

1 **Supplementary Information**

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3 **Intrinsic short-tailed azole resistance in mucormycetes is due to an**
4 **evolutionary conserved aminoacid substitution of the lanosterol 14 α -**
5 **demethylase.**

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23 **Table S1.** *In vitro* antifungal susceptibility of *Mucor circinelloides*,
 24 *Rhizopus microsporus*, and *Rhizopus arrhizus* against fluconazole and
 25 triadimenol.

Species (n. strains tested)	Median MIC (range) ^a (mg/L)	
	FLC	TDM
<i>R. arrhizus</i> (n=6)	>64.00 (>64.00)	>16.00 (>16.00)
<i>R. microsporus</i> (n=6)	>64.00 (>64.00)	>16.00 (>16.00)
<i>M. circinelloides</i> (n=6)	>64.00 (>64.00)	>16.00 (>16.00)

Table legend. ^a FLC and TDM susceptibility according to CLSI standard microbroth dilution method; MIC. minimum inhibitory concentration; FLC, fluconazole; TDM, triadimenol.

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29 **Table S2.** Accession numbers corresponding to fungal LDMs aligned with the CYP51 F5
 30 and CYP51 F1 consensus sequences of Mucorales species *Rhizopus arrhizus*, *Rhizopus*
 31 *microsporus*, and *Mucor circinelloides*.

species	LDM Genbank accession number
<i>Saccharomyces cerevisiae</i>	NP_011871
<i>Histoplasma capsulatum</i>	AAU01158.1
<i>Coccidioides posadasii</i>	KMM65971.1
<i>Coccidioides immitis</i>	XP_001246802.1
<i>Pneumocystis carinii</i>	AAO38776.1
<i>Scedosporium apiospermum</i>	KEZ44212.1
<i>Zymoseptoria tritici</i>	AKA94187.1
<i>Uncinula necator</i>	O14442*
<i>Aspergillus fumigatus</i>	ACF18581.1
<i>Aspergillus flavus</i>	XP_002375123
<i>Candida albicans</i>	ADO21804.1
<i>Candida glabrata</i>	XP_445876.1
<i>Candida tropicalis</i>	AMR44154.1
<i>Candida dubliniensis</i>	AAK57519.1
<i>Malassezia globosa</i>	XP_001730619.1
<i>Cryptococcus neoformans var. grubii</i>	AEX20237.1
<i>Cryptococcus gattii</i>	AEQ63274.1
<i>Mucor ambiguus</i> CYP51 F5/CYP51 F1	A0A0C9M5D0*/A0A0C9M5R4*
<i>Parasitella parasitica</i> CYP51 F5/CYP51 F1	A0A0B7NND2*/ A0A0B7NSN9*
<i>Absidia glauca</i> CYP51 F5/CYP51 F1	ABSGL_12517.1 scaffold 12955*/ABSGL_12606.1 scaffold 13066*

32 **Table legend.** LDM. lanosterol 14- α demethylase; * accession number retrieved from UniProt
 33 database (<http://www.uniprot.org/uniprot>).

Table S3. Conditions for the q-PCRs developed for CYP51 F5, CYP51 F1.

qPCR gene target	species	direction	primer sequence (5' to 3')	amplicon size (bp)	targeted triplet	spanned CDS region	translated aa	LDM location
CYP51 F5	<i>R. arrhizus</i>	For	GGACCCGACGGKAACCAATT	208	TTY	positions 385 to 387	phenylalanine	129
		Rev	ATTCCTCGATRATCAGCGGC					
	<i>R. microsporus</i>	For	AAGCGCGTCACTGCATGCCT	196				
		Rev	ACGACTCGATATTTAAYCCGCT					
	<i>M. circinelloides</i>	For	CTCTTGGGTYCCCATCATGG	202				
		Rev	GTTGTAAGCAGCAGCAGCAG					
CYP5 F1	<i>R. arrhizus</i>	For	TGAATGCAGACGGAAACCAA	247	TAY	positions 379 to 381	tyrosine	127
		Rev	CCCGTAGGCTTCTTGTAGTTTT					
	<i>R. microsporus</i>	For	ATATGGTTGGCCGACGTGTT	223				
		Rev	GGAACGTGTTGACGGAATGC					
	<i>M. circinelloides</i>	For	CCGTTCTCTTGGGTGCTGAT	203				
		Rev	GGGCACATGTTGACGRAARC					
Actin	all 3 species*	For	TCCTGGTATTGCGAYCGT	128	N.A	positions 388 to 390		130
		Rev	GAGARGCYAARATGGAACCA					

Table legend. The primers, amplicon sizes, targeted triplets, the regions where these are located and the position of each respective translated amino acid used to confirm the expression of both CYP51 genes are presented. The qPCR primers and amplicon sizes used for the actin as normalization gene are also depicted. qPCR, quantitative reverse transcription PCR; for, forward; rev, reverse; bp, base pairs; CDS, coding sequence; aa, amino acid; LDM, lanosterol 14- α demethylase; N.A. not applicable; **M. circinelloides*, *R. microsporus*, and *R. arrhizus*.

