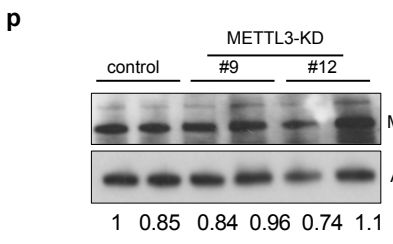
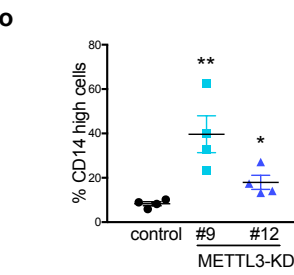
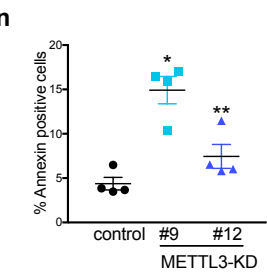
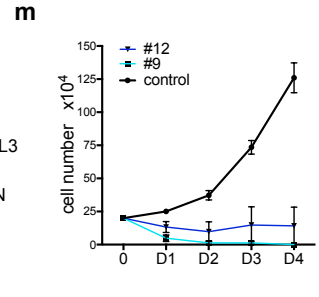
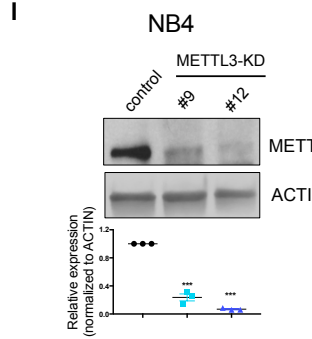
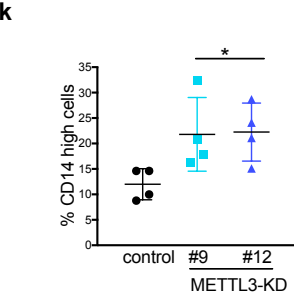
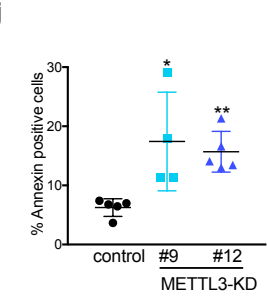
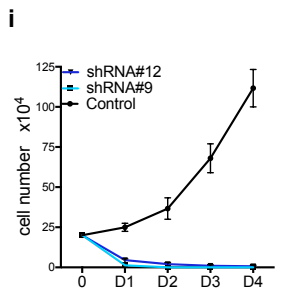
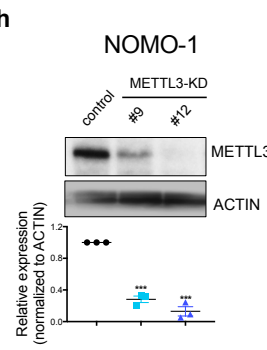
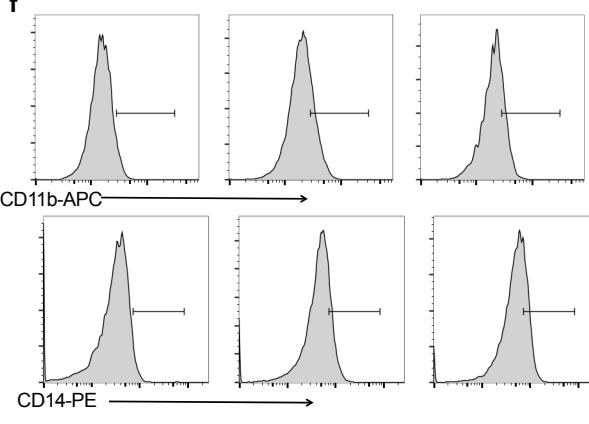
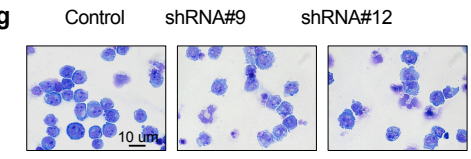
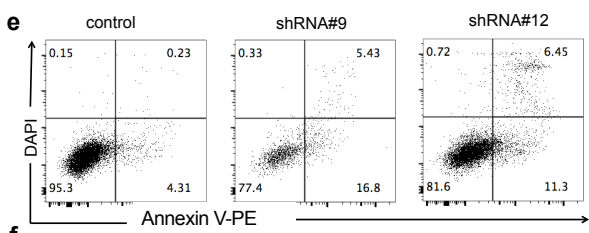
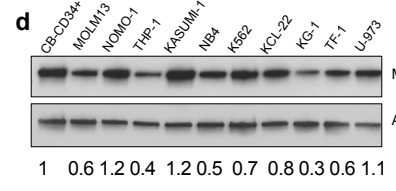
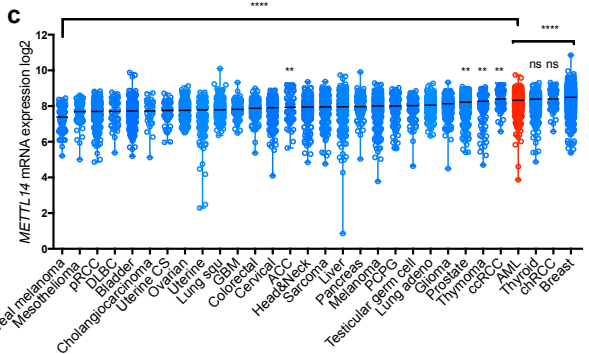
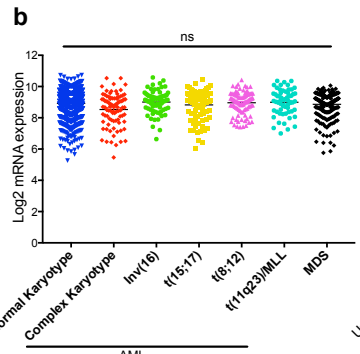
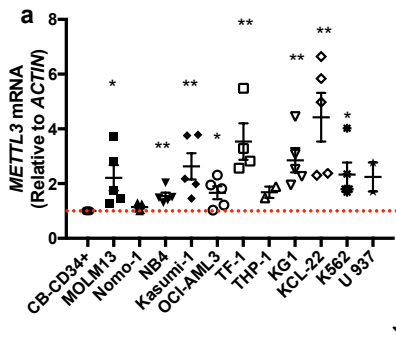


**Supplementary Figure 1**

$m^6A$  is required to maintain hematopoietic stem/progenitor cells in an undifferentiated state.

(a) Representative TLC images of **Figure 1b**. (b) Colony forming assay of HSPCs in **Figure 1(a-e)**.  $10^4$  CD34<sup>+</sup> cells were plated from control or *METTL3* knockdown cells four days post-transduction. The total number of colony forming units (CFUs) was scored two weeks after plating. Macrophage (light blue), Erythroid (red), GEMM = Granulocyte/Erythrocyte/Macrophage/Megakaryocyte (dark blue), and GM = Granulocyte/Macrophage (grey). n=3 independent experiments. \*\* p < 0.01 two tailed *t*-test. (c-d) Myeloid differentiation of HSPCs in **Figure 1(a-e)**. Representative FACS analysis used to quantify differentiation of HSPCs in **Figure 1e**. (e) Morphological analysis of HSPCs in **Figure 1(a-e)**. Cytospins of control and *METTL3* knockdown normal hematopoietic cells were prepared and stained with GIEMSA. (f) Expression of stem/progenitor cells CD34<sup>+</sup> surface marker. Representative FACS analysis used to quantify differentiation of HSPCs in **Supplemental figure 1i**. (g) Representative TLC images of **Figure 1f**. (h) Colony forming assay of HSPCs in **Figure 1 (f-h)**. n=2 independent experiments. (i) Quantitative summary of FACS analysis of CD34<sup>+</sup> expression in control and *METTL3*-knockdown and in HSPCs transduced with EV and wild type *METTL3* and *METTL3-CD*. \*p < 0.05, \*\*p < 0.01, two-tailed *t* test. (j) Immunoblot analysis of *METTL3* expression in HSPCs in **figure 1 (f-h)**. Top: representative immunoblot images. Bottom:

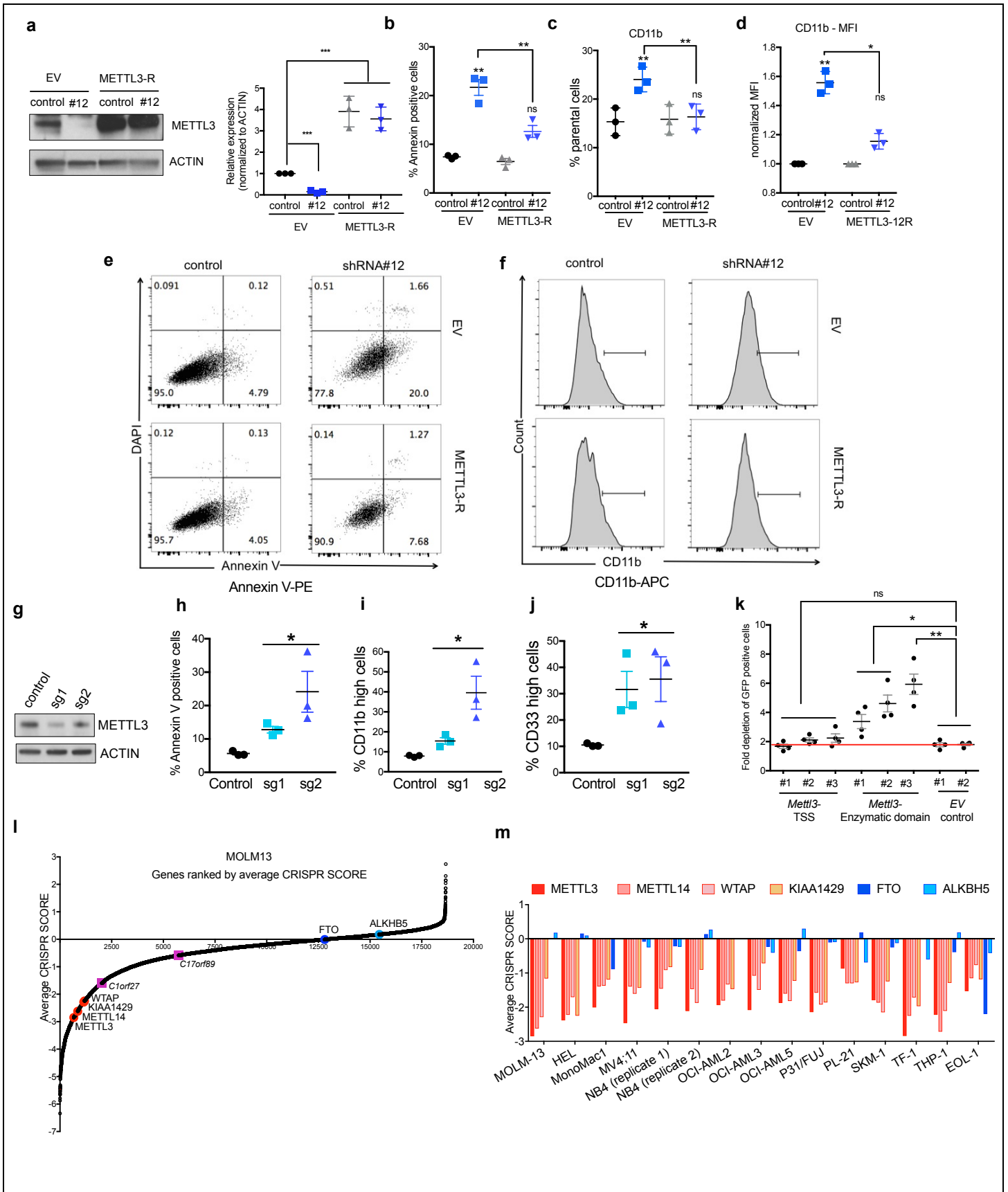
quantitative summary of the immunoblots. n=3 independent experiments; error bars, s.e.m. \* p<0.05, \*\*p<0.01,\*\*\*p<0.001 two-tailed *t* test(k) Representative FACS analysis used to quantify differentiation of HSPCs in **Figure 1h**. (l) *Mettl3* mRNA expression in mouse hematopoietic cells. Bone marrow cells from n=4 wild type C57BL/6J mice were harvest. Cells from different population were sorted based on surface markers previously reported<sup>1</sup>. HSC: Hematopoietic stem cells; MPP: Multi-potent progenitor; GMP: Granulocyte Macrophage progenitor; CMP: Common Myeloid progenitor; MEP: Megakaryocyte-Erythrocyte progenitor; Myeloid cells (Gr1+Mac1+); B cells (B220+) and T cells (IgM+). n=4 independent experiments; error bars, s.e.m. \* p<0.05, \*\*p<0.01,\*\*\*p<0.001 two-tailed *t* test



## Supplementary Figure 2

### METTL3 is essential for acute myeloid leukemia cells

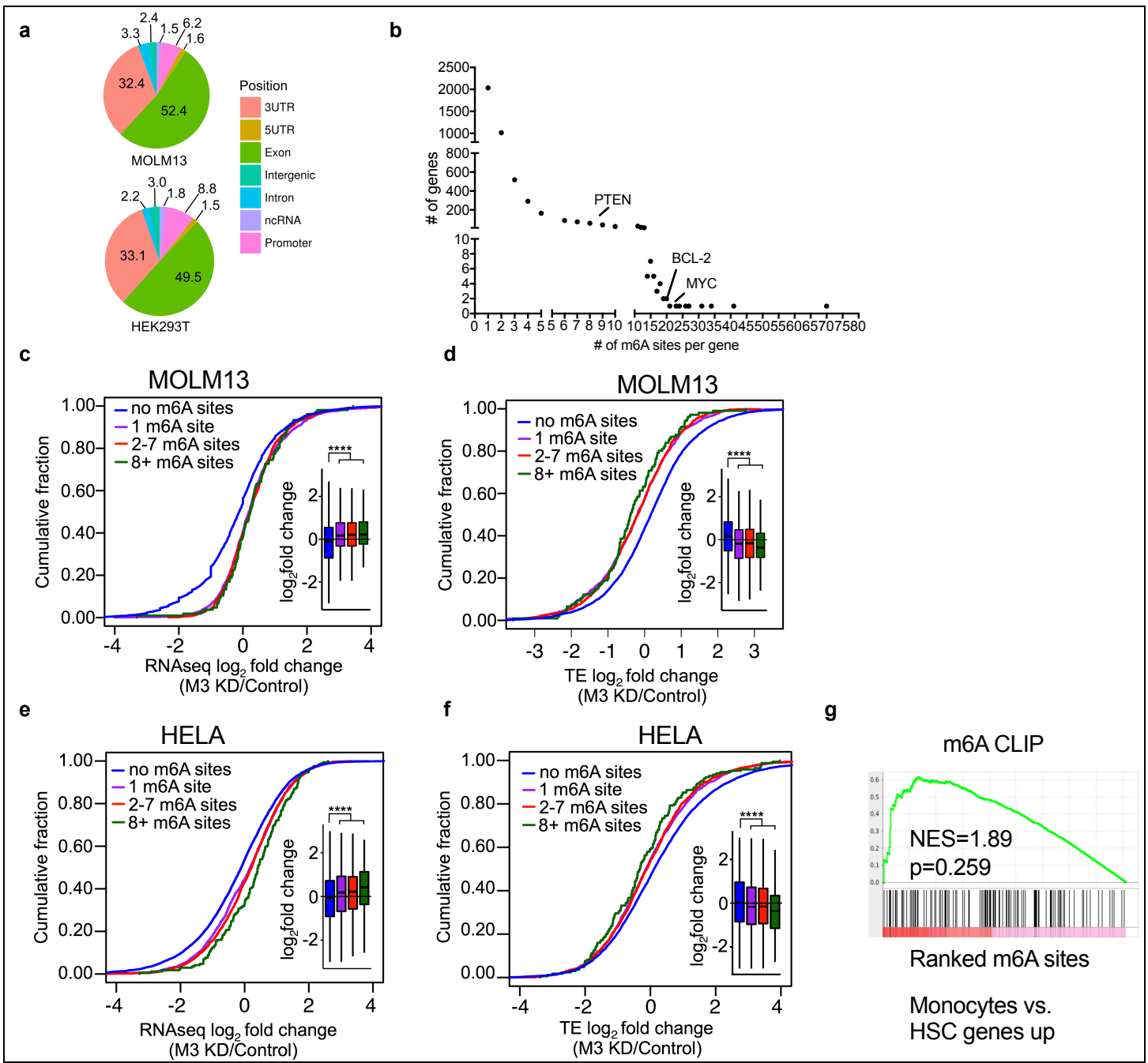
(a) mRNA expression of *METTL3* in 11 AML cell lines versus control HSPCs. RNA was extracted from cells and *METTL3* mRNA was quantified by qPCR. *ACTIN* serves as control. (b) *METTL3* mRNA expression AML patients. The graph shows the log<sub>2</sub> expression of *METTL3* from patients with various subtypes of AML and MDS. Normal karyotype, n=989; Complex karyotype, n=87; AML Inv(16), n=77; AML t(15;17), n=87; AML t(8;12), n=98, AML t(11q23)/MLL, n=58; MDS, n=228. (BloodPool data of *METTL3* probe 209265\_s\_at from U133 Plus 2.0 array)<sup>2</sup>. (c) *METTL14* mRNA expression in AML compared to other cancers. \*\*\*\* p<0.00001, \*\* p<0.01 ns not significant, ANOVA with multiple comparisons. (d) *METTL14* protein levels in normal HSPCs and AML cell lines. Quantitative summary is shown. (e) Representative FACS analysis used to quantify apoptosis of MOLM13 cells in **Figure 2f**. (f) Representative FACS analysis used to quantify differentiation of MOLM13 cells in **Figure 2g**. (g) Morphological analysis of HSPCs in **Figure 2(g)**. Cytospins of control and *METTL3* knockdown MOLM13 cells were prepared and stained with GIEMSA. (h-o) *METTL3* depletion multiple AML cell lines induces differentiation and apoptosis. Immunoblot of *METTL3* expression four days post-transduction with shRNA lentivirus in NOMO-1 cells (h) and NB4 cells (i). Quantitative summary is shown. Knockdown of *METTL3* reduced proliferation in NOMO-1 cells (j) and NB4 cells (m). NOMO-1 and NB4 cells had increased apoptosis (j and n, respectively) and increased myeloid differentiation (k and o, respectively) following *METTL3* depletion. n=3 independent experiments; error bars, s.e.m. \* p<0.05, \*\*p<0.01, \*\*\*p<0.001 two-tailed t test. (p) Immunoblot analysis of MOLM13 *METTL3* knockdown cells that outgrow in moribund leukemic mice. Leukemia cells were sorted for human CD45 positive (a marker for hematopoietic cells) from leukemic mice in **Figure 2h**. *ACTIN* serves as loading control. Quantitative summary is shown.



