

## Supplementary Information

### Two independent modes of chromatin organization revealed by cohesin removal

Wibke Schwarzer, Nezar Abdennur, Anton Goloborodko, et al.

#### Supplementary Tables.

Supplementary Table 1 – Summary statistics of read pairs for TCC (Hi-C) libraries in this study.

Supplementary Table 2 – Statistics of Hi-C libraries in Schmitt et.al, 2016<sup>27</sup>, an example of a contemporary study on primary tissues.

Supplementary Table 3 – Statistics of Hi-C libraries in other cohesin depletion studies.

Supplementary Table 4 - The list of enriched GO terms for differentially expressed genes, with a Benjamini-Hochberg p-value < 0.05.

Supplementary Table 5 – Public data sources

#### Supplementary Data 1.

Original scans of western blots. The selected regions for the different panels shown on Fig.1b are boxed in red. Additional biological replicates of the two conditions can be seen (samples 1 and 4)

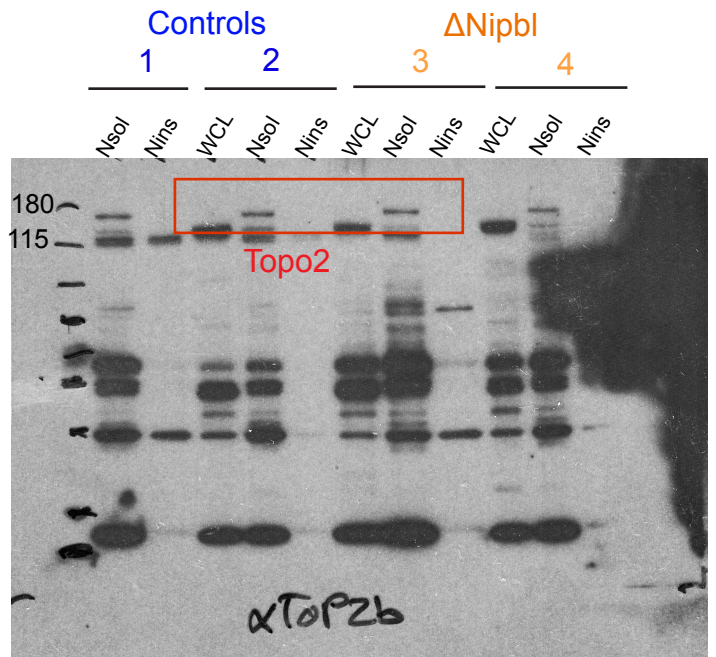
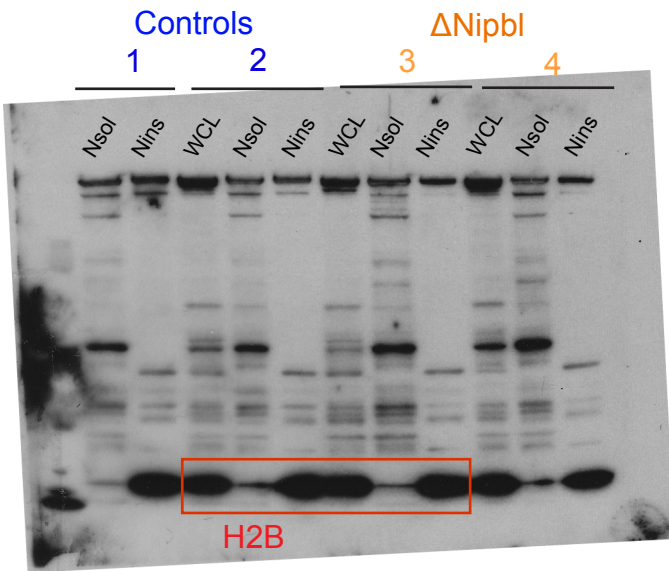
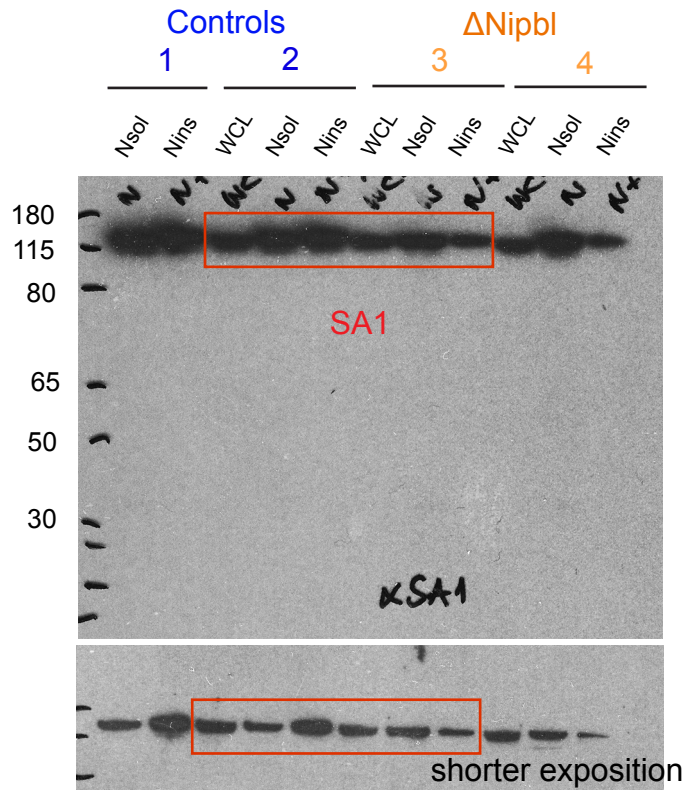
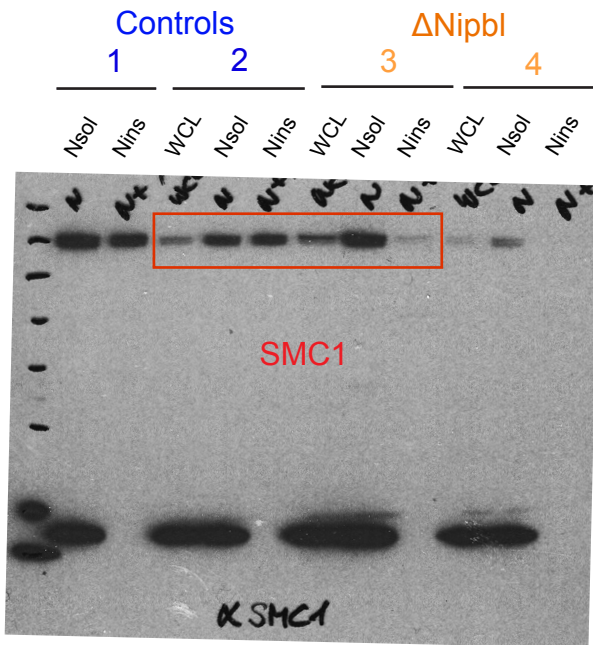
#### Supplementary Data 2.

Experimental Hi-C maps show extensive similarities between replicates. (a) The curves of contact frequency  $P(s)$  vs genomic distance  $s$ . (b) Pearson correlation coefficients between *cis* eigenvectors at 100kb. (c) The average Hi-C maps around 102 500kb-600kb peaks. (d) Compartmentalization saddle plots.

