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Electronic Supplementary Information for:

Bodipy-C₆₀ Triple Hydrogen Bonding Assembly as Heavy atom-free Triplet Photosensitizers: Preparation and Study of the Singlet/triplet Energy Transfer

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Fig. S1 ¹H NMR of **2** (400 MHz, Acetone- d_6).



Fig. S2 TOF MS EI+ of **2**.



Fig. S3 ¹H NMR of **3** (500 MHz, CDCl₃).



Fig. S4 TOF MS EI⁺ of 3.





Fig. S6 TOF MS EI^+ of **5**.



Fig. S7 1 H NMR of 6 (500 MHz, CDCl₃).



Fig. S8 ¹H NMR of **7** (400 MHz, CDCl₃).



Fig. S9 TOF MS EI⁺ of **7**.



Fig. S10 ¹H NMR of **8** (400 MHz, CDCl₃).



Fig. S11 TOF HRMS (MALDI) ESI of 8.



Fig. S12 ¹H NMR of **C-1** (500 MHz, DMSO-d₆).



Fig. S13 ¹C NMR of **C-1** (100 MHz, DMSO-d₆).



Fig. S14 TOF HRMS ESI of C-1.



Fig. S15 ¹H NMR of **9** (400 MHz, CDCl₃).







Fig. S18 TOF HRMS (MALDI) ESI of 12.



ppm

Fig. S19 ¹H NMR of **14** (400 MHz, CDCl₃).



Fig. S20 ¹H NMR of **15** (400 MHz, CDCl₃).



Fig. S21 ¹H NMR of **16** (400 MHz, CDCl₃).



Fig. S22 TOF HRMS (MALDI) ESI of 16.



Fig. S23 ¹H NMR of **18** (400 MHz, CDCl₃).



Fig. S24. TOF MS EI^+ of **18**.



Fig. S25. ¹H NMR of **19** (500 MHz, CDCl₃).



Fig. S26. TOF MS EI+ of **19**.



Fig. S27 ¹H NMR of B-1 (500 MHz, DMSO- d_6).



ppm

Fig. S28 ¹³C NMR of **B-1** (100 MHz, CDCl₃).



Fig. S29 TOF HRMS MALDI of B-1.



Fig. S30 ¹H NMR of **B-1-Me** (500 MHz, CDCl₃).



ppm

Fig. S31 ¹³C NMR of **B-1-Me** (100 MHz, CDCl₃).



Fig. S32 TOF HRMS MALDI of B-1-Me.



Fig. S33 ¹H NMR of **B-2** (500 MHz, CDCl₃).



Fig. S34 ¹³C NMR of **B-2** (100 MHz, CDCl₃).







Fig. S36 1 H NMR of B-2-Me (400 MHz, CDCl₃).



ppm

Fig. S37¹³C NMR of **B-2-Me** (100 MHz, CDCl₃).



Fig. S38 TOF HRMS MALDI of B-2-Me.



Fig. S39 ¹H NMR of **B-3** (500 MHz, CDCl₃).



ppm

Fig. S40. ¹³C NMR of **B-3** (100 MHz, CDCl₃).



Fig. S41 TOF HRMS MALDI of B-3.



Fig. S42 ¹H NMR of **B-3-Me** (500 MHz, CDCl₃).





ppm

Fig. S43. ¹³C NMR of **B-3-Me** (100 MHz, CDCl₃).



Fig. S44. TOF HRMS MALDI of B-3-Me.

2.0 Synthetic details

Synthesis of Compound 2. 2-Chloro-acetyl chloride (0.36 mL, 4.6 mmol) was added dropwise into the solution of 2-amino-6-acetylamidopyridine (0.7 g, 4.6 mmol) in dry CH_2Cl_2 (30 mL) during 10 min, and then the mixture was kept at room temperature (RT) for 2 h. The resulted solution was poured into saturated aqueous Na₂CO₃ solution (30 mL). The CH_2Cl_2 layer was separated and washed with saturated aqueous NaCl solution. The organic layer was dried over Na₂SO₄. Solvent was removed under reduced pressure. A white solid was obtained. Yield:). Yeild: 492.0 mg (46.8%). ¹H NMR (400 MHz, CD_3COCD_3): 9.25 (s, 1H), 7.95–7.75 (m, 3H), 4.34 (s, 2H), 2.05 (s, 3H). TOF HRMS ESI⁺: *m/z* calcd for [C₉H₁₀N₃O₂Cl]⁺: 227.0462; found: 227.0461.

