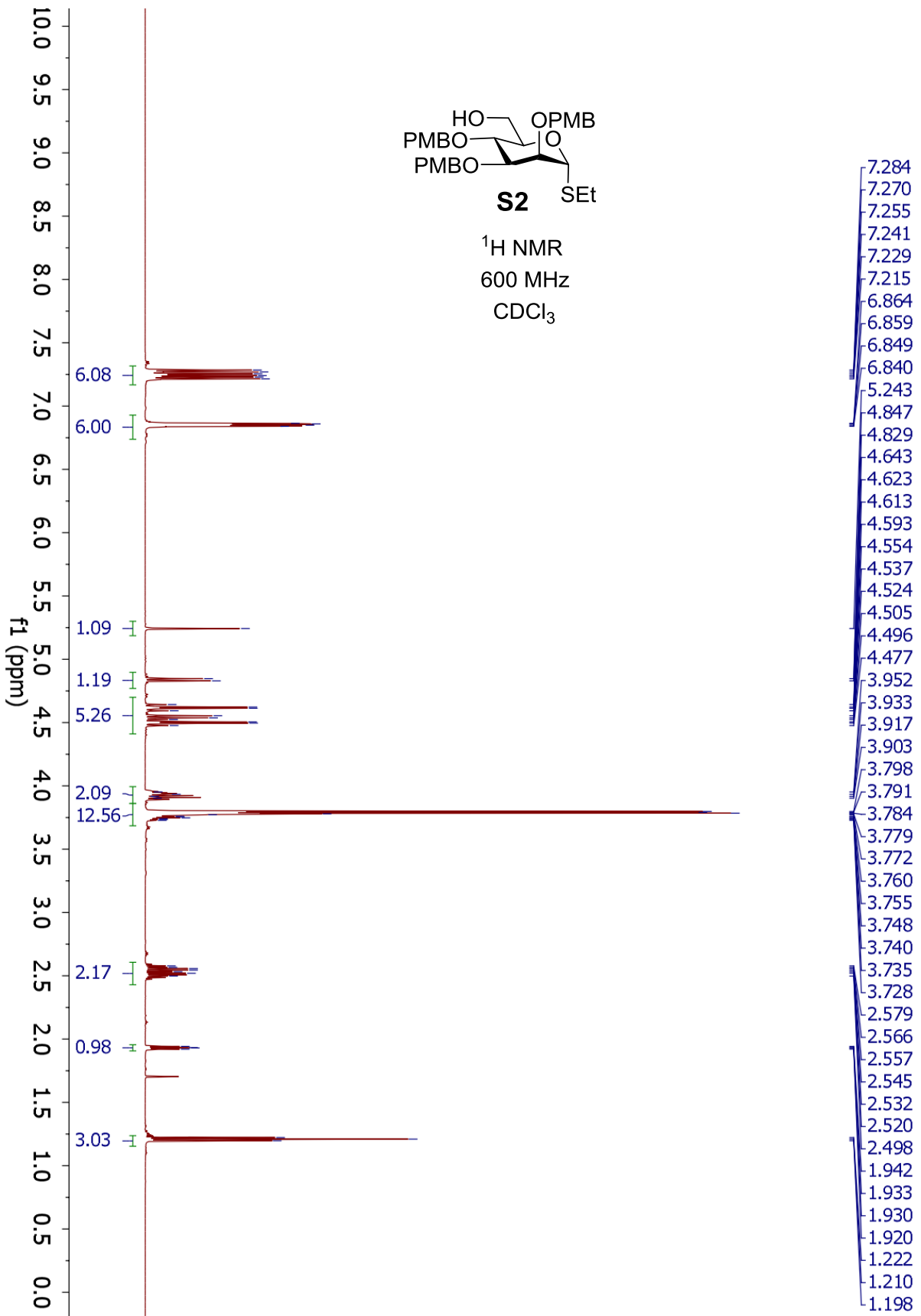


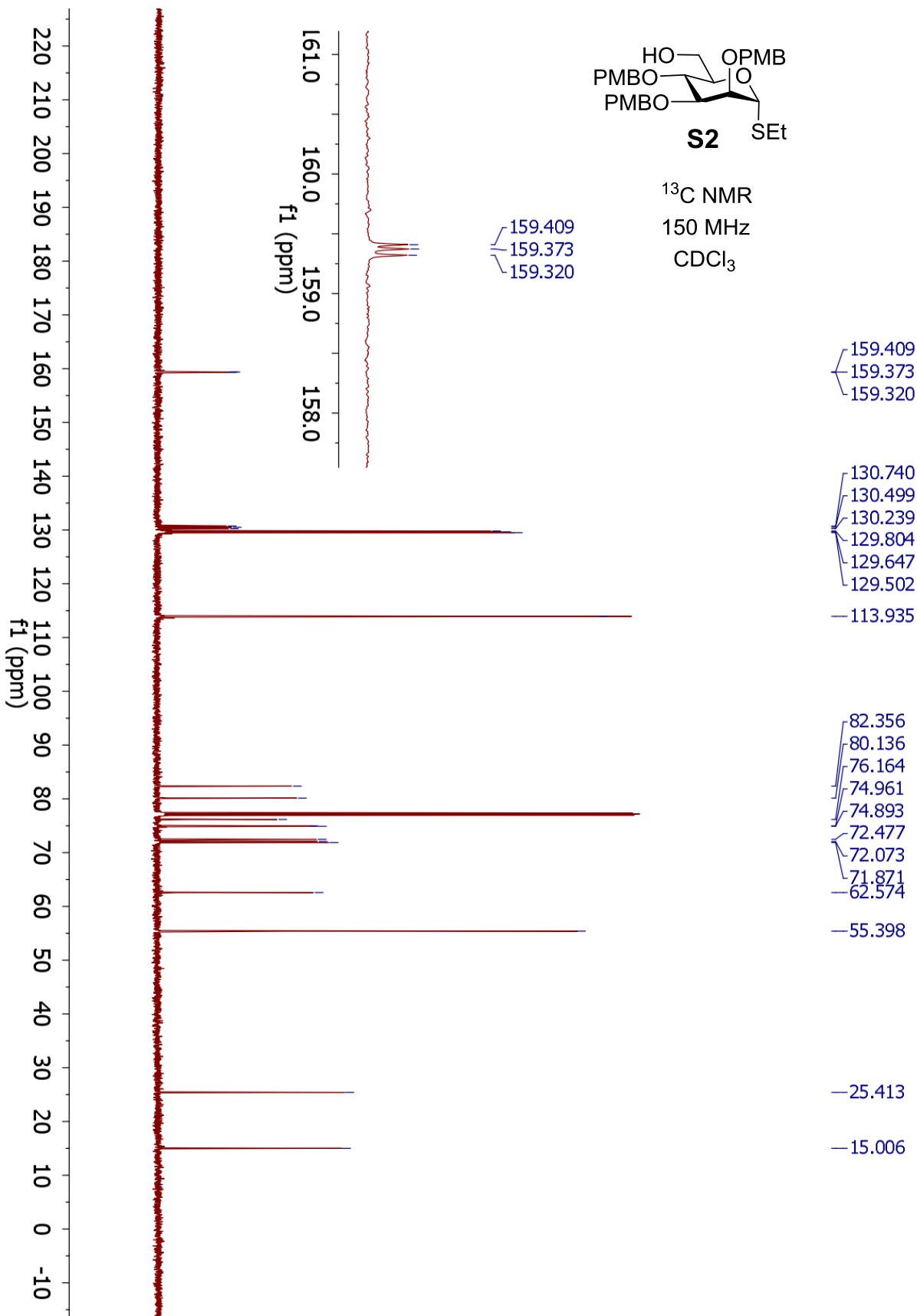
^1H - ^{13}C HMQC
600/150 MHz
 CDCl_3

1	11645.7	77.220	34.3
2	11613.7	77.008	35.0
3	11581.7	76.796	34.7
4	8326.9	55.214	13.0









Sample Name :

Data Collected on:

a600-vnmr5600

Archive directory:

/home/vnmr1/vnmr5600/probes/probe_calibs

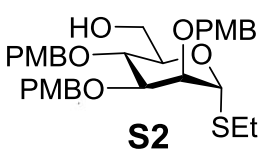
Sample directory:

Fidfile: gcosy

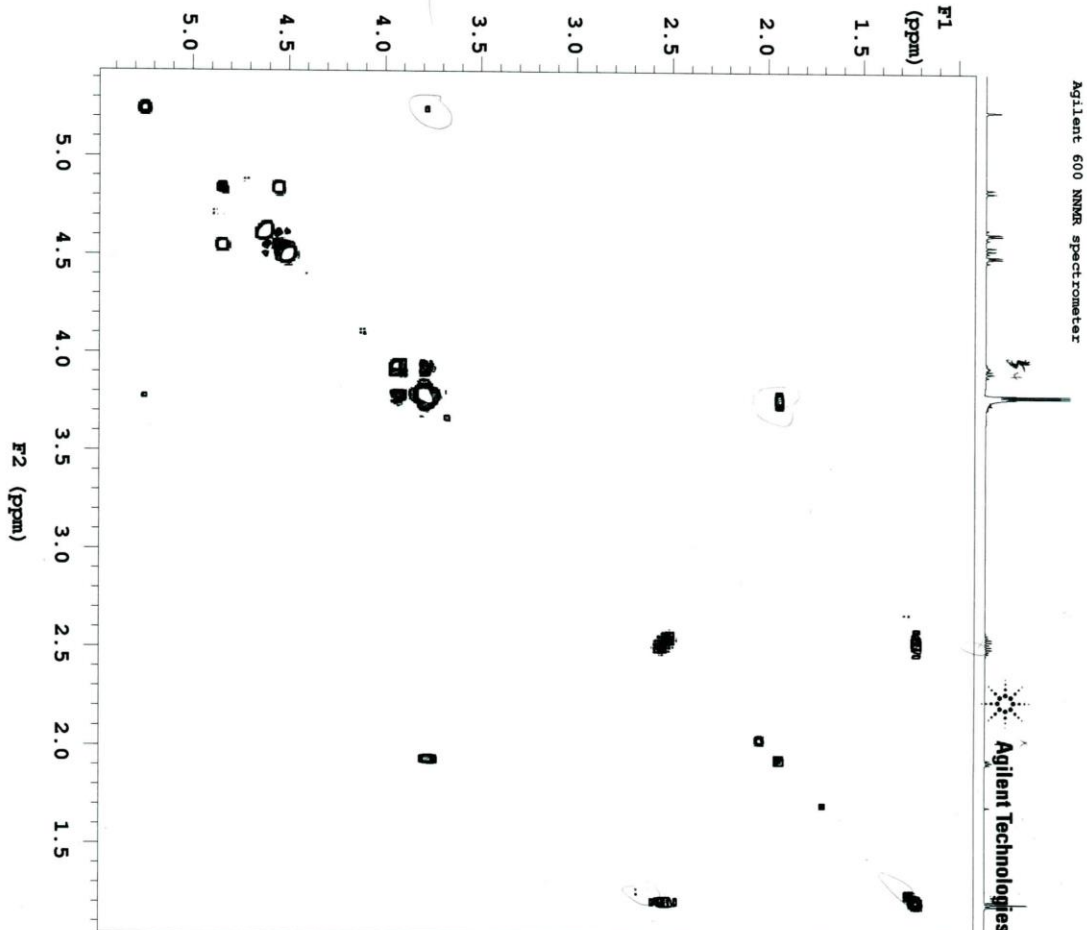
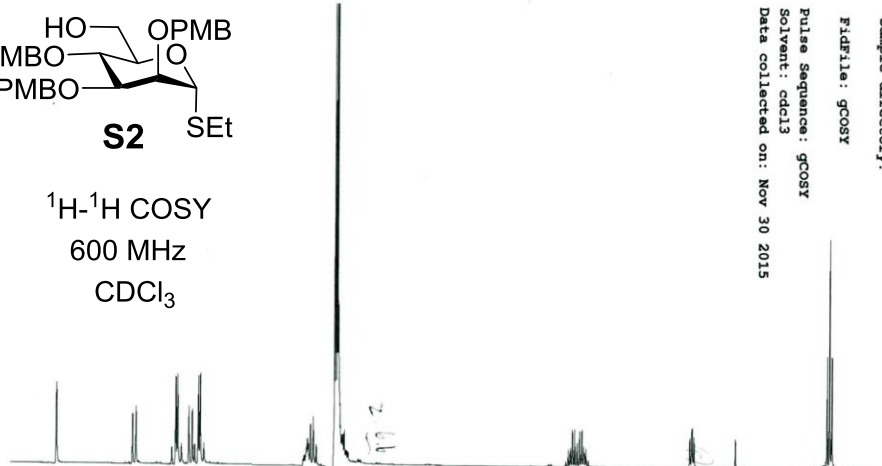
Pulse Sequence: gcosy

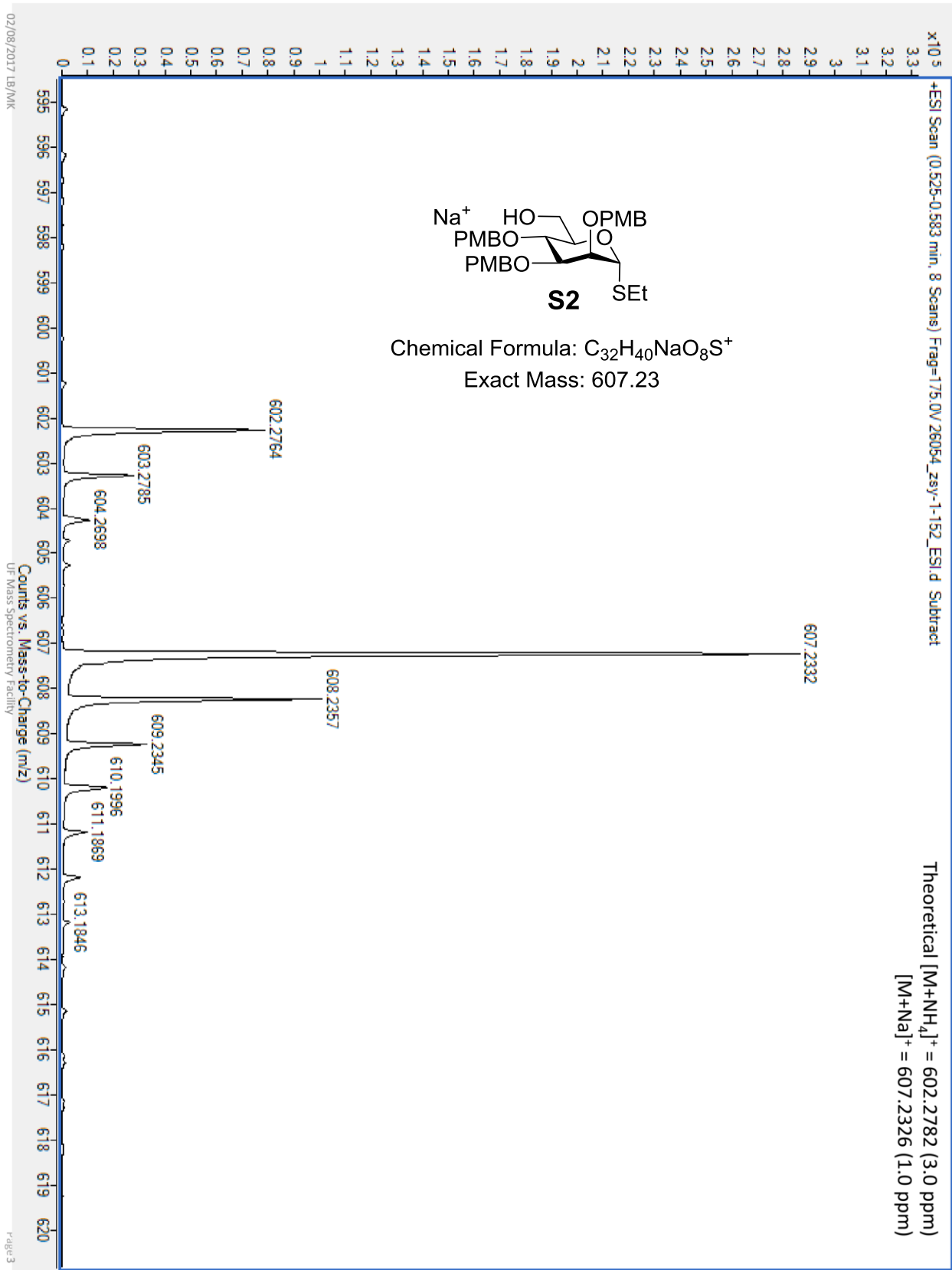
Solvent: cdcl3

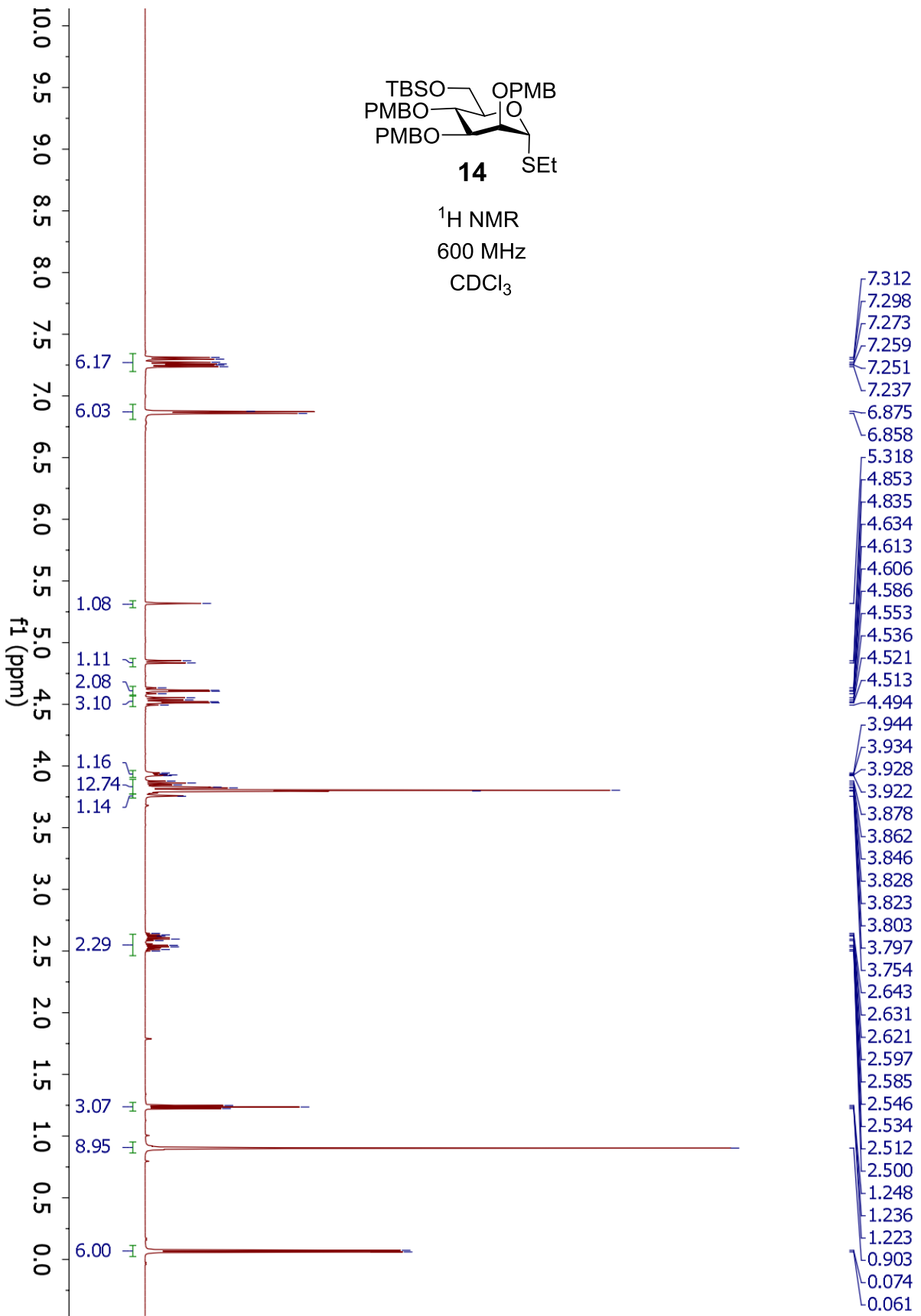
Data collected on: Nov 30 2015

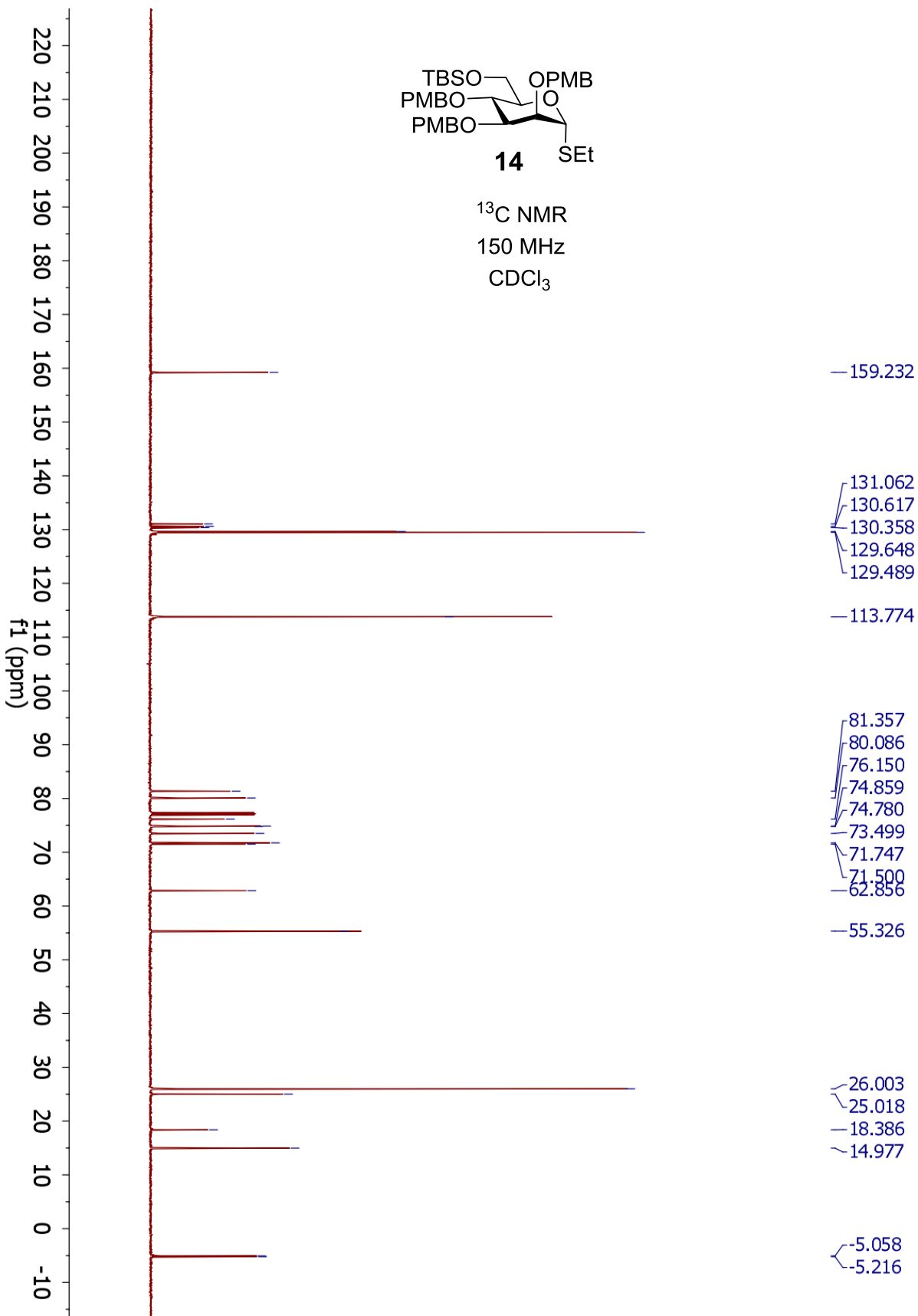


$^1\text{H}-^1\text{H}$ COSY
600 MHz
 CDCl_3









Sample Name:

Data Collected on:

a600-nmr600

Archive directory:

/home/vmari/vmrzsys/probes/probe_calibs

Sample directory:

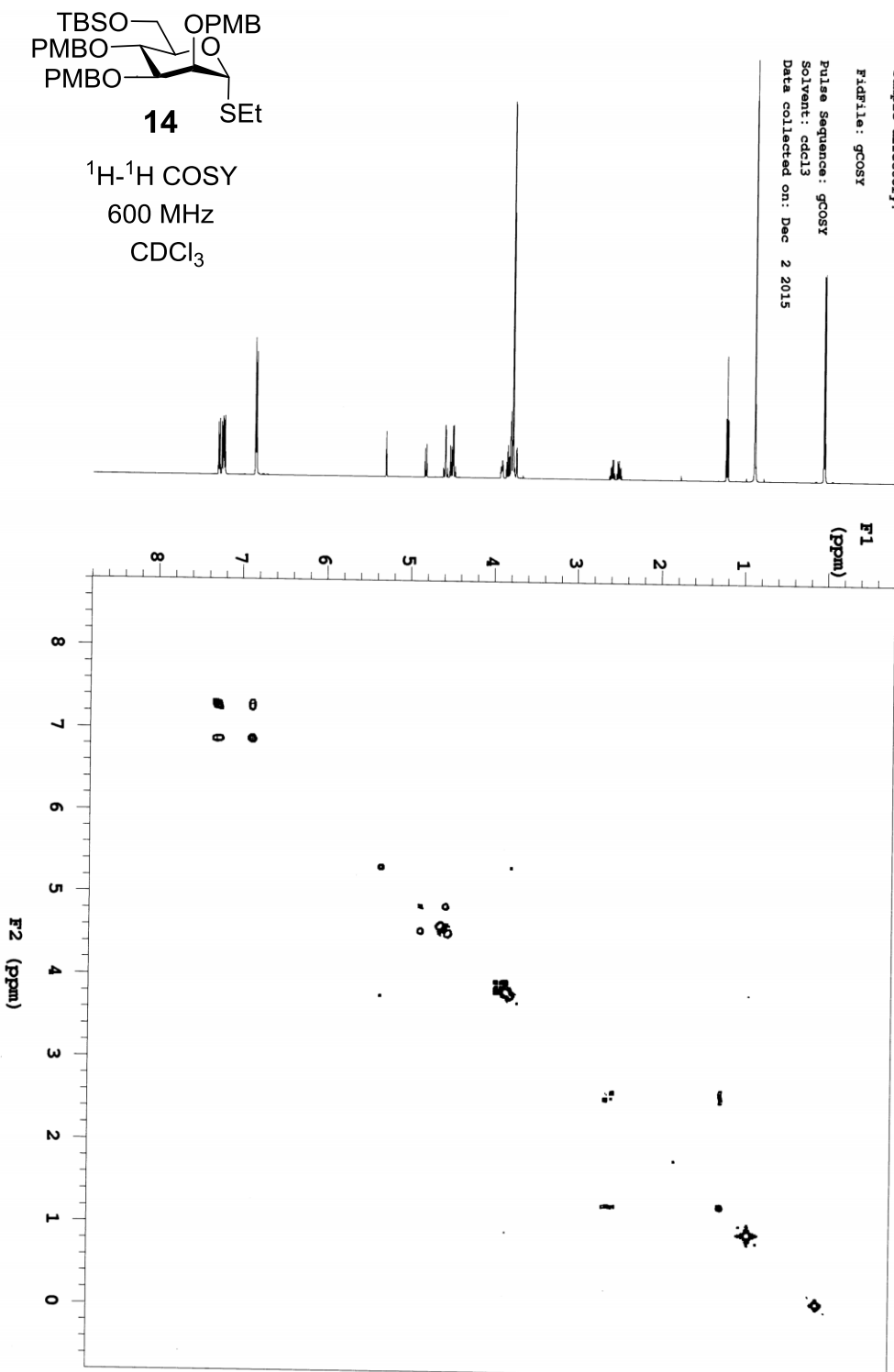
Fidfile: gCOSY

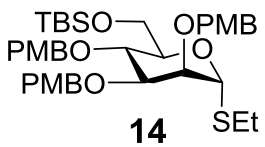
Pulse Sequence: gCOSY

Solvent: cdcl3

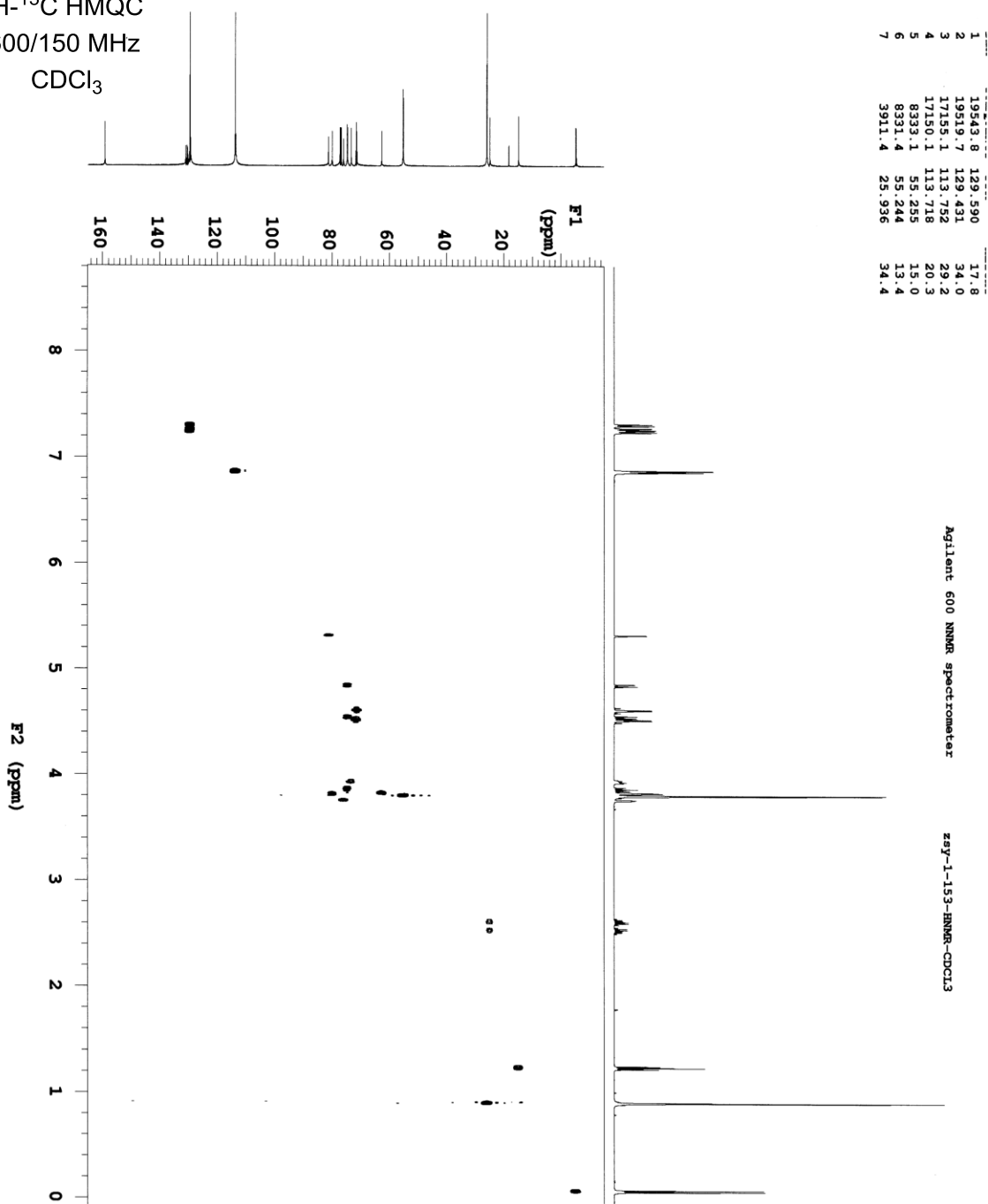
Data collected on: Dec 2 2015

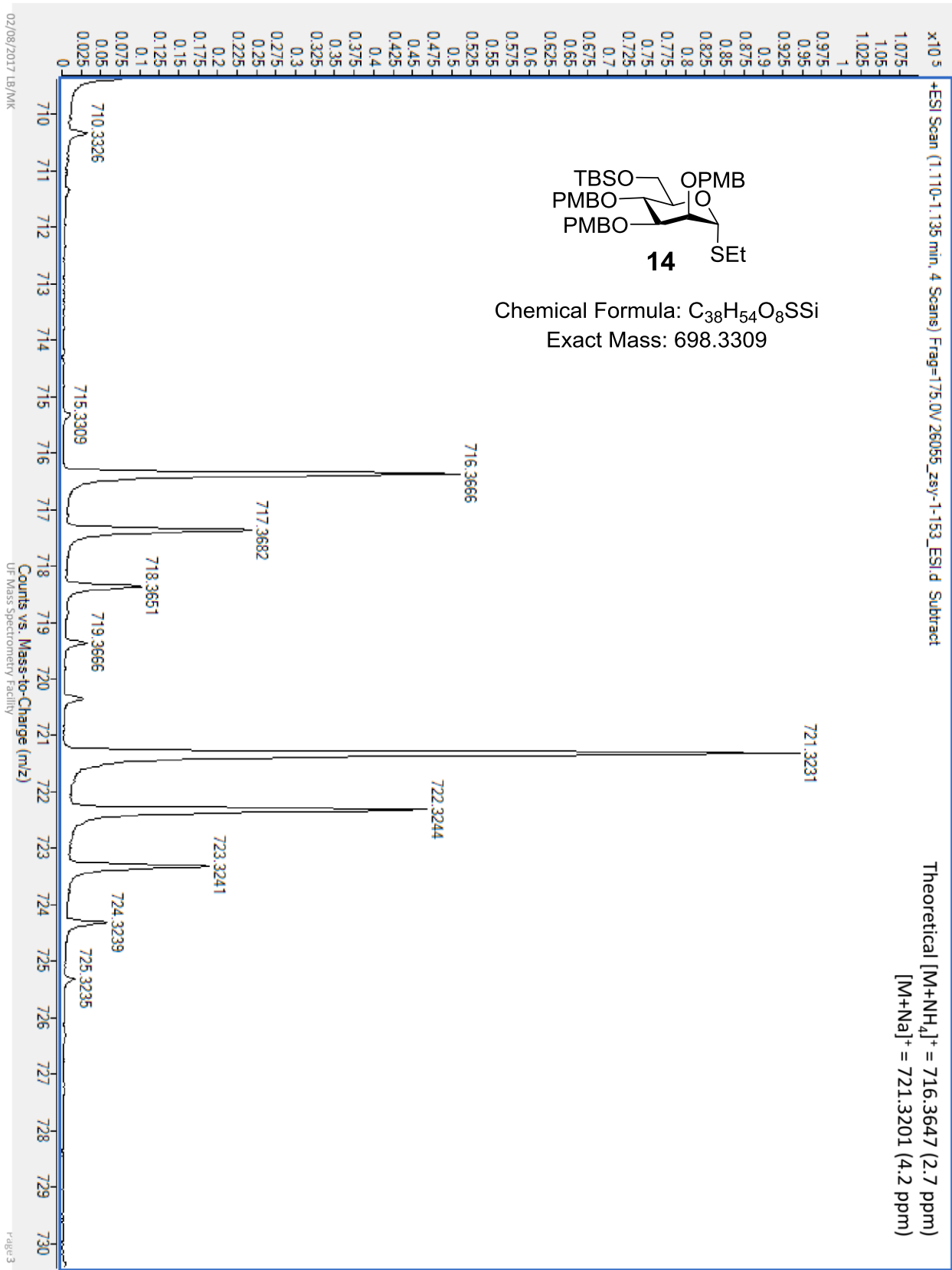
Agilent 600 NMR spectrometer

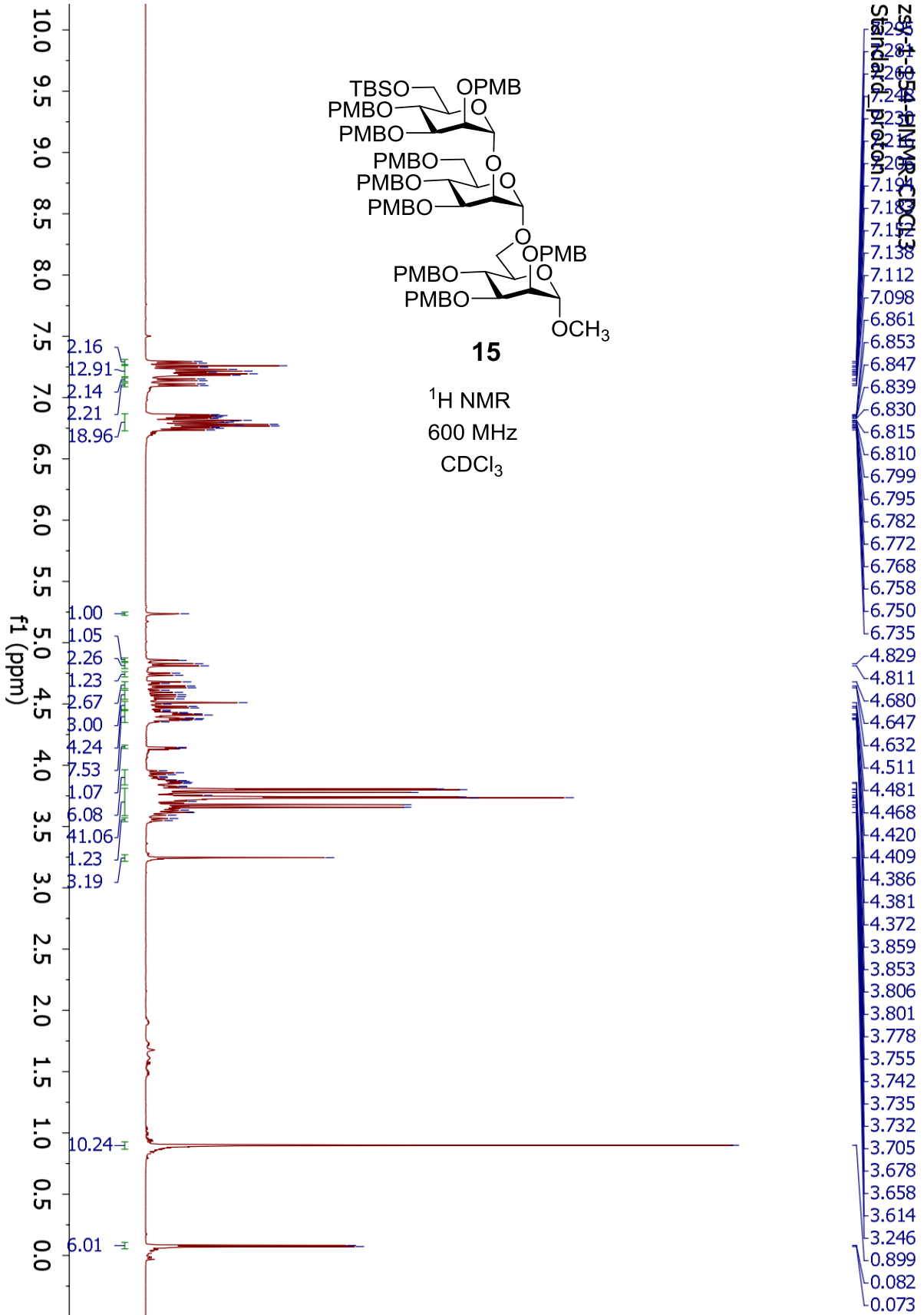


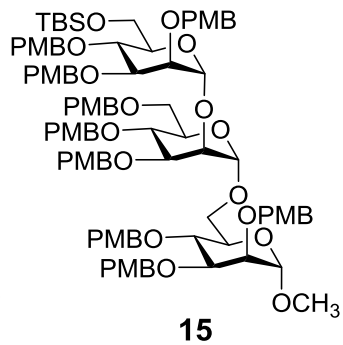


¹H-¹³C HMQC
 600/150 MHz
 CDCl₃



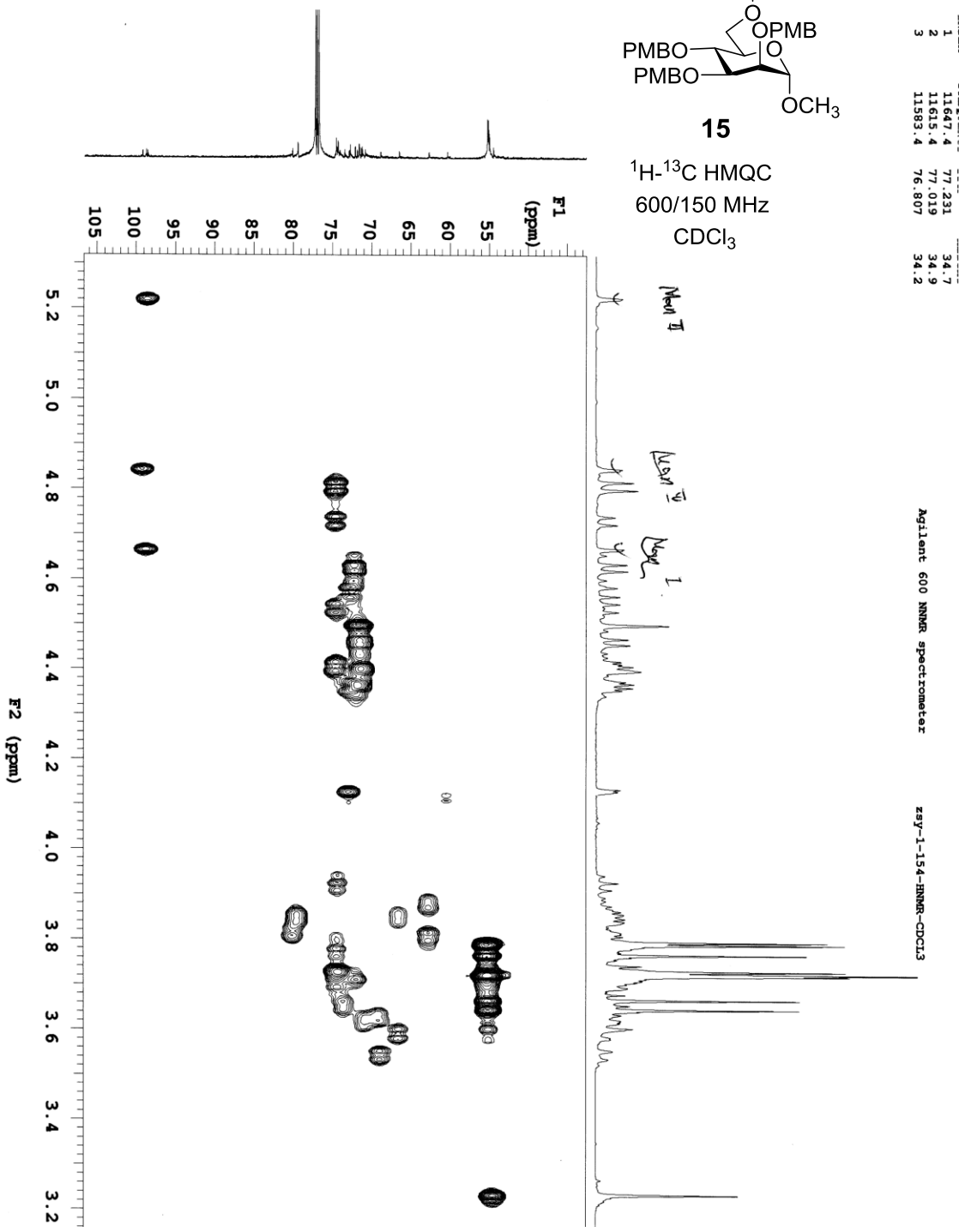






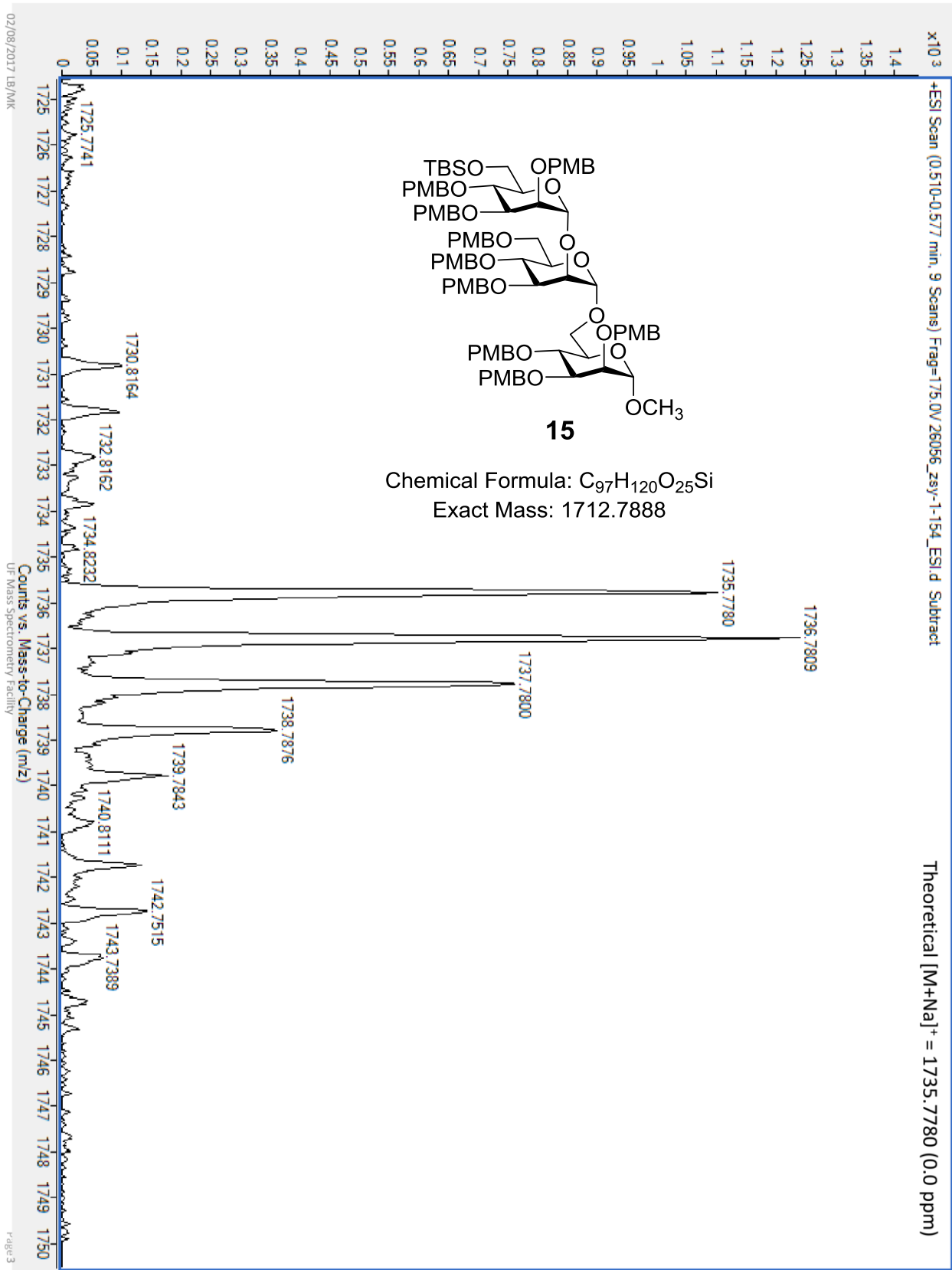
^1H - ^{13}C HMQC
600/150 MHz
 CDCl_3

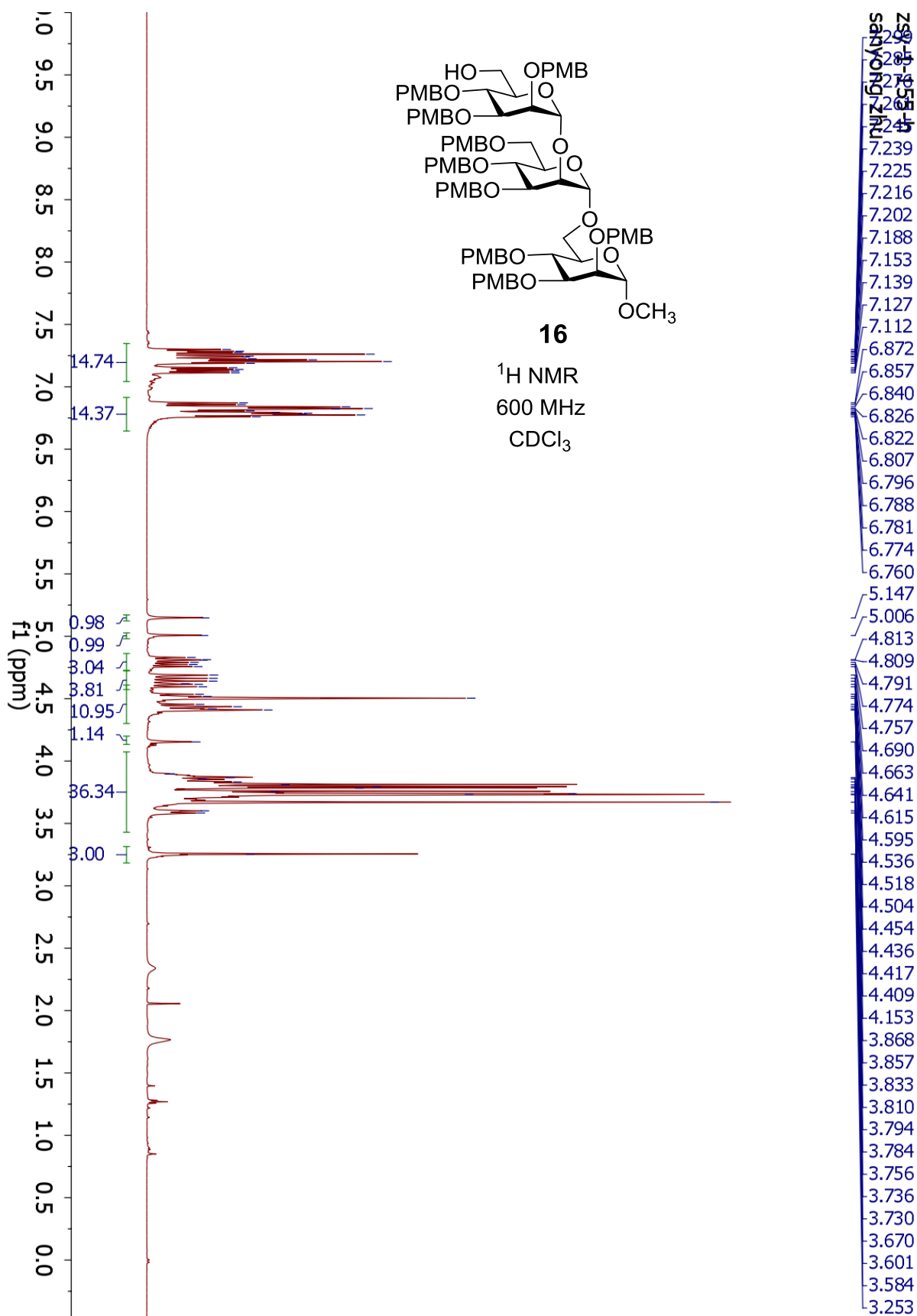
1	11647.4	77.231	34.7
2	11615.4	77.019	34.9
3	11583.4	76.807	34.2