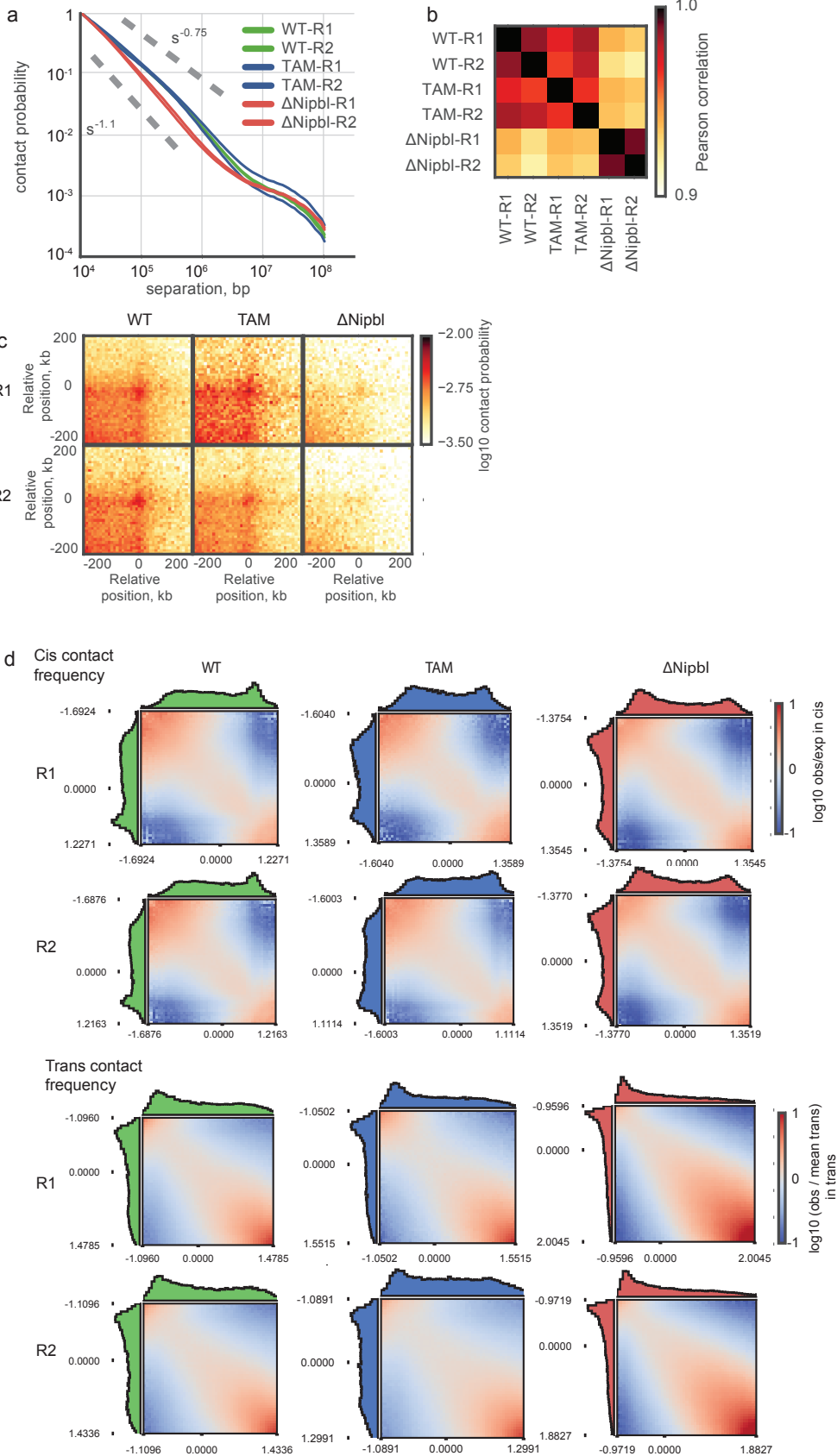
#### Supplementary Data 3.

Consistent differences in compartment eigenvector upon Nipbl deletion (a) The genome-wide *cis* eigenvectors at 100kb: WT (green), TAM (blue),  $\Delta$ Nipbl (red) are shown. (b) Zoom-in of the region shaded in grey in (a) using 20kb eigenvectors. In (a-b), the curves are shown for each of two replicates of each condition.