Synthesis of Compound 3. NaN₃ (45.5 mg, 0.7 mmol) and compound 2 (113.5 mg, 0.5 mmol) was dissolved in DMF (15 mL). The mixture was stirred at 70 °C for 5 h. Then the reaction mixture was washed with water (3× 50 mL). The mixture solution was extracted with dichloromethane (2× 30 mL). The organic layer was dried over Na₂SO₄. Solvent was removed under reduced pressure. White solid was obtained. Yeild: 103.2 mg (88.2%). ¹H NMR (500 MHz, CDCl₃): 7.90–7.79 (m, 3H), 4.19 (s, 2H), 2.26 (s, 3H). TOF HRMS ESI⁺: m/z calcd for [C₉H₁₀N₆O₂+Na]⁺:257.0763; found: 257.0762.

Synthesis of Compound 5. A mixture of *p*-hydroxybenzaldehyde (2.44 g, 20 mmol), 3-bromo-1propyne (3.54 g, 30 mmol) and potassium carbonate (4.15 g, 30 mmol) were dissolved in ethanol (150 mL), then the reaction mixture was refluxed for 6 h. The mixture was concentrated under reduced pressure. The residul was purified by column chromatography (silica gel, CH₂Cl₂). Yield: 2.88 g (60.0 %). ¹H NMR (400 MHz, CDCl₃): 9.91 (s, 1H), 7.88–7.85 (m, 2H), 7.12–7.08 (m, 2H), 4.79 (d, 2H), 2.58 (t, 1H, J = 4.0 Hz). Mp 76.6–77.8 °C. TOF HRMS ES⁺: m/z calcd ([C₁₀H₈O₂+H]⁺): 161.0597; found: 161.0594.

Synthesis of Compound 6. NaBH₄ (304 mg, 8 mmol) and compound 5 (500 mg, 3.1 mmol) were mixed in THF/CH₃OH(V/V=1/1, 50 mL). The mixture was stirred at RT for 5 min. Then the solvent was evaporated under reduced pressure. The crude product was purified by column chromatography (silica gel, CH₂Cl₂). Colourless oil was obtained. Yield: 0.48g (81.8 %). ¹H NMR (500 MHz, CDCl₃): 7.30 (d, 2H, J = 8.5 Hz), 6.97 (d, 2H, J = 8.7 Hz), 4.68 (s, 2H), 4.60 (s, 2H), 2.51 (s, 1H), 1.91 (s, 2H).

Synthesis of Compound 7. Compound 6 (0.21 mg, 1.3 mmol) was dissolved in CH₂Cl₂ (20 mL) and pyridine (270 μ L, 2.6 mmol) was added under Ar atmosphere. The mixture was cooled on an ice bath and a solution of methyl malonyl chloride (270 μ L, 2.6 mmol) in CH₂Cl₂ (10 mL) was added dropwise. The mixture was stirred for 12 h at RT. The crude product was purified by column chromatography (silica gel, CH₂Cl₂) to afford of compound 6 as colourless oil. Yield: 303.7 mg (84.6 %). ¹H NMR (500 MHz, CDCl₃): 7.32 (d, 2H, *J* = 8.5 Hz), 6.97 (d, 2H, *J* = 8.7 Hz), 5.12 (s, 2H), 4.69 (s, 2H), 4.20–4.16 (m, 2H), 3.39 (s, 2H,), 2.52 (s, 1H), 1.24 (t, 2H, *J* = 7.0 Hz). TOF HRMS EI⁺: *m/z* calcd ([C₁₅H₁₆O₅]⁺): 276.0998; found: 276.0990.

