Supplemental material

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Figure S1. **CRISPR screen overview and validation of SETDB1 as a target.** (A) CRISPR negative selection screen flowchart showing low MOI transduction with the sgRNA library into THP-1–Cas9 cells. Reference cells were collected at day 7, and cells were also analyzed at day 21 by next generation sequencing. (B) Western blotting for SETDB1 in THP-1–Cas9 cells after treatment with two different *SETDB1*-specific sgRNAs or NTC for 5 d. n = 2 blots shown. (C) THP-1–Cas9 colony growth after 28 d in methocult, after treatment with three different *SETDB1*-targeting sgRNAs, a positive control sgRNA against PLK1, or two different control NTC sgRNAs. (D) Quantification of colony numbers from C. n = 2 biological replicates, and error bars represent standard deviation. (E) RNA-seq correlation plot comparing gene expression fold change values at day 4 for cells treated with *SETDB1* sgRNA 6 or 9, relative to NTC sgRNA. n = 3 biological replicates for each treatment condition; $R^2 = 0.7814$.



Figure S2. Loss of SETDB1 leads to loss of viability and induction of viral response genes and induction of TEs. (A) Western blotting of THP-1 cells treated with SETDB1-specific or NTC-inducible shRNA THP-1 cells with or without 7 d of doxycycline treatment. (B) Taqman qRT-PCR assay for SETDB1 mRNA levels with SETDB1-specific or NTC shRNAs 5 d after treatment with doxycycline. (C) THP-1 viability as measured by CellTiter-Glo after 14 d of exposure to induced SETDB1-specific or NTC shRNAs. (D) Caspase 3/7 apoptosis assay on inducible shRNA cell lines treated with doxycycline for 14 d. (E–H) Expression of ERV3-1 endogenous retrovirus and ISGs 5 d after treatment with doxycycline. (I) Taqman qRT-PCR assay for ERV3-1 and IFIT1-3 mRNA levels after treatment with SETDB1-specific or NTC sgRNAs for 7 d. (B–I) n = 3 experiments, and Student's t-tests were performed. All error bars represent standard deviation. (B–I) ***, P < 0.001; ****, P < 0.001. (J) An RNA-seq volcano plot depicting expression changes of TEs in THP-1-Cas9 cells at day 4 after treatment with a representative SETDB1-specific sgRNA versus NTC. n = 3 for all samples; EdgeR GLM log₂ fold change.



Figure S3. Integrative genomics browser view of SETDB1 deletions caused by sgRNA 6 and strand-specific analysis of TE transcripts. (A) IGV view RNAseq data confirm deletions in SETDB1 transcripts caused by SETDB1 sgRNA 6 at day 4 after treatment with SETDB1 sgRNAs. (B–D) Percentage of strands expressed for significantly up-regulated ERV, LINE-1, and Satellite subfamily members to assess bidirectional transcription after treatment with NTC or SETDB1 sgRNA after 4 and 7 d of treatment. n = 3 biological replicates, and all error bars represent standard deviation.



Figure S4. ChIP-seq on SETDB1 mutant cells shows some loss of H3K9me3 at zinc-finger genes, but not IFIT genes. (A and B) H3K9me3 ChIP-seq density plots showing H3K9me3 peaks at IFIT (A) and zinc-finger genes (B) at 6 d after treatment with SETDB1 sgRNAs or NTC. Y axes are 0–25, and regions are ~150 kb.



Figure S5. **Cell death in SETDB1 mutant cells is dependent on the viral sensing machinery.** (A) Viability of cells treated with five different sgRNAs/target for MDA5, MAVS, DDX58 (RIG-I), or NTC control for 2 d, followed by 7 d of treatment with SETDB1 sgRNA 6 or an NTC sgRNA. Mean represents viability of cells treated with five different MAVS or MDA5 sgRNAs and SETDB1 sgRNA 6). *, P < 0.05; **, P < 0.01. (B and C) FACS assays confirming loss of MDA5 and MAVS with distinct, target-specific sgRNAs 14 d after transduction with SETDB1 sgRNA 6. (D) Indel rate in THP-1–Cas9 cells treated with RIG-1 sgRNAs 14 d after transduction with SETDB1 sgRNA 6.