# Supplementary Data 1

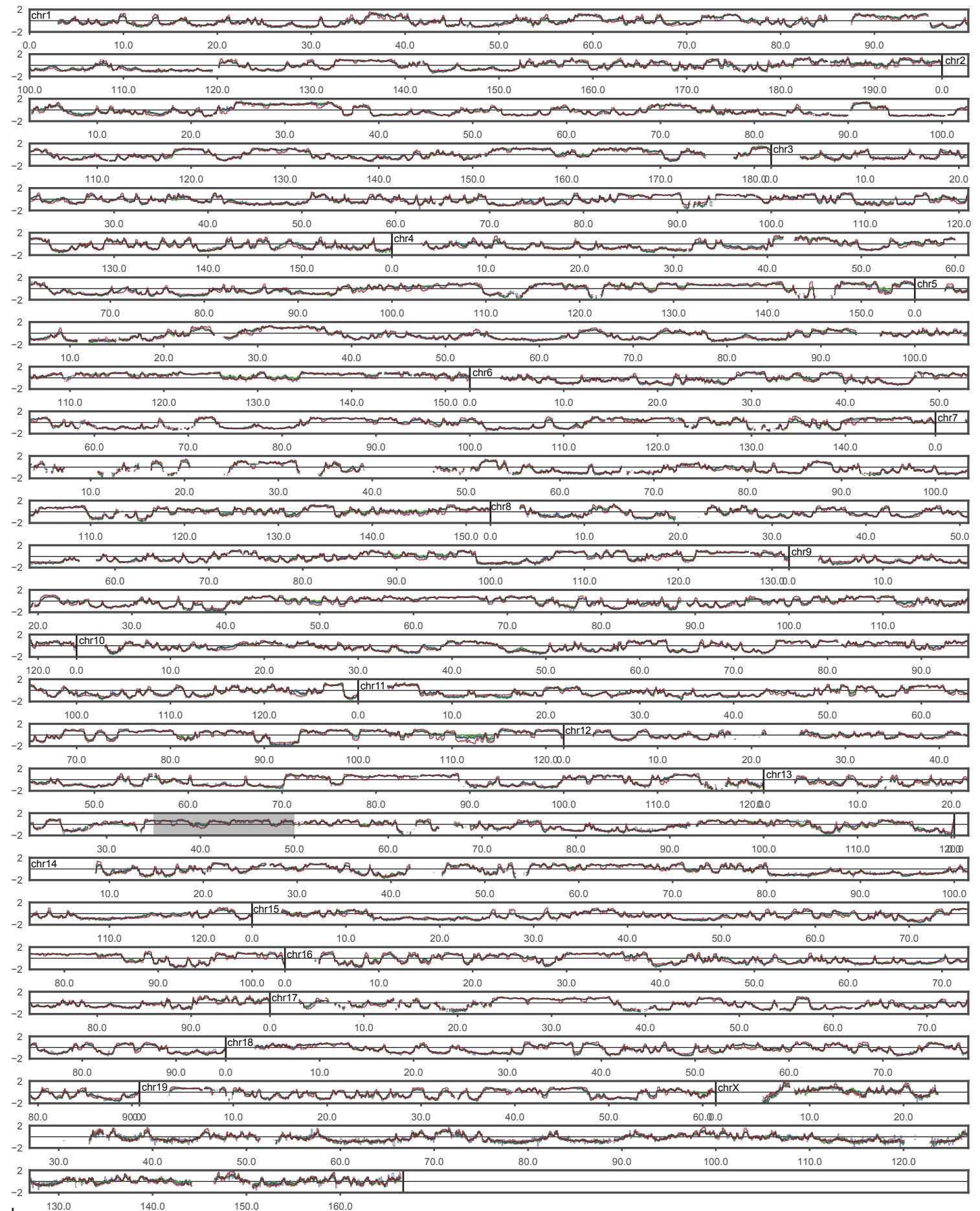


# Supplementary Data 2

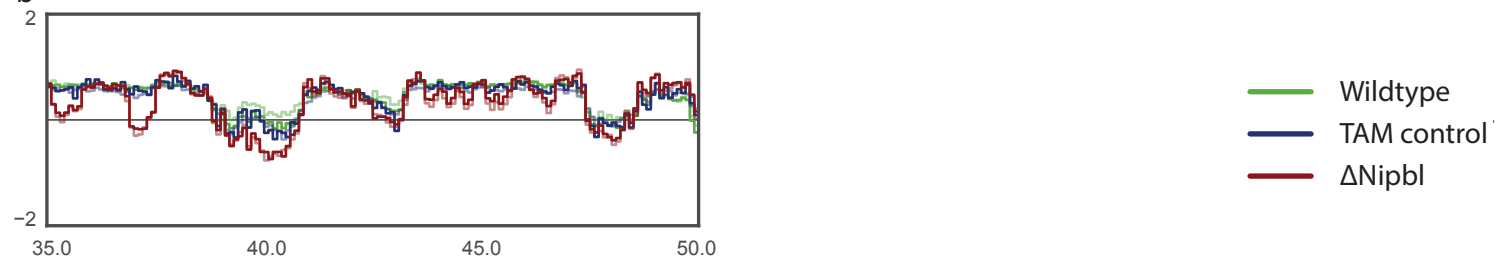


# Supplementary Data 3

a



b



**Supplementary Table 1 – Summary statistics of read pairs for TCC (Hi-C) libraries in this study.**

<b>Replicate</b>	<b>codename</b>	<b>Raw read pairs</b>	<b>Mapped read pairs</b>	<b>Filtered read pairs</b>	<b>Cis read pairs, separation &gt;10kb</b>
WT-R1	014637Untr	139,343,707	75,465,383	51,745,705	35,529,637
WT-R2	019440Untr	114,258,179	60,638,254	43,097,410	28,704,961
TAM-R1	014199TAMcontr	94,032,309	53,285,208	14,530,511	9,870,917
TAM-R2	014399TAMcontr	134,045,253	72,789,581	51,281,408	34,490,881
$\Delta$ Nipbl-R1	014200Nipbl	131,409,162	66,635,374	42,405,784	26,633,610
$\Delta$ Nipbl-R2	016350Nipbl	133,914,046	69,894,091	50,527,104	31,561,513