Synthesis of Compound 8. Iodine (62 mg, 0.25 mmol) were added to a solution of C_{60} (182.6 mg, 0.27 mmol) in toluene (60 mL). A solution of compound 7 in toluene (150 mL) was added and the mixture was degassed with Ar. A solution of 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU) (72 µL, 0.27 mmol) in toluene (20 mL) was added dropwise within one hour and the reaction mixture was stirred over night. The crude product was purified with column chromatography (silica gel, DCM) to yield 126.6 mg of gray soild, Yield: 51 %. ¹H NMR (500 MHz, CDCl₃): 7.47 (d, 2H, *J* = 8.6 Hz), 7.01 (d, 2H, *J* = 8.7 Hz), 5.47 (s, 2H), 4.71 (s, 2H), 4.50–4.46 (m, 2H), 2.52 (s, 1H), 1.38 (t, 2H, *J* = 7.1 Hz). HRMS (MALDI): m/z calcd for $[C_{75}H_{14}O_5]^+$: 994.0841; found: 994.0844.

Synthesis of Compound 9. Under Ar atmosphere, *p*-hydroxybenzaldehyde (4.88 g, 40 mmol) and anhydrous K₂CO₃ (11.06 g, 80 mmol) were mixed in dry ethanol (150 mL). The mixture was stirred at 70 °C for 0.5 h. The mixture was then cooled to RT and 1,2-dibromoethane (14.9 g, 80 mmol) was added using syringe. The mixture was stirred at 79 °C for 8 h. The mixture was then cooled to RT. K₂CO₃ was removed by filtration and the filtrate was evaporated to dryness under reduced pressure. The product was purified by column chromatography (silica gel, CH₂Cl₂) to afford colourless oil. Yield: 4.32 g (47.2%). ¹H NMR (400 MHz, CDCl₃): 9.90 (s, 1H), 7.87–7.85 (m, 2H), 7.04–7.01 (m, 2H), 4.40 (t, 2H, J = 12.0 Hz), 3.68 (t, 2H, J = 12.0 Hz). TOF HRMS ESI⁺: m/z calcd for [C₉H₉BrO₂+H]⁺: 228.9859; found: 228.9688.

Synthesis of Compound 10. A mixture of compound 8 (1.65g, 7.19 mmol) and NaN₃ (0.6 g, 9.46 mmol) in DMF (20 mL) was heated at 100 °C. After 10 h, the mixture was poured to water . The aqueous phased was extracted with CH_2Cl_2 . The solvent was evaporated under reduced pressure and the azide compound was obtained as colourless oil. Yield: 1.36 g (99 %). The compound was used without any further purification.

Synthesis of Compound 11. Under N₂ atmosphere, a mixture of compound **10** (859.5 mg, 4.5 mmol) and 2,4-dimethylpyrrole (2.0 mL, 945 mg, 9.0 mmol) in dry CH₂Cl₂ (100 mL) was stirred at RT. Under ice-cold condition, trifluoracetic acid (TFA. 0.1 mL) was added. The mixture was stirred at RT over night. A solution of DDQ (1.02 g, 4.5 mmol) in THF (40 mL) was added, then the mixture was stirred at RT for 7 h. Triethylamine (13.5 mL) was added into the mixture dropwise with ice bath cooling, and the mixture was stirred for 0.5 h. BF₃·Et₂O (13.5 mL) was added dropwise via syringe. The reaction mixture was stirred at RT over night. The solution was concentrated under reduced pressure and the residual was poured into water (200 mL), the mixture was stirred for 24 h. The mixture was extract with CH₂Cl₂ and the organic layer was dried over anhydrous MgSO₄. The solvent was evaporated under reduced pressure. The crude product was purified using column chromatography (silica gel, CH₂Cl₂ : hexane = 1:1, v/v) to give **11** as red power . Yield: 214.5 mg (11.6%). ¹H NMR (400 MHz, CDCl₃): 7.21 (d, 2H, *J* = 12.0 Hz), 7.04 (d, 2H, *J* = 8.0 Hz), 5.98 (s, 2H,), 4.21 (t, 2H, *J* = 8.0 Hz), 3.66 (t, 2H, *J* = 8.0 Hz), 2.55 (s, 6H), 1.43 (s, 6H). TOF HRMS ESI⁺: *m*/z calcd for [C₂₁H₂₂BF₂N₅O+H]⁺: 410.2073; found: 410.1938.

