

Supporting Information Appendix

Structure and function of yeast Atg20, a sorting nexin that facilitates autophagy induction

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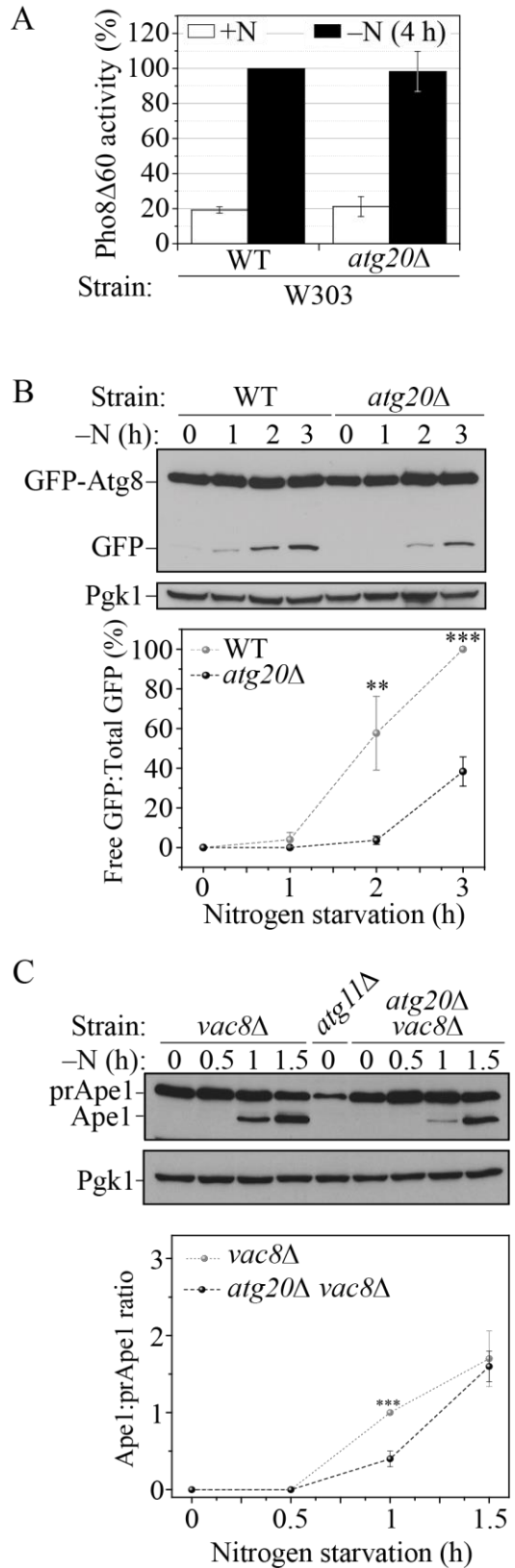
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Fig. S1. Atg20 is essential for the efficient initiation of nonselective autophagy. Wild-type and *atg20* Δ cells were monitored using the (A) Pho8 Δ 60 assay and (B) GFP-Atg8 processing assay (strains ZFY202 and YAB87). (C) The *vac8* Δ and *atg20* Δ *vac8* Δ strains were examined for prApe1 maturation (strains HPY081 and HPY082) as described in the legend for Figure 1C.



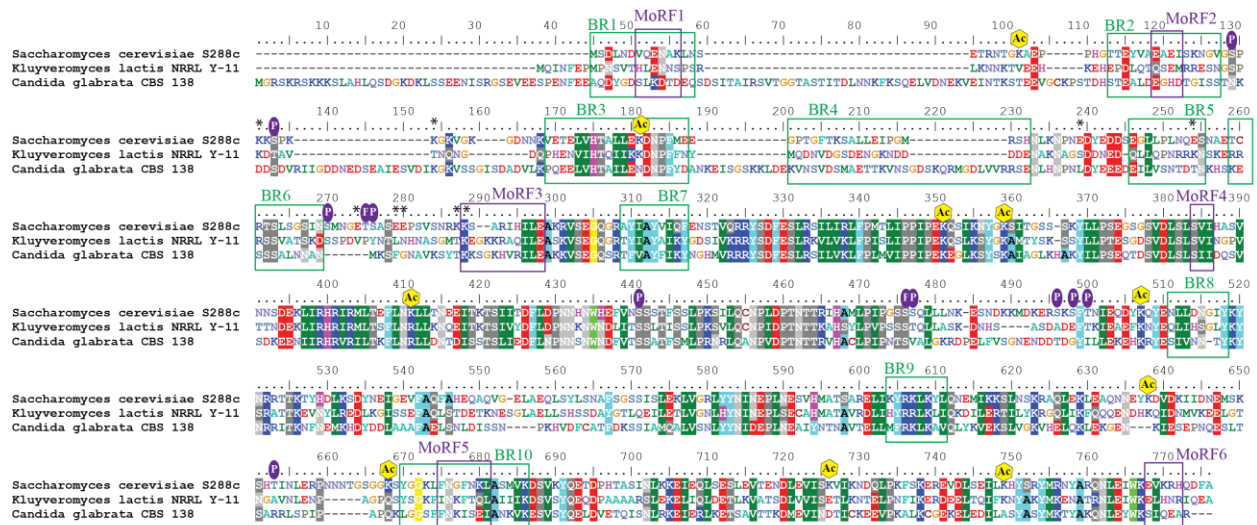


Fig. S2. Multiple sequence alignment of Atg20 proteins from three fungi that are often used in autophagy studies. Phosphorylation (P) and acetylation (Ac) sites for the *S. cerevisiae* sequence are marked. Predicted BR and MoRF regions from the *S. cerevisiae* sequence are boxed in green and purple respectively. Black asterisks show the amino acid residues mutated in the Atg20[Aroma] mutant.