**Supplementary Table 2 – Statistics of Hi-C libraries in Schmitt et.al, 2016, an example of a contemporary study on primary tissues.**

<b>Study</b>	<b>Raw read pairs</b>	<b>Filtered pairs</b>	<b>Cis read pairs, separation &gt;10kb</b>
Bladder, R1	233,041,838	52,456,550	24,121,444
Bladder, R2	236,250,557	43,395,294	19,659,511
Adrenal Gland, R1	181,680,303	48,951,415	22,449,886
Psoas Muscle, R1	196,863,028	36,856,215	13,566,008
Psoas Muscle, R2	95,110,351	25,776,091	10,410,829
Pancreas, R1	160,416,883	49,627,891	21,200,091
Pancreas, R2	12,147,138	3,538,979	1,367,214
Pancreas, R3	128,745,528	15,683,831	7,787,867
Lung, R1	308,335,703	85,597,004	24,417,652
Hippocampus, R1	187,658,680	64,151,770	29,220,961
Right Ventricle, R1	672,319,838	179,219,920	63,091,486
Small Bowel, R1	177,864,628	58,017,249	21,073,296
Spleen, R1	378,529,216	107,599,392	33,032,593
Ovary, R1	201,215,217	70,283,904	20,909,766
Dorsolateral Prefrontal Cortex, R1	165,728,687	52,713,868	25,426,433