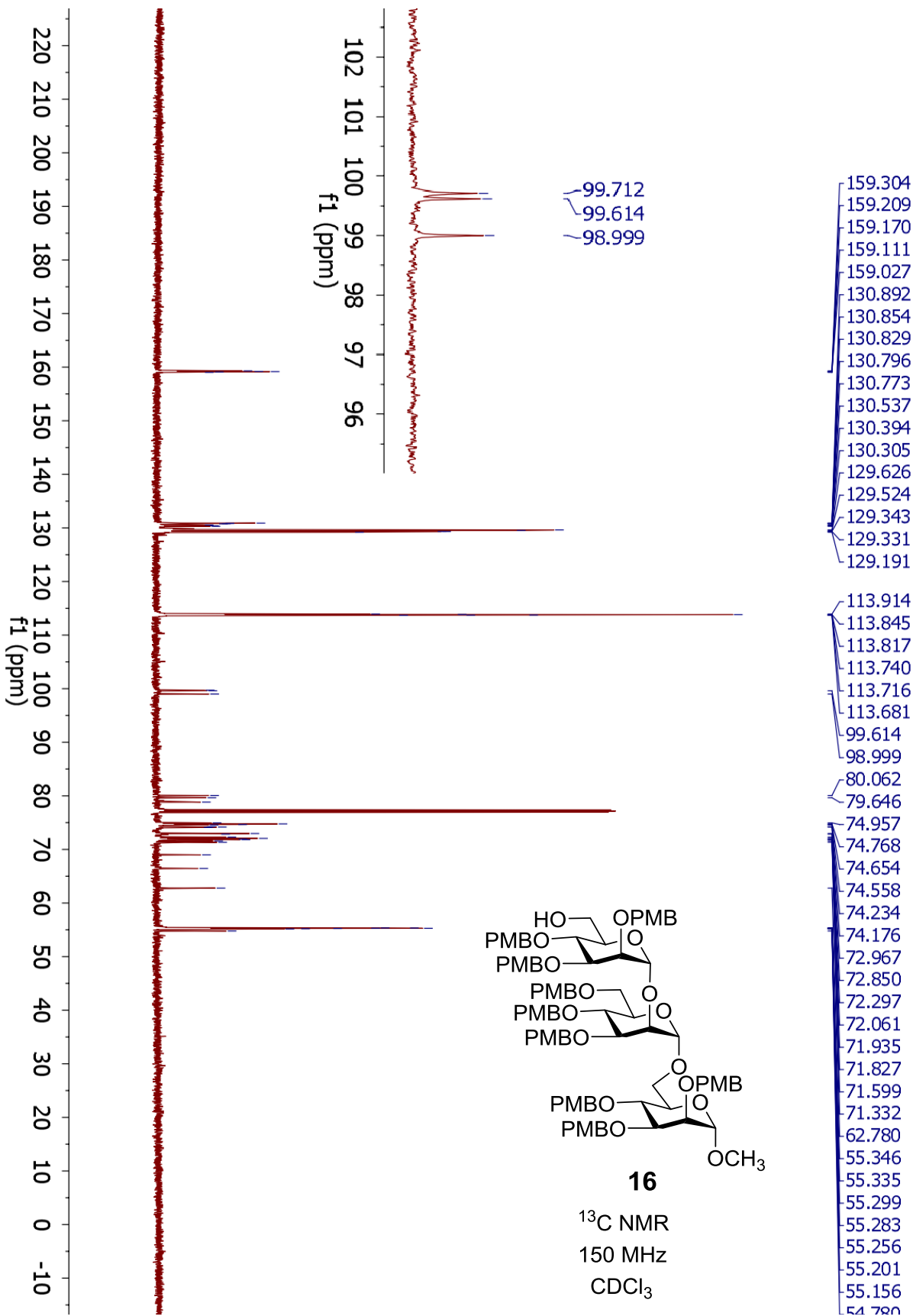


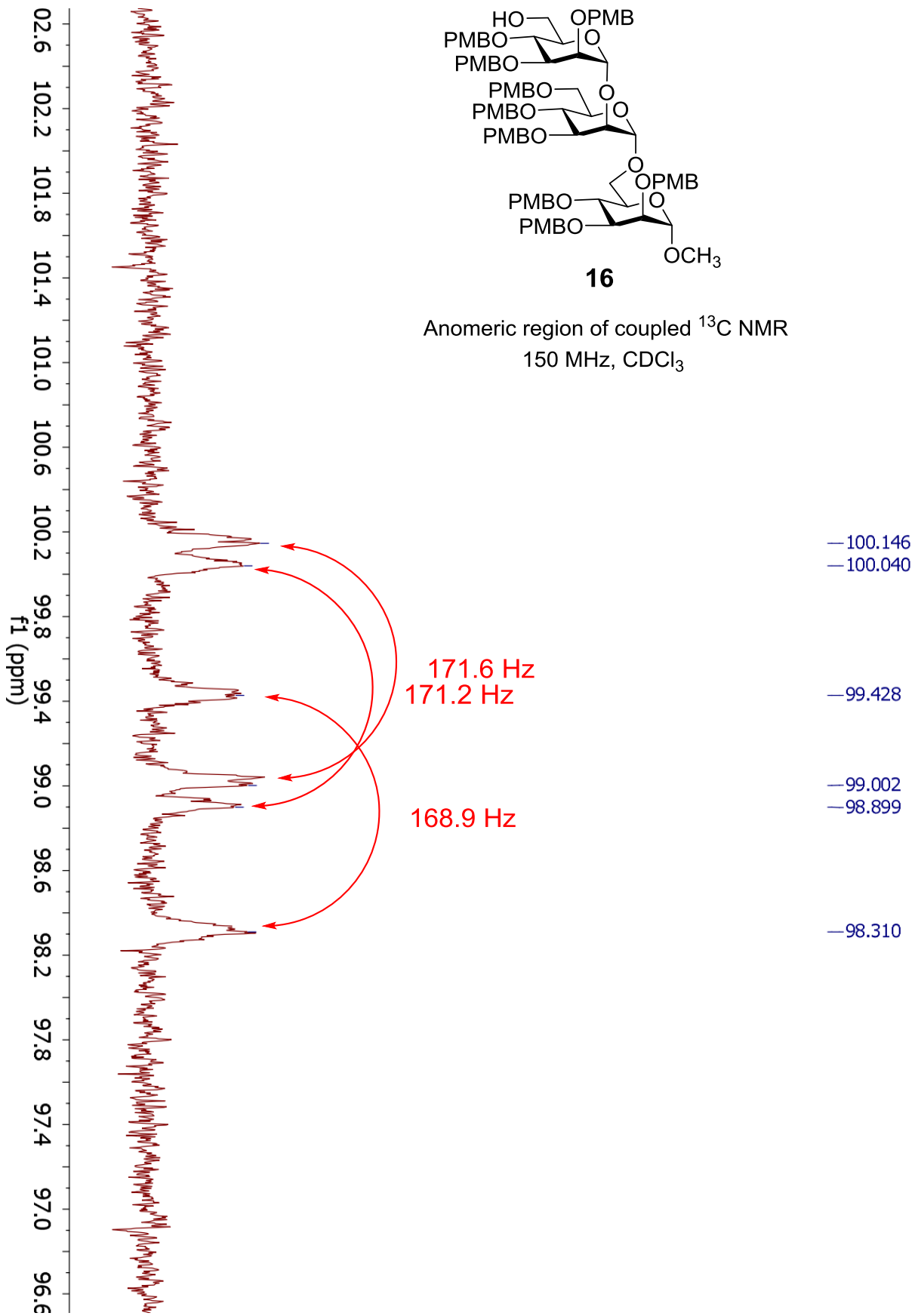
Agilent 600 NMR spectrometer

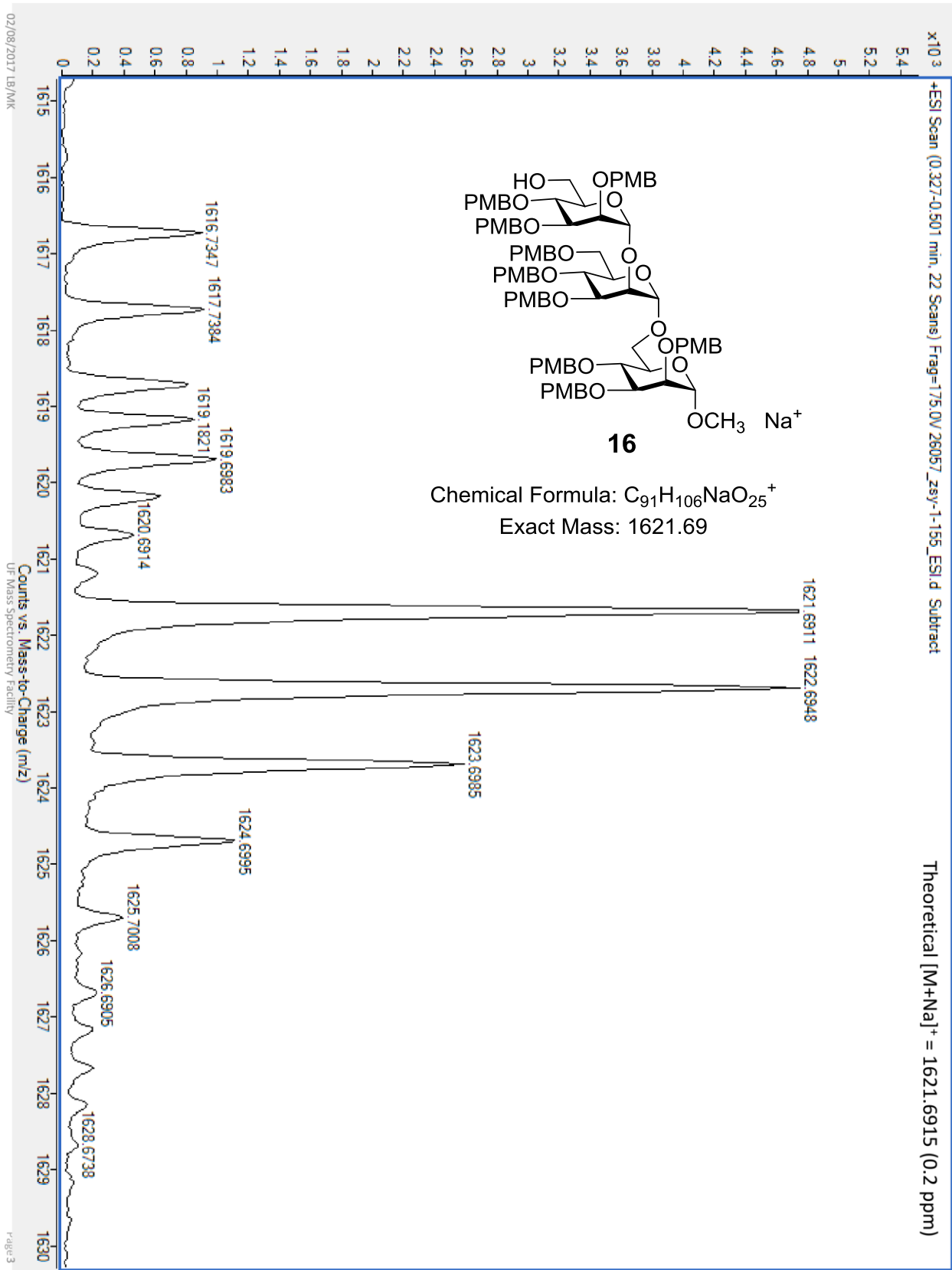
zsy-1-154-NMR-CDCl3

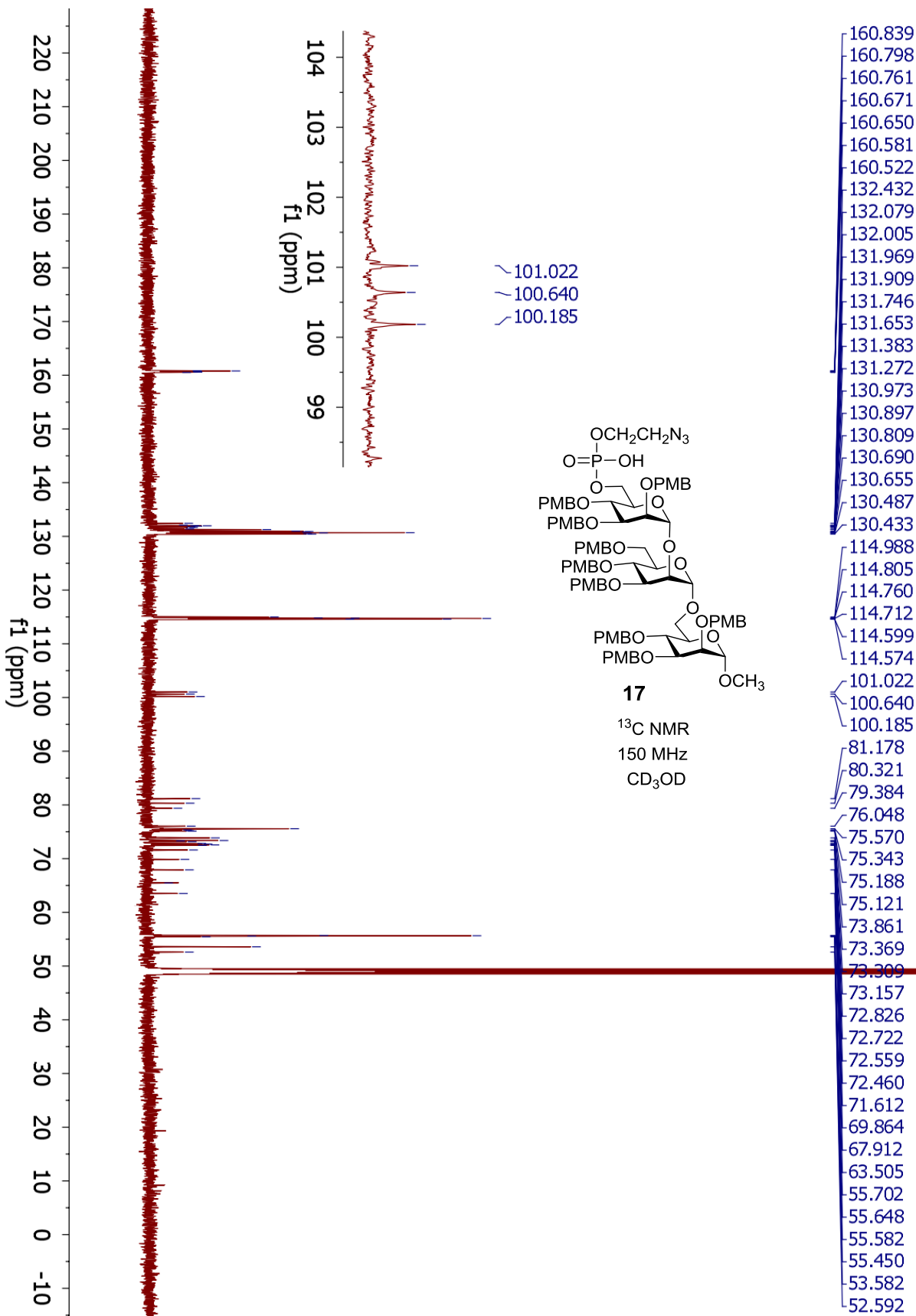


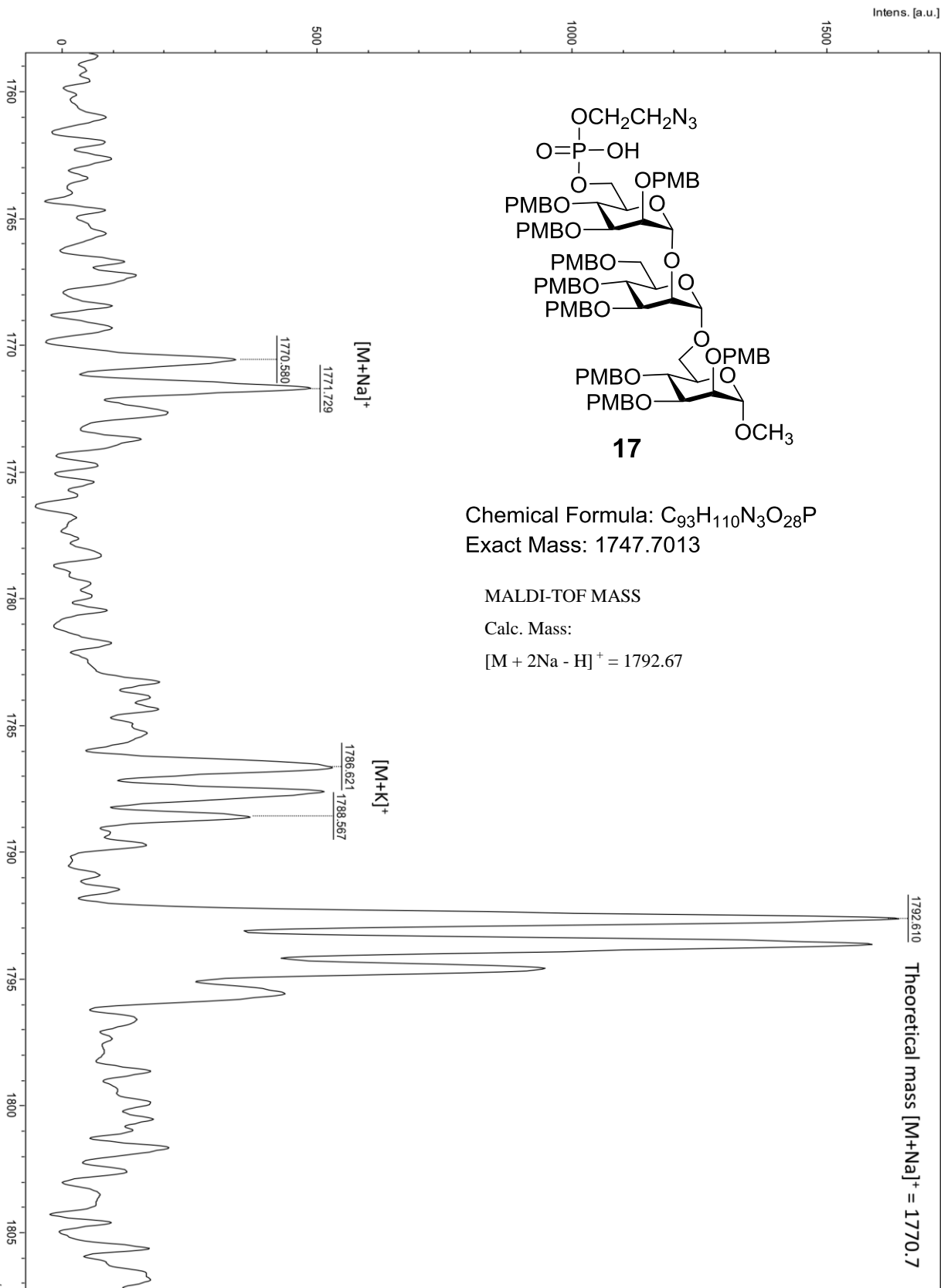


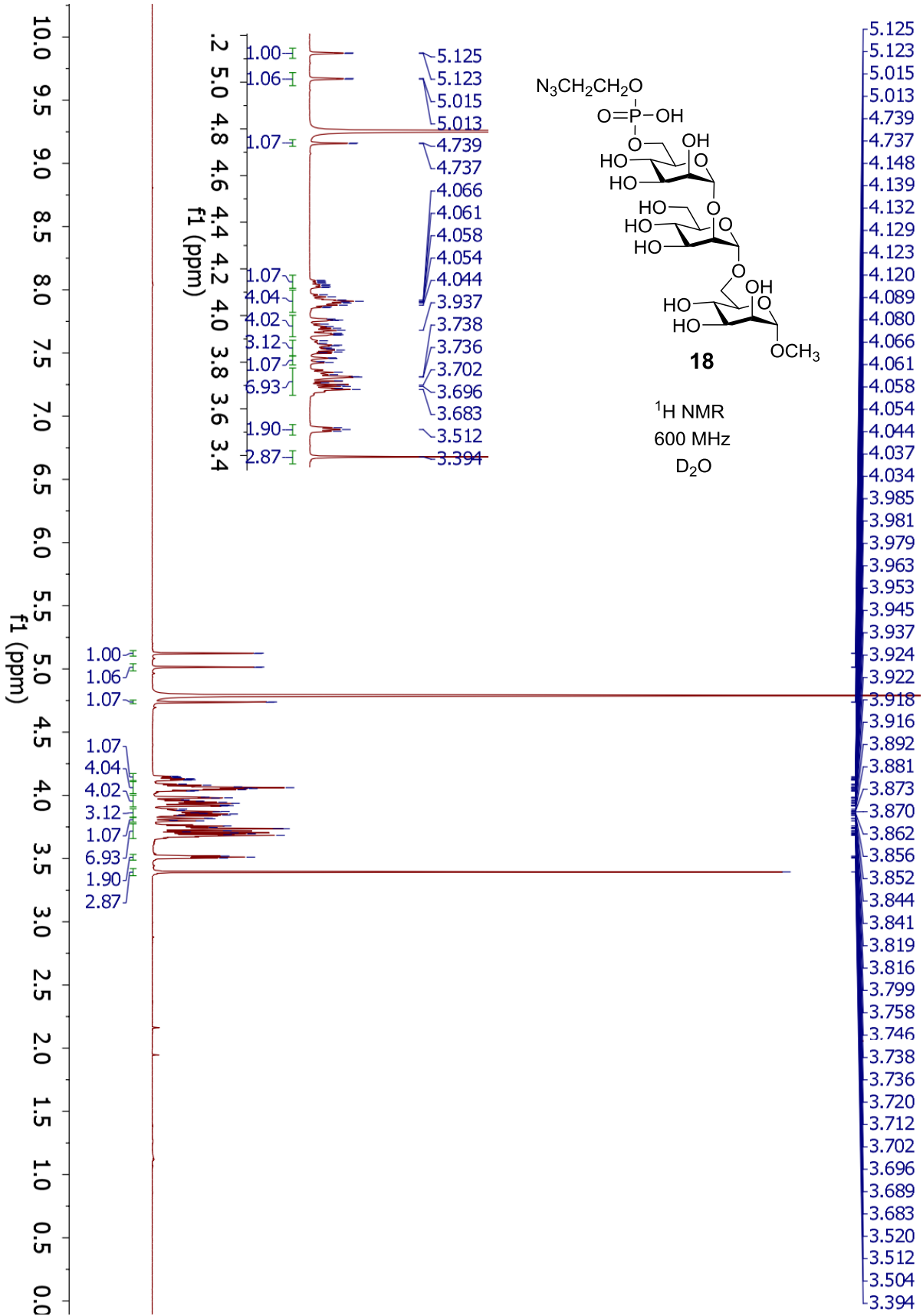


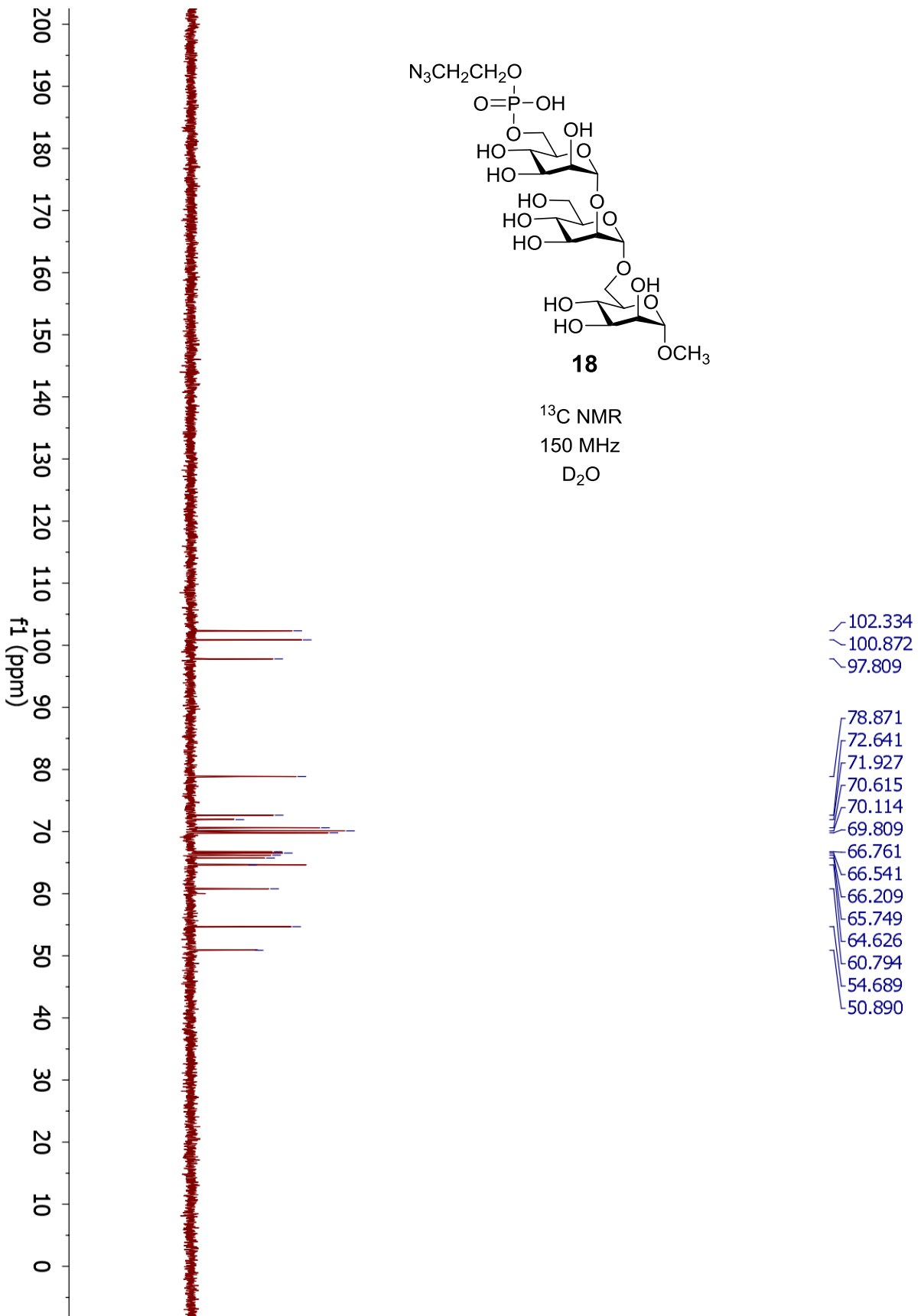


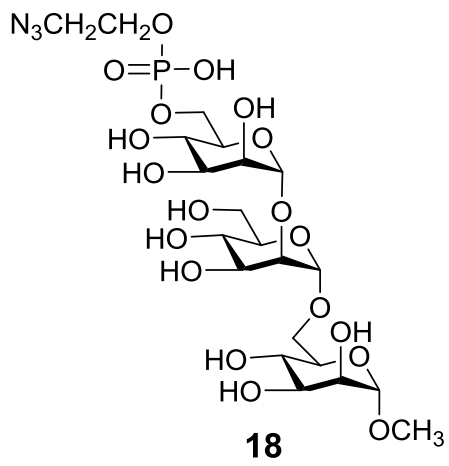
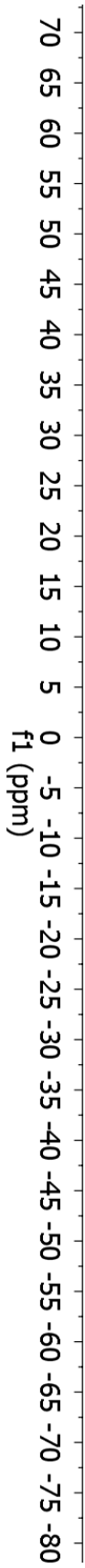










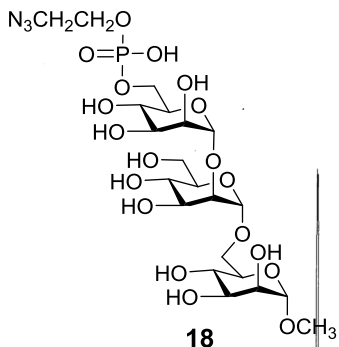


³¹P NMR
160 MHz
D₂O

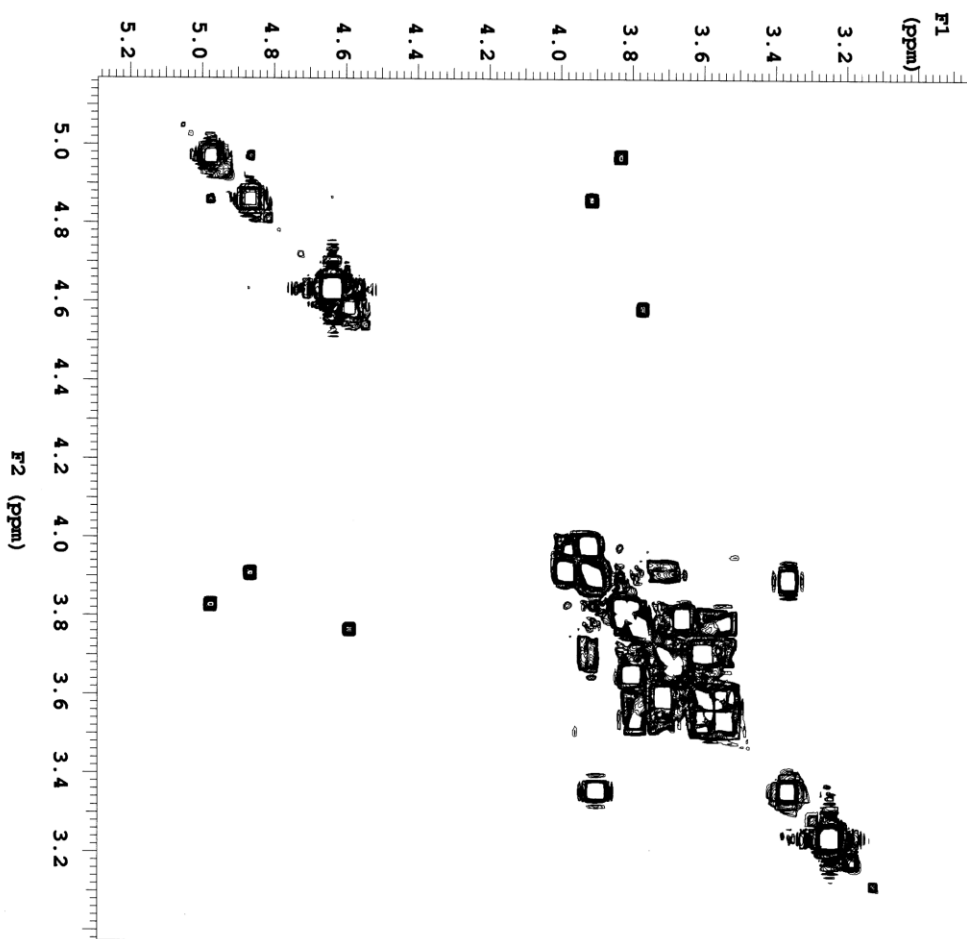
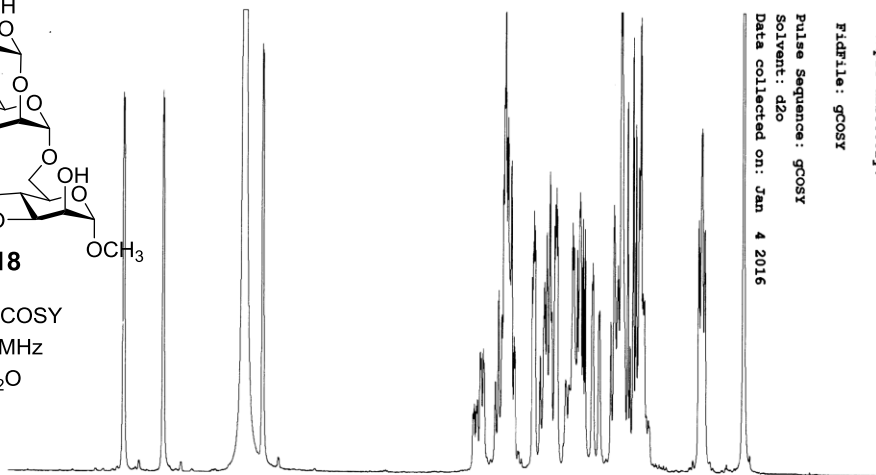
-0.300

Sample Name:
 Data collected on: a600-ymms600
 Archive directory: /home/ymml/ymmsys/probes/probe_caliba
 Sample directory:

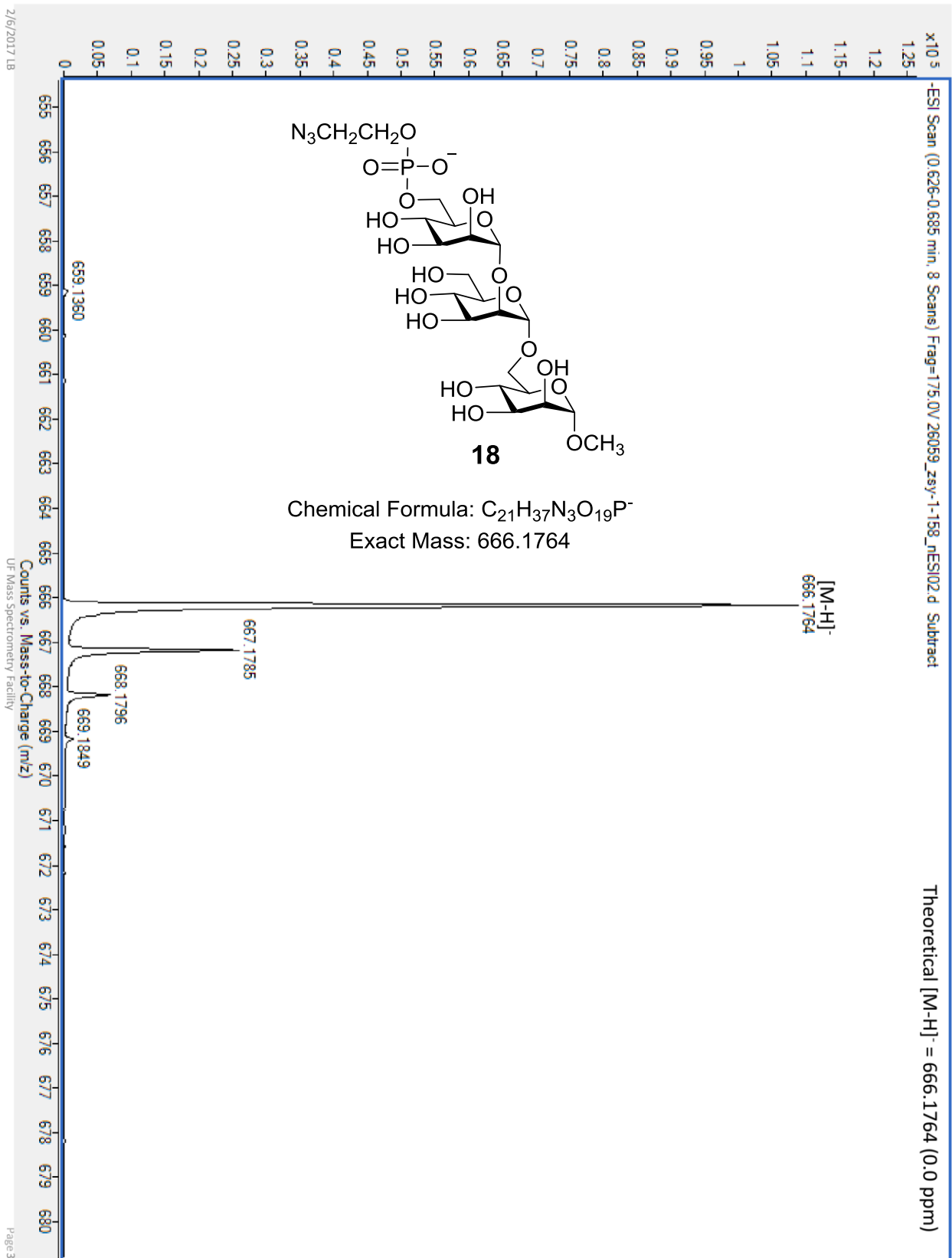
Fidfile: gcosy
 Pulse sequence: gcosy
 Solvent: d2o
 Data collected on: Jan 4 2016

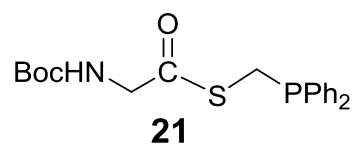


¹H-¹H COSY
 600 MHz
 D₂O

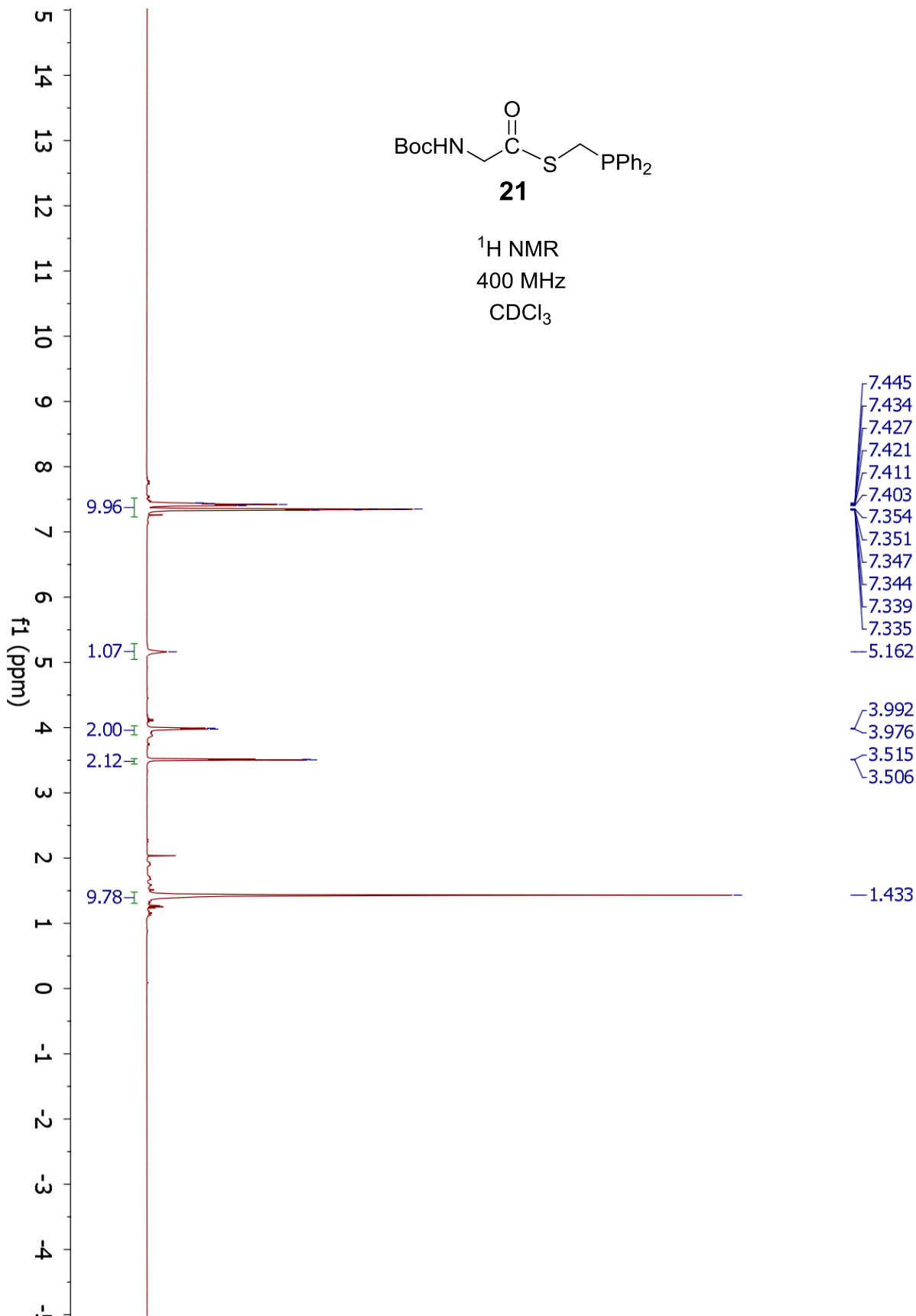


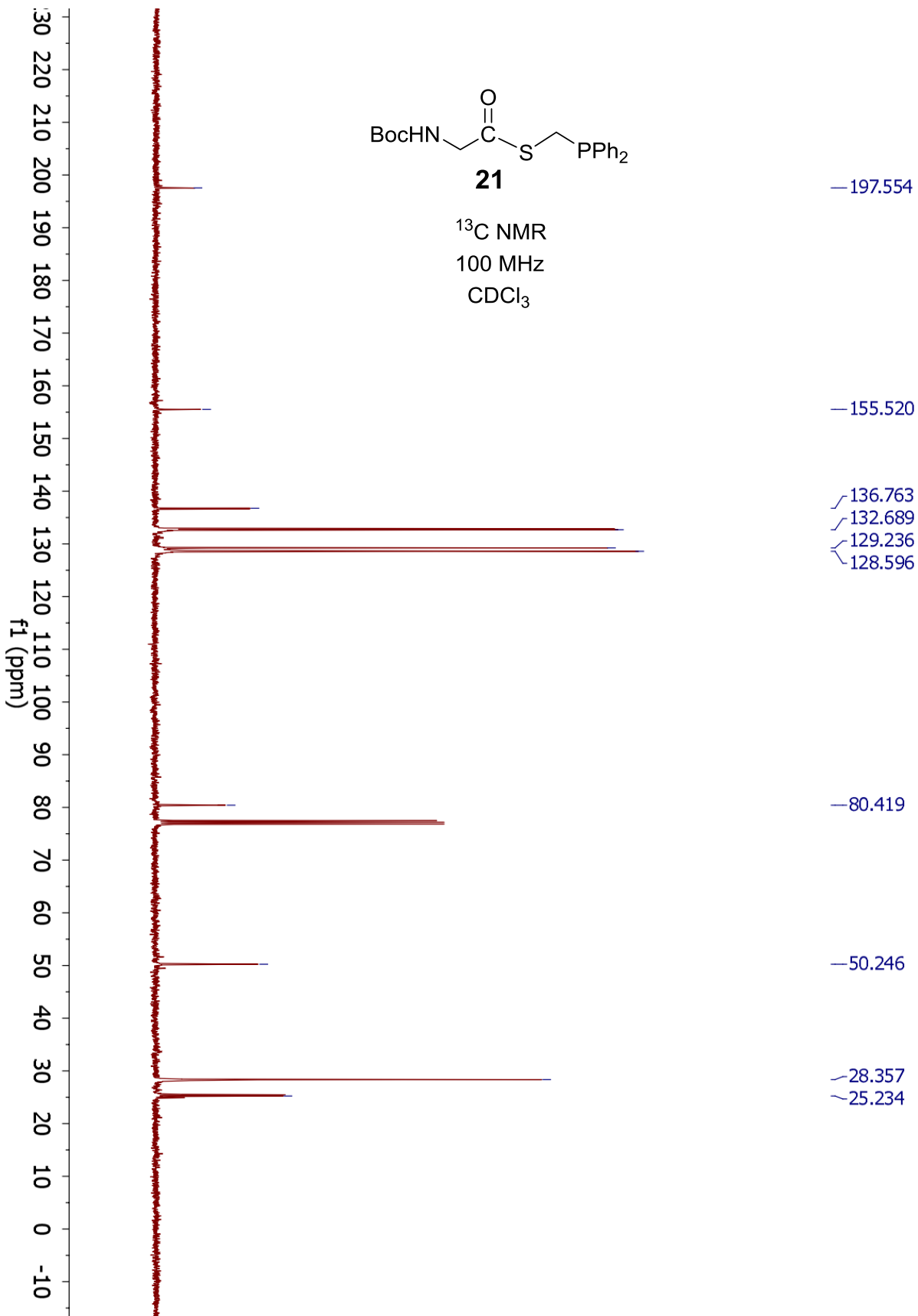
Agilent 600 NMR spectrometer
 H₂O C-1 A-1
 CH₂N₃
 CH₂N₃
 Agilent Technologies