### Supplementary Figure 3

#### METTL3 is required for survival of acute myeloid leukemia cells

(a) Immunoblot of control or knockdown MOLM13 cells previously transduced with empty vector (EV) or METTL3-12R resistant to targeting by shRNA#12. Top: representative immunoblot images. Bottom: quantitative summary of the immunoblots. n=3 independent experiments; error bars, s.e.m. \* p<0.05, \*\*p<0.01, \*\*\*p<0.001 two-tailed *t* test. (b-d) Overexpression of the shRNA-resistant METTL3-12R rescues survival and differentiation defects of MOLM13 cells depleted of endogenous METTL3. Quantitative summary of Annexin V staining (b), percentage of CD11b cells (c) and MFI of CD11b expression (d). n=3 independent experiments, error bars, s.e.m. \* p<0.05, \*\*p<0.001 two-tailed *t* test. (e-f) Representative FACS analysis used to quantify apoptosis of MOLM13 cells in **Supplemental Figure 2b** and differentiation in **Supplemental Figure 2c-d**. (g-j) CRISPR/Cas9 mediated- deletion of METTL3 in MOLM13 cells. (g) Immunoblot showing METTL3 expression in MOLM13 cells upon CRISPR/Cas9-mediated deletion with two independent sgRNAs (sg1 and sg2) three days post transduction. ACTIN serves as loading control. Cells were assayed for Annexin V positivity and (h) and differentiation (i-j) as previously described with *METTL3* shRNAs. n=3 independent experiments; error bars, s.e.m. \* p<0.05, \*\*p<0.001, \*\*\* p <0.0001 two-tailed *t* test. (k) The enzymatic domain of METTL3 is essential for AML cell viability. Mouse MLL-AF9 NRAS<sup>G12D</sup> constitutively expressing Cas9 (RN2-Cas9) cells were transduced with guide RNAs (gRNAs) that specifically target regions of METTL3. Targeted regions include the transcription start site (TSS, black bars), METTL3 enzymatic domain (blue bars), or an empty virus control (grey bars). Viability was measured as the fold depletion of GFP positive cells (gRNAs transduced cells) beginning two days post-transduction. n=4 independent experiments, error bars, s.e.m. \* p<0.05, \*\*p<0.001 two-tailed *t* test. (l) CRISPR score rank of members of the RNA methylation “writers” complex and “erasers” in MOLM13 cells. CRISPR score (CS) is the average log<sub>2</sub> fold-change in the abundance of all single guide (sg)RNAs targeting the gene after 14 population doublings<sup>3</sup>. Members of the “writer” complex METTL3, METTL14, WTAP, and KIAA1429 were highlighted in red. The “erasers” were highlighted in blue. C1orf27 and C17orf89, highlighted in purple, were genes previously reported to be essential for survival of leukemia cells. (m) CRISPR score of members of the RNA methylation “writers” complex and “erasers” across all 14 tested leukemia cell lines<sup>3</sup>.



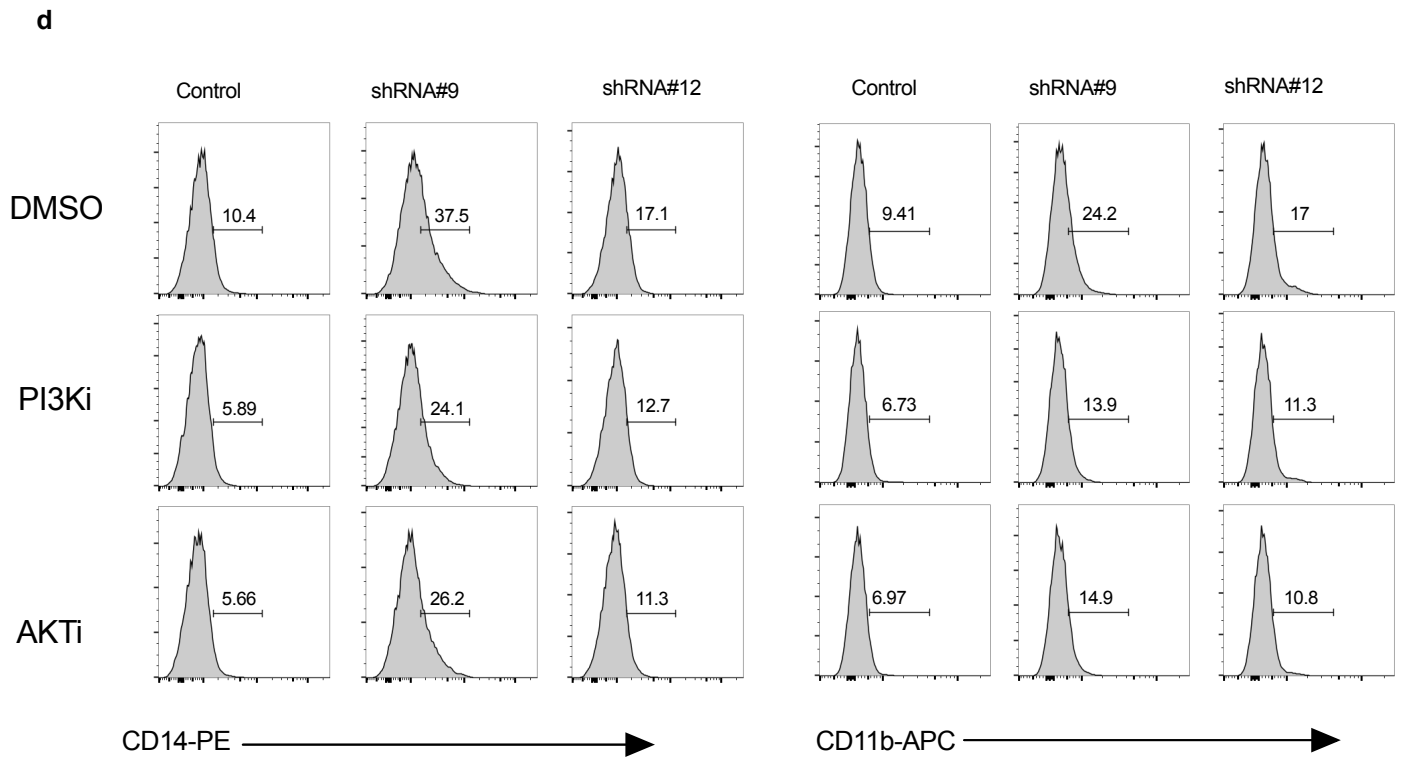
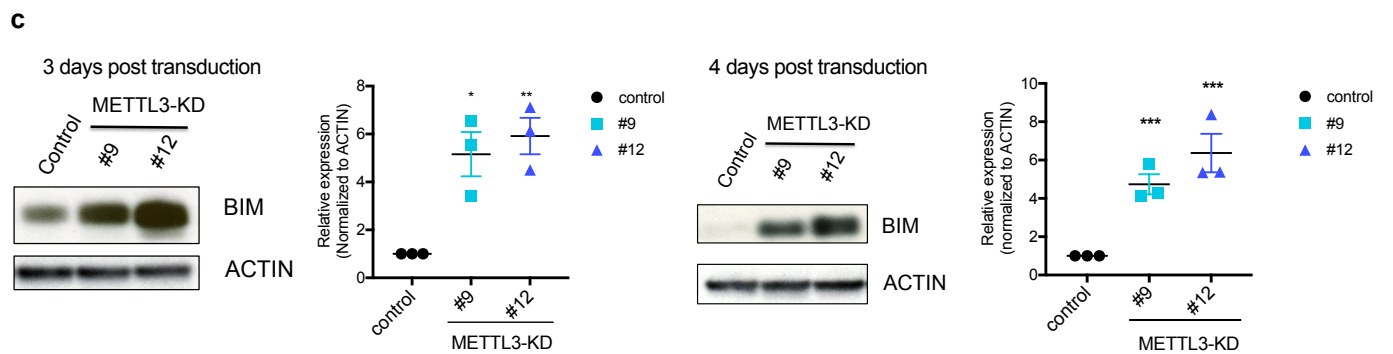
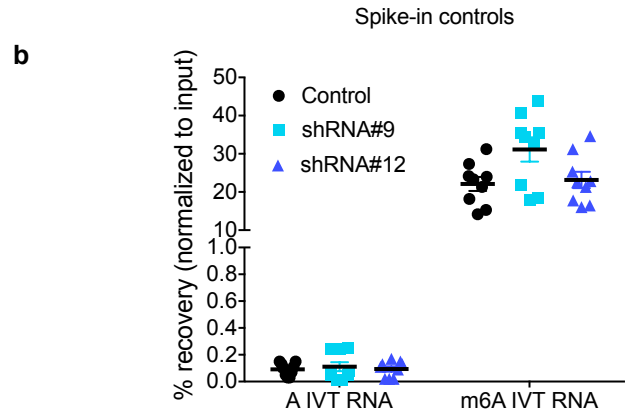
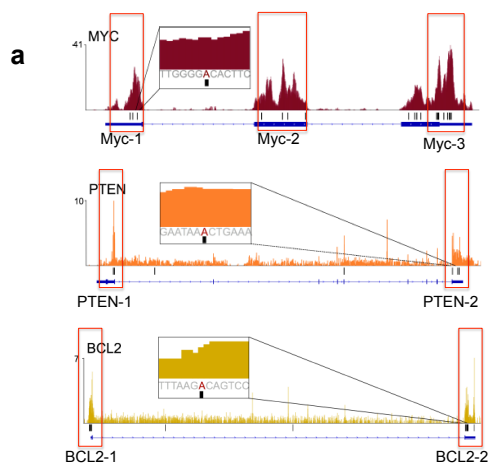
**Supplementary Figure 4**