**Supplementary Table 3 – Statistics of Hi-C libraries in other cohesin depletion studies.**

<b>Study</b>	<b>Filtered read pairs</b>	<b>Cis read pairs, separation &gt;10kb</b>
Seitan et al, 2013, WT, R1	51,337,902	12,676,930
Seitan et al, 2013, WT, R2	96,784,574	28,679,223
Seitan et al, 2013, $\Delta$ Rad21, R1	57,060,666	14,219,809
Seitan et al, 2013, $\Delta$ Rad21, R2	96,784,574	22,594,749
Zuin et.al, 2014, Control, R1	120,366,592	48,221,154
Zuin et.al, 2014, Control, R2	134,277,024	46,378,835
Zuin et.al, 2014, Rad 21 cleaved, R1	113,394,855	41,623,810
Zuin et.al, 2014, Rad 21 cleaved, R2	159,090,030	51,870,950
Sofueva et.al, 2012, Control NSC, R1	16,886,965	12,761,787
Sofueva et.al, 2012, $\Delta$ Rad21 NSC Lox, R1	69,788,779	52,087,400
Sofueva et.al, 2012, $\Delta$ Rad21 NSC Lox, R2	63,347,539	47,760,318
Sofueva et.al, 2012, Control AST, R1	29,246,903	21,885,386
Sofueva et.al, 2012, $\Delta$ Rad21 AST Lox, R1	24,710,993	15,866,342
Sofueva et.al, 2012, $\Delta$ Rad21 AST Lox, R2	20,287,207	13,980,244

**Supplementary Table 4 - The list of enriched GO terms for differentially expressed genes, with a Benjamini-Hochberg p-value < 0.05.**

Term	Category	Name	Regulated	Count	%
GO:0060271	BP	cilium morphogenesis	up	16	3.6
GO:0070062	CC	extracellular exosome	up	107	23.8
GO:0042995	CC	cell projection	up	36	8.0
GO:0005929	CC	cilium	up	18	4.0
GO:0005930	CC	axoneme	up	10	2.2
GO:0032869	BP	cellular response to insulin stimulus	down	14	2.4
GO:0006629	BP	lipid metabolic process	down	35	6.0
GO:0005615	CC	extracellular space	down	83	14.0
GO:0016021	CC	integral component of membrane	down	231	39.1
GO:0005783	CC	endoplasmic reticulum	down	88	14.9
GO:0005576	CC	extracellular region	down	84	14.2
GO:0005789	CC	endoplasmic reticulum membrane	down	54	9.1
GO:0043231	CC	intracellular membrane-bounded organelle	down	48	8.1
GO:0016020	CC	membrane	down	273	46.2
GO:0016324	CC	apical plasma membrane	down	25	4.2
GO:0005887	CC	integral component of plasma membrane	down	51	8.6
GO:0020037	MF	heme binding	down	21	3.6
GO:0016491	MF	oxidoreductase activity	down	47	8.0
GO:0052689	MF	carboxylic ester hydrolase activity	down	11	1.9



