

SUPPLEMENTAL MATERIAL

Peterson et al., <https://doi.org/10.1084/jem.20170347>

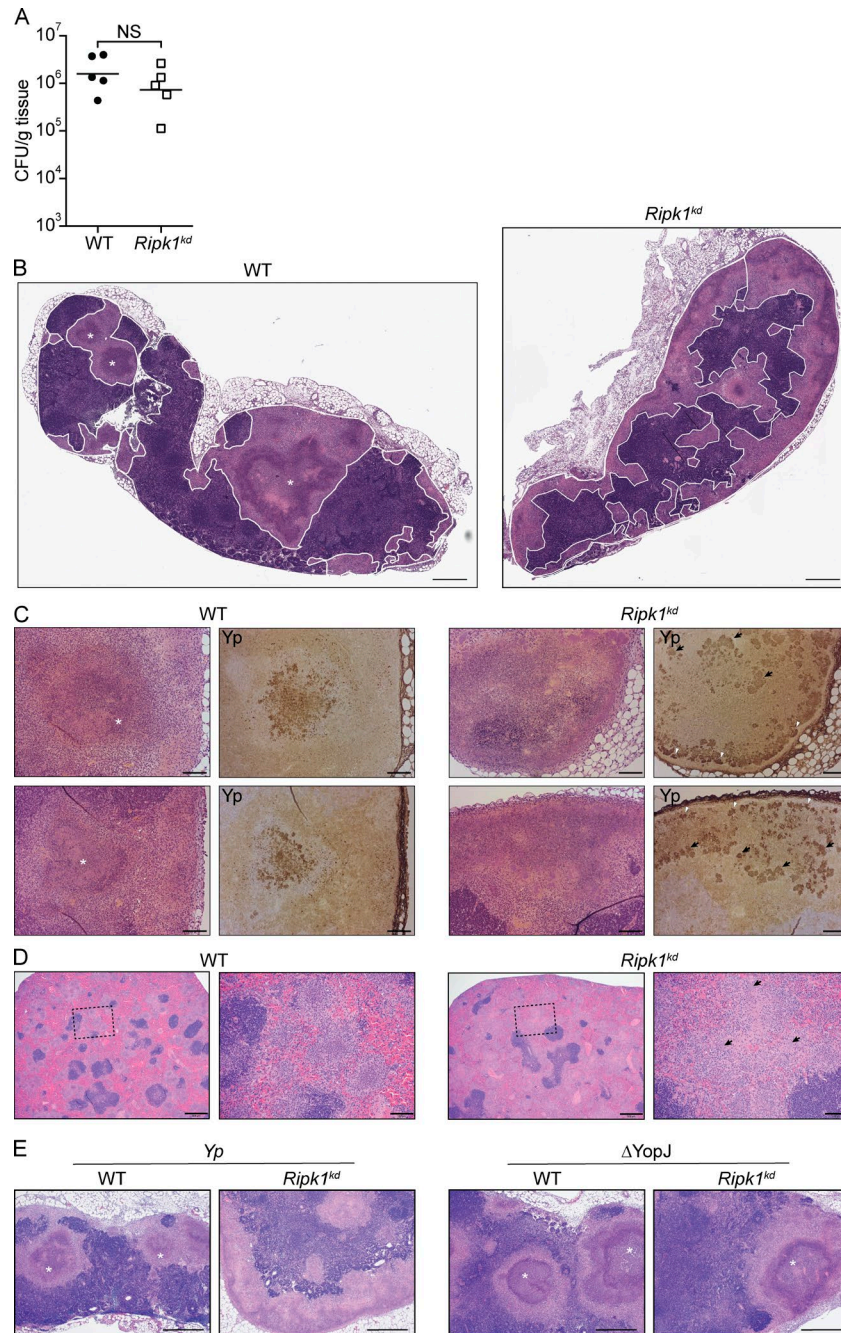


Figure S1. **RIPK1 kinase activity limits tissue pathology during in vivo *Yersinia* infection.** (A) WT and *Ripk1<sup>kd</sup>* mice were infected with  $2 \times 10^4$  CFUs *Yp* by intraperitoneal injection and splenic bacterial burdens were measured on day 3 postinfection. (B) H&E staining of MLN sections from WT and *Ripk1<sup>kd</sup>* mice infected with *Y. pseudotuberculosis* (*Yp*) were used to quantify percentage effacement. Bars, 500  $\mu$ m. Total area and effaced areas were traced on ImageJ software by a pathologist blinded to sample identification. Results are shown in Fig. 2 G. (C) Representative H&E and immunohistochemical staining for *Yersinia* from WT and *Ripk1<sup>kd</sup>* MLNs. In WT mice (left), bacteria are primarily located in pyogranulomas, characterized by central cores of cell debris surrounded by dense cuffs of neutrophils and epithelioid macrophages (asterisks). In *Ripk1<sup>kd</sup>* mice (right), many bacterial colonies are scattered throughout areas lymphoid tissue effacement (arrows) and border the subcapsular sinus (arrowheads). Bars, 100  $\mu$ m. (D) Representative H&E staining of sections from WT and *Ripk1<sup>kd</sup>* spleens, with increased numbers of bacterial colonies present in infected *Ripk1<sup>kd</sup>* spleens (arrows). Bars: (left) 500  $\mu$ m; (right, magnified image from area designated by dotted box) 100  $\mu$ m. (E) Representative H&E staining of sections from MLN of WT and *Ripk1<sup>kd</sup>* mice infected with wild-type (*Yp*) or YopJ-deficient ( $\Delta$ YopJ) *Yersinia*. While WT mice infected with either *Yp* or  $\Delta$ YopJ and *Ripk1<sup>kd</sup>* mice infected with  $\Delta$ YopJ contain bacteria within pyogranulomas (white asterisks), *Ripk1<sup>kd</sup>* mice infected with *Yp* fail to contain bacteria and have scattered bacterial colonies throughout the MLN. Bars, 500  $\mu$ m. Data in all panels are representative of two independent experiments. NS, not significant by Student's *t* test.