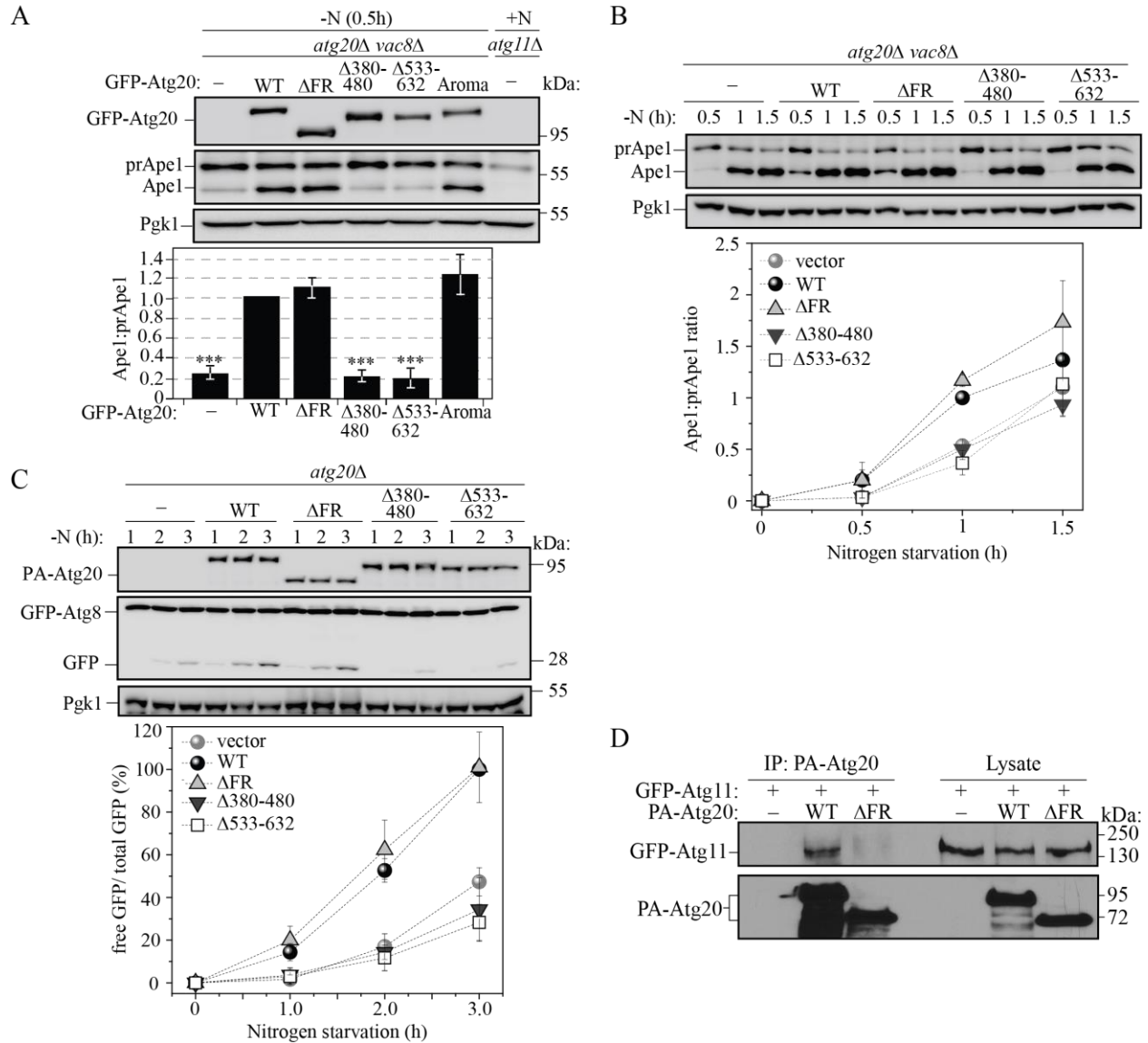


Fig. S3. Functionality and binding properties of Atg20 mutants. (A) The prApe1 maturation in *atg20Δ vac8Δ* (HPY079) cells transformed with the plasmids pCuGFP(426), pCuGFP-Atg20(426), pCuGFP-Atg20[Δ FR](426), pCuGFP-Atg20[$\Delta 380-480$](426), pCuGFP-Atg20[$\Delta 533-632$](426) or pCuGFP-Atg20[Aroma](426), cultured in rich selective medium and then nitrogen starved for 0.5 h. Statistical significance was tested by unpaired two-tailed Student's t-test. The p values less than 0.005 were considered to be significant (***). (B) Kinetics of prApe1 maturation after the shift to nitrogen starvation medium for the indicated times. Deletion mutants of Atg20 were compared to empty vector and wild-type Atg20. (C) Kinetics of GFP-Atg8 processing during nitrogen starvation for the indicated times. The *atg20Δ* (D3Y009)

cells were transformed with the plasmid pCuGFP-Atg8 (426) and the plasmid pCuPA(424), pCuPA-Atg20(424), pCuPA-Atg20[Δ FR](424), pCuPA-Atg20[Δ 380-480](424), or pCuPA-Atg20[Δ 533-632](424). Error bars represent standard deviation from 3 independent experiments.

(D) Deletion of Atg20 FR weakens the interaction between Atg20 and Atg11. The plasmids pCuPA(424) or pCuPA-Atg20(424) were transformed into *atg20* Δ (D3Y009) cells and co-expressed with a plasmid encoding GFP-Atg11 (pCuGFP-Atg11; 416) under the *CUP1* promoter. The large GFP tag at Atg11 completely abolished interaction with the deletion mutant of Atg20. Cells were cultured in SMD media and cell lysates were prepared and incubated with IgG-Sepharose for affinity purification. The proteins were separated by SDS-PAGE and detected with monoclonal antibody that recognizes HA or PA or GFP.

