

## **Effect of the acylation of TEAD4 on its interaction with co-activators YAP and TAZ**

Yannick Mesrouze<sup>#a</sup>, Marco Meyerhofer<sup>#a</sup>, Fedir Bokhovchuk<sup>a</sup>, Patrizia Fontana<sup>a</sup>, Catherine Zimmermann<sup>a</sup>, Typhaine Martin<sup>a</sup>, Clara Delaunay<sup>a</sup>, Aude Izaac<sup>b</sup>, Joerg Kallen<sup>b</sup>, Tobias Schmelzle<sup>a</sup>, Dirk Erdmann<sup>a</sup> and Patrick Chène\*<sup>a</sup>.

<sup>a</sup>Disease Area Oncology and <sup>b</sup>Chemical Biology & Therapeutics, Novartis Institutes for Biomedical Research, Basel, Switzerland.

# These 2 authors have contributed equally to this work.

### **Legend to the supplementary figures**

**Supplementary figure S1.** Global view of the X-ray structure for myristoylated human hTEAD4<sup>217-434</sup> in complex with YAP<sup>60-100</sup>. The coordinates have been deposited in the PDB databank (PDB access code = 5O AQ). hTEAD4<sup>217-434</sup> and hYAP<sup>60-100</sup> are represented by green and orange ribbons, respectively. The covalently bound myristate moiety is in purple. hTEAD4<sup>217-434</sup> has a similar structure to hTEAD1 and mTEAD4 in previously published structures [1, 2]. hYAP<sup>60-100</sup> contacts hTEAD4<sup>217-434</sup> via two distinct secondary structure elements ( $\alpha$ -helix and  $\Omega$ -loop) as already described [1, 2].

**Supplementary figure S2.** Stability of the immobilized TEAD4 proteins in SPR. The filled circles at cycle 0 are the calculated  $R_{\max}^{\text{theo}}$  (see text for explanation). These values vary

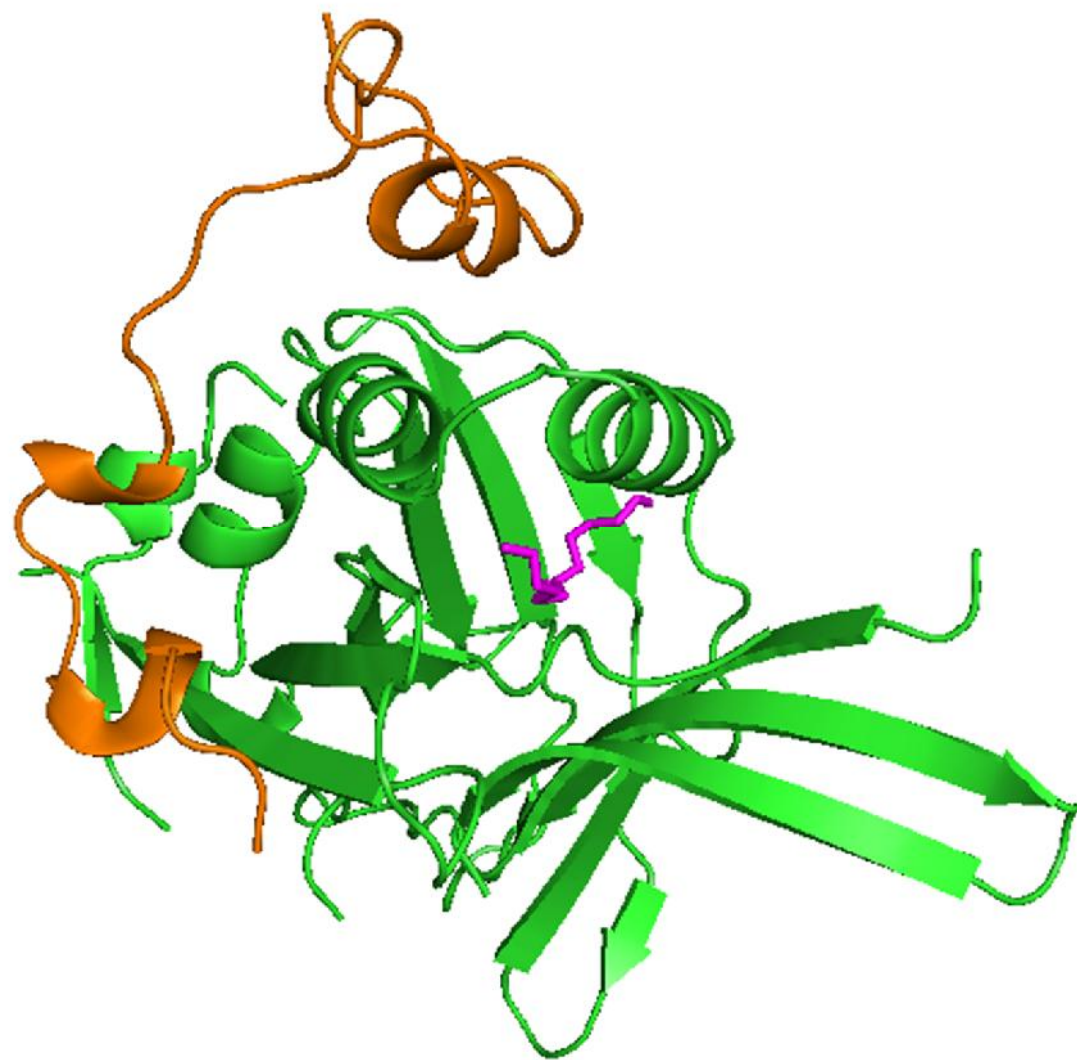
between experiments, because different levels of TEAD4 have been immobilized. At the beginning and the end of every dose range experiment with a peptide to determine its  $K_d$ , a measurement with a saturating concentration of TAZ<sup>14-56</sup> (1  $\mu$ M) to estimate the stability of the immobilized TEAD4 protein was conducted. The filled circles represent the signal measured during these cycles. These experiments were performed at 10 and 25°C. The signal measured at 25°C (red filled circles) during these test cycles with Non-Acyl-TEAD4 and Cys367Ser decreased more rapidly than the signal measured with Acyl-TEAD4. This suggests a higher instability of the non-acylated proteins. At 10°C (blue filled circles), all 3 proteins were more stable, and the decrease in signal over the course of an experiment was largely reduced.

