

## SUPPLEMENTARY INFORMATION

**Supplementary Table 1. Primary and secondary antibodies used in this study.**

Antibodies	Species	Company	Catalog number	Dilution
<i>Primary</i>				
<i>Oct3/4</i>	Rabbit	MBL	PM048	1/200
<i>Sox2</i>	Goat	R&D Systems	AF2018	1/100
<i>GATA6</i>	Goat	R&D Systems	AF1700	1/100
<i>RFP</i>	Rabbit	MBL	PM005	1/100
<i>EGFP</i>	Rabbit	MBL	#598	1/100
<i>EGFP</i>	Rat	Nacalai	#04404-84	1/100
<i>Lefty</i>	Goat	R&D Systems	AF746	1/50
<i>Cer11</i>	Rat	R&D Systems	225807	1/50
<i>Nodal</i>	Rabbit	Abcam	Ab39953	1/50
<i>Secondary</i>				
Alexa 488 anti-rabbit IgG	Donkey	Molecular Probes	A21206	1/200
Alexa 568 anti-rat IgG	Donkey	Molecular Probes	A11057	1/200
Alexa 594 anti-rat IgG	Donkey	Molecular Probes	A21209	1/200
Alexa 633 anti-goat IgG	Donkey	Molecular Probes	A21082	1/200

**Supplementary Table 2. The numbers of PrE cells and DVE cells in *Lefty* mutant embryos.**

**(a)**

Genotype	DAPI (cells)	PrE (cells)	<i>Lefty1(mVenus)</i> (cells)
<i>Lefty1</i> <sup>+/+</sup>	125	40	4
	136	40	4
	134	41	5
<i>Lefty1</i> <sup>+/-</sup>	131	36	4
	145	49	7
	132	48	7
	99	29	5
	132	43	7
	128	37	10
<i>Lefty1</i> <sup>-/-</sup>	117	34	9
	130	37	6
	138	48	16
	100	35	3
	113	51	10
	116	24	4

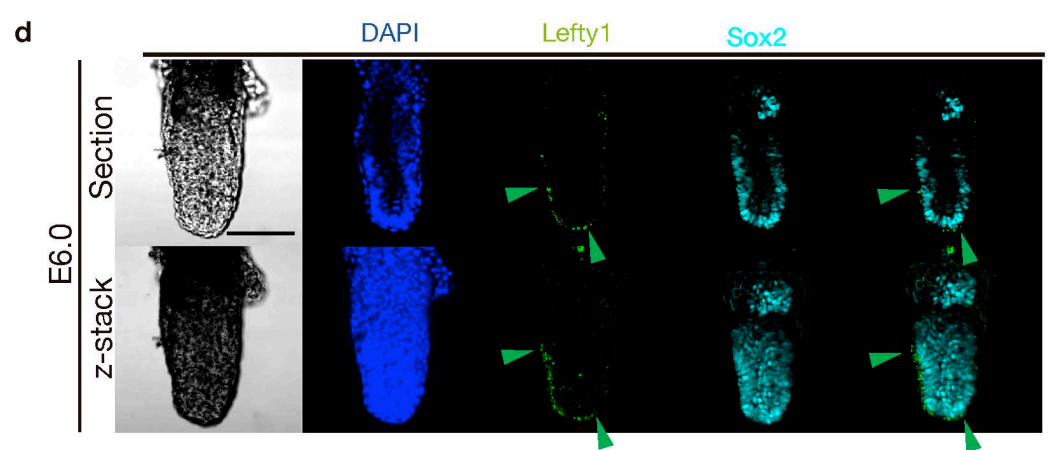
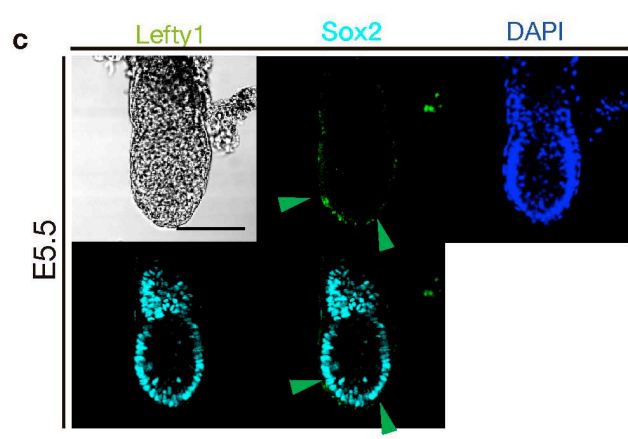
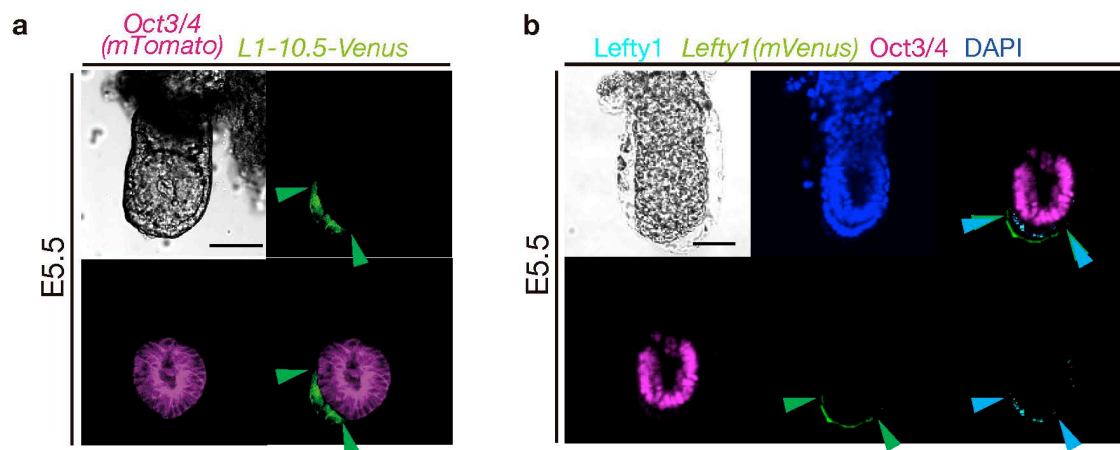
(b)