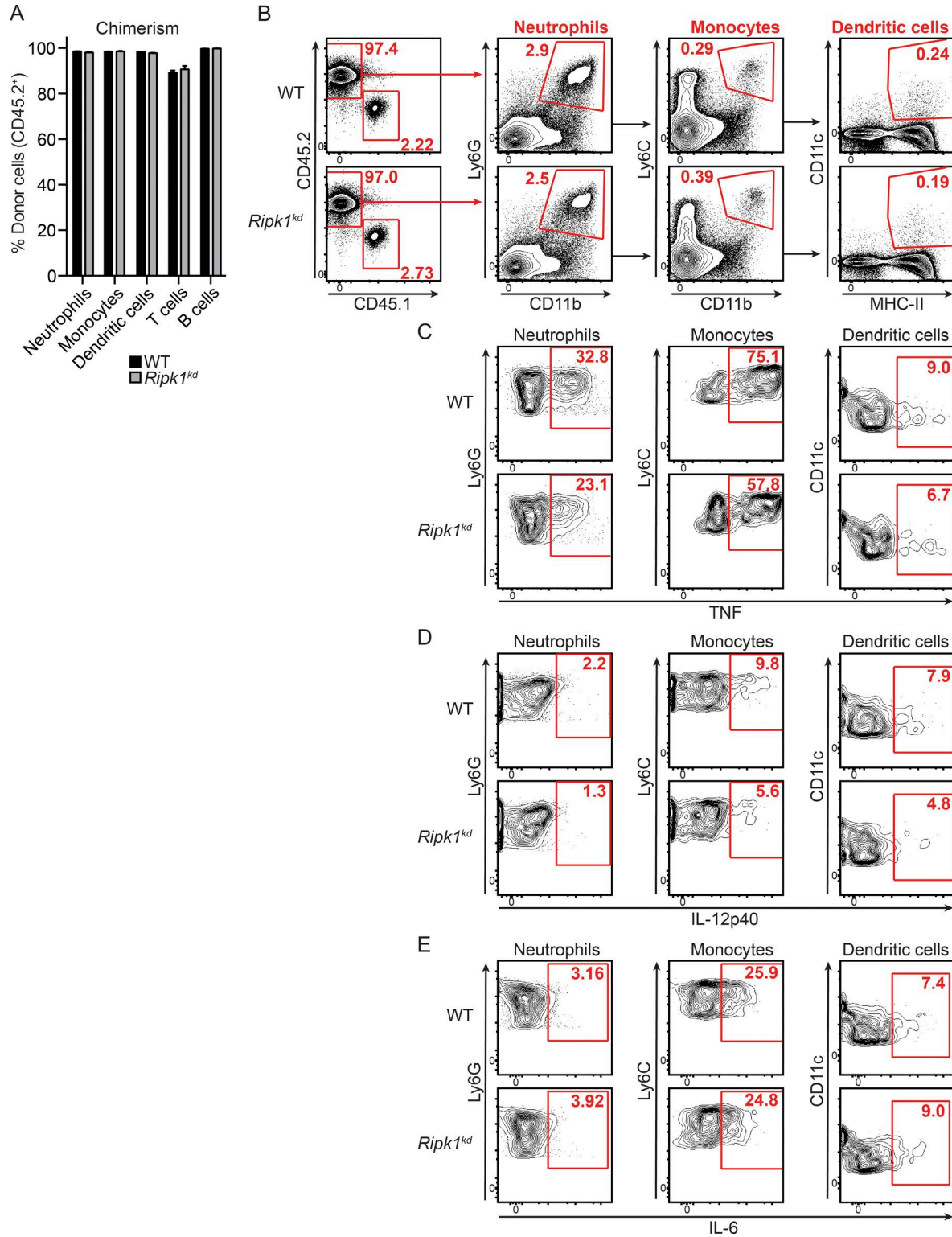


Figure S2. **Gating strategy for flow cytometry and BM chimerism.** Flow cytometry was performed on MLN samples from WT and *Ripk1<sup>kd</sup>* BM chimeras on day 5 postinfection by oral gavage with  $1-2 \times 10^8$  CFUs *Y. pseudotuberculosis*. (A and B) Singlet, live cells were gated to identify the percentage of donor BM-derived (CD45.2<sup>+</sup>) neutrophils (CD11b<sup>hi</sup>Ly6G<sup>hi</sup>), monocytes (CD11b<sup>hi</sup>Ly6C<sup>hi</sup>Ly6G<sup>-</sup>), and DCs (MHC-II<sup>+</sup>CD11c<sup>+</sup>). Data are representative of three independent experiments ( $n = 7$  mice per group). (C-E) TNF (C), IL-12p40 (D), and IL-6 (E) production by gated cell populations was measured by intracellular cytokine staining. Flow plots are representative of data shown in Fig. 3 (E-G) and consistent across three independent experiments.