Synthesis of Compound 12. Compound 11 (40 mg, 0.1 mmol) was dissolved in dry DMF (5 mL). Then benzaldehyde (42.4 mg 0.4 mmol) was added, followed by acetic acid (3 drops) and piperidine (3 drops). Under Ar atmosphere, the mixture was subjected to microwave irradiation (8 min, 150 °C). Then the solution was poured into ice water, and the mixture was extracted with CH₂Cl₂. The organic layer was dried over Na₂SO₄ and evaporated to dryness. The residule was purified by column chromatography (silica gel, ethyl acetate/hexanes, 1/4, v/v) to give a blue solid. Yield: 12.3 mg (21.0 %). ¹H NMR (500 MHz, CDCl₃): 7.76 (s, 1H), 7.73 (s, 1H), 7.64 (d, 4H, *J* = 7.4 Hz), 7.42–7.39 (m, 4H),

7.34–7.24 (m, 6H), 7.06 (d, 2H, J = 7.6 Hz), 6.65 (s, 2H), 4.23 (t, 2H, J = 4.9 Hz), 3.68 (t, 2H, J = 4.8 Hz), 1.51 (m, 5H). TOF HRMS (MALDI) ESI⁺: m/z calcd for $[C_{35}H_{30}N_5OBF_2]^+$: 585.2511; found: 585.2516.

Synthesis of Compound 14. A mixture of carbazole (1.161 g, 6.95 mmol) and NaH (0.695 g, 17.35 mmol, 60% oil dispersion) in dry DMF (10 mL) was stirred at RT for 1 h. n-Butyl bromide (0.472 g, 3.47 mmol) was added, and the mixture was stirred for 1 h. The mixture was then poured into water (150 mL), and the precipitate was filtered, dissolved in dichloromethane (50 mL), dried over Na₂SO₄, and evaporated to dryness. The crude product was purified by column chromatography (silica gel, petroleum ether), to give **14** as white solid. Yield: 1.47 g (95%). ¹H NMR (400 MHz, CDCl₃): 8.10 (d, 2H, J = 8 Hz), 7.44 (d, 1H, J = 7.2 Hz), 7.39 (d, 2H, J = 8 Hz), 7.21 (t, 2H, J = 7.2 Hz), 4.27 (t, 2H, J = 7.2 Hz), 1.87–1.79 (m, 2H), 1.43–1.33 (m, 2H), 0.92–0.90 (m, 2H). TOF HRMS ESI⁺: *m/z* calcd for [C₁₆H₁₇N+H]⁺: 224.1434; found: 224.1435.

Synthesis of Compound 15. POCl₃ (4.2 mL) was added dropwise into DMF (2 mL) while the mixture was cooled with ice-bath. Then the temperature of the mixture was warmed to RT. The reaction mixture was stirred at RT for 2 h. 9-Butyl-9*H*-carbazole (2.00 g, 8.96 mmol) was added into the reaction mixture at 60 °C. After 6 h, the reaction was quenched with saturated NaHCO₃ solution (100 mL). Then the mixture was stirred for 1 h. The precipitate was collected with filtration and the solid was washed with cold ethanol (50 mL). The crude product was purified by column chromatography (silica gel, CH₂Cl₂ : petroleum ether = 1:1, v/v) to give the 9-butyl-9H-carbazole-3-carbaldehyde as colourless oil. Yield: 1.58 g (70%). ¹H NMR (400 MHz, CDCl₃): 10.08 (s, 1H), 8.58 (s, 1H), 8.14 (d, 2H, *J* = 7.8 Hz), 8.00 (d, 2H, *J* = 8.5 Hz), 7.52 (t, 1H, *J* = 8.3 Hz), 7.45 (d, 2H, *J* = 8.6 Hz), 7.31 (t, 1H, *J* = 7.2 Hz), 4.30 (t, 2H, *J* = 7.1 Hz), 1.89–1.81 (m, 2H), 1.44–1.35 (m, 2H), 0.95 (t, 3H, *J* = 8.7 Hz). TOF HRMS ESI⁺: *m*/z calcd for [C₁₇H₁₇NO+H]⁺: 252.1383; found: 252.1381.