Genotype	DAPI (cells)	PrE (cells)	<i>Lefty1(mVenus)</i> (cells)
<i>Lefty1,2</i> <sup>+/+</sup>	97	26	3
	146	45	5
	128	34	5
	94	23	3
	133	29	4
	132	29	6
	112	34	3
<i>Lefty1,2</i> <sup>+/-</sup>	145	41	9
	149	48	10
	138	40	6
	146	46	9
	88	24	6
	123	33	6
	92	19	4
	137	36	5
	101	29	7
	104	28	5
	131	34	5
	143	38	4
	144	32	7
	131	44	7
	117	31	7
	143	44	9
	143	46	5
	137	43	5
	153	47	7
	135	37	7
	137	36	8
	184	54	12
	176	44	12
	170	46	13
146	31	6	
110	32	4	
<i>Lefty1,2</i> <sup>-/-</sup>	147	43	16
	122	38	14
	148	33	16
	147	46	13
	101	30	7
	138	40	15

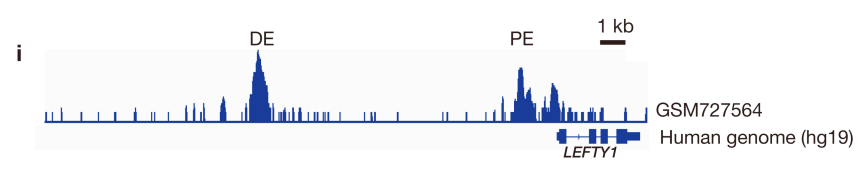
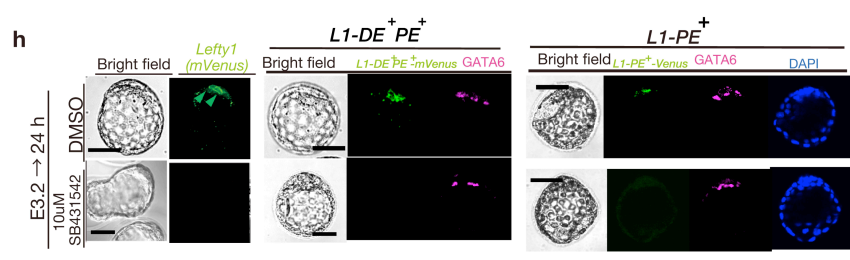
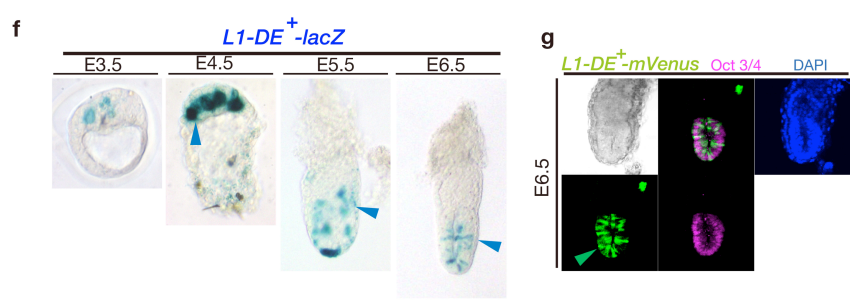
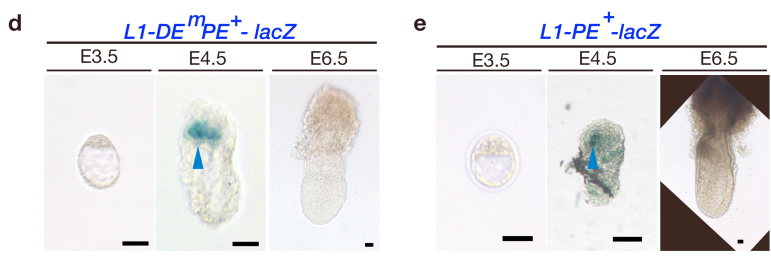
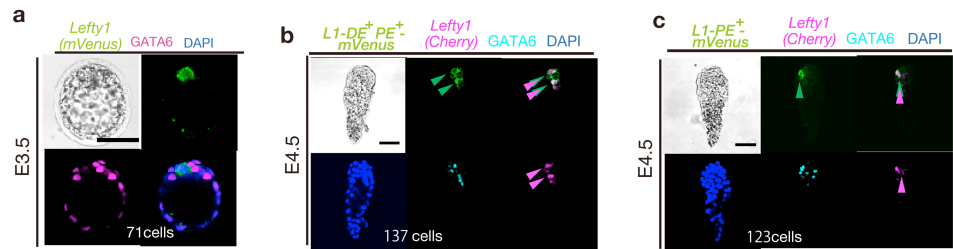
(c)

Genotype	DAPI+ (cells)	oct3/4+ (cells)
<i>Lefty1,2</i> <sup>+/+</sup>	149	58
	122	39
<i>Lefty1,2</i> <sup>+/-</sup>	125	30
	124	48
	136	42
	151	32
	123	43
	148	30
	124	45
<i>Lefty1,2</i> <sup>-/-</sup>	135	43
	132	40
	117	31
	130	44

Footnotes to Table 2: The numbers of DAPI<sup>+</sup> cells, PrE cells, mVenus<sup>+</sup> and Oct3/4<sup>+</sup> cells are summarized for individual embryos with the indicated genotype. The Table corresponds to Figure 4.