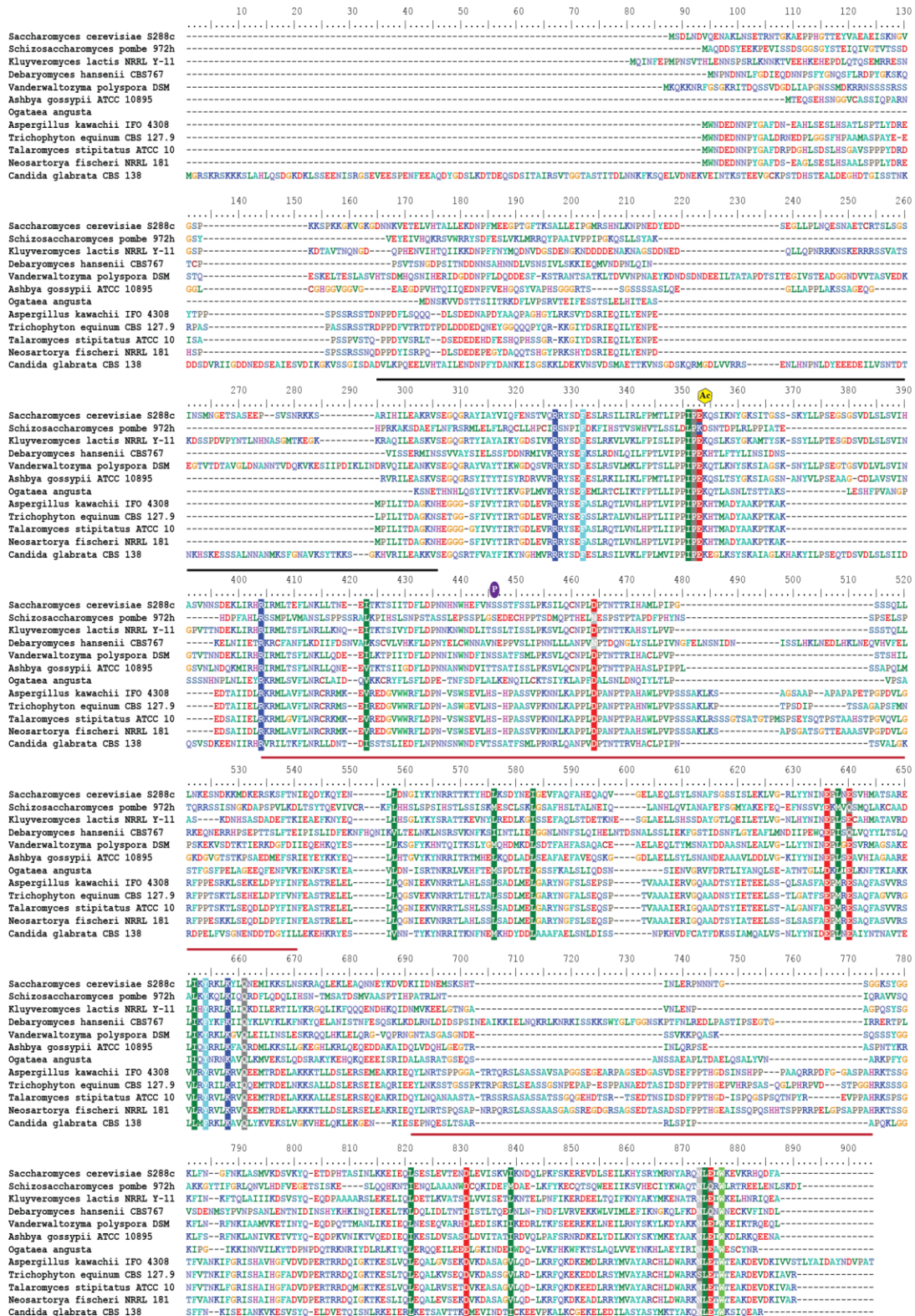


Fig. S4. Multiple sequence alignment of Atp2L proteins from various fungi. The alignment was created in the BioEdit Sequence Alignment Editor (1). The most conserved acetylated lysine

(K218) and phosphorylated serine (S307) in *S. cerevisiae* are marked. The black and red lines above the sequences denote the span of the PX and BAR domain in *S. cerevisiae*, respectively.

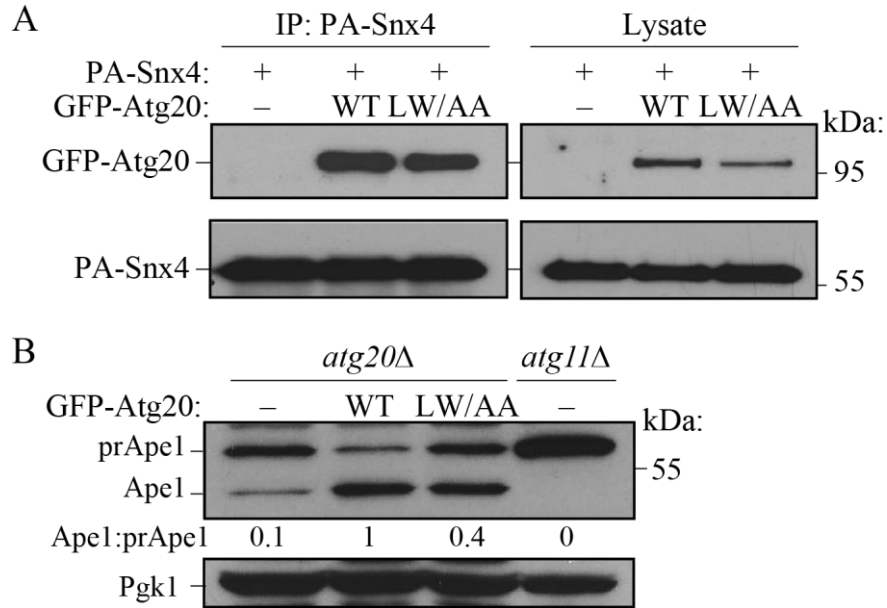


Fig. S5. Probing the conserved motif $_{626}\text{NLExW}$ in Atg20. (A) The plasmids pCuGFP(426), pCuGFP-Atg20(426) or pCuGFP-Atg20[L627A/W630A](426) were transformed into MKO (YCY123) cells and co-expressed under the *CUP1* promoter with a plasmid encoding PA-Snx4 (pCuPA-Snx4; 424). Cells were cultured in SMD, and cell lysates were prepared and incubated with IgG-Sepharose for affinity purification. The proteins were separated by SDS-PAGE and detected with monoclonal antibody that recognizes GFP or PA. (B) PrApe1 processing assay. The *atg20Δ* (D3Y009) strain was transformed with the plasmid pCuGFP(426), pCuGFP-Atg20(426) or pCuGFP-Atg20[L627A/W630A](426). The *atg11Δ* strain (SEY6210) was used as a negative control. Cells were cultured in rich selective medium and cell lysates were TCA precipitated. Proteins were detected by Ape1 and Pgk1 antibody after separation on SDS-PAGE.

phosphorylated serine (S+80) or threonine (T+80) is confirmed by *b* and/or *y* ions that are highlighted in red and blue color.