Table S4. Oligonucleotide primers used for amplification and sequencing of the pairs of paralogous CYP51 genes in three Mucorales species.

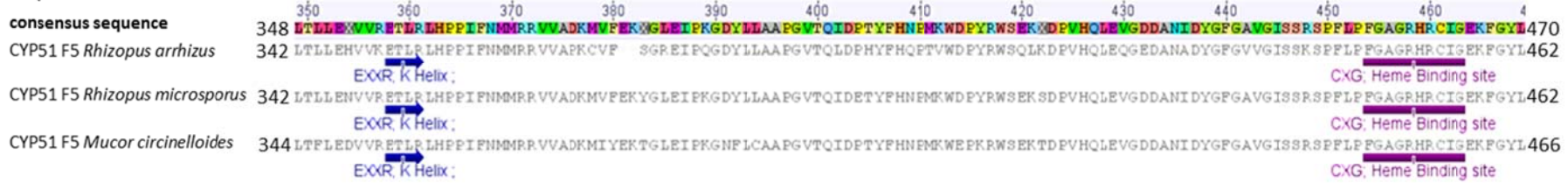
primer name	species	locus ^a	direction	sequence (5' to 3')	AT (°C)	amplicon size (bp)
Muci1_Fw	<i>M. circinelloides</i>	CYP51A F1	For	ACC TCA CTC CCC TCT CTC TCC A	58	1800
Muci1_R1			Rev	CGG TGA CCT TCT TGC CGA CCA		
Muci1_F2			For	TCG AGT TGG CCG CTC GTC CT		
Muci1_c2			Rv	CAG GCA CAA CAA CCA TAG AAG TGT		
Muci2_Fw		CYP51B F5	For	ACC GAT GGT GAC ACG CCG TA	60	1967
Muci2_F1.1			For	TGG GCC CTG ATG GCA ACC AA		
Muci2_F2.1			For	TCT GCT CTC CCT GAC CAC CA		
Muci2_R1			Rev	TGT AGG AAT CGA CGA CAC CAG TCT		
Muci2_Rv			Rev	GCG CAC ATG TAT TTG TCC AGT AGC		
Rmicr_CYP51A_FW	<i>R. microsporus</i>	CYP51A F1	For	CAG ACC CGT CCG TTA CAC AA	56	1779
Rmicr_CYP51A_F2			For	AAC GCA AAG CAG AAT TTG GC		
Rmicr_CYP51A_R1			Rev	AGA CAA CTT CAG GGC CAA ACA		
Rmicr_CYP51A_F3			For	GGA ACA CCC TTA CCT GAT CAT CA		
Rmicr_CYP51A_F4			For	GCG TGC CTA AGC CTG ACT AT		
Rmicr_CYP51A_R2			Rev	CAG AGG TGT GTT GAC CAC CA		
Rmicr_CYP51A_RV			Rev	TGC TCC GTC ACT ACA CAT GT		
Rmic_Fw		CYP51B F5	For	TGC CGG GCT CGA TGA CCG AT	60	1568
Rmic_F1			For	AAGCGCGTCACTGCATGCCT		
Rmic_R1			Rev	TCT GCA ACG ACA CGG CGC AT		
Rmic_F2			For	TGG TCC AGA GGT TCG TAA CGC		
Rmic_Rv			Rev	CGG TGG CGT CCA GCA CCA AA		
Rdel_Cyp51A_1775fw	<i>R. arrhizus</i>	CYP51A F5	For	GTT CGA TCT GCT ACA AAC GCC	58	2032

Rdel2.F2		For	AAA CGC GTC ACG GCC TGC TT		
Rdel2.R1		Rev	CCA GGC GGC AGT GGT GGA AG		
Rdel2.F3		For	TAC GG CCT CCC GTA CCC TGC		
Rdel2.Rv		Rev	ACA GGG TTC CAT CCG TTC CAG T		
Rdel_CYP51A_19851v		Rev	TTG ATG CTG ATG GCG CTC TAT		
Rdel_F1	<i>CYP51B</i> F1	For	TGC AAA TGG AGC CGC GAC GA	58	1932
Rdel_R1		Rev	TCG GAT GAG TTG TGT TAT CGC TTC		
Rdel_F1.1		For	TGC TCC TCA CAG CGT ATT CAT GG		
Rdel_F2.1		For	CGT ACC TGA CCA TCA CAT TGC CG		
Rdel_Rv		Rev	ACG TCA CAT AGT TAA ACA GCC A		