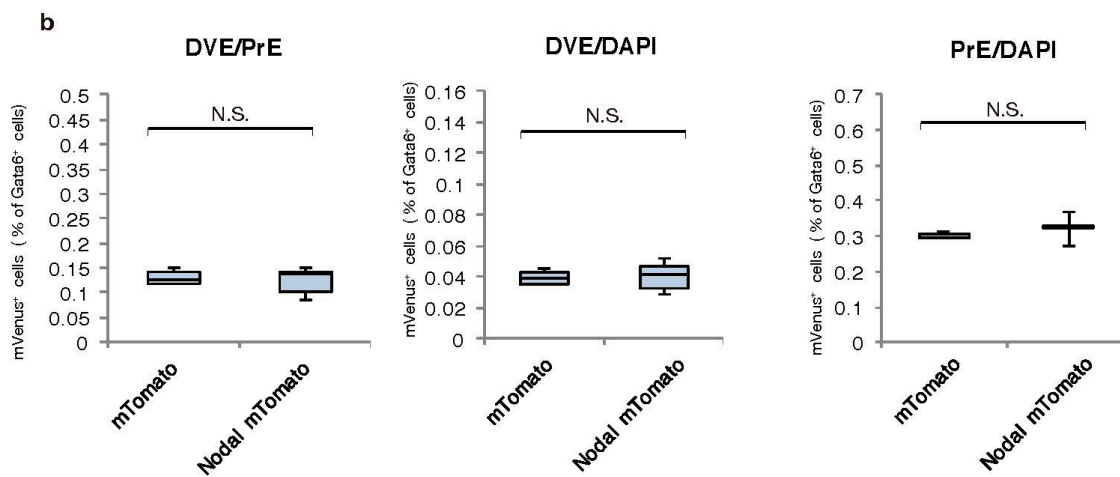
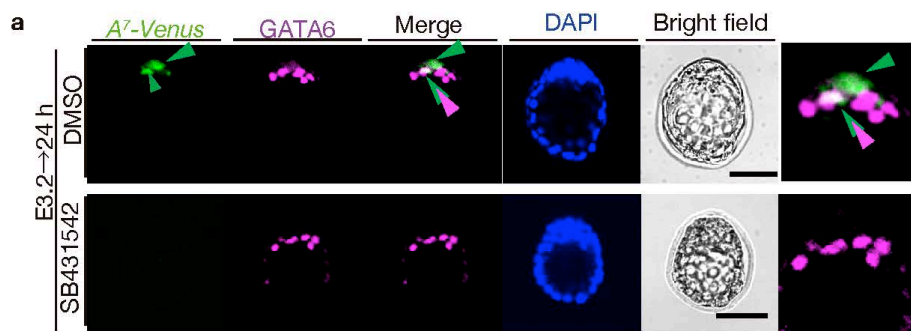


**Supplementary Figure 1. Epiblast does not contribute to DVE.** (a) An E5.5 mouse embryo harboring *LI-10.5-Venus* and *Oct3/4(mTomato)* BAC transgenes were examined for Venus and mTomato immuno-fluorescence. Note that epiblast cells (marked with mTomato expressed under the control of the *Oct3/4* promoter) do not contribute to DVE (labeled with Venus expressed under the control of the 10.5-kb upstream region of *Lefty1*). Arrowheads indicate DVE region. (b) An E5.5 embryo harboring *Lefty1(mVenus)* was examined for Venus, endogenous *Lefty1* and *Oct3/4* immuno-fluorescence. Note that *Lefty1*<sup>+</sup> cells and *Oct3/4*<sup>+</sup> cells are segregated. (c) An E5.5 wild-type embryo was stained for endogenous *Lefty1* and *Sox2*. (d) An E6.0 wild-type embryo was immunostained for endogenous *Lefty1* and *Sox2*. *Sox2*<sup>+</sup> cells are negative for *Lefty1*. All scale bars, 200  $\mu$ m.



**Supplementary Figure 2. *Lefty1* expression in L1<sup>epi</sup> cells and L1<sup>dve</sup> cells is regulated by Nodal-Foxh1 signaling.** (a) An E3.5 embryo harboring a *Lefty1(mVenus)* BAC transgene was examined for mVenus and GATA6 expression by immunofluorescence analysis. Scale bar, 50  $\mu$ m. Note that the mVenus<sup>+</sup> cell (L1<sup>epi</sup> cell) is negative for GATA6. (b) An E4.5 embryo harboring *LI-DE<sup>+</sup>PE<sup>+</sup>-mVenus* and *Lefty1(Cherry)* BAC transgenes was examined for mVenus, Cherry, and GATA6 