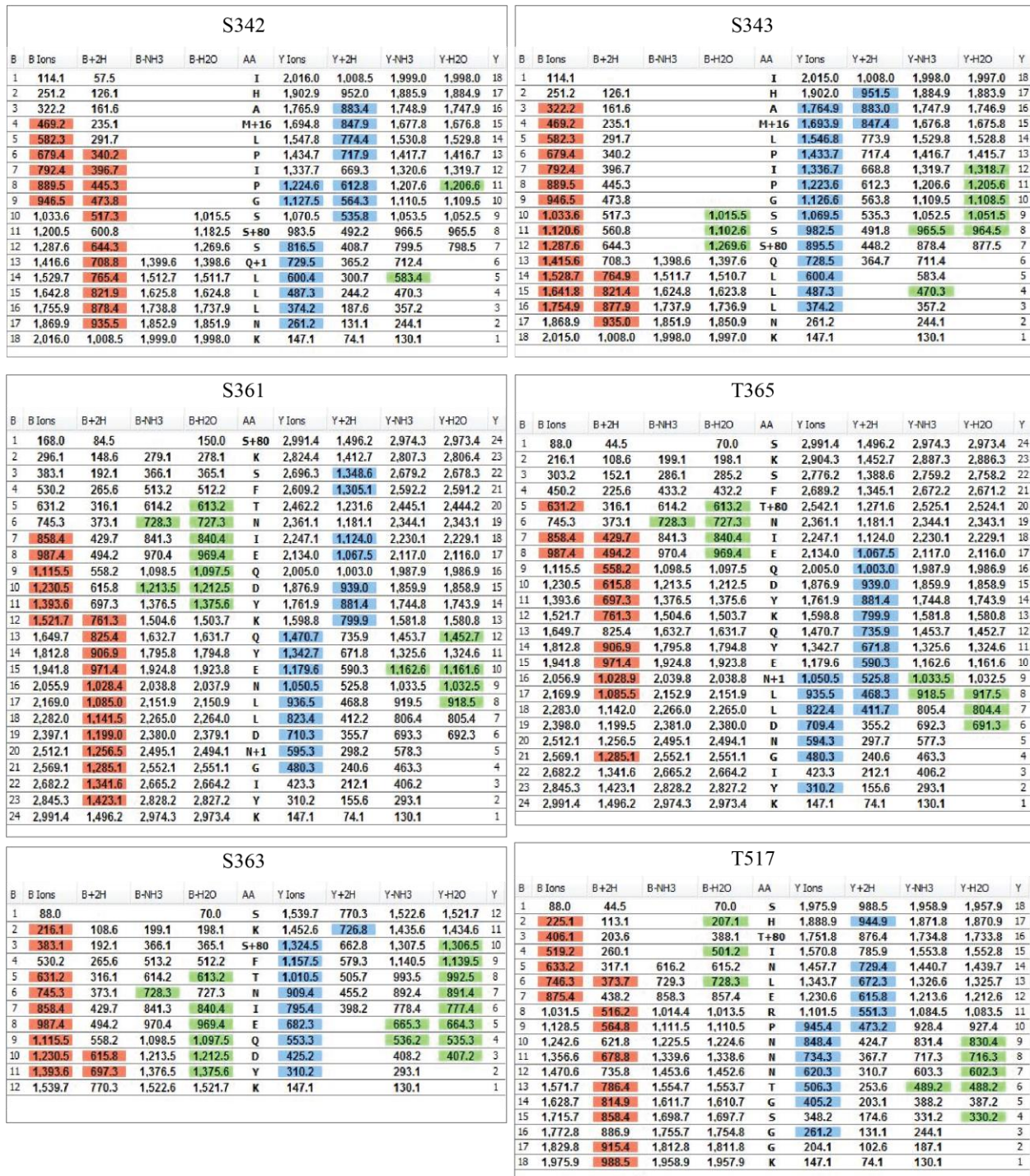


Fig. S9. Fragmentation tables as they appear in the Scaffold program for phosphorylated serines and threonines (S342, S343, S361, S363, T365, and T517) in Atg20. The presence of phosphorylated serine (S+80) or threonine (T+80) is confirmed by *b* and/or *y* ions that are

highlighted in red and blue color. Phosphorylation on Ser361 was confirmed by peptide mass only.

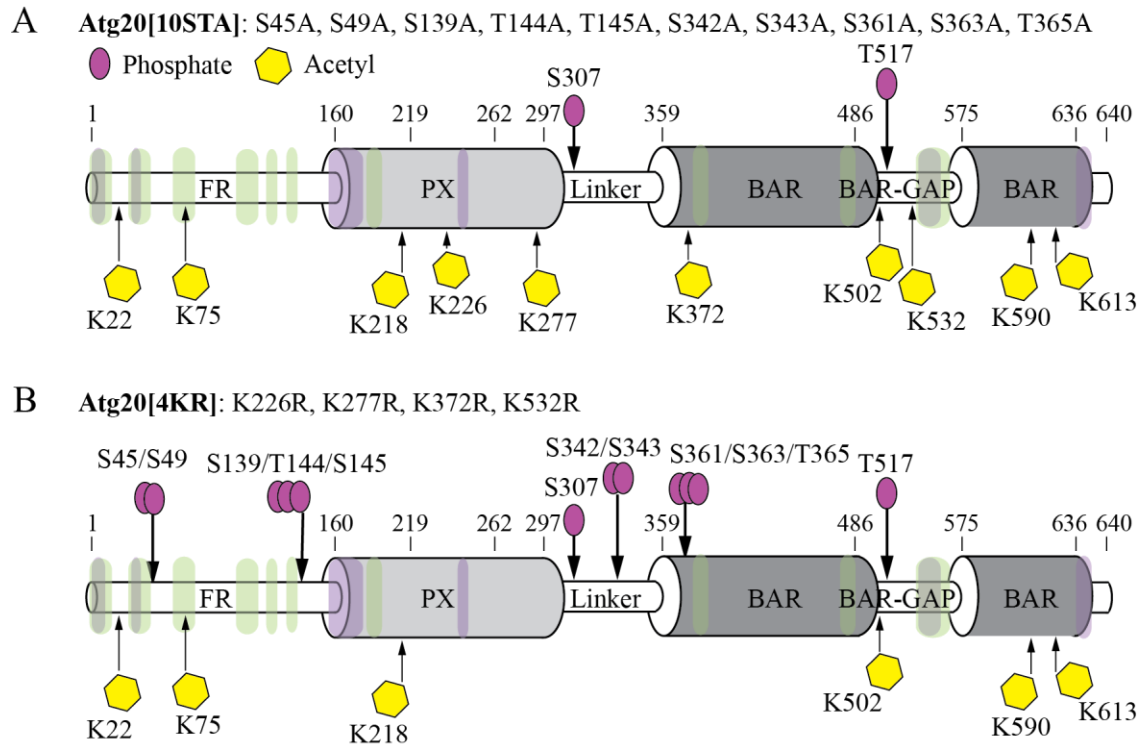


Fig. S10. Schematic representations of Atg20[10STA] and Atg20[4KR]. The representations depict which post-translational modifications were removed and which remained in the mutated Atg20[10STA] (A) and Atg20[4KR] (B) proteins.