Table legend: ^a locus names *CYP51A* (F1) and *CYP51B* (F5) were attributed according to Nelson's database, as a result of BLAST analysis against P450 protein families. For, forward; Rev, reverse; AT, annealing temperature °C; bp. base pairs; primers shown in bold were used for both DNA amplification and Sanger sequencing.

1 **Figure S1.**

A)



B)

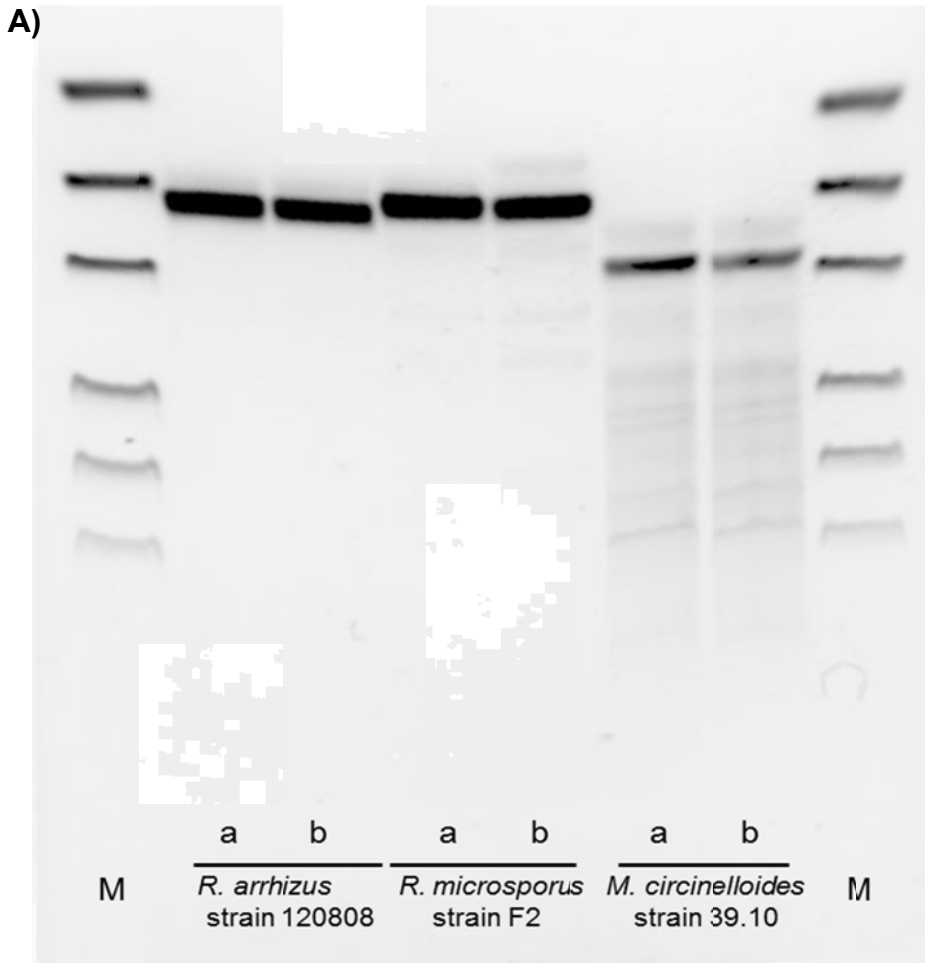


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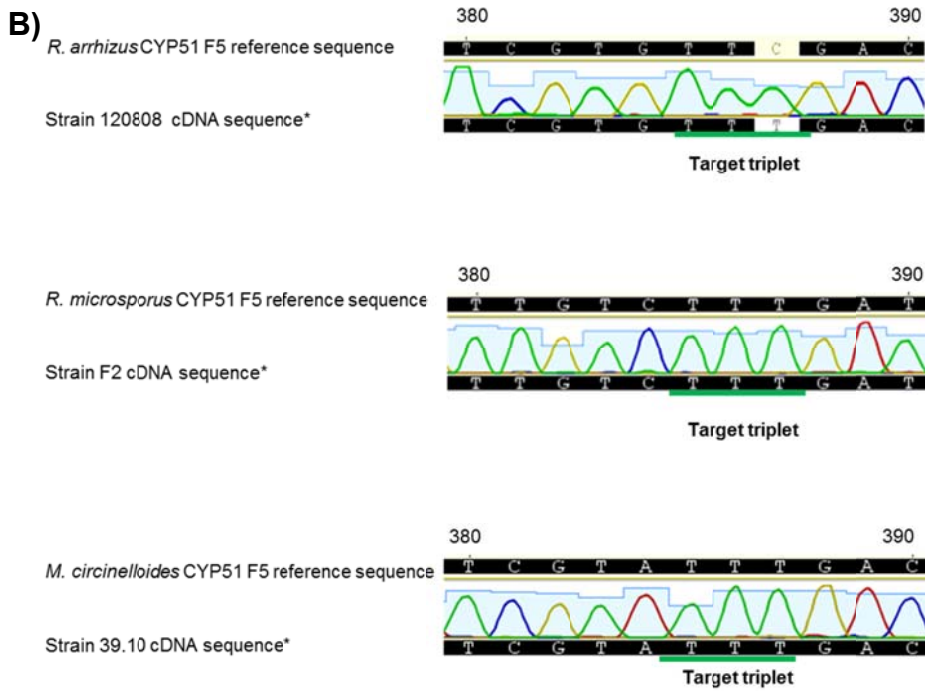
3 **Figure legend.** Sequence alignment of the two orthologous paralogous CYP51 genes showing consensus sequences retrieved
 4 for the species *R. arrhizus*, *R. microspores* and *M. circinelloides* and the location of the conserved motifs EXXR (ETLR in both
 5 protein sequences) marked in blue, and CXG (CIG in both protein sequences) marked in purple.

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7 **Figure S2.**



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10 **Figure legend. A)** A 2% w/v agarose gel depicting cDNA products from three isolates
11 of the species *R. arrhizus*, *R. microsporus*, and *M. circinelloides*, (CBS strain 120808,
12 clinical isolate F2 and clinical isolate 39.10, respectively) obtained from the qPCR
13 performed for expression of *CYP51 F5*; **a**, cDNA product obtained after 48 h growth
14 without exposure to the ITC, and **b** cDNA product obtained after exposure for 48h to
15 0.50 mg/L of ITC; M, 100 bp marker. **B)** Sequence analysis of each cDNA product
16 shown in **A)** after ITC exposure. The sequence obtained using just the forward primer
17 is shown (both primers were used for sequence analysis and assembly). The target
18 triplet (positions 385 to 387) in the *CYP51 F5* CDS of the three species is underlined