**Supplementary figure S3.** Thermal shift assay of refolded and non-refolded Cys367Ser. Ref-Cys367Ser (**A**) and non-refolded Cys367Ser (Non-Ref-Cys367Ser, **B**) possess a His<sub>6</sub>-tag at their N-terminus in addition to an Avi-tag. The thermograms were obtained in a fluorescence-based thermal denaturation assay in the presence of 1-2  $\mu$ M protein. The melting temperatures ( $T_m$ ) were obtained by plotting the first derivative of the fluorescence emission (F) as a function of the temperature (-dF/dT). The curve minimum corresponds to  $T_m$ . The indicated  $T_m$  have been measured for the experiments depicted on the figure. These  $T_m$  are slightly different from the  $T_m$  presented on Table 2 because these later are the average values obtained from several independent measurements.

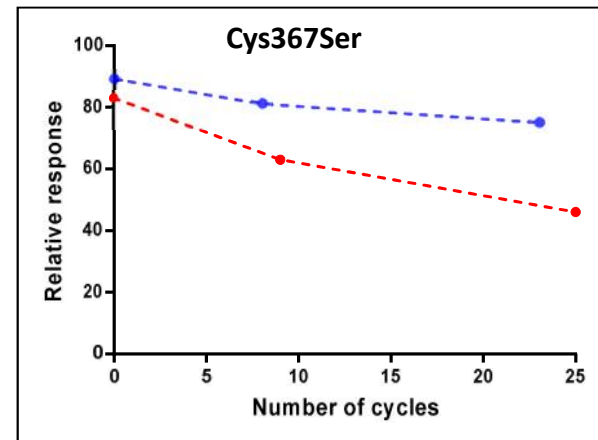
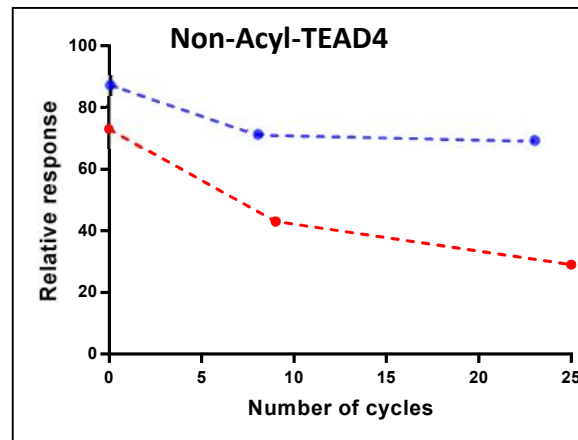
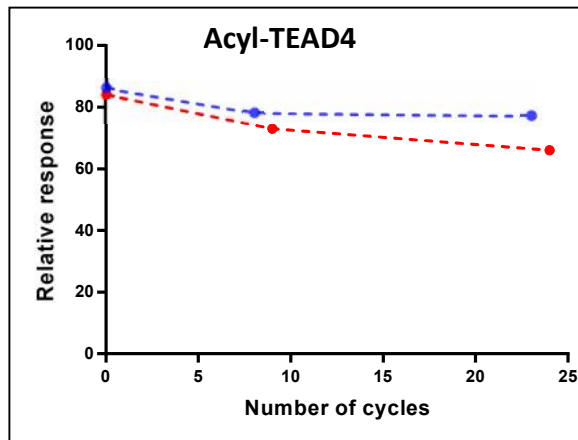
## References

- [1] Chen L, Loh PG, Song H. Structural and functional insights into the TEAD-YAP complex in the Hippo signaling pathway. *Protein Cell*. 2010;1:1073-83.
- [2] Li Z, Zhao B, Wang P, Chen F, Dong Z, Yang H, et al. Structural insights into the YAP and TEAD complex. *Genes & Dev*. 2010;24:235-40.

Supplementary Fig. S1



## Supplementary Fig. S2



SPR conducted at 10°C (●) or 25°C (●)

## Supplementary Fig. S3