immunofluorescence. Note that Venus<sup>+</sup> cells at this stage (L1<sup>dve</sup> cells) are positive for Cherry and GATA6 (arrowheads). Scale bar, 50  $\mu$ m. (c) An E4.5 embryo harboring *LI-PE<sup>+</sup>-mVenus* and *Lefty1(Cherry)* BAC transgenes was examined for mVenus, Cherry, and GATA6 immunofluorescence. Note that mVenus<sup>+</sup> cells at this stage (L1<sup>dve</sup> cells) are positive for Cherry and GATA6. Scale bar, 50  $\mu$ m. (d–f) Embryos harboring *LI-DE<sup>m</sup>PE<sup>+</sup>-lacZ* (d), *LI-PE<sup>+</sup>-lacZ* (e), or *LI-DE<sup>+</sup>-lacZ* (f) were stained with X-gal at the indicated stages. Arrowheads indicate X-gal + cells in EPI. (g) An E6.5 embryo harboring *LI-DE<sup>+</sup>Venus* was examined for Venus and endogenous *Oct3/4(mTomato)*. Note that Venus<sup>+</sup> cells are positive in EPI cells (Oct3/4<sup>+</sup>). Arrowheads indicate Venus + cells in EPI. (h) E3.2 embryos harboring *Lefty1(mVenus)*, *LI-DE<sup>+</sup>PE<sup>+</sup>-mVenus*, or *LI-PE<sup>+</sup>-Venus* transgenes were cultured for 24 h with 10  $\mu$ M SB431542 (Sigma #S4317) or dimethyl sulfoxide (DMSO) vehicle and were then examined for GATA6 as well as mVenus or Venus immune-fluorescence. Arrowheads indicate transgene expression in L1<sup>epi</sup> cells of control embryos. Scale bars, 50  $\mu$ m. (i) Localization of Foxh1 binding sites in the *LEFTY1* gene of human embryonic stem cells by ChIP-seq analysis with antibodies to Foxh1<sup>#1</sup>. The Foxh1 binding peaks are located at the DE and PE regions.