**Mapping m<sup>6</sup>A in MOLM13 at single-nucleotide resolution**

(a) The distribution of m<sup>6</sup>A in MOLM13 cells compared to the distribution of HEK293T cells (b) Plot showing the number of m<sup>6</sup>A sites per gene (x-axis) versus the number of genes (y-axis). (c) Correlation between the number of m<sup>6</sup>A sites and mRNA expression. Transcripts were binned based on the number of m<sup>6</sup>A sites: no sites, 1 site, 2-7 sites, or 8+ sites and analyzed their expression. n=3 independent replicates per condition, \*\*\*\* p<2.2e<sup>-16</sup>, one-way ANOVA with Tukey's post-hoc test. (d) Translational efficiency is correlated with the number of m<sup>6</sup>A sites per transcript. Transcripts were binned based on the number of m<sup>6</sup>A sites: no sites, 1 site, 2-7

sites, or 8+ sites and analyzed their translational efficiency. n=3 independent replicates per condition, \*\*\*\*  $p < 2.2e^{-16}$ , one-way ANOVA with Tukey's post-hoc test. (e) Correlation between m<sup>6</sup>A sites and mRNA expression after *METTL3* knockdown in HeLa cells. n=1 independent replicate per condition, \*\*\*\*  $p < 2.2e^{-16}$ , one-way ANOVA with Tukey's post-hoc test. (f) Correlation between m<sup>6</sup>A sites and translational efficiency (TE) after *METTL3* knockdown in HeLa cells. n=1 independent replicate per condition, \*\*\*\*  $p < 2.2e^{-16}$ , one-way ANOVA with Tukey's post-hoc test. (g) The Gene set promoting monocyte differentiation enrichment analysis for m6A ranked lists.





### Supplementary Figure 5

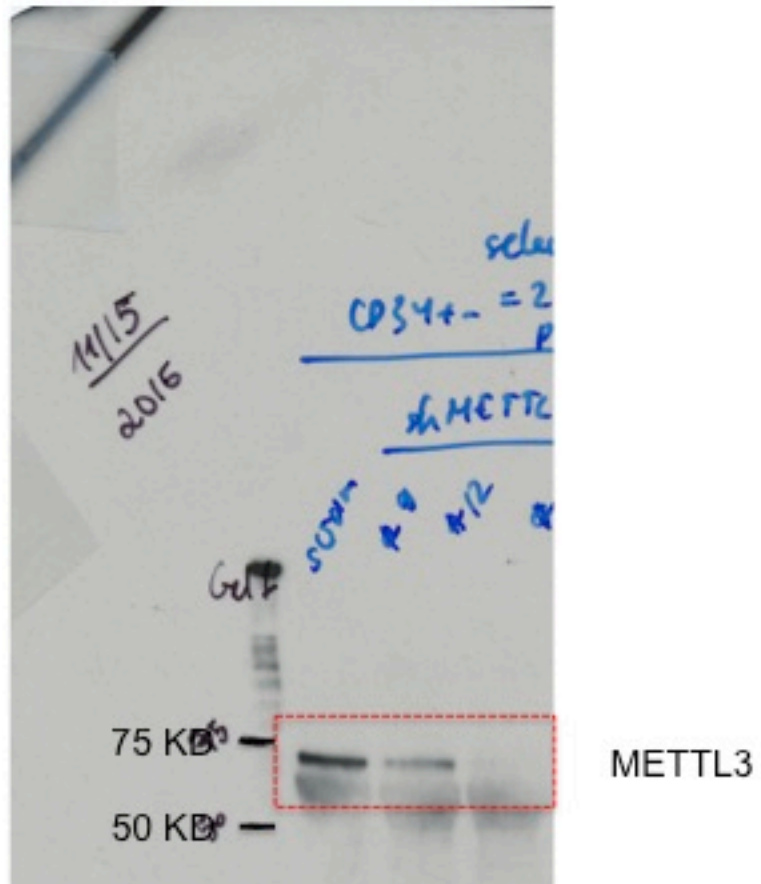
m<sup>6</sup>A modulates expression of targets c-MYC, PTEN and BCL2 and attenuation of the AKT pathway.

(a) m<sup>6</sup>A sites in *MYC*, *PTEN*, and *BCL2*. Reads (unique tags per millions) from miCLIP in MOLM13 with called m<sup>6</sup>A sites indicated with black vertical bars. Zoomed-in areas show individual m<sup>6</sup>A sites called based on C→T transitions. (b) Validation of meRIP-qPCR with spike-in RNA controls. Negative and positive control RNAs were in vitro transcribed with either ATP or m<sup>6</sup>ATP in the reaction. 100 ng of each RNA were added to poly-A+ purified RNA from MOLM-13 cells immediately before immunoprecipitation with an anti-m<sup>6</sup>A antibody. Plots represent the recovery of the control RNAs after immunoprecipitation as a percentage of the total input. (c) Immunoblot analysis for the protein level of BIM upon METTL3 depletion in MOLM13 cells. The panels are representative blots from three days or four days post-transduction of MOLM13 cells with 2 shRNAs targeting METTL3. ACTIN serves as loading control. Left: representative immunoblot images. Right: quantitative summary of the immunoblots. n=3 independent experiments; error bars, s.e.m. \* p<0.05, \*\*p<0.01, \*\*\*p<0.001 two-tailed *t* test. (d) Inhibition of the AKT pathway inhibits myeloid differentiation of MOLM13 METTL3 knocked down cells. Representative FACS analysis used to quantify differentiation of MOLM13 cells in **Figure 4g-h**.

#### Reference:

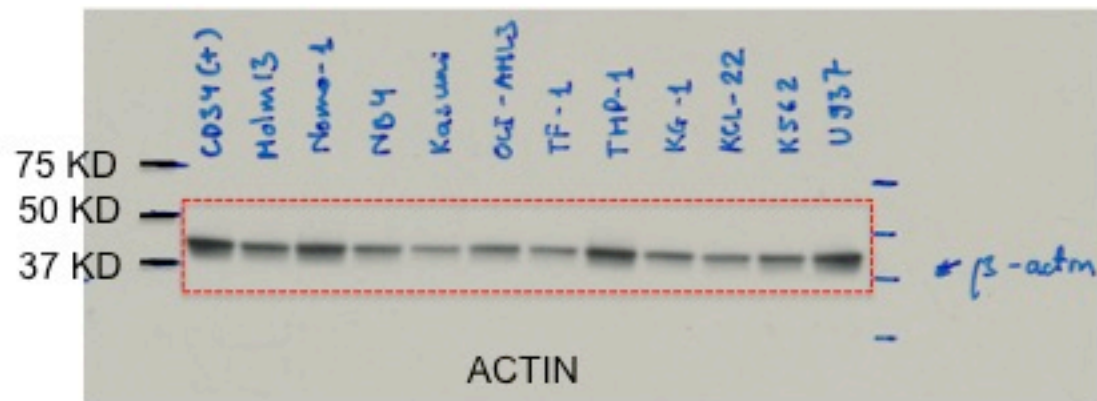
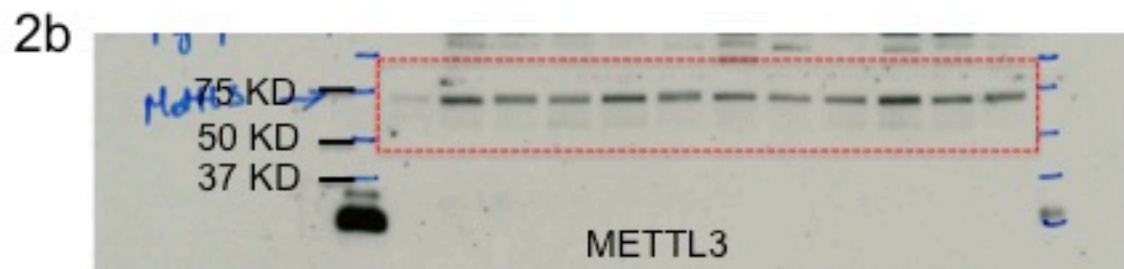
1. Kharas, M.G., *et al.* Musashi-2 regulates normal hematopoiesis and promotes aggressive myeloid leukemia. *Nat Med* **16**, 903-908 (2010).
2. Bagger, F.O., *et al.* BloodSpot: a database of gene expression profiles and transcriptional programs for healthy and malignant haematopoiesis. *Nucleic acids research* **44**, D917-924 (2016).
3. Wang, T., *et al.* Gene Essentiality Profiling Reveals Gene Networks and Synthetic Lethal Interactions with Oncogenic Ras. *Cell* **168**, 890-903 e815 (2017).