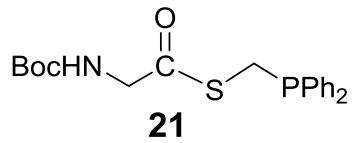




¹H NMR
400 MHz
CDCl₃

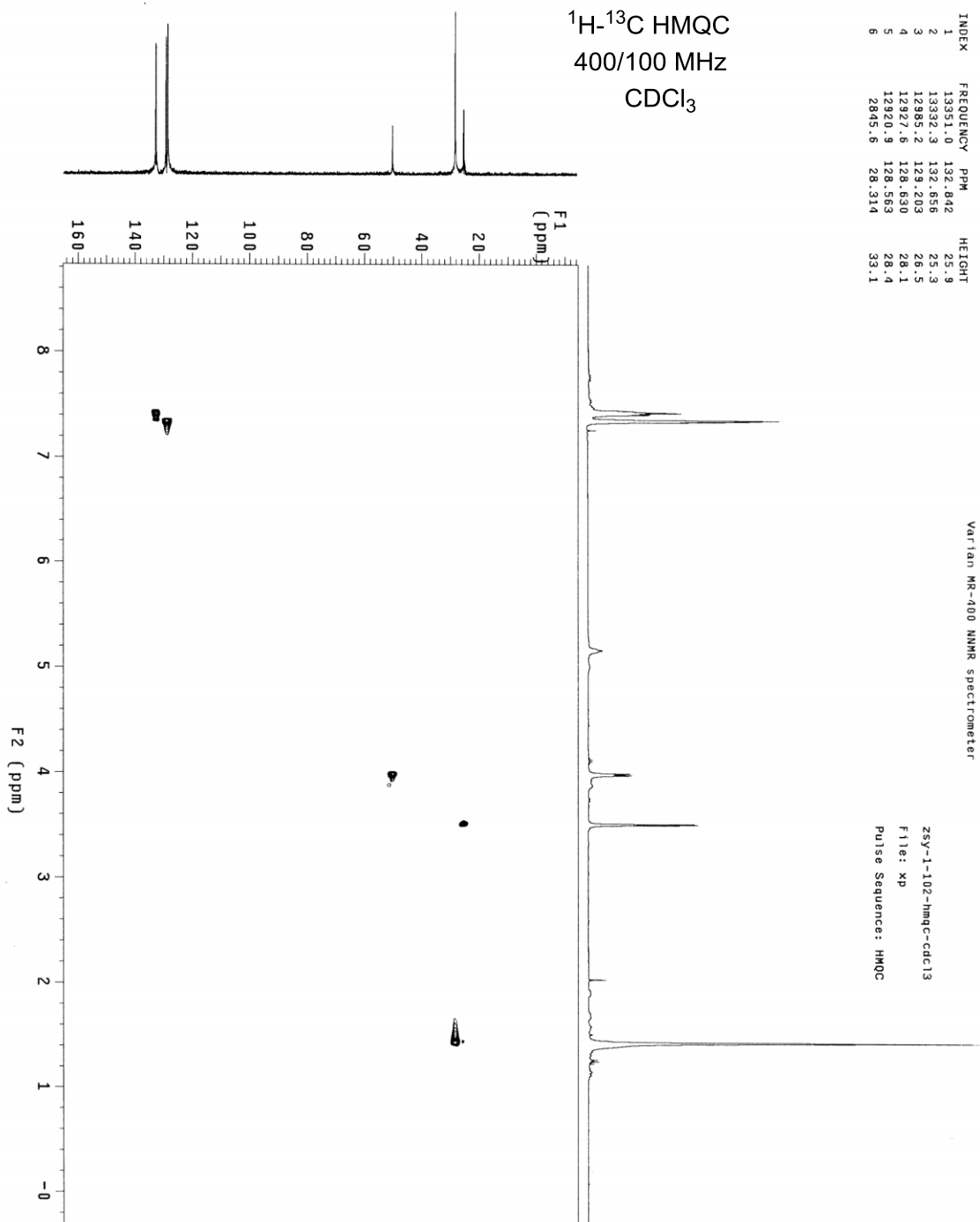


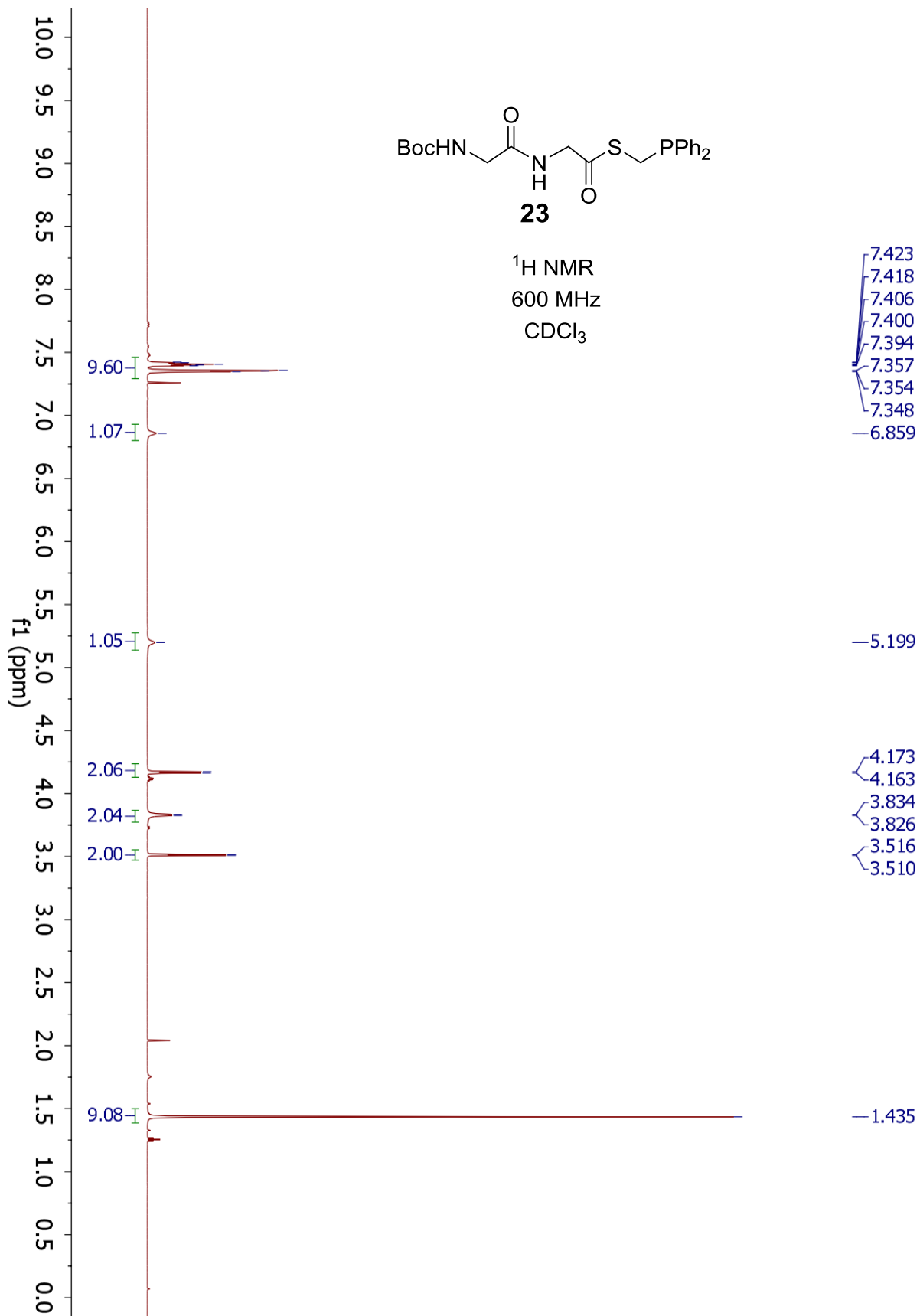


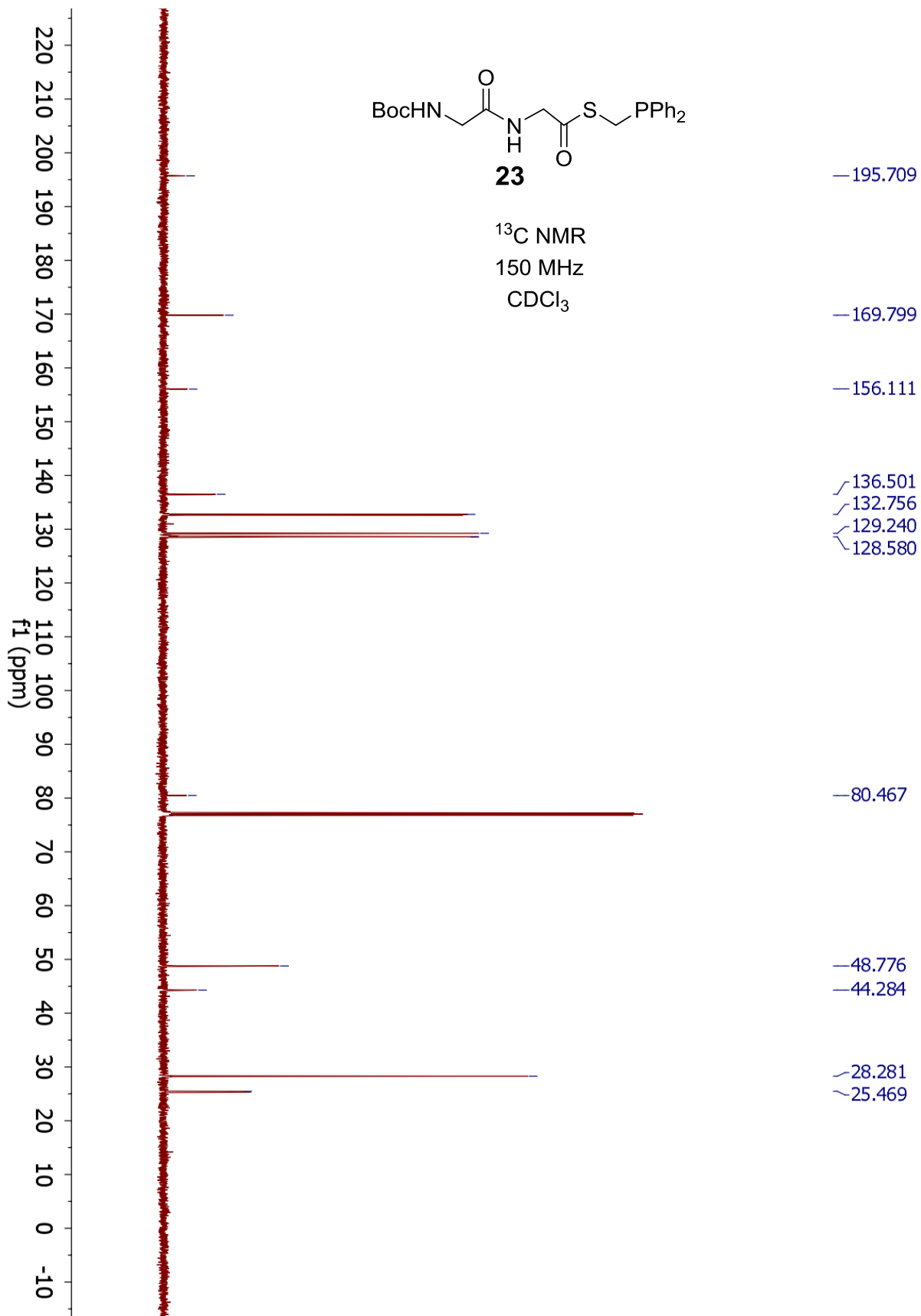


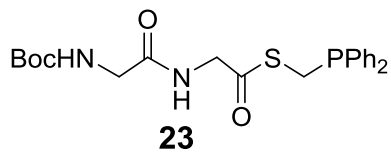
¹H-¹³C HMQC
400/100 MHz
CDCl₃

INDEX	FREQUENCY PPM	HEIGHT
1	13951.0	25.9
2	13332.3	25.3
3	12985.2	26.5
4	12927.6	28.1
5	12920.9	28.4
6	2845.6	33.1



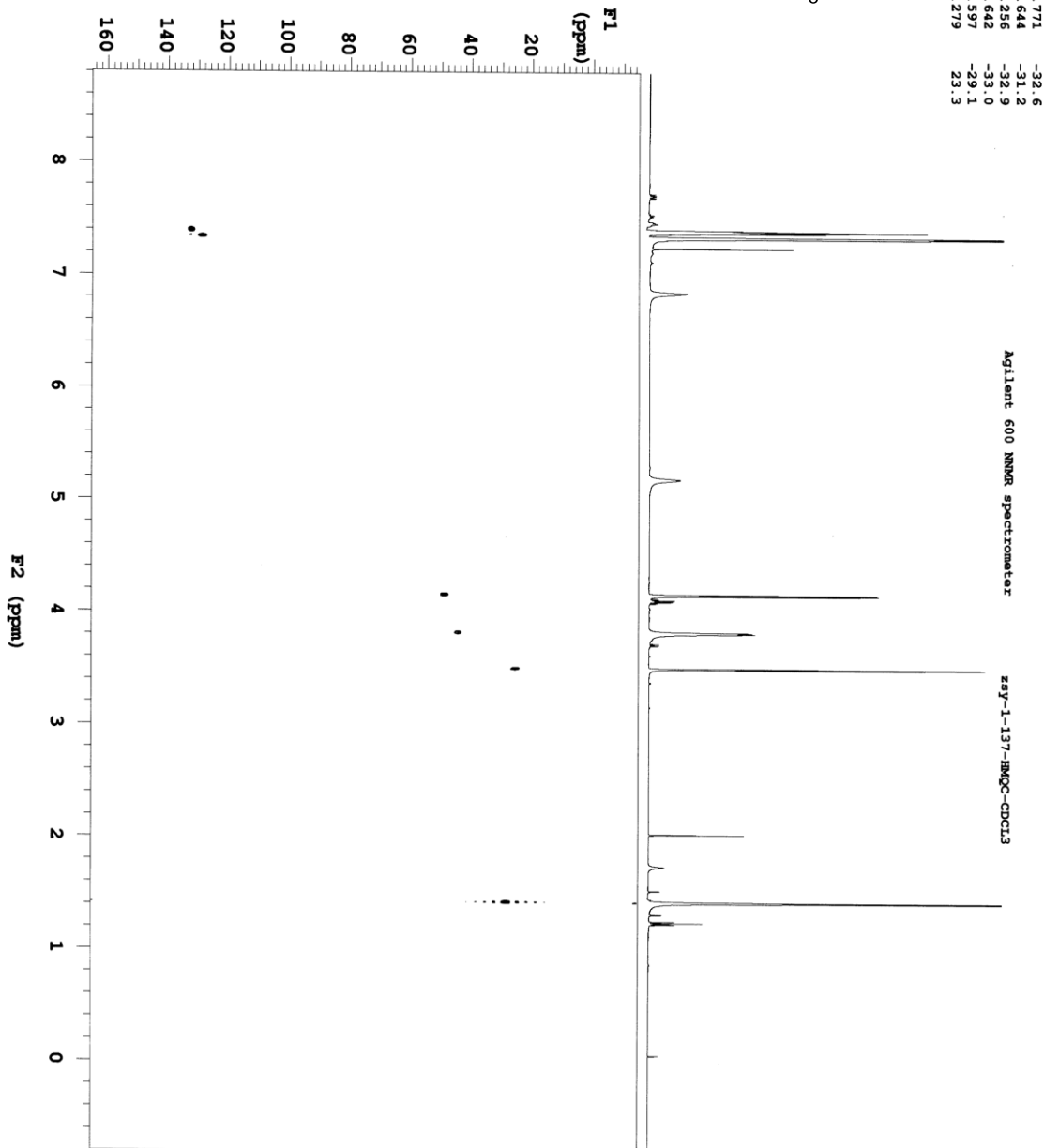


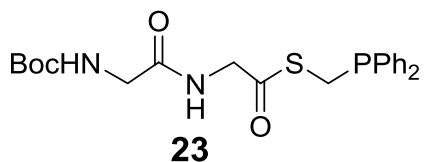




^1H - ^{13}C HMQC
600/150 MHz
 CDCl_3

1	20023.4	132.771	-32.6
2	20004.4	132.644	-31.2
3	19493.3	129.256	-32.9
4	19400.7	128.642	-33.0
5	19394.0	128.597	-29.1
6	4264.8	28.279	23.3





Chemical Formula: C₂₂H₂₇N₂O₄PS
 Exact Mass: 446.1429

HR ESI-TOF MS

Calc. Mass:

[M + H]⁺ = 447.1507

[M + Na]⁺ = 469.1327

[M + K]⁺ = 485.1066

Elemental Composition Report

Single Mass Analysis

Tolerance = 10.0 PPM / DBE: min = -1.5, max = 100.0

Element prediction: Off

Number of isotope peaks used for i-FIT = 6

Monoisotopic Mass, Even Electron Ions

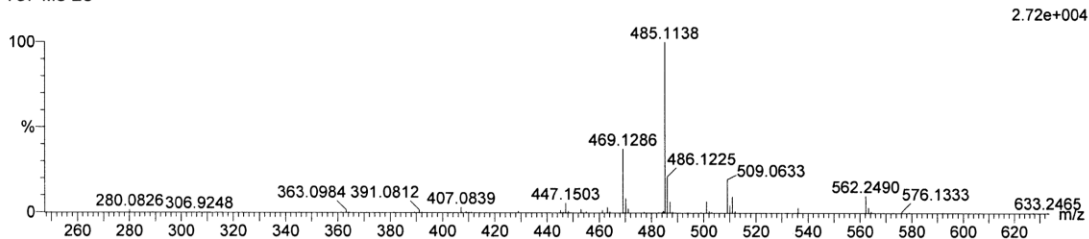
934 formula(e) evaluated with 4 results within limits (all results (up to 1000) for each mass)

Elements Used:

C: 22-22 H: 0-40 N: 0-5 O: 0-10 Na: 0-1 31P: 0-1 S: 0-1 39K: 0-1