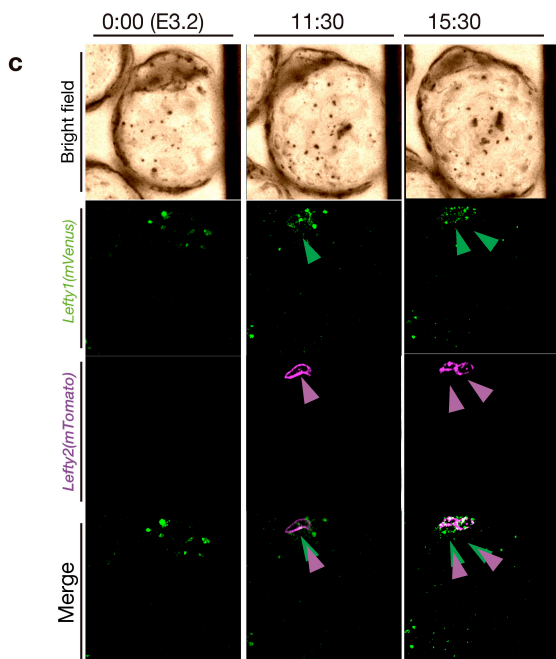
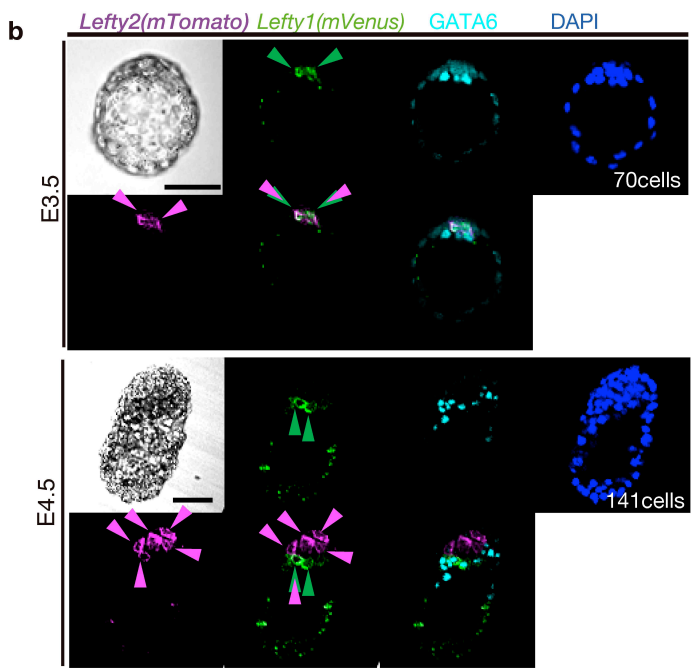
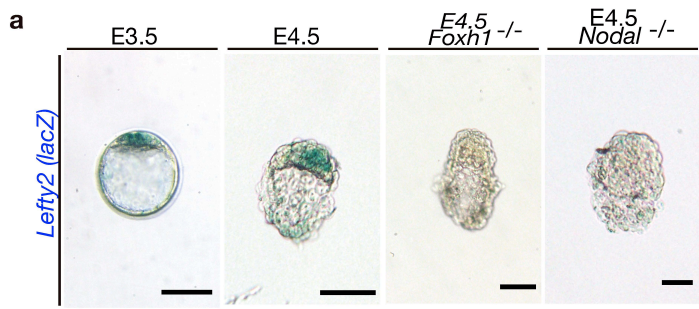