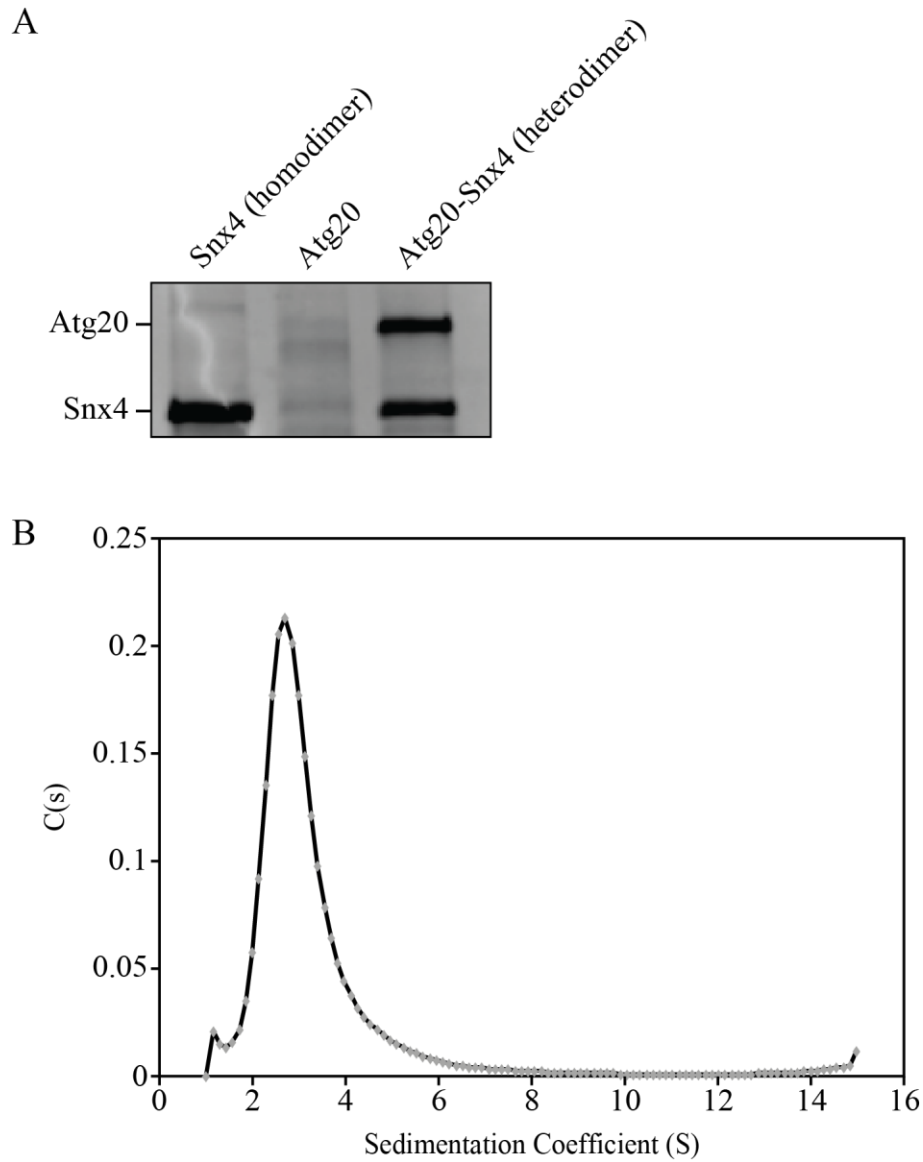
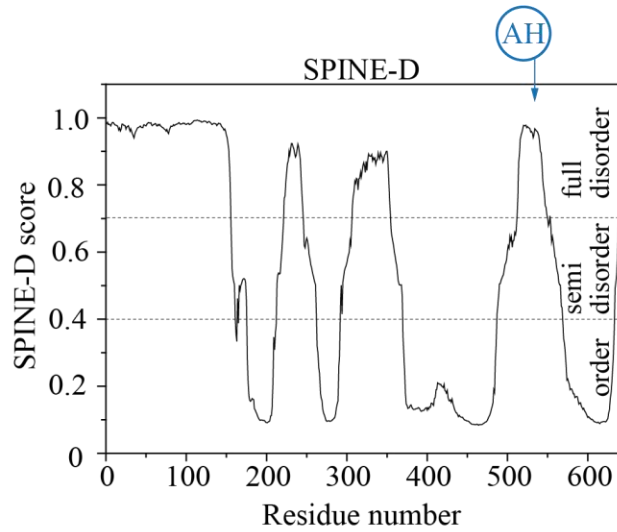


Fig. S11. Structural analysis of the sorting nexins Atg20 and Snx4. (A) SDS-PAGE gel showing the capture of full-length Atg20 only as a heterodimer with full-length Snx4. The full-length Snx4 can also form homodimers, but no monomeric or dimeric Atg20 can be captured. (B) The AUC data for the Snx4₄₂₁₋₄₂₃ homodimer. The molecular mass determined by dynamic light scattering for this homodimer is 93.3 kDa, which is very similar to the expected molecular mass of 93.7 kDa.



MFDp2

GREEN letters represent residues predicted as ordered
RED letters correspond to predicted disordered residues

SEQ Predicted disorder content: 35.47 %; # of disorder segments: 2

10	20	30	40	50	60	70	80
MSDLNDVQEN	AKLNSETRNT	GKAEPPHGT	EYVAEAEISK	NGVGSPPKSP	KKGKVGKGDN	NKVETELVHT	ALLEKDNPEM
90	100	110	120	130	140	150	160
EEGPTGFTKS	ALLEIPGMRS	HNLKPNEDY	EDDSEGLLPL	NQESNAETCR	TSLSGSINSM	NGETSASEEP	SVSNRKK SAR
170	180	190	200	210	220	230	240
THILEAKRVS	EGQGRAYIAY	VIQFENSTVO	RRYSDFESLR	SILIRLFPMT	LIPPIPEKQS	TKNYGKSITG	SSSKYLLPSE
250	260	270	280	290	300	310	320
GSGVDLSLS	VIHAVNNSD	EKLIRHRIRM	LTEFLNKLIT	NEEITKTSII	TDFLDPNNHN	WHEFVNSST	FSSLPKSILQ
330	340	350	360	370	380	390	400
CNPLDPTNTT	RIHAMPLPIG	SSSQLLLNKE	SNDKKMDKER	SKSFTNIEQD	YKQYENLLDN	GIYKYNRRTT	KTYHDLKSDY
410	420	430	440	450	460	470	480
NEIGEVFAQF	AHEQAQVDEL	AEQLSYLSNA	FSGSSISLEK	LVGRLYYNIN	EPLNESVHMA	TSARELIK YR	KLKYLQNEMI
490	500	510	520	530	540	550	560
KKSLNSKRAQ	LEKLEAQNE	YKDVOKTIDN	EMSKSHTINL	ERPNNNTGSG	GKSYGGKLEN	GENKLASMYK	DSVLYQETDB
570	580	590	600	610	620	630	640
HTASINLKKE	IEQLSESLEV	TENDLEVISK	VIKNDQLPKF	SKEREVDLSE	ILKHYSRYMR	NYARQNL EIW	KEVKRHQDFA