S.Zhu ZSY-1-137 in AcN Cone(V)50

LCT Premier KD128
 TOF MS ES+

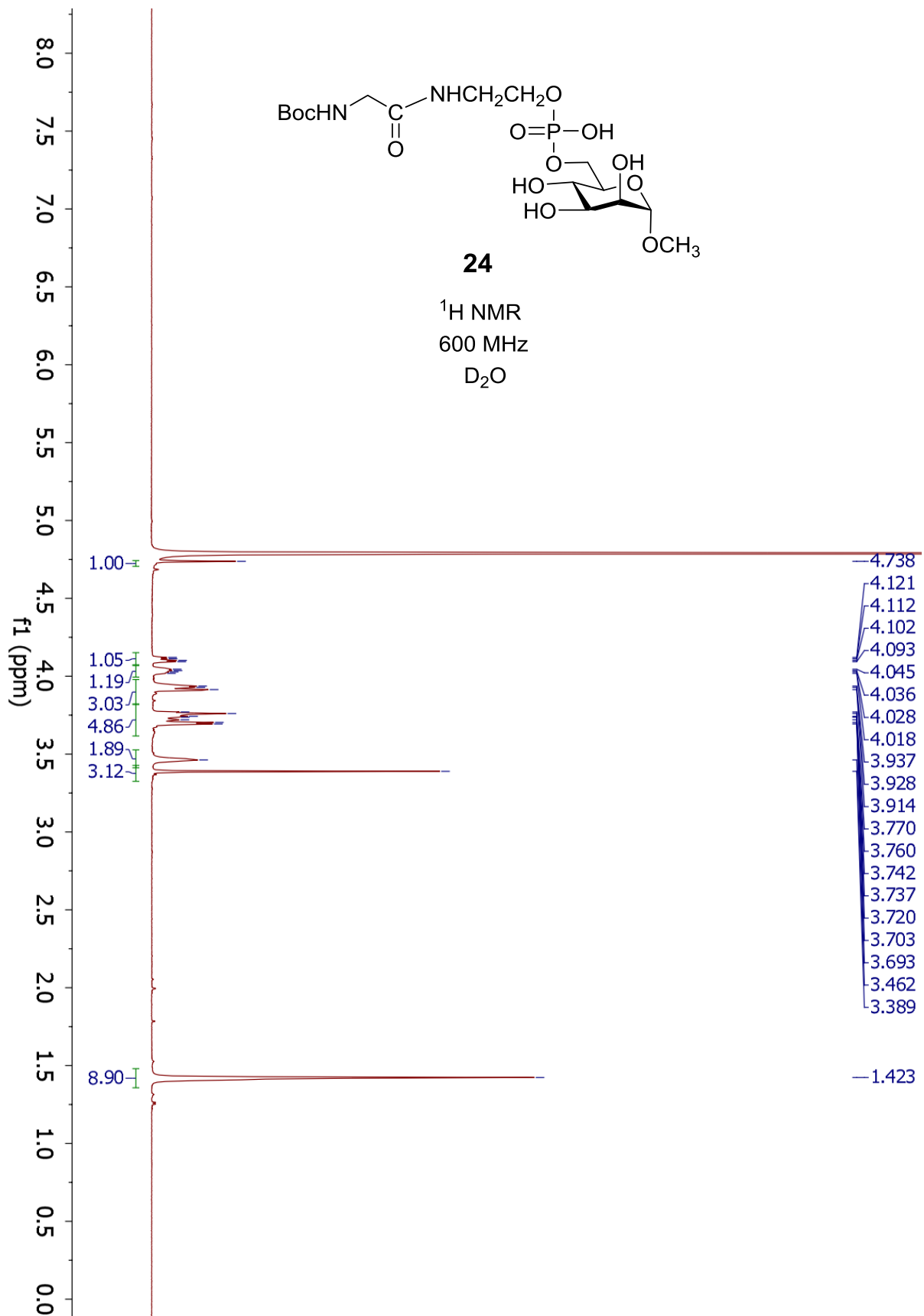


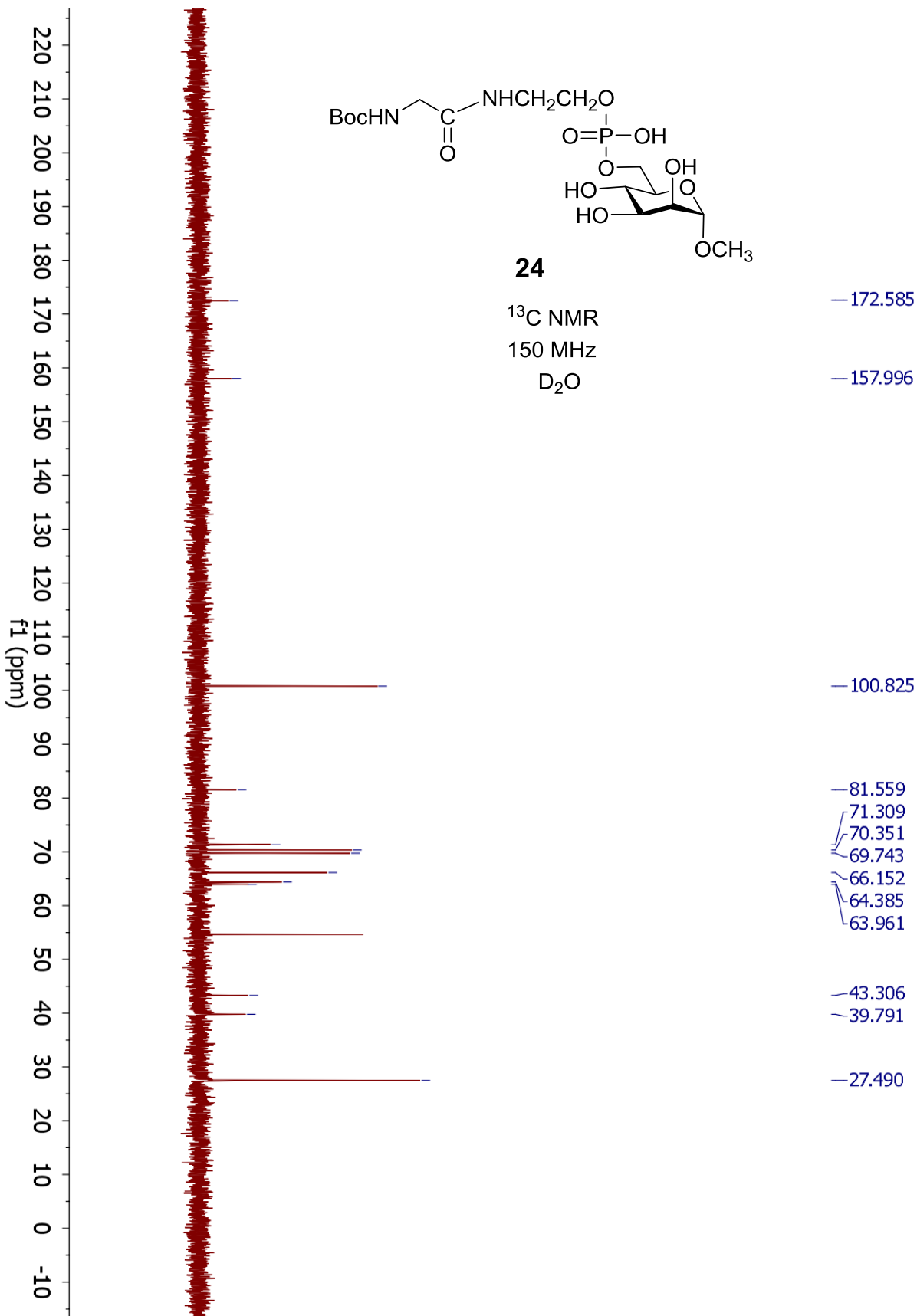
Minimum:

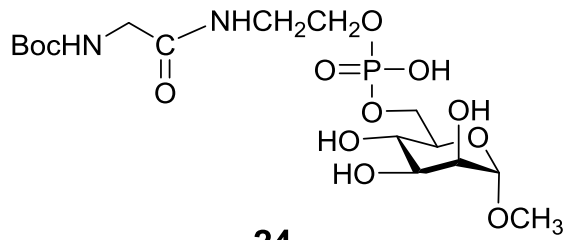
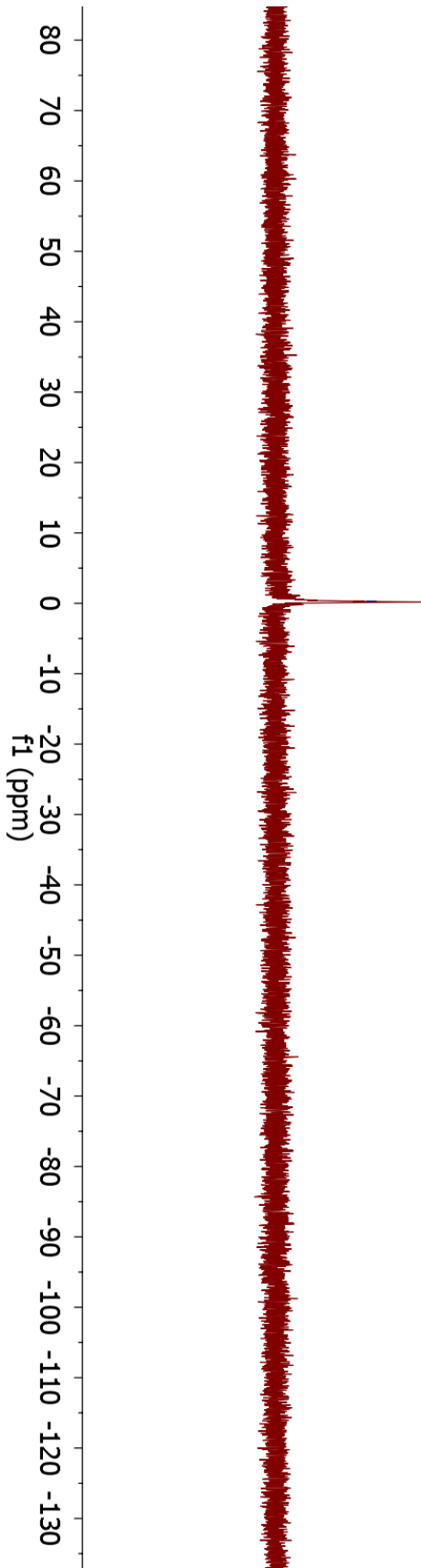
Maximum: 3.0 10.0 -1.5

Mass Calc. Mass mDa PPM DBE i-FIT i-FIT (Norm) Formula

447.1503	447.1507	-0.4	-0.9	10.5	39.9	1.4	C22 H28 N2 O4 31P S
447.1485	1.8	4.0	8.5	39.8	1.3	C22 H29 N2 O2 Na S 39K	
447.1525	-2.2	-4.9	6.5	40.0	1.5	C22 H33 O3 31P S 39K	
447.1467	3.6	8.1	12.5	39.8	1.3	C22 H24 N4 O3 Na S	







24

³¹P NMR
160 MHz
D₂O

-0.287

Sample Name:

Data collected on:

a600-nmr600

Archive directory:

/home/vmr1/vmr5ys/probes/probe_calibs

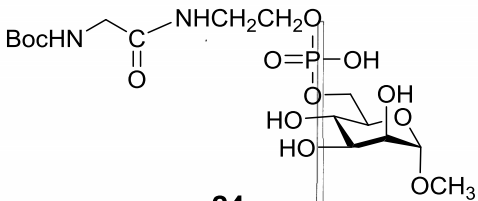
Sample directory:

Fidfile: gcosy

Pulse Sequence: gcosy

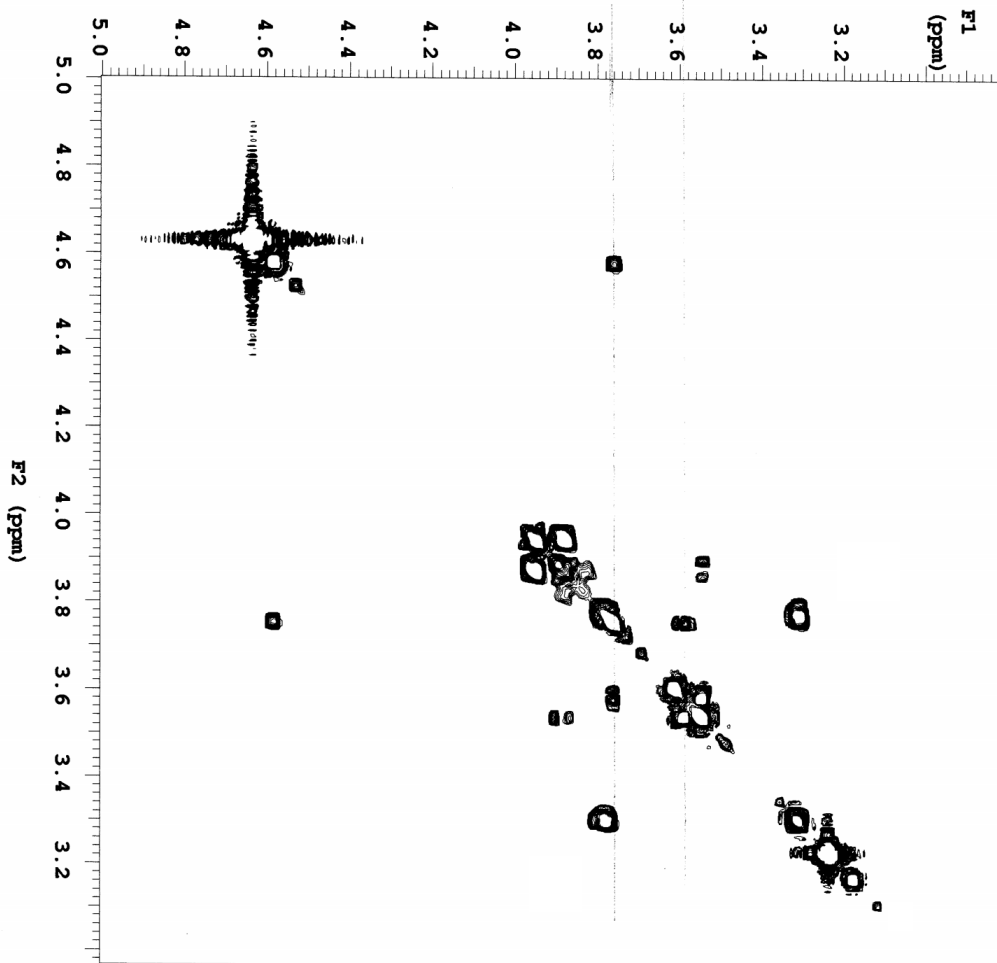
Solvent: d2o

Data collected on: Oct 21 2015



24

¹H-¹H COSY
600 MHz
D₂O



Elemental Composition Report

Single Mass Analysis

Tolerance = 5.0 PPM / DBE: min = -1.5, max = 100.0

Element prediction: Off

Number of isotope peaks used for i-FIT = 6

Monoisotopic Mass, Even Electron Ions

238 formula(e) evaluated with 1 results within limits (all results (up to 1000) for each mass)