1a

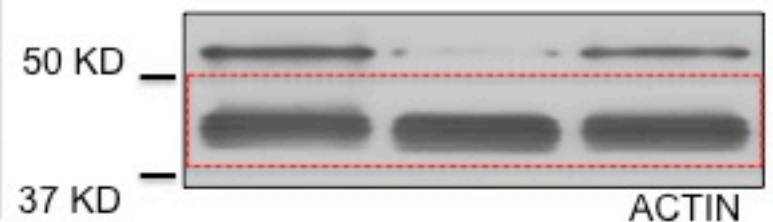
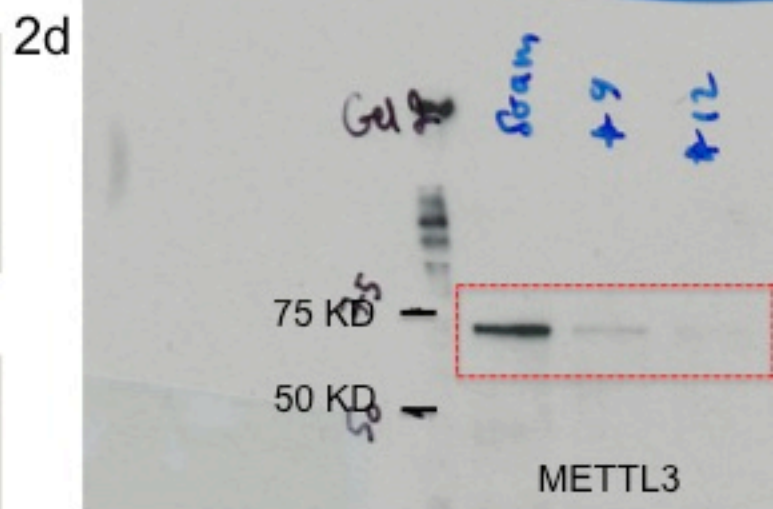


METTL3 KD in CD34+ cells

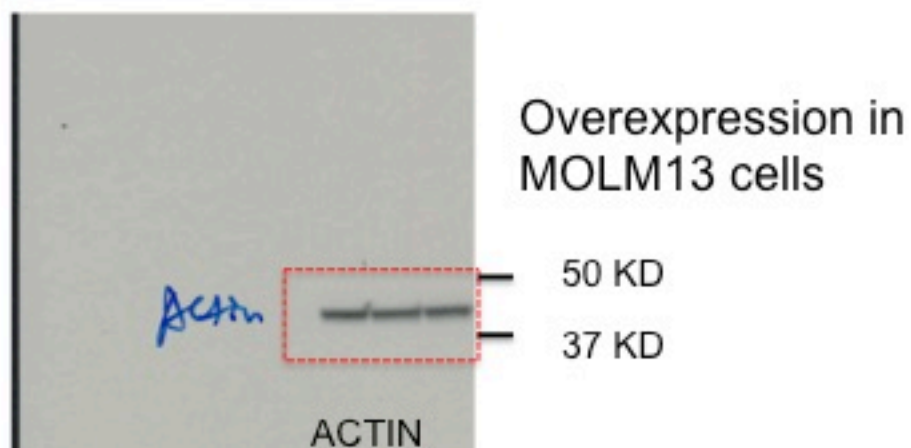
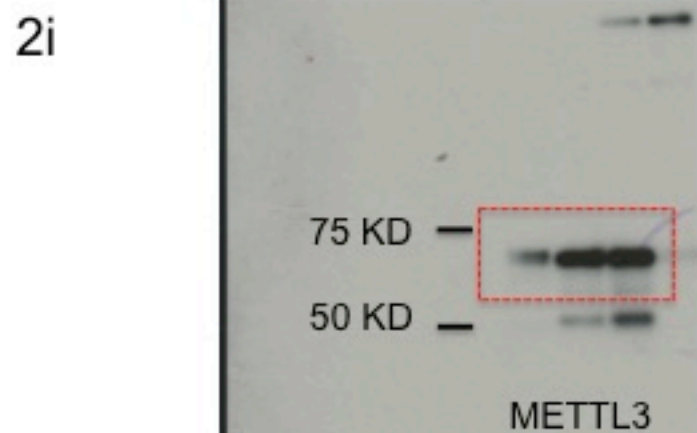
Figure 1



Cell panel



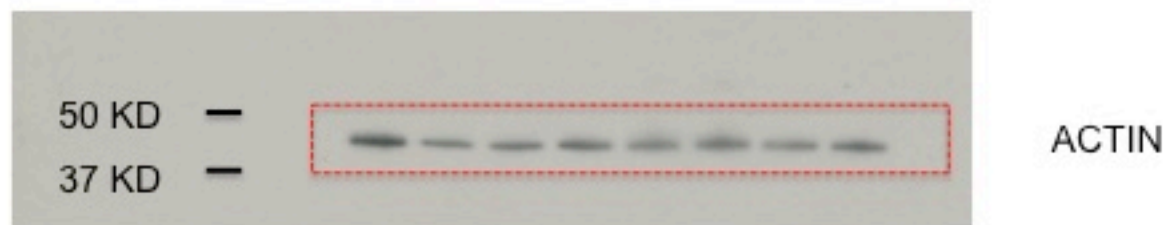
METTL3 KD in MOLM13 cells



Overexpression in  
MOLM13 cells

Figure 2

2k



human patients panel

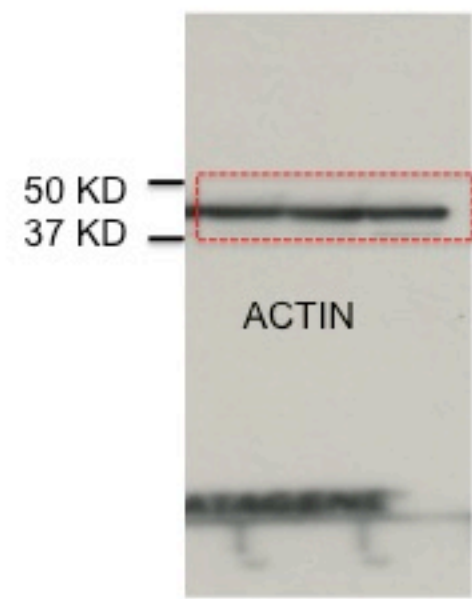
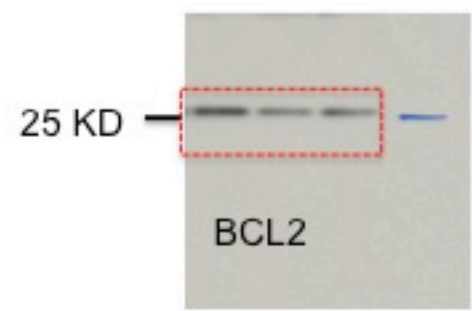
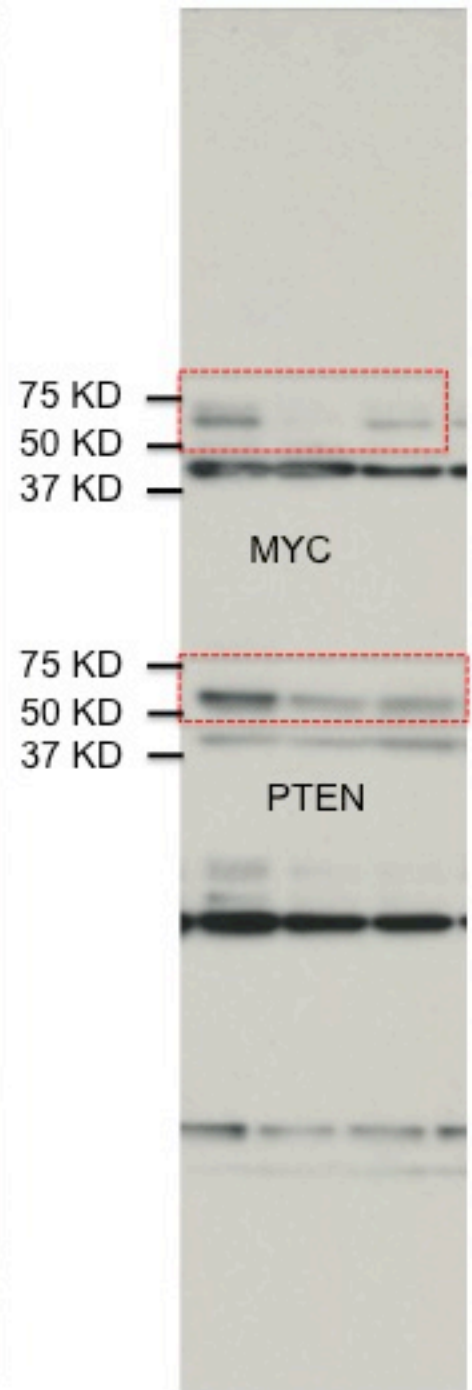
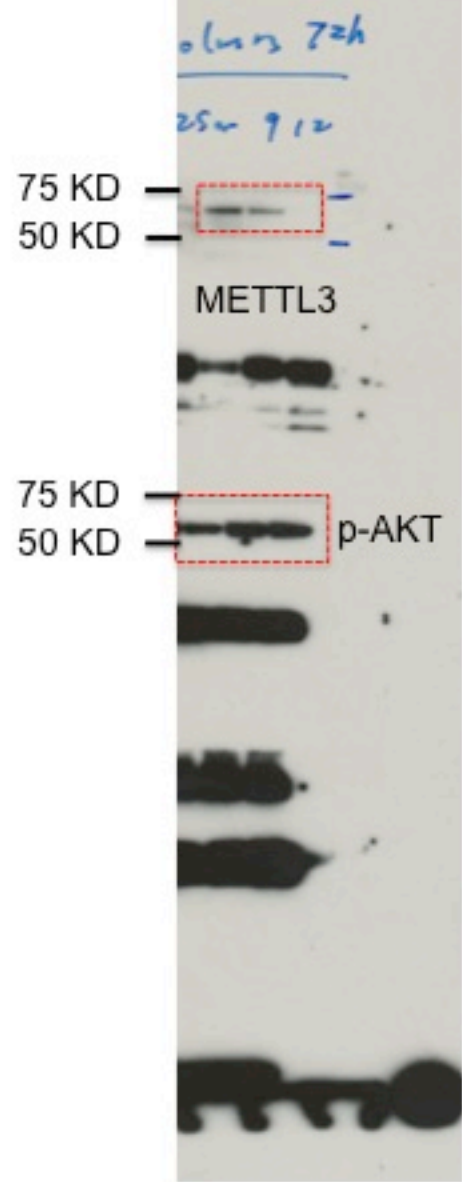
2n