AH

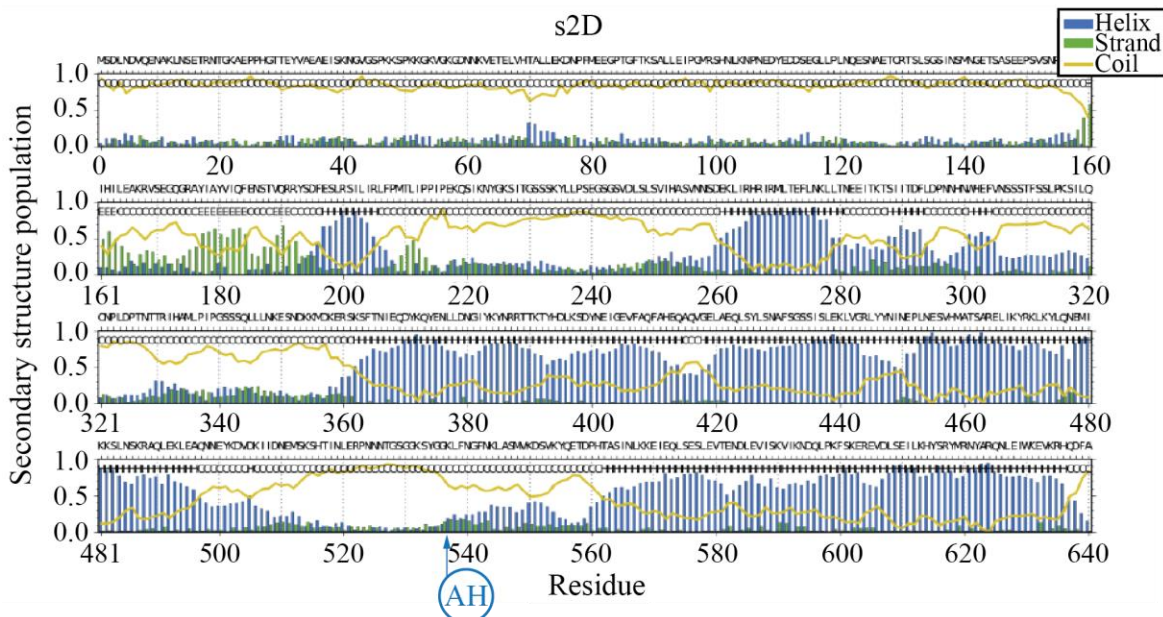


Fig. S12. Analysis of the amino acid sequence of Atg20 by predictors (SPINE-D, MFDp2, and s2D; (2-4)) for intrinsically disordered protein regions. The position of the putative amphipathic helix (AH) in Atg20 is indicated by a blue arrow.