19 in green. For a better visualization the image was color-invented.

20 **Figure S3.** *Rhizopus arrhizus* and *R. microsporus* strains exhibit variable *in vitro* minimum inhibitory concentrations for voriconazole
 21 according to CLSI microbroth dilution method.

Phenotype	Species/strain number ²	Lanosterol 14- α demethylase F5 partial sequence, mapped to reference
VCZ MIC ¹ (mg/L)		
n.a.	<i>Saccharomyces cerevisiae</i> ERG11 ³	130 T T P V F G K G V I Y D C P N S R L M E Q 150
4.00	<i>R. arrhizus</i> / 120808 LDM F5	T K Y V F G N D I V F D T A H S I F M E Q
16.00	<i>R. arrhizus</i> / 136236 LDM F5	T K Y V F G N D I V F D T A H S V F M E Q
>16.00	<i>R. arrhizus</i> / 136239 LDM F5	T K Y V F G N D I V F D T A H S V F M E Q
4.00	<i>R. microsporus</i> / 71.11 LDM F5	T K Y V F G N D V V F D T A H S I F M E Q
8.00	<i>R. microsporus</i> / AS 40 LDM F5	T K Y V F G N D V V F D T A H S I F M E Q
16.00	<i>R. microsporus</i> / 30.10 LDM F5	T K Y V F G N D V V F D T A H S I F M E Q

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23 **Figure legend.** Voriconazole (VCZ) minimum inhibitory concentrations (MIC¹) were determined according to CLSI standard
 24 microbroth dilution method ^[54] ; ² strain information is given in a previous published study ^[32]; the partial LDM F5 sequence with amino
 25 acid substitution Y140F (*S. cerevisiae* numbering) is mapped to the reference *S. cerevisiae* LDM ³ (accession number given in
 26 Supplementary Table S2): LDM F5, lanosterol 14- α demethylase primary sequence translated from CYP51 F5 gene and named
 27 according to Nelson's P450 Database ²⁷; location of the amino acid substitution in all Mucorales strains highlighted in bold; n.a. not
 28 applicable.