METTL3 KD in human AML patient

Figure 2

4c



shRNA-3 days after transduction

Figure 4

4d

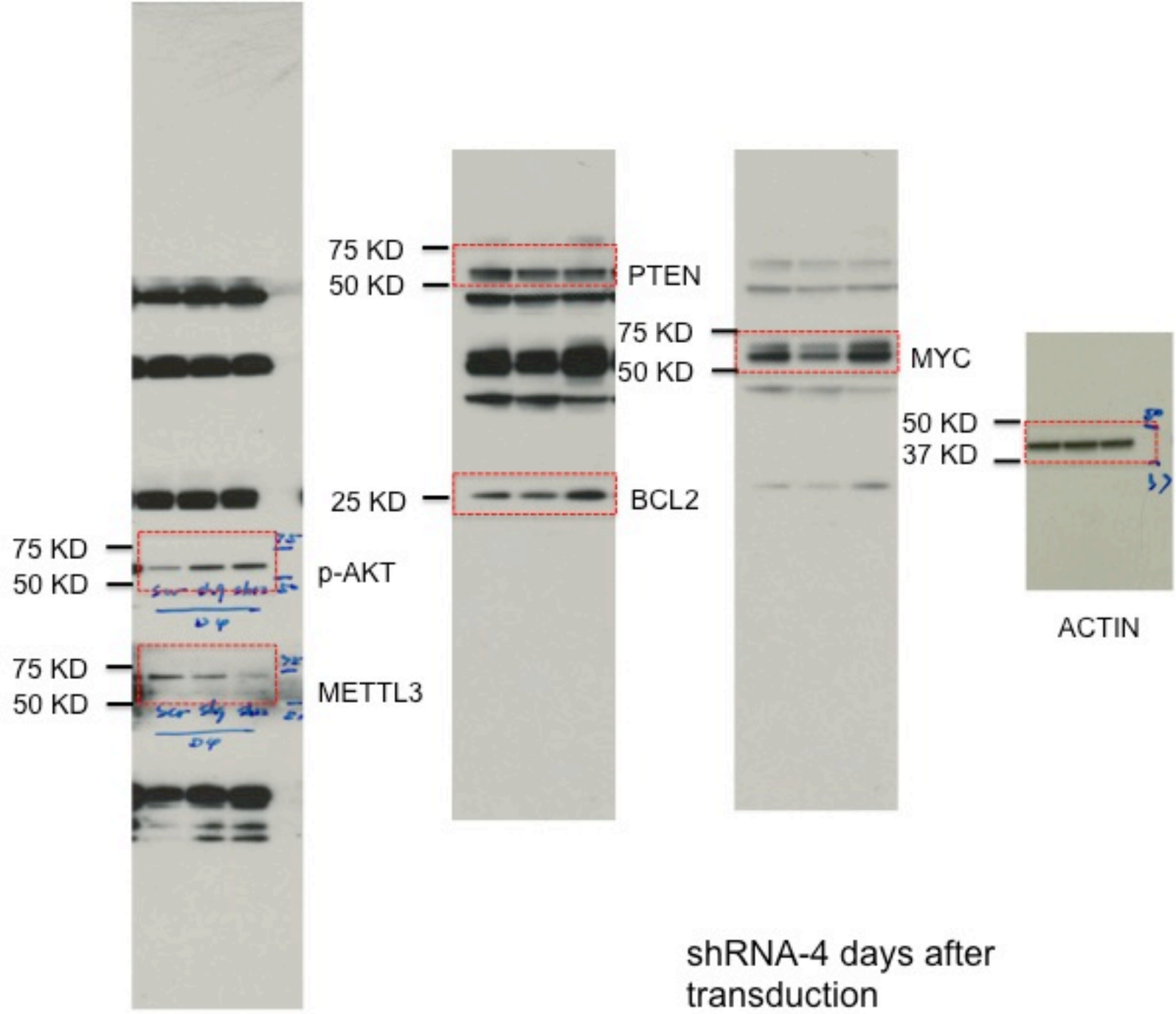
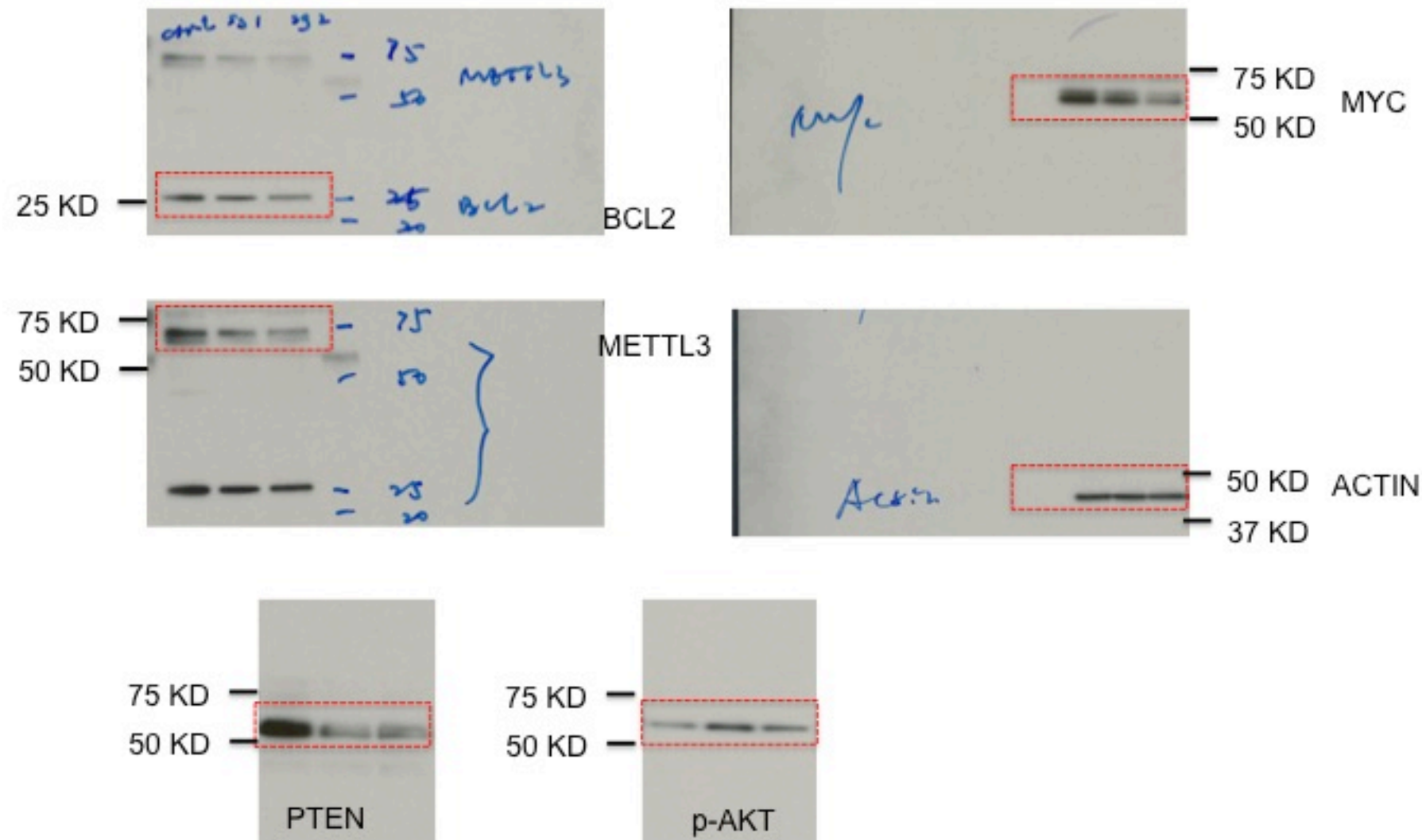