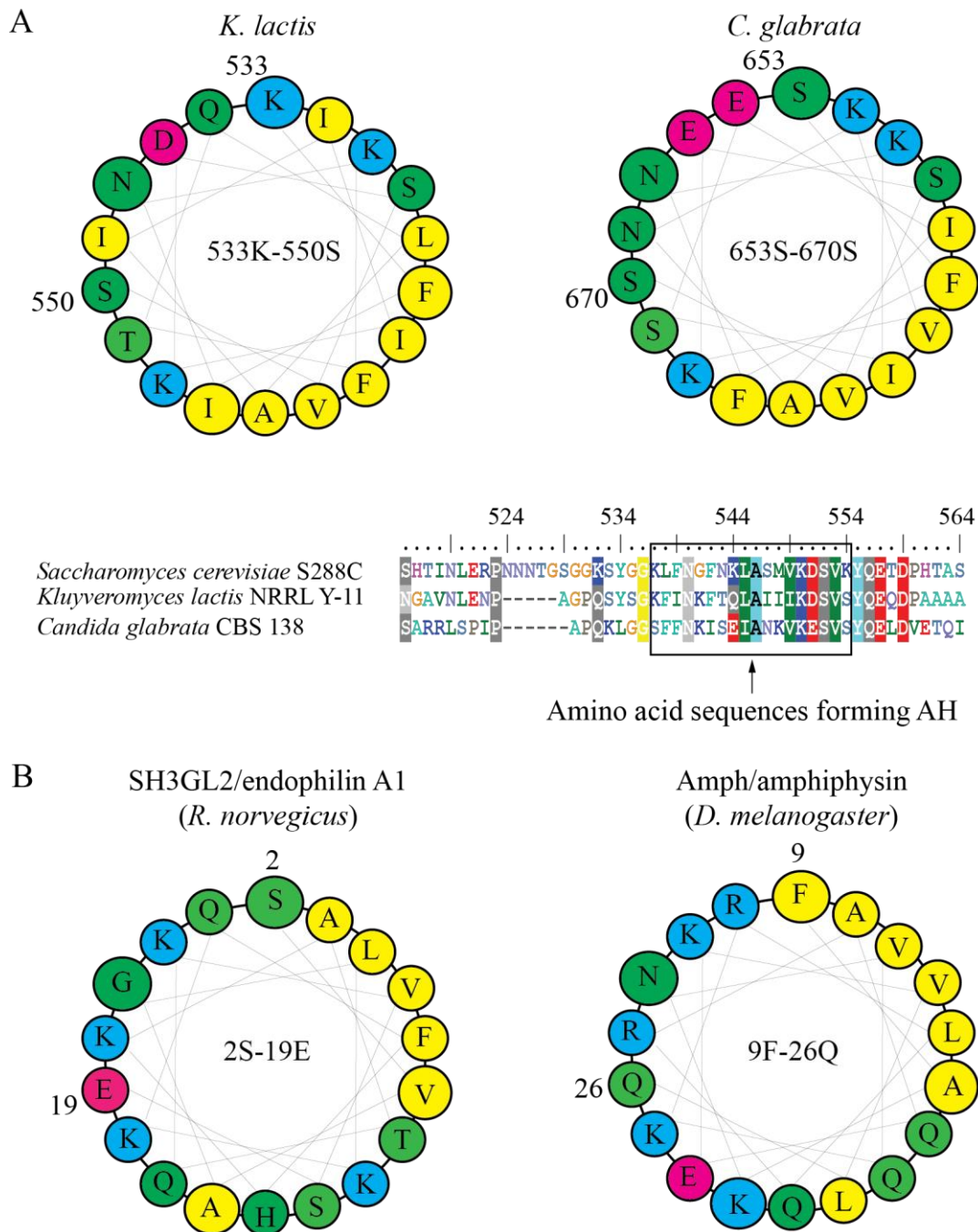


Fig. S13. Comparison of predicted amphipathic helices in fungal Atg20 homologs and BAR domain-containing proteins. (A) AH in Atg20 of *K. lactis* and *C. glabrata* formed by amino acid sequences homologous to the amino acid sequence forming AH in Atg20 of *S. cerevisiae*. (B) AH in SH3GL2/endophilin A1 and Amph/amphiphysin (5) are shown for comparison with those in (A).

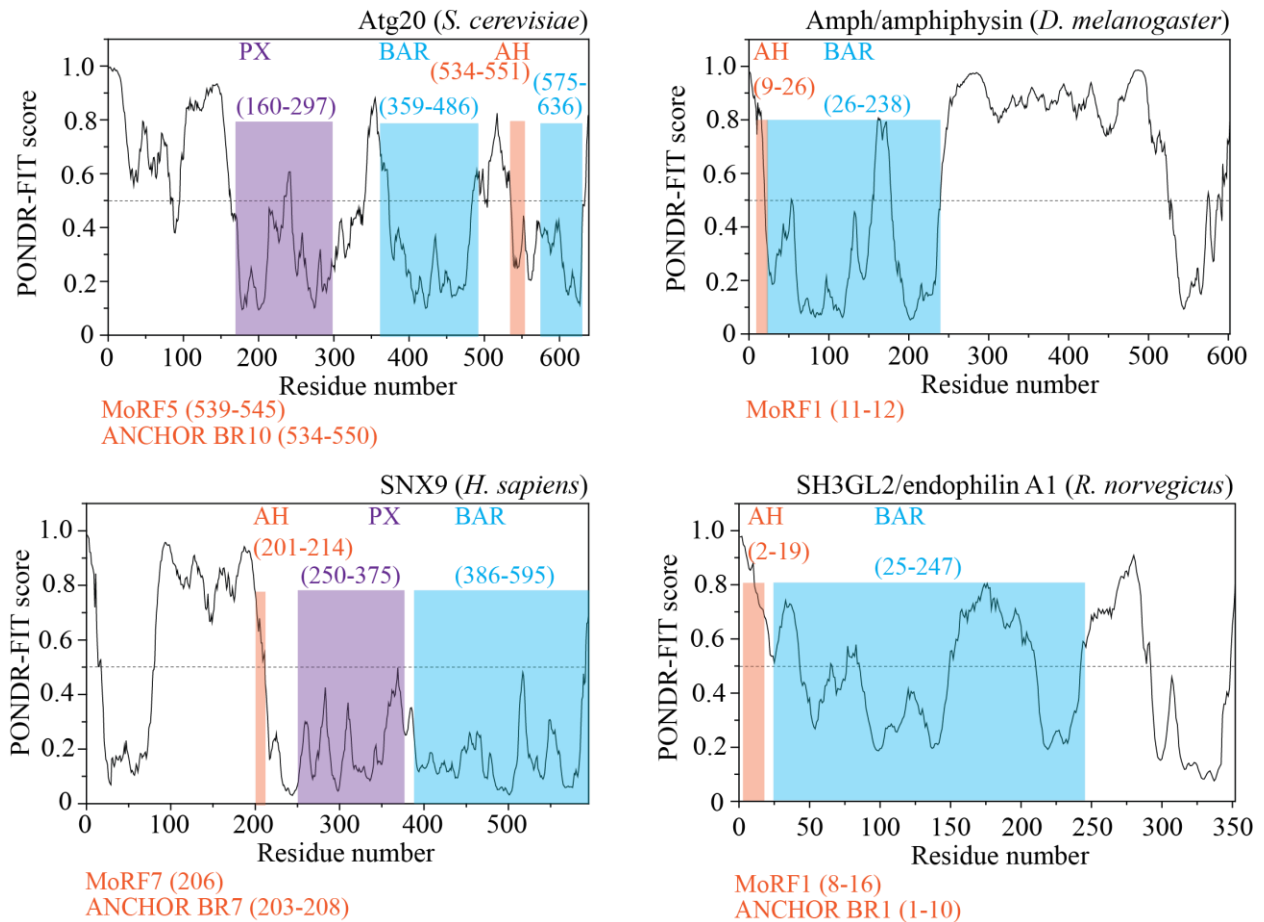


Fig. S14. Comparative bioinformatics analysis of the amino acids sequences of the BAR proteins that encompass a membrane-inducible AH. The consensus sequences for the BAR and PX domains, as determined by protein-protein BLAST, are indicated in blue and purple, respectively. The position of a particular AH within an IDPR determined by PONDR-FIT is shown in red. The results of the algorithms MoRFPred and ANCHOR (if any), which determine the position of a foldable element (termed molecular recognition feature or ANCHOR disordered binding domain) within an IDPR, is indicated in red for each BAR protein under the corresponding diagram.