Synthesis of Compound 16. Compound **15** (69 mg, 0.3 mmol) was dissolved in dry DMF (5 mL). Then compound **11** (109.8 mg 0.3 mmol) was added, followed by acetic acid (10 drops) and piperidine (10 drops). The mixture was put under an Ar atmosphere before it was subjected to microwave

irradiation (5 min, 150 °C). After the microwave irradiation, the solution was poured into ice water and the mixture was extracted with CH₂Cl₂ (3×30 mL). The organic layer was dried over Na₂SO₄ and evaporated to dryness, the mixture was purified by column chromatography (silica gel, ethyl acetate/hexanes, 1/15, v/v) to give blue solid. Yield: 42 mg (22.1 %). ¹H NMR (400 MHz, CDCl₃): 8.29 (s, 1H), 8.17 (d, 1H, J = 7.8 Hz), 7.78–7.70 (m, 2H), 7.48 (t, 2H, J = 8.3 Hz), 7.40 (t, 2H, J = 8.8 Hz), 7.28–7.22 (m, 3H), 7.06 (d, 1H, J = 8.7 Hz), 6.67 (s, 1H), 6.00 (s, 1H), 4.31 (t, 2H, J = 7.1 Hz), 4.22 (t, 2H, J = 5.0 Hz), 3.67 (t, 2H, J = 5.0 Hz), 2.63 (s, 3H), 1.91–1.83 (m, 2H), 1.51 (s, 3H), 1.45 (s, 3H), 1.42–1.38 (m, 2H), 0.96 (t, 3H, J = 7.4 Hz). HRMS (MALDI): m/z calcd for [C₃₈H₃₇N₆OF₂B]⁺: 642.3065; found: 642.3090.

Synthesis of Compound 17. Under Ar atmosphere, thymine (500 mg, 4 mmol) and K₂CO₃ (828 mg, 6 mmol) were dissolved in dry DMF (6 mL). Then a solution of propargyl bromide (1.19 g 10 mmol) in *n*-hexane (0.8 mL) was added dropwise. The reaction mixture was stirred over night. The mixture was poured into ice water. The mixture was extracted with CH₂Cl₂, dried over Na₂SO₄ and evaporated to dryness. The mixture was purified by column chromatography (silica gel, ethyl acetate/hexanes, 1/2, v/v) to give a white solid. Yield: 123.3 mg (18.8 %). ¹H NMR (400 MHz, CDCl₃): 9.04 (s, 1H), 7.25 (s, 1H), 4.55 (d, 2H, J = 29.5 Hz), 2.47 (t, 1H, J = 2.5 Hz), 1.96 (s, 3H). TOF HRMS ESI⁺: *m/z* calcd for [C₈H₈N₂O]⁺:164.0586; found: 164.0593.

Synthesis of Compound 19. Compound **18** (100 mg, 0.61 mmol) was dissolved in dry DMF (5 mL). Then NaH (152 mg 1.3 mmol) was added under ice-bath cooling, followed by lithium bromide (156.6 mg, 1.8 mmol). The mixture was kepted in ice-bath for 20 min. Then methyl iodide (260 mg, 1.8 mmol) was added and the mixture was stirred at RT overnight. The solution was poured into ice water. The mixture was extracted with CH_2Cl_2 , the organic layer was dried over Na_2SO_4 and evaporated to dryness. The crude product was purified by column chromatography (silica gel, dichloromethane/methnol, 40/1, v/v) to give white solid. Yield: 50.8 mg (46.7 %). ¹H NMR (500 MHz, CDCl₃): 7.24 (s, 1H), 4.57 (s, 2H), 3.37 (s, 3H), 2.45 (s, 1H), 1.97 (s, 3H,). TOF HRMS ESI⁺: *m/z* calcd for $[C_9H_{10}N_2O_2]^+$: 178.0472; found: 178.0473.

3.0 Abosption and Emssion details



Fig. S45 UV-vis absorption spectra of (a) **B-1-Me**, **C-1** and **C-1/B-1-Me**=1/1, (b) **C-1** solution $(1.0 \times 10^{-5} \text{ M})$ in toulene with increasing of **B-1-Me**. c = $1.0 \times 10^{-5} \text{ M}$ in toluene. 20 °C.



Fig. S46 UV-vis absorption spectra of (a) **B-2**, **C-1** and **C-1/B-2=**1/1, (b) **C-1** solution (1.0×10^{-5} M) in toulene with increasing of **B-2**. c = 1.0×10^{-5} M in toluene. 20 °C.