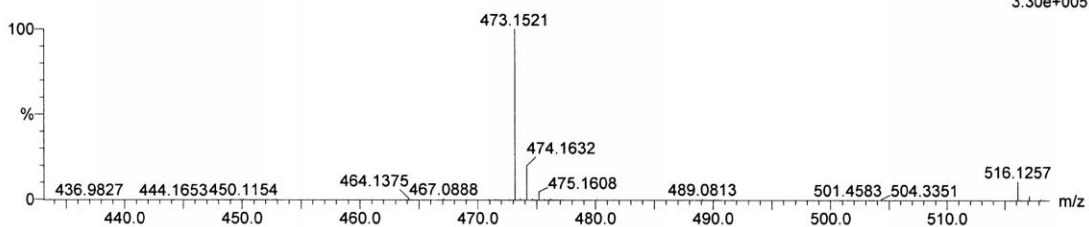
Elements Used:

C: 16-16 H: 0-35 N: 0-5 O: 0-20 31P: 0-1

S.Zhu ZSY-1-136 inMeOH Cone(V)50

LCT Premier KD128

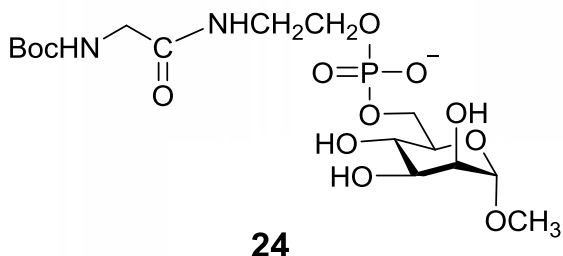
TOF MS ES-



Minimum:

Maximum: 3.0 5.0 -1.5 100.0

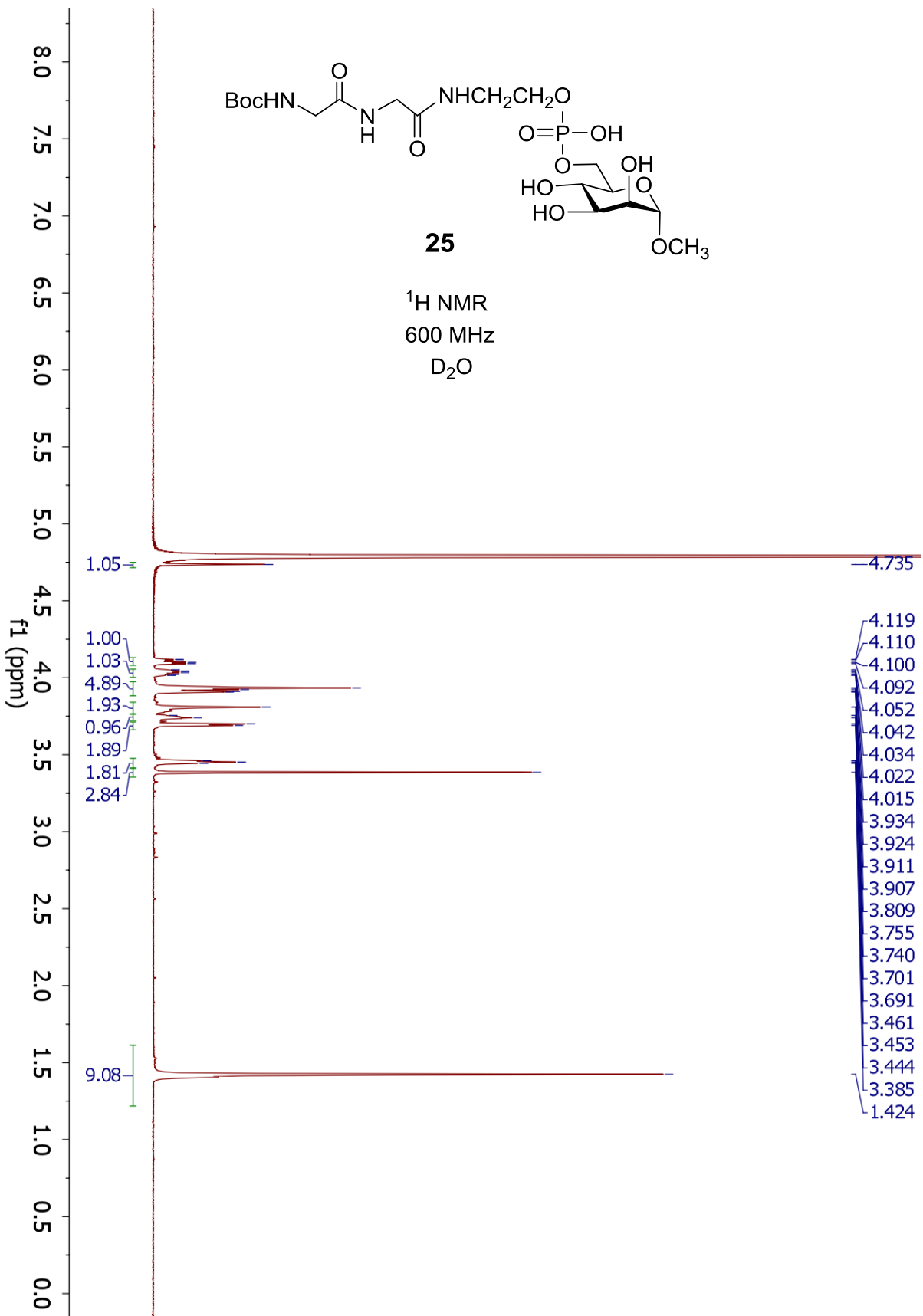
Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	i-FIT (Norm)	Formula
473.1521	473.1536	-1.5	-3.2	3.5	125.1	0.0	C16 H30 N2 O12 31P

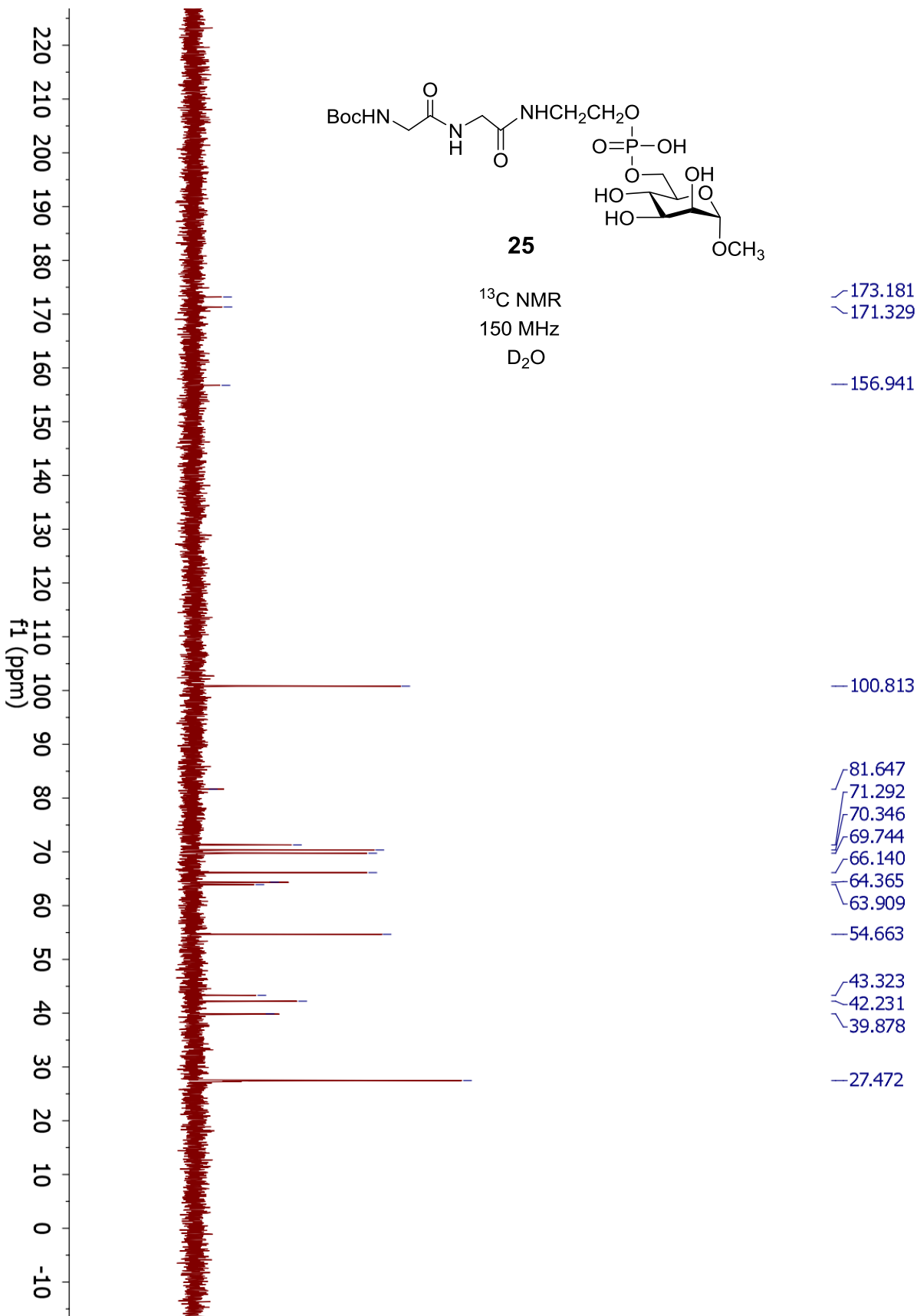


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Chemical Formula: C₁₆H₃₀N₂O₁₂P⁻

Exact Mass: 473.1542





Sample Name:

Data Collected on:

600-nmr600

Archive directory:

/home/vmml/vmrmsys/probes/probe_calibs

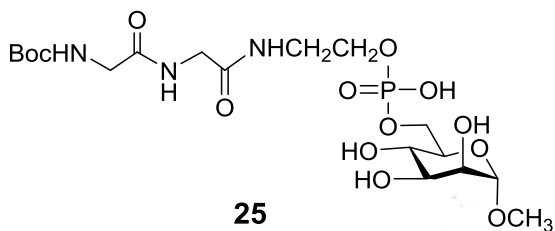
Sample directory:

F1F1file: rzy-1-139-cosy-D20

Pulse Sequence: COSY

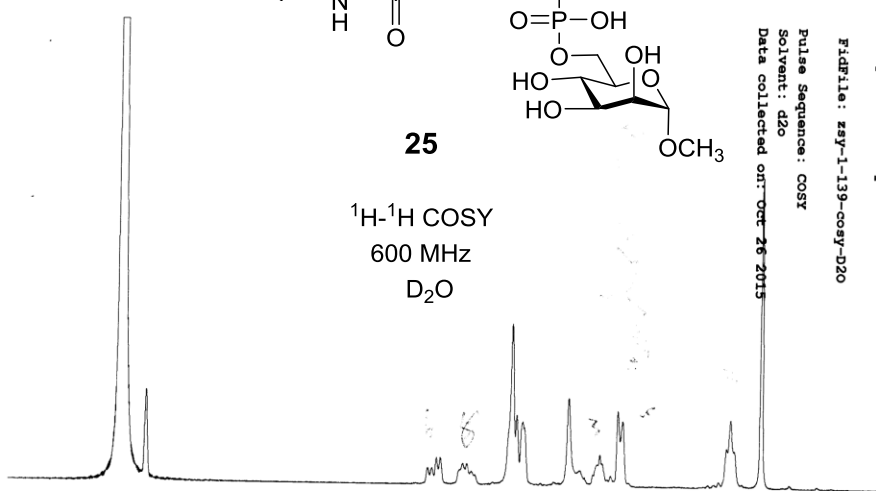
Solvent: d2o

Data collected on: Oct 26 2015



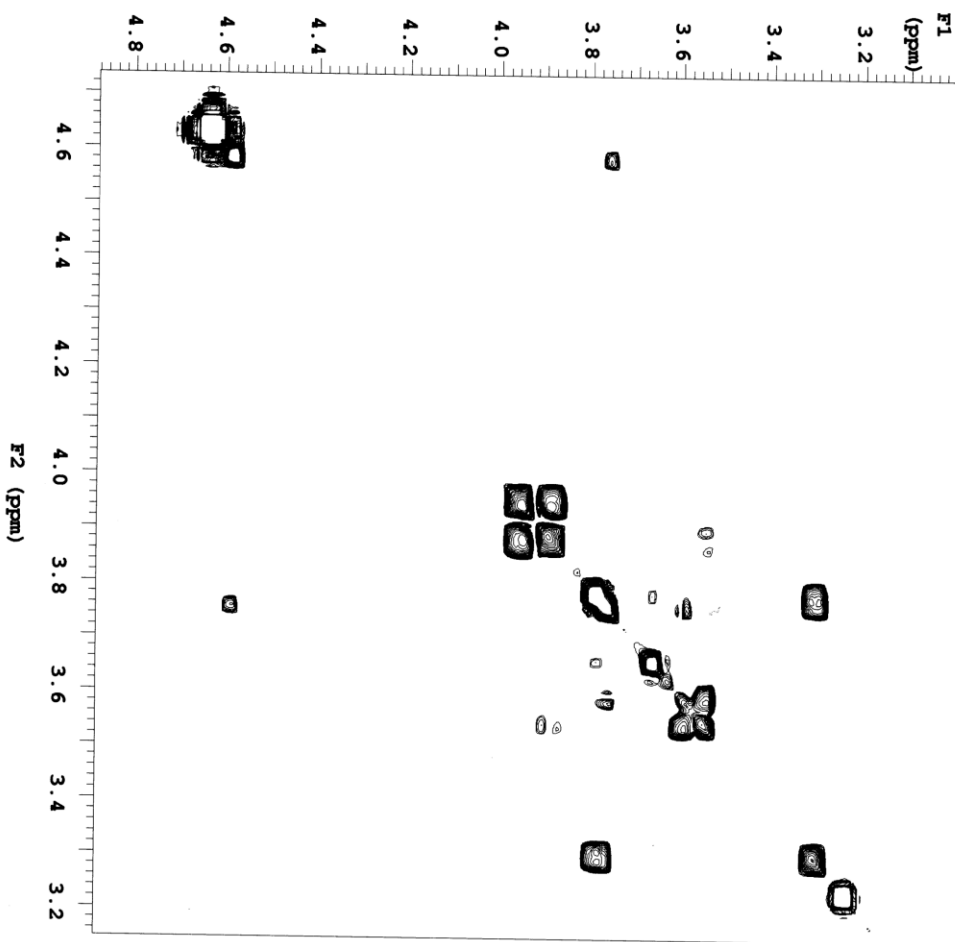
25

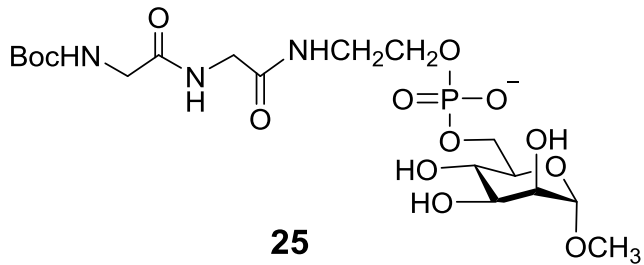
¹H-¹H COSY
600 MHz
D₂O



Agilent 600 NMR spectrometer

Agilent Technologies





Chemical Formula: $C_{18}H_{33}N_3O_{13}P^-$
 Exact Mass: 530.1756

Elemental Composition Report

Single Mass Analysis

Tolerance = 10.0 PPM / DBE: min = -1.5, max = 100.0

Element prediction: Off

Number of isotope peaks used for i-FIT = 6

Monoisotopic Mass, Even Electron Ions

182 formula(e) evaluated with 1 results within limits (all results (up to 1000) for each mass)

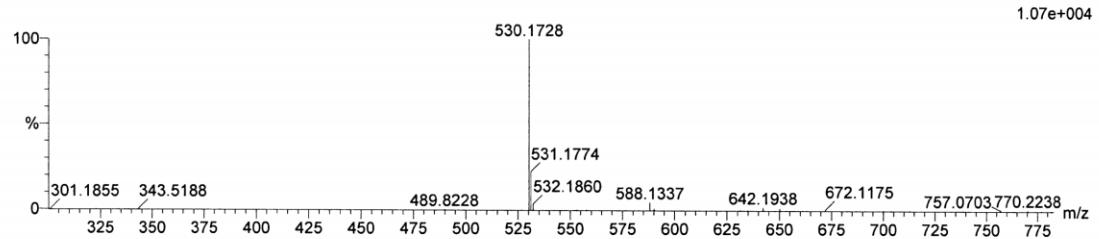
Elements Used:

C: 18-18 H: 0-40 N: 0-5 O: 0-15 31P: 0-1

LCT Premier KD128

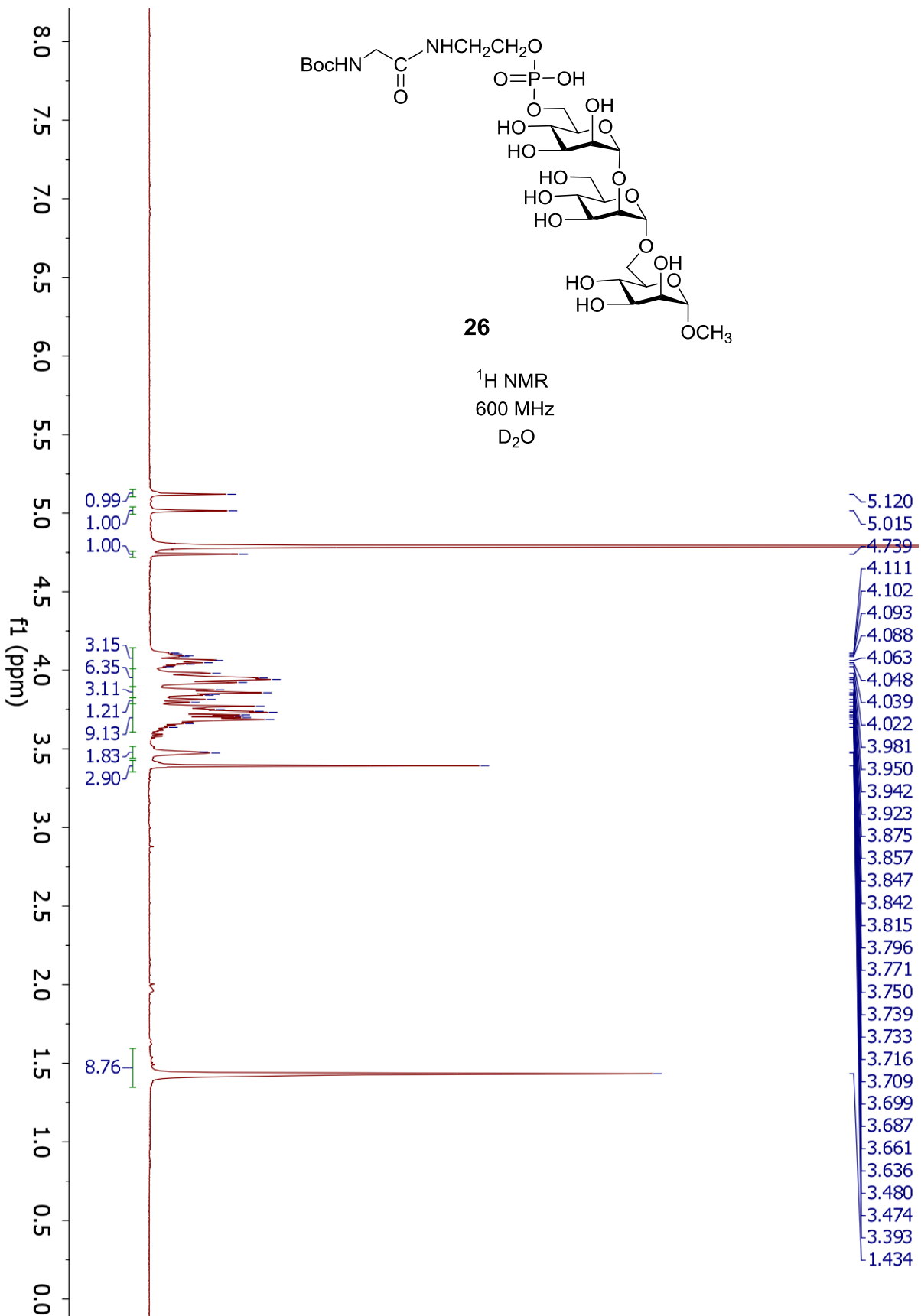
TOF MS ES-

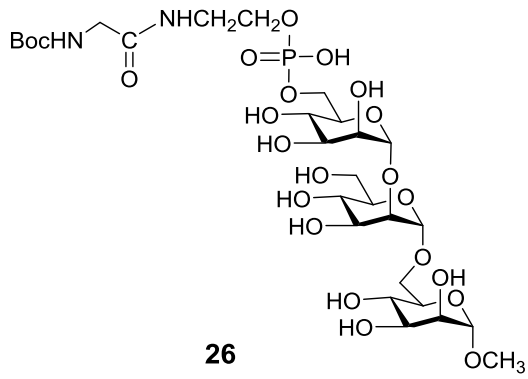
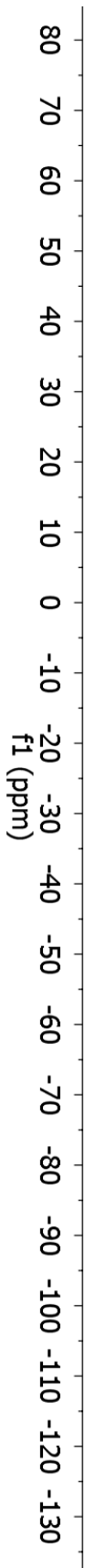
S.Zhu ZSY-1-139 in MeOH Cone(V)50