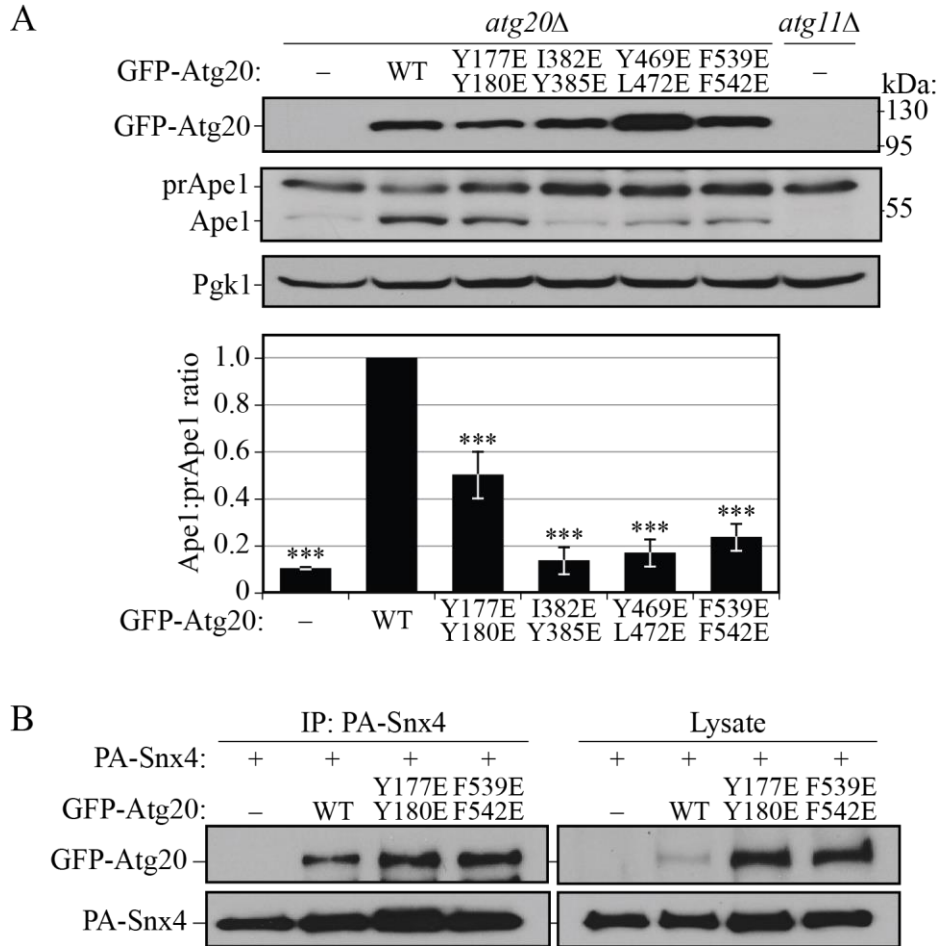


Fig. S15. Analysis of mutations in the PX, BAR, and BAR-GAP domains of Atg20. (A) The structural mutants of Atg20 are defective in the Cvt pathway. The *atg20Δ* strain (D3Y009) was transformed with the plasmid pCuGFP(426), pCuGFP-Atg20(426), pCuGFP-Atg20[Y177E Y180/E](426), pCuGFP-Atg20[I382E Y385E](426), pCuGFP-Atg20[Y469E L472E](426) or pCuGFP-Atg20[F539E F542E](426). The *atg11Δ* (SEY6210) strain was used as a negative control. GFP, Ape1 and Pgk1 (loading control) were detected from the cell lysate by immunoblotting. Columns are averages and error bars represent standard deviation from 3 independent experiments. Statistical significance was tested by unpaired two-tailed Student's t-test. The p values less than 0.005 were considered to be significant (***). (B) PA-Snx4 co-precipitates all variants of GFP-Atg20. The plasmids pCuGFP(426), pCuGFP-Atg20(426), pCuGFP-Atg20[Y177E Y180/E](426), or pCuGFP-Atg20[F539E F542E](426) were transformed into MKO (YCY123) cells and co-expressed under the *CUPI* promoter with the pCuPA-Snx4(424) plasmid. Cells were cultured in SMD, and cell lysates were prepared and incubated

with IgG-Sepharose for affinity purification. The proteins were separated by SDS-PAGE and detected with monoclonal antibody that recognizes GFP or PA. The F539E F542E mutation in MoRF5/BR10 does not weaken the interaction between Atg20 and Snx4.

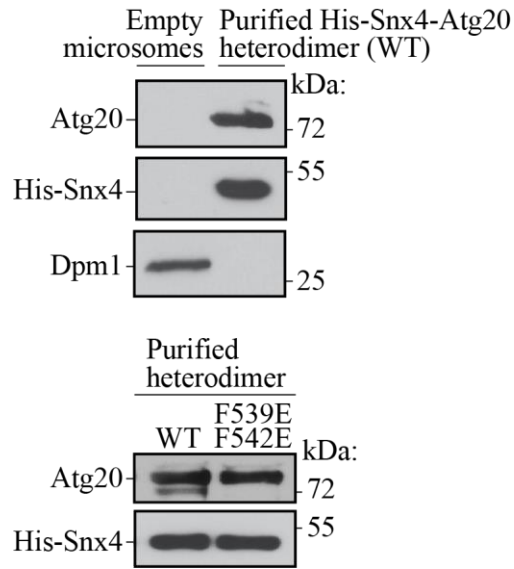


Fig. S16. Analysis of the microsomes and protein input for the *in vitro* membrane-binding assay. Upper panel, western blots showing the presence of Atg20 and His-Snx4 in the recombinant purified heterodimer and the presence of the endoplasmic reticulum marker protein Dpm1 in microsomes isolated from *snx4Δ atg20Δ* (SEY6210) cells; lower panel, western blots showing an equal input of purified heterodimers (wild-type or mutant) that were used in the reconstitution experiment.

Table S1. *S. cerevisiae* strains used in this study.

Name	Genotype	Reference
CWY230	<i>SEY6210 vac8Δ::KAN</i>	(6)
D3Y009	<i>SEY6210 atg20Δ::HIS</i>	This study
HPY017	<i>SEY6210 atg24Δ::HIS, atg20Δ::LEU2</i>	This study
HPY063	<i>WLY176 atg20Δ::LEU2</i>	This study
HPY079	<i>CWY230 atg20Δ::HIS3</i>	This study
HPY081	<i>W303-1B vac8Δ::URA3</i>	This study
HPY082	<i>W303-1B vac8Δ::URA3 atg20Δ::KAN</i>	This study
SEY6210	<i>MATα leu2-3,112 ura3-52 his3-Δ200 trp1-Δ901 lys2-801 suc2-Δ9 GAL</i>	(7)
W303-1B	<i>MATα leu2-3,112 ura3-1 his3-11,15 trp1-1, ade2-1 can1-100</i>	(8)
WLY176	<i>SEY6210 pho13Δ pho8Δ60</i>	This study
YAB87	<i>ZFY202 atg20Δ::KAN</i>	This study
YCY123	<i>SEY6210 atg1Δ, 2Δ, 3Δ, 4Δ, 5Δ, 6Δ, 7Δ, 8Δ, 9Δ, 10Δ, 11Δ, 12Δ, 13Δ, 14Δ, 16Δ, 17Δ, 18Δ, 19Δ, 20Δ, 21Δ, 23Δ, 24Δ, 27Δ, 29Δ,</i>	(9)
YTS147	<i>SEY6210 atg11Δ::LEU2</i>	This study
ZFY202	<i>W303-1B pho13Δ pho8Δ60</i>	This study

SI References

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