Fig. S47 UV-vis absorption spectra of (a) **B-2-Me**, **C-1** and **C-1/B-2-Me**=1/1, (b) **C-1** solution $(1.0 \times 10^{-5} \text{ M})$ in toulene with increasing of **B-2-Me**. c = $1.0 \times 10^{-5} \text{ M}$ in toluene. 20 °C.



Fig. S48 UV-vis absorption spectra of (a) **B-3**, **C-1** and **C-1/B-3** = 1/1, (b) **C-1** solution $(1.0 \times 10^{-5} \text{ M})$ in toulene with increasing of **B-3**. $c = 1.0 \times 10^{-5} \text{ M}$ in toluene. 20 °C.



Fig. S49 UV-vis absorption spectra of (a) **B-3-Me**, **C-1** and **C-1/B-3-Me** =1/1, (b) **C-1** solution (1.0×10^{-5} M) in toulene with different concentration **B-3-Me** solution. c = 1.0×10^{-5} M in toluene. 20 °C.



Fig. S50 (a) B-1 and B-1-Me dependence of I_0/I on the concentration of C-1; (b) Emission intensity of B-1 and B-1-Me at constant concentration of 10^{-5} M in toluene, *versus* increasing concentration of [C-1] at 517 nm following excitation at 475 nm.



Fig. S51 Fluorescence of (a) **B-2** and (b) **B-2-Me** solution $(1.0 \times 10^{-5} \text{M})$ in toluene quenched by C-1 solution in toluene ($\lambda_{ex} = 600 \text{ nm}$).



Fig. S52 (a) B-2 and B-2-Me dependence of I_0/I on the concentration of C-1, (b) Emission intensity of B-2 at constant concentration of 10^{-5} M in toluene, *versus* increasing concentration of [C-1] at 642 nm following excitation at 600 nm.



Fig. 53 Fluorescence titration (a) **B-3** and (b) **B-3-Me** solution $(1.0 \times 10^{-5} \text{M})$ in toulene quenched by **C-1** solution in toluene ($\lambda_{\text{ex}} = 560 \text{ nm}$). (b) Emission intensity of **B-3** at constant concentration of 10^{-5} M in toluene, *versus* increasing concentration of [**C-1**] at 611 nm following excitation at 560 nm.



Fig. S54 Fluorescence of (a) B-1, B-1 + 12eq C-1 + 100 μ L HFTP and B-1 + 12eq C-1 ($\lambda_{ex} = 475$ nm), (b) B-2, B-2 + 12eq C-1 + 100 μ L HFTP and B-2 + 12eq C-1 ($\lambda_{ex} = 600$ nm), (c) B-3, B-3 + 12eq C-1 + 100 μ L HFTP and B-3 + 12eq C-1 ($\lambda_{ex} = 560$ nm). In toluene. 20 °C.



Fig. S55 The emssion spectra of C-1. c = 1.0×10^{-5} M in toluene ($\lambda_{ex} = 331$ nm). 20 °C.



4.0 Transient absorption details

Fig. S56 Nanosecond time-resolved transient absorption difference spectra with different delay times of compound **C-1** with adding compound **B-1-Me**. (a) 0 eq **B-1-Me**, (b) 0.5 eq **B-1-Me**, (c) 1 eq **B-1-Me**. (d) In different concentration of **B-1-Me** with delay times at 0 μ s. C = 1.0×10⁻⁵ M, 20 °C. In toluene after pulsed excitation at 495 nm under N₂.



Fig. S57 Nanosecond time-resolved transient absorption difference spectra of compound C-1 with adding compound B-1-Me. Decay trace at 720 nm. In toluene after pulsed excitation at 495 nm under N₂. 0eq: 42.8 μ s, 0.5eq: 42.8 μ s, 1 eq: 46.3 μ s.



Fig. S58 Nanosecond time-resolved transient absorption difference spectra with different delay times of compound **C-1** with adding compound **B-2**. (a) 0 eq **B-2**, (b) 0.25 eq **B-2**, (c) 0.5 eq **B-2**, (d) 1 eq **B-2**. C = 1.0×10^{-5} M, 20 °C. In toluene after pulsed excitation at 620 nm under N₂.