Figure 4

4e

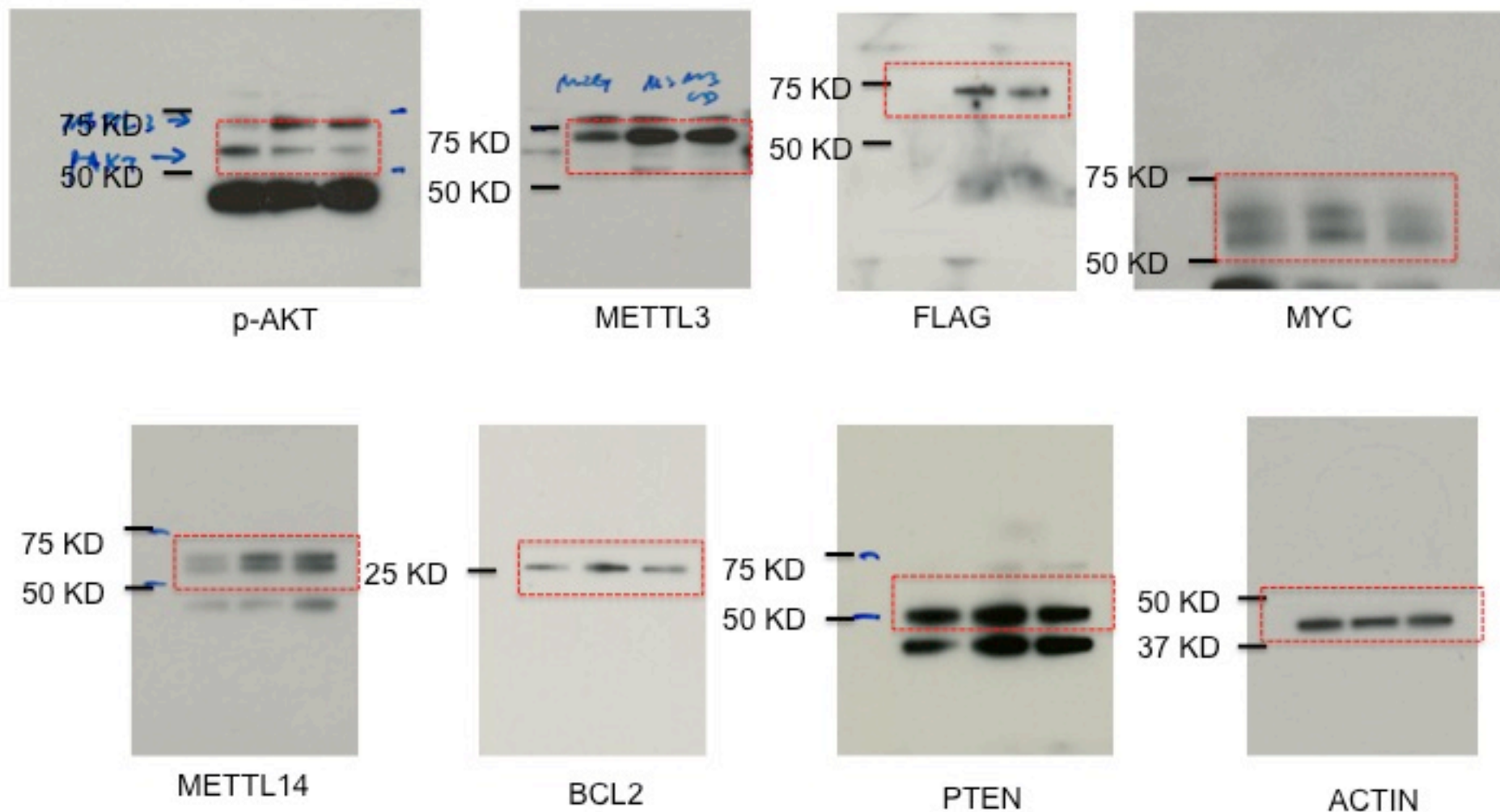


sgRNA-3days after transduction

Figure 4



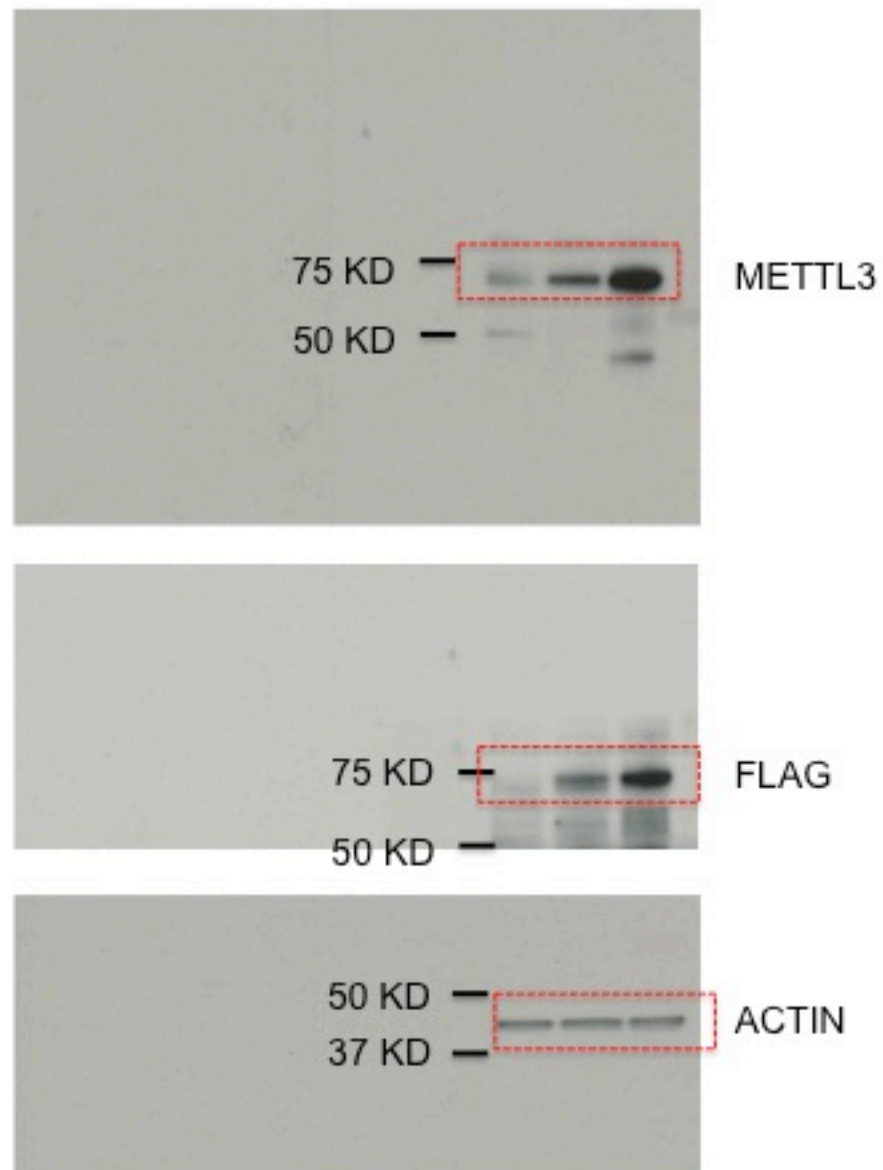
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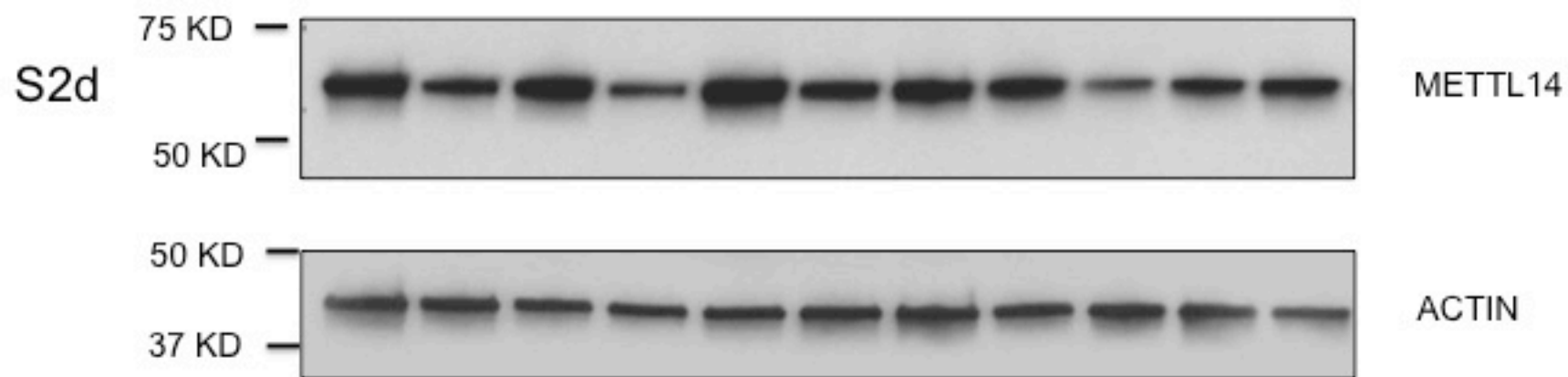