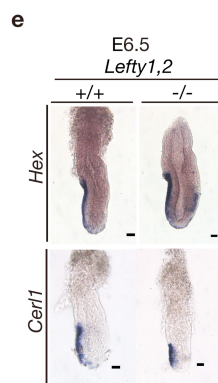
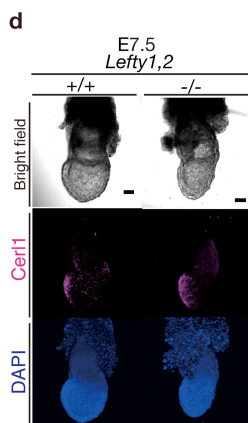
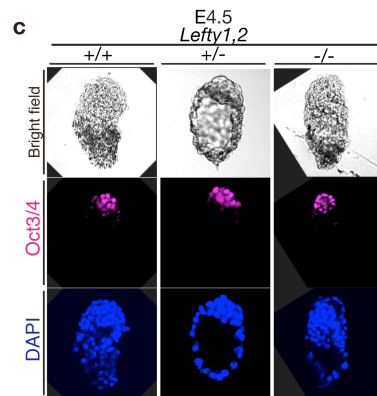
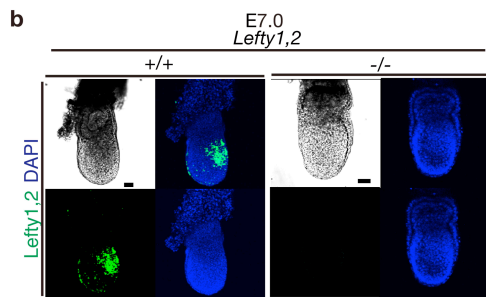
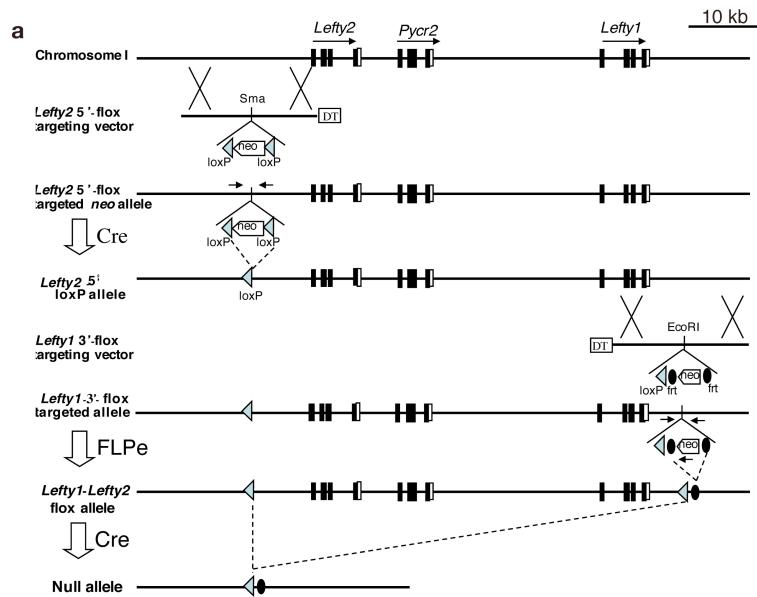
**c**

injected mRNA	DAPI(cells)	PrE (cells)	<i>Lefty1(mVenus)</i> (cells)
<i>mTomato</i>	142	42	5
	118	35	5
	131	40	6
	143	42	5
	128	40	5
<i>Nodal mRNA + mTomato mRNA</i>	135	50	7
	140	46	4
	154	50	5
	129	42	6
	121	33	5