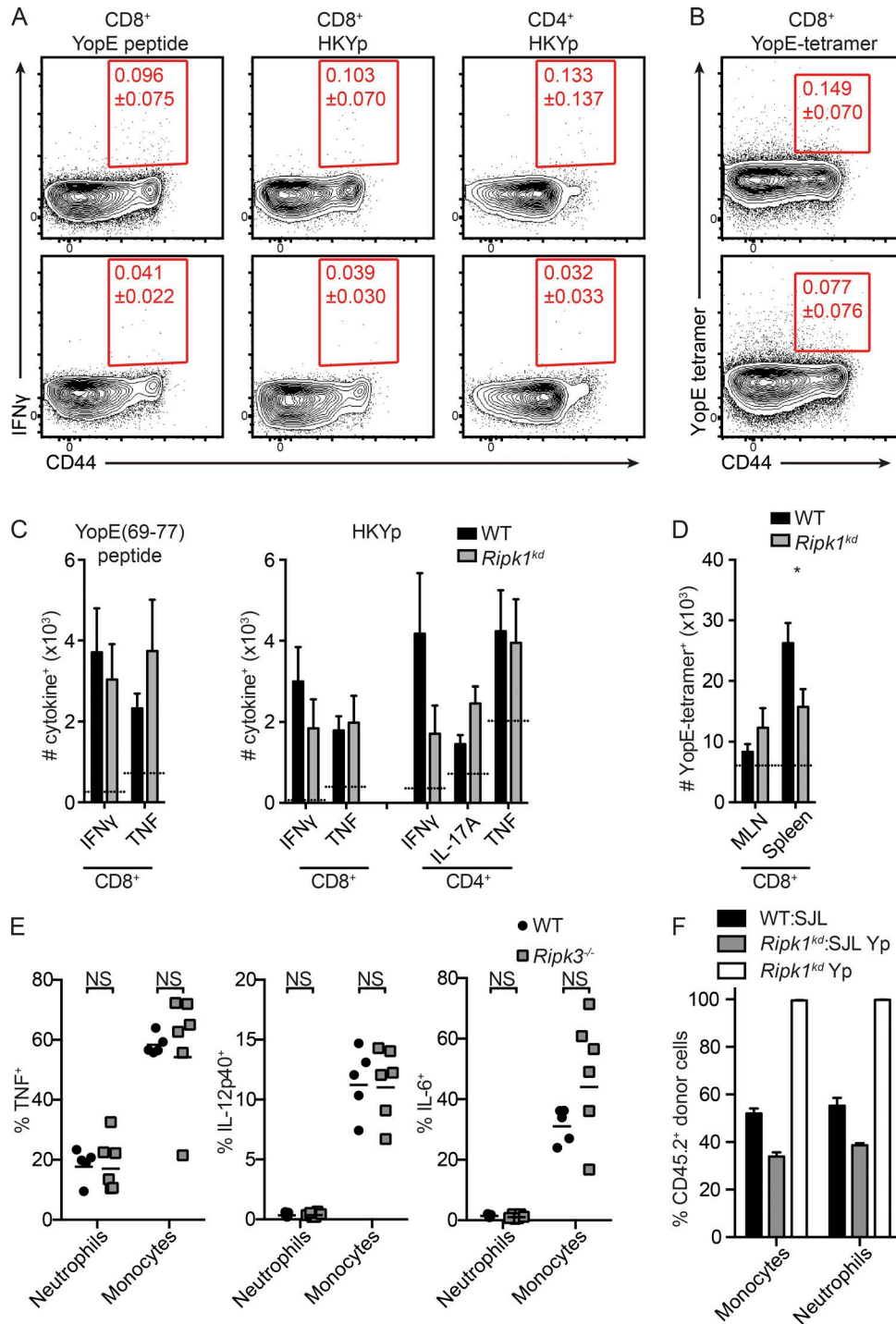


Figure S3. **RIPK1-dependent cytokine production in response to *Yp* infection is independent of adaptive immunity and RIPK3.** (A–D) T cell response on day 5 postinfection. (A) Representative flow plots showing intracellular cytokine staining for IFN- $\gamma$  produced by CD8<sup>+</sup> T cells stimulated with YopE<sub>69–77</sub> peptide and both CD8<sup>+</sup> and CD4<sup>+</sup> T cells stimulated with heat-killed *Yp* (HKYp). (B) Representative flow plots showing YopE<sub>69–77</sub>-tetramer (Lin et al., 2011)-positive CD8<sup>+</sup> T cells. (C and D) Total number of cells gated as in A and C and similarly for TNF or IL-17A production. Numbers in flow plots and bar graphs show mean and SD. Data are representative of two independent experiments ( $n = 6–7$  animals per group). Statistical differences determined by Student's *t* test. \*,  $P < 0.05$ . (E) WT and *Ripk3*<sup>-/-</sup> mice were infected with  $2 \times 10^8$  CFUs *Y. pseudotuberculosis* by oral gavage, and cytokine expression by MLN neutrophils and monocytes was measured on day 5 postinfection. Line represents the mean. Data are representative of three independent experiments. NS, not significant by Student's *t* test. (F) Percentage of CD45.2<sup>+</sup> donor BM-derived neutrophils and monocytes. Data are representative of three independent experiments ( $n = 7$  mice per group).

## REFERENCE

Lin, J.S., E.M. Szaba, L.W. Kummer, B.A. Chromy, and S.T. Smiley. 2011. *Yersinia pestis* YopE contains a dominant CD8 T cell epitope that confers protection in a mouse model of pneumonic plague. *J. Immunol.* 187:897–904. <http://dx.doi.org/10.4049/jimmunol.1100174>