Minimum: -1.5
 Maximum: 3.0 10.0 100.0

Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	i-FIT (Norm)	Formula
530.1728	530.1751	-2.3	-4.3	4.5	38.2	0.0	C18 H33 N3 O13 31P

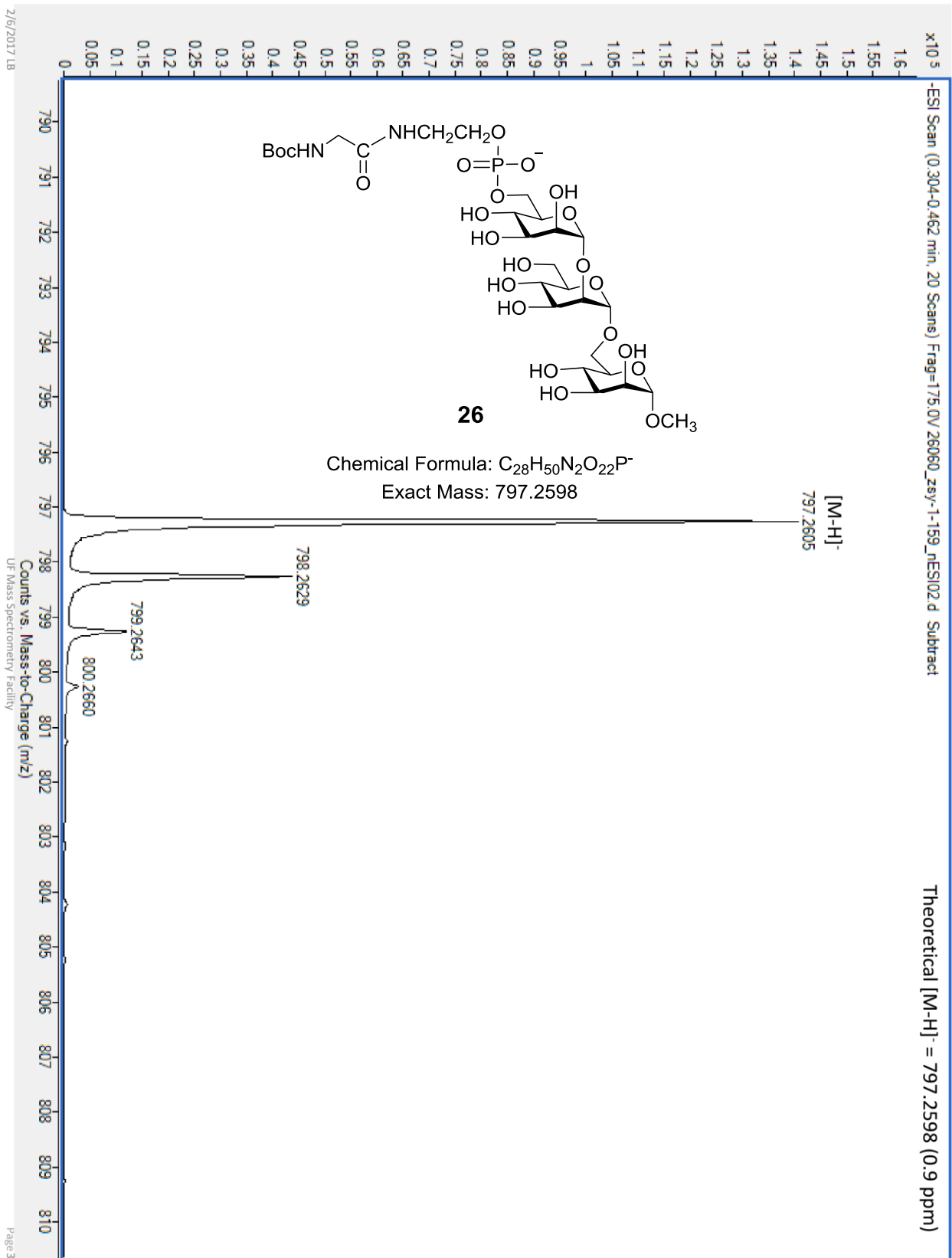


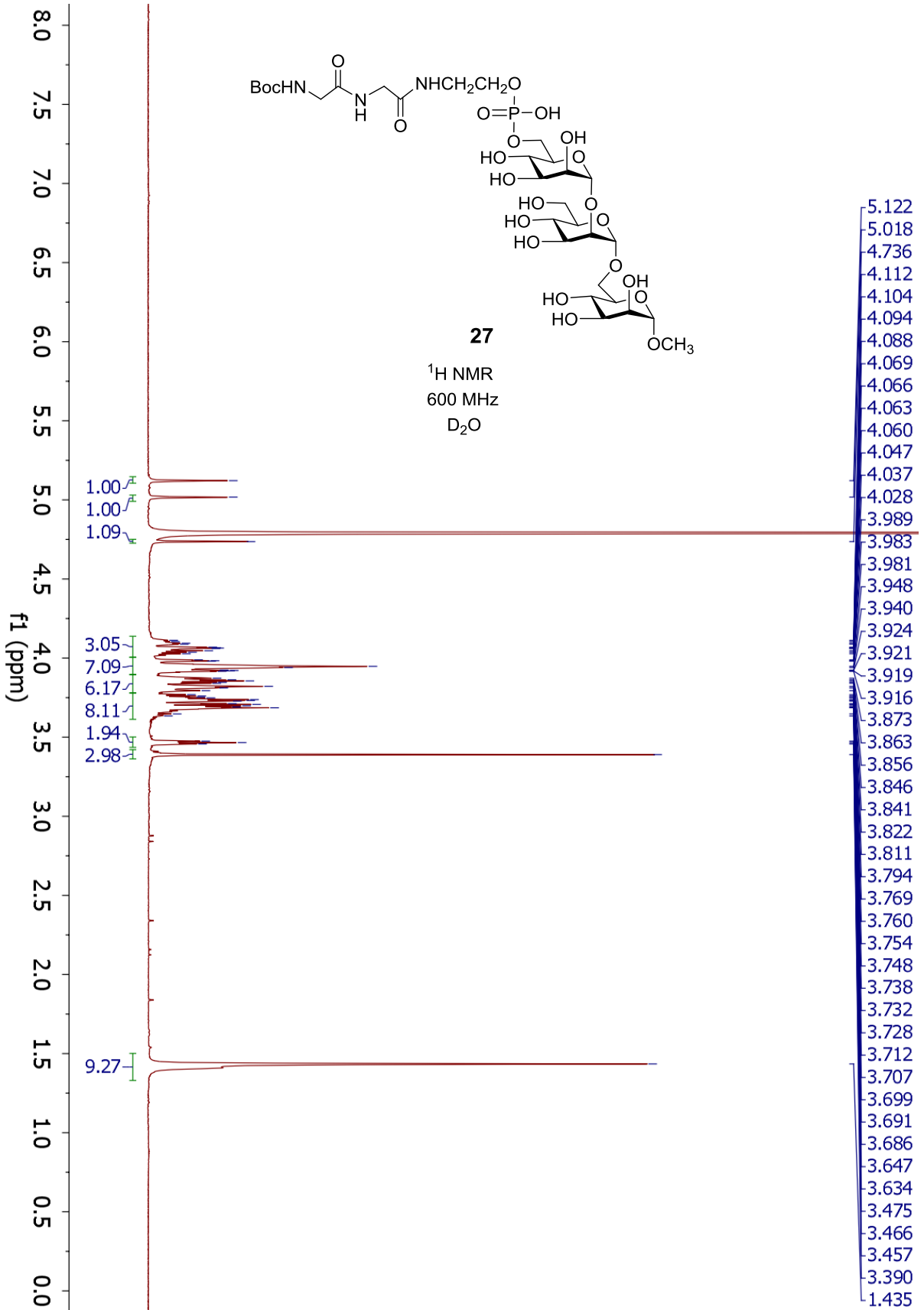


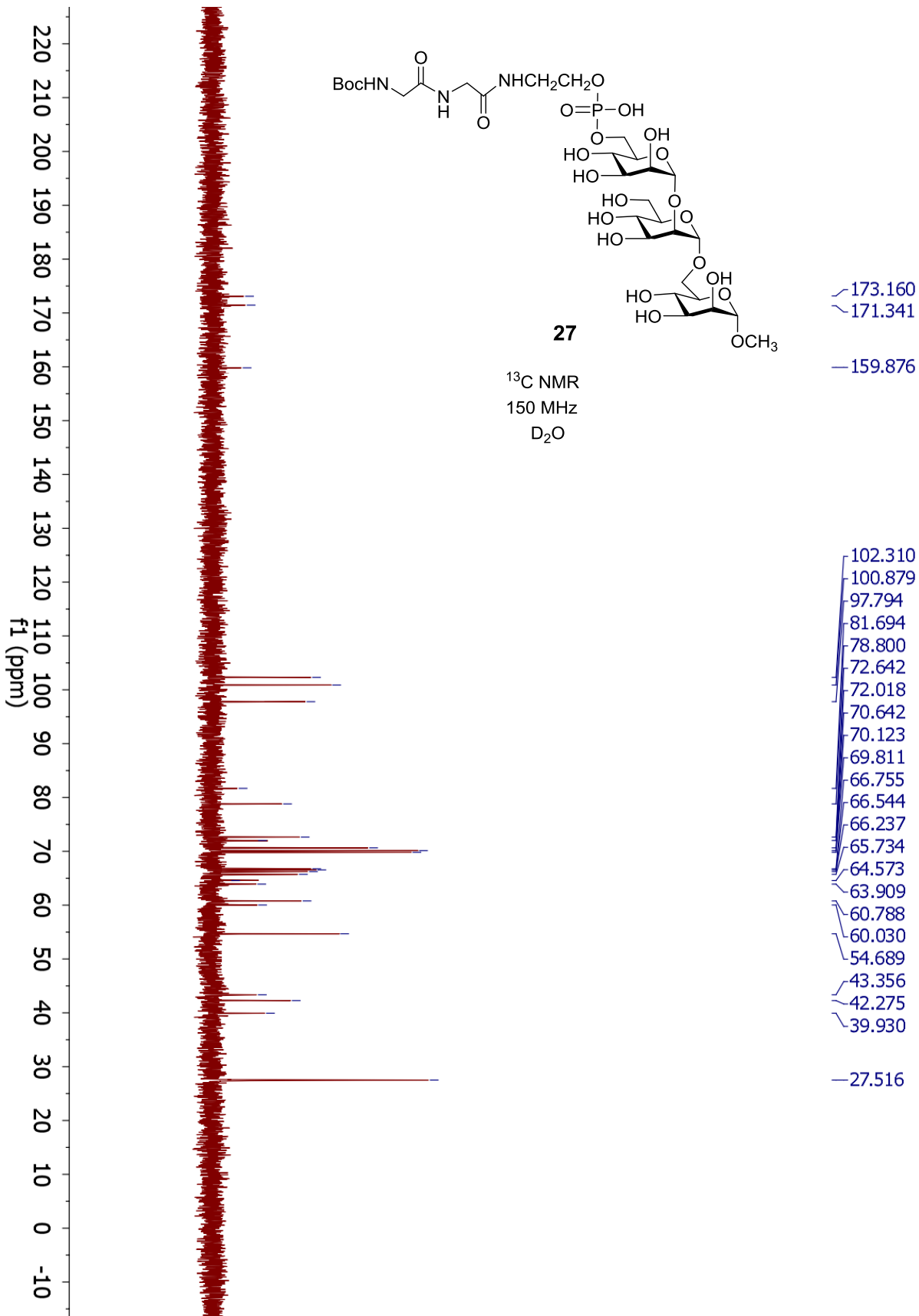
26

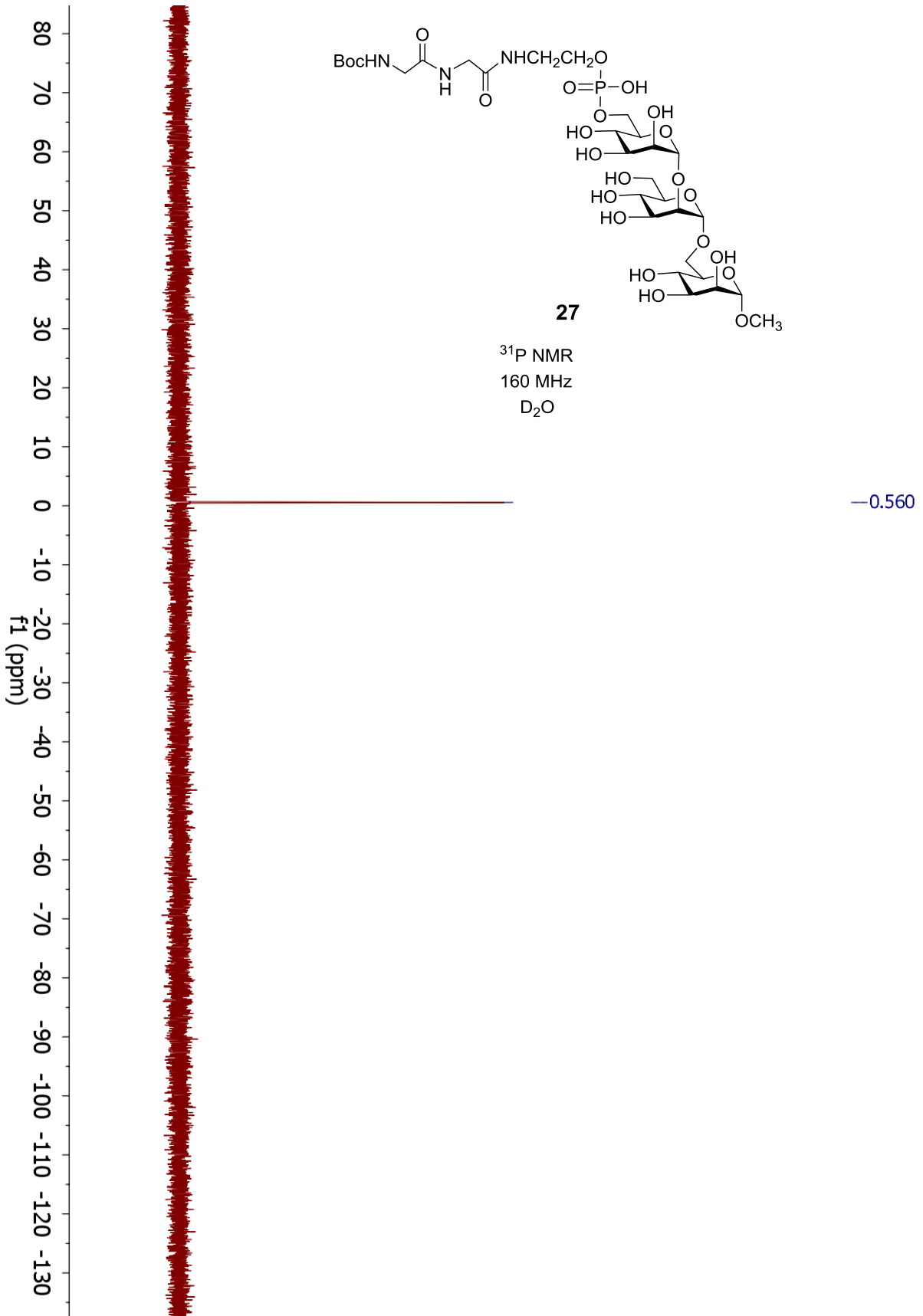
³¹P NMR
160 MHz
D₂O

-0.504

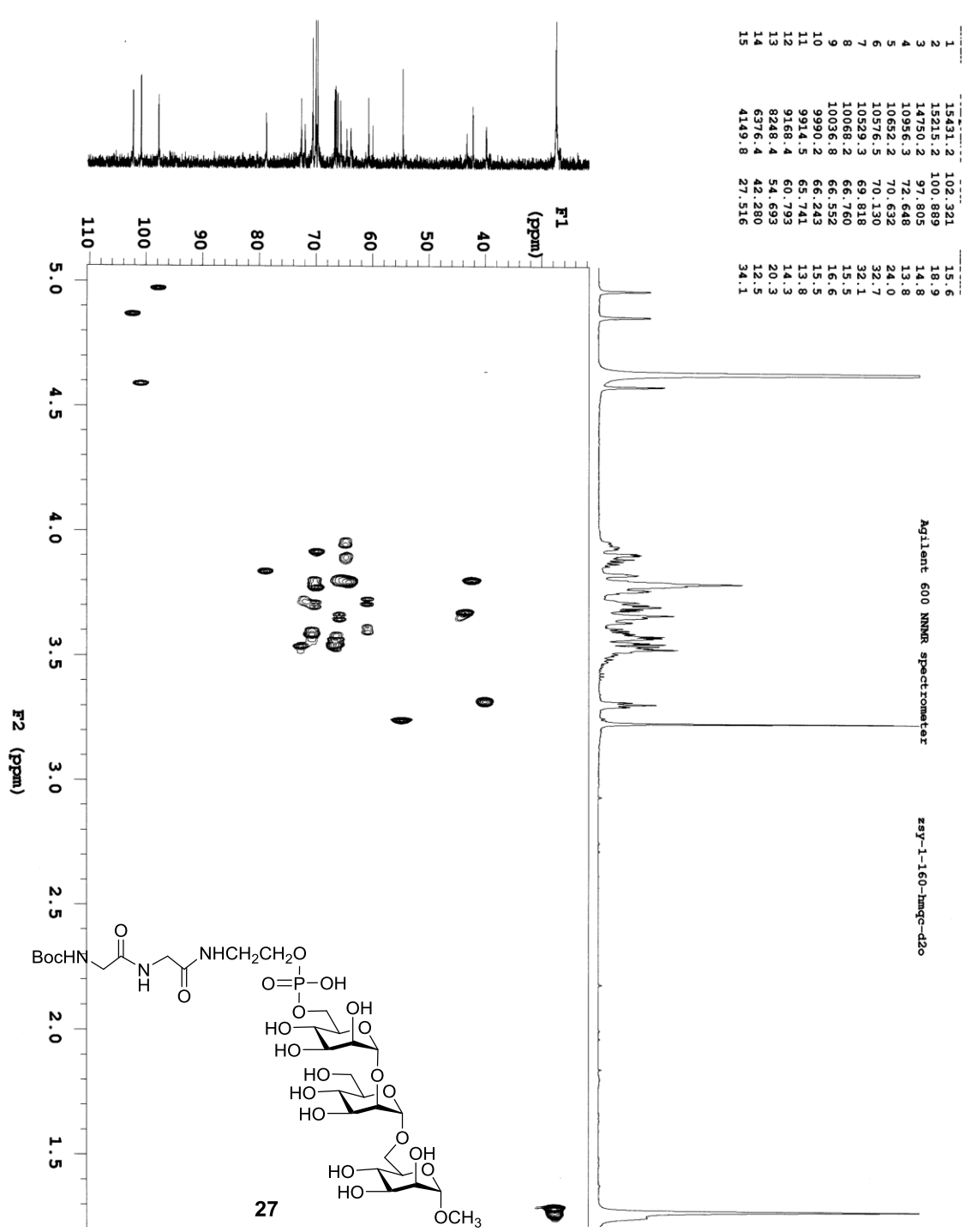




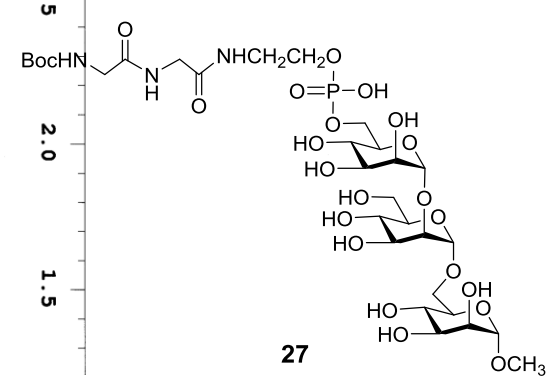




1	15431.2	102.321	15.6
2	15215.2	100.889	18.9
3	14750.2	97.805	14.8
4	10956.3	72.648	13.8
5	10652.2	70.632	24.0
6	10576.5	70.130	32.7
7	10529.3	69.818	32.1
8	10068.2	66.760	15.5
9	10036.8	66.552	15.6
10	9990.2	66.243	15.5
11	9914.5	65.741	13.8
12	9168.4	60.793	14.3
13	8248.4	54.693	20.3
14	6376.4	42.280	12.5
15	4149.8	27.516	34.1



Agilent 600 NMR spectrometer
 zxy-1-160-1mqc-d2o



¹H-¹³C HMQC
 600/150 MHz
 D₂O