**Supplementary Figure 3. Nodal-Foxh1 signaling activity as revealed with the *A<sub>7</sub>-Venus* transgene.** (a) E3.2 embryos harboring the *A<sub>7</sub>-Venus* transgene were cultured for 24 h in the presence of 10  $\mu$ M SB431542 or DMSO vehicle and were then subjected to immunofluorescence staining for Venus and GATA6 as well as to staining of nuclei with DAPI. Venus<sup>+</sup> cells in the control embryo are either GATA6<sup>+</sup> or GATA6<sup>-</sup> (arrowheads), presumably corresponding to L1<sup>dve</sup> and L1<sup>epi</sup> cells, respectively. Note that Venus expression is abolished by SB431542. Scale bars, 50  $\mu$ m. (b) Data are presented as box-and-whisker plots (first and third quartile, the line represents the median; whiskers: minimum to maximum). The number of DVE cells as a percentage of PrE cells, the number of DVE cells as a percentage of total cells and the number of PrE cells as a percentage total cells are summarized for embryos harboring a *Lefty1(mVenus)* BAC transgene, injected with *mTomato* mRNA with or without *Nodal* mRNA. NS, not significant (*t*-test).. (c) the numbers of DAPI<sup>+</sup> cells, PrE cells, and mVenus<sup>+</sup> cells are summarized for *Nodal* mRNA-injected and *mTomato* mRNA-injected embryos.



**Supplementary Figure 4. *Lefty2* expression overlaps in part with that of *Lefty1* and is regulated by Nodal-Foxh1 signaling.** (a) Expression of *Lefty2* as monitored with a *Lefty2(lacZ)* BAC transgene. *Lefty2* was expressed in the ICM at E3.5 and in the epiblast at E4.5. Such expression of the transgene was lost in *Foxh1*<sup>-/-</sup> or *Nodal*<sup>-/-</sup> embryos. Scale bars, 50 μm. (b) Embryos harboring both *Lefty1(mVenus)* and *Lefty2(mTomato)* BAC transgenes were examined for mVenus, mTomato, and GATA6 expression at E3.5 and E4.5. Note that *Lefty1* and *Lefty2* are expressed in the same cells at E3.5 (arrowheads indicated mTomato + cells in magenta and mVenus+ cells in green). At E4.5, *Lefty1* is expressed in GATA6<sup>+</sup> cells whereas *Lefty2* is expressed mostly in GATA6<sup>-</sup> cells. Scale bars, 50 μm. (c) An E3.2 embryo harboring both *Lefty1(mVenus)* and *Lefty2(mTomato)* BAC transgenes was cultured for 15.5 h. Fluorescence of mVenus and mTomato was recorded at the indicated times. Note that *Lefty1* and *Lefty2* are coexpressed at the indicated times in a subset of ICM cells (arrowheads).



**Supplementary Figure 5. Generation of *Lefty1,2*<sup>-/-</sup> mice and development of the A-P axis in such embryos at E6.5 and E7.5.** (a) Strategy for generation of a *Lefty1,2* double-mutant allele. (b) *Lefty1,2*<sup>+/+</sup> and *Lefty1,2*<sup>-/-</sup> embryos at E7.0 were stained with antibodies that recognize both Lefty1 and Lefty2 proteins. Note that Lefty proteins are absent in the *Lefty1,2*<sup>-/-</sup> embryo. Scale bars, 50 μm. (c) *Lefty1,2*<sup>+/+</sup>, *Lefty1,2*<sup>+/-</sup>, and *Lefty1,2*<sup>-/-</sup> embryos at E4.5 were stained for Oct3/4. Note that Oct3/4<sup>+</sup> cells are maintained in the *Lefty1,2*<sup>-/-</sup> embryo. (d) *Lefty1,2*<sup>+/+</sup> and *Lefty1,2*<sup>-/-</sup> embryos at E7.5 were subjected to immunofluorescence staining for Cer11 (a marker for AVE). Scale bars, 50 μm. (e) Expression of *Cer11* and *Hex* in E6.5 *Lefty1,2*<sup>+/+</sup> and *Lefty1,2*<sup>-/-</sup> embryos as determined by *in situ* hybridization. Scale bars, 50 μm.

### Supplementary References

#1. Kim, S.W. *et al.* Chromatin and transcriptional signatures for Nodal signaling during endoderm formation in hESCs. *Dev Biol* **357**, 492-504 (2011).