**Supplementary Table 5 – Public data sources**

Data type	Description	Cell-type	Ref.	URL
Hi-C data	Hi-C raw reads	Mouse ESC, NSC, AST	18	<a href="https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE49018">https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE49018</a>
	Hi-C raw reads	Mouse Developing Thymocytes	19	<a href="https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE48763">https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE48763</a>
	Hi-C raw reads	Human HEK293T	17	<a href="https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE44267">https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE44267</a>
Peaks	Annotations from Hi-C	Mouse CH12-LX	8	<a href="ftp://ftp.ncbi.nlm.nih.gov/geo/series/GSE63nnn/GSE63525/suppl/GSE63525_CH12-LX_HiCCUPS_looplist.txt.gz">ftp://ftp.ncbi.nlm.nih.gov/geo/series/GSE63nnn/GSE63525/suppl/GSE63525_CH12-LX_HiCCUPS_looplist.txt.gz</a>
CTCF M1 motif	PWM for mouse		56	<a href="ftp://ftp.ebi.ac.uk/pub/databases/vertebrategenomics/FOG03/calls/CTCF_canonical_species">ftp://ftp.ebi.ac.uk/pub/databases/vertebrategenomics/FOG03/calls/CTCF_canonical_species</a>
Lamin-B1 DamID	Normalized signal	Mouse ESCs	37	<a href="http://hgdownload.cse.ucsc.edu/goldenPath/mm9/database/laminB1_AC.txt.gz">http://hgdownload.cse.ucsc.edu/goldenPath/mm9/database/laminB1_AC.txt.gz</a>
	Normalized signal	NPCs		<a href="http://hgdownload.cse.ucsc.edu/goldenPath/mm9/database/laminB1_EF.txt.gz">http://hgdownload.cse.ucsc.edu/goldenPath/mm9/database/laminB1_EF.txt.gz</a>
	Normalized signal	Astrocytes		<a href="http://hgdownload.cse.ucsc.edu/goldenPath/mm9/database/laminB1_ES.txt.gz">http://hgdownload.cse.ucsc.edu/goldenPath/mm9/database/laminB1_ES.txt.gz</a>
	Normalized signal	MEFs		<a href="http://hgdownload.cse.ucsc.edu/goldenPath/mm9/database/laminB1_NP.txt.gz">http://hgdownload.cse.ucsc.edu/goldenPath/mm9/database/laminB1_NP.txt.gz</a>
	HMM state calls		38	<a href="https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE17051">https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE17051</a>
ChromHMM	Chromatin state segmentation trained on ENCODE mouse ChIP-seq data	Mouse Liver, Adult8wks	55	<a href="https://github.com/gireeshkbogu/chromatin_states_chromHMM_mm9">https://github.com/gireeshkbogu/chromatin_states_chromHMM_mm9</a>
ENCODE Histone ChIP-seq	ChIP-seq, peak and bigwig files Input H3K27ac H3K27me3 H3K4me3 H3K4me1 H3K9me3 H3K36me3 H3K79me2	Mouse Liver, Adult8wks	45 and ENCODE	<a href="http://hgdownload.cse.ucsc.edu/goldenPath/mm9/encodeDCC/wgEncodeLicrHistone/">http://hgdownload.cse.ucsc.edu/goldenPath/mm9/encodeDCC/wgEncodeLicrHistone/</a>
ENCODE TFBS ChIP-seq	ChIP-seq, Peak and bigwig files Input PolII CTCF	Mouse Liver, Adult8wks	45 and ENCODE	<a href="http://hgdownload.cse.ucsc.edu/goldenPath/mm9/encodeDCC/wgEncodeLicrTfbs/">http://hgdownload.cse.ucsc.edu/goldenPath/mm9/encodeDCC/wgEncodeLicrTfbs/</a>
ENCODE DNase-seq	DNase I HS, bigwig	Mouse Liver, Adult8wks	45 and ENCODE	<a href="http://hgdownload.cse.ucsc.edu/goldenPath/mm9/encodeDCC/wgEncodeUwDnase/">http://hgdownload.cse.ucsc.edu/goldenPath/mm9/encodeDCC/wgEncodeUwDnase/</a>
ENCODE RNA-seq	Poly-A RNA-seq, bigwig	Mouse Liver, Adult8wks	45 and ENCODE	<a href="http://hgdownload.cse.ucsc.edu/goldenPath/mm9/encodeDCC/wgEncodeUwRnaSeq/">http://hgdownload.cse.ucsc.edu/goldenPath/mm9/encodeDCC/wgEncodeUwRnaSeq/</a>
	Poly-A RNA-seq, bigwig	Mouse Liver, Adult8wks		<a href="http://hgdownload.cse.ucsc.edu/goldenPath/mm9/encodeDCC/wgEncodeCshLongRnaSeq/">http://hgdownload.cse.ucsc.edu/goldenPath/mm9/encodeDCC/wgEncodeCshLongRnaSeq/</a>