Overexpression of METTL3  
and METTL3-CD

Figure 4

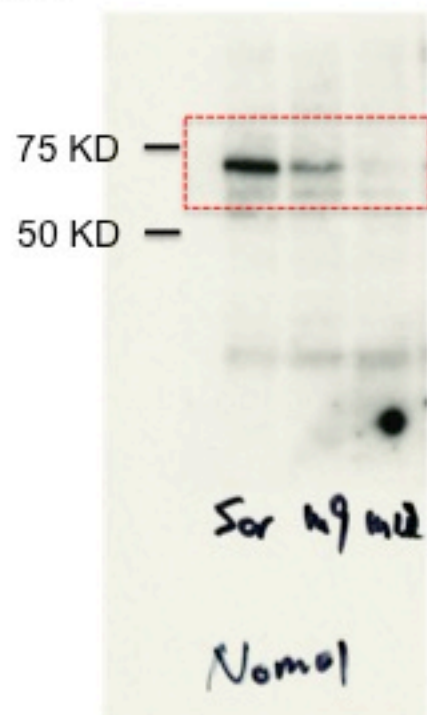
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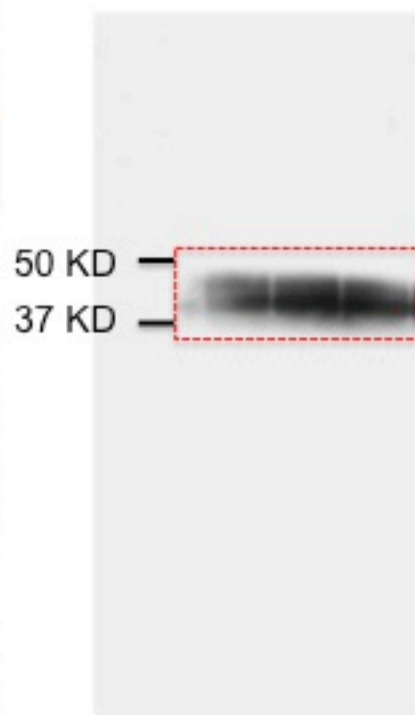


Supplementary Fig. 2

S2h



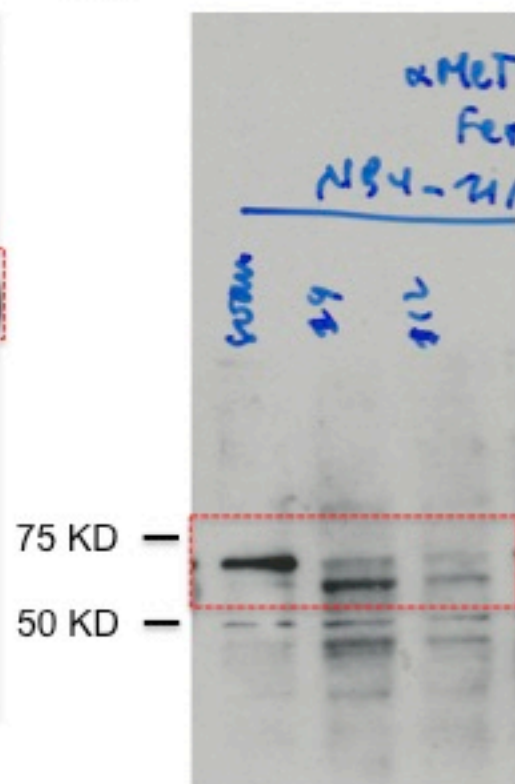
METTL3



ACTIN

NOMO-1 cells

S2I



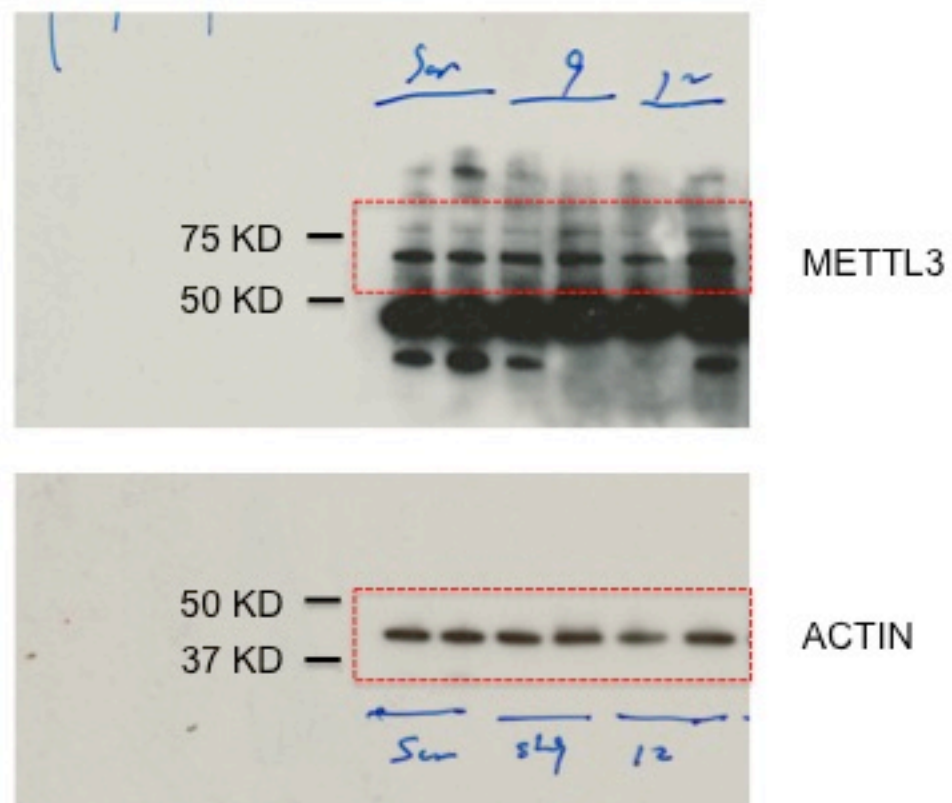
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NB4 cells



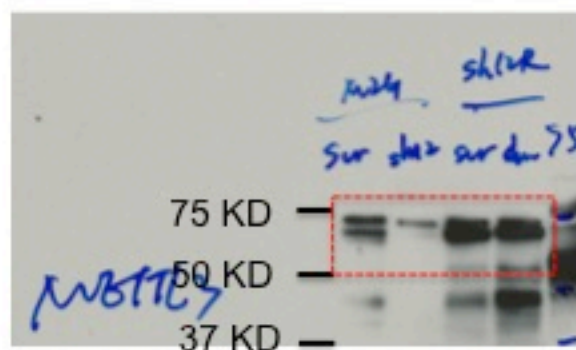
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S2p

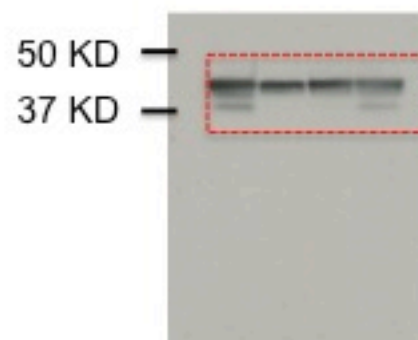


Supplementary Fig. 2

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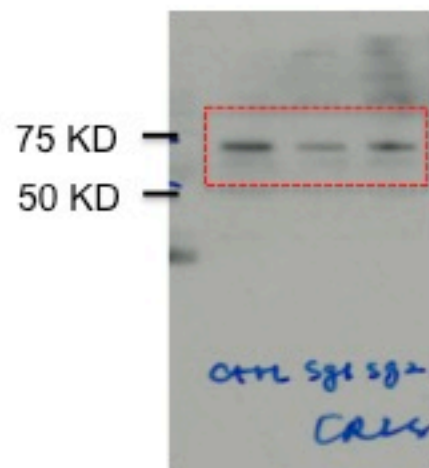


METTL3

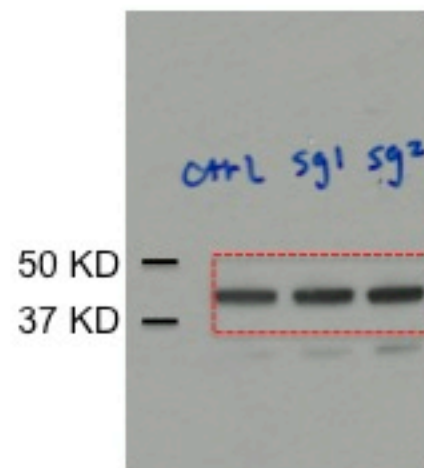


ACTIN

S3g



METTL3



ACTIN

S5c