Fig. S59 Nanosecond time-resolved transient absorption difference spectra of compound C-1 with adding compound B-2. (a) Decay trace at 630 nm, (b) with delay times at 0 μ s. C = 1.0×10⁻⁵ M, 20 °C. In toluene after pulsed excitation at 620 nm under N₂.

0 eq: - µs, 0.25eq: 312.4 µs, 0.5 eq: 250.7 µs, 1 eq: 185.5 µs.



Fig. S60 Nanosecond time-resolved transient absorption decay trace at 630 nm of mixture solution of C-1 and 0.5eq B-2-Me. 20 °C. $c = 1.0 \times 10^{-5}$ M, In toluene after pulsed excitation at 425nm and 620 nm under N₂.



Fig. S61 Nanosecond time-resolved transient absorption difference spectra with different delay times of compound **C-1** with adding compound **B-2-Me**. (a) 0 eq **B-2-Me**, (b) 0.5 eq **B-2-Me**, (c) 1 eq **B-2-Me**, (d) Decay trace at 630 nm. $c = 1.0 \times 10^{-5}$ M, 20 °C. In toluene after pulsed excitation at 425 nm under N₂.



Fig. S62 Nanosecond time-resolved transient absorption decay trace at 500 nm: (a) **B-1**, (b) **B-1-Me**. 20 °C. $c = 1.0 \times 10^{-5}$ M, In toluene after pulsed excitation at 495 nm under N₂.



Fig. S63 Nanosecond time-resolved transient absorption decay trace at 630 nm: (a) **B-2**, (b) **B-2-Me**. 20 $^{\circ}$ C. c = 1.0×10⁻⁵ M, In toluene after pulsed excitation at 620 nm under N₂.



Fig. S64 Nanosecond time-resolved transient absorption decay trace at 593 nm: (a) **B-3**, (b) **B-3-Me**. 20 $^{\circ}$ C. c = 1.0×10⁻⁵ M, In toluene after pulsed excitation at 590 nm under N₂.



Fig. S65. Nanosecond time-resolved transient absorption difference spectra and decay curves of compounds. (a) **C-1** under nitrogen. (b) Decay curve of **C-1** at 720 nm in air, after pulsed excitation at 410 nm. the absorption for **C-1** at 410 nm is 0.096 in toluene, 20 °C.



Fig. S66. Nanosecond time-resolved transient absorption difference spectra and decay curves of compounds. (a) $C-1/B-1(c_{c-1}:c_{B-1}=1:1)$ under nitrogen. (b) Decay curve of $C-1/B-1(c_{c-1}:c_{B-1}=1:1)$ at 720 nm in air, after pulsed excitation at 410 nm. the absorption for $C-1/B-1(c_{c-1}:c_{B-1}=1:1)$ at 410 nm is 0.096 in toluene, 20 °C.

5.0 The detailed data of electrochemistry.

Calculation of the free energy changes of the electron transfer

The free energy changes of the electron transfer process (charge seperation, CS), can be calculated with the Weller equation (eq. 1 and eq.2).

$$\Delta G^{0}_{CS} = e[E_{OX} - E_{RED}] - E_{00} + \Delta G_{S}$$
 (Eq. 1)

$$\Delta G_{\rm S} = -\frac{e^2}{4\pi\varepsilon_{\rm S}\varepsilon_{\rm 0}R_{\rm CC}} - \frac{e^2}{8\pi\varepsilon_{\rm 0}} \left(\frac{1}{R_{\rm D}} + \frac{1}{R_{\rm A}}\right) \left(\frac{1}{\varepsilon_{\rm REF}} - \frac{1}{\varepsilon_{\rm S}}\right)$$
(Eq. 2)

Where ΔG_S is the static Coulombic energy, which is desribed by eq. 2. e = electronic charge, E_{OX} = half-wave potential for mone-electron oxidation of the electron-donor unit, E_{RED} = halfwave potential for one-electron reduction of the electron-acceptor unit; note herein the anodic and cathodic peak potentials were used because in some cases the oxidation is iiriversible therefore the formal potential $E_{1/2}$ cannot be derived; E_{00} = energy level approximated with the fluorescence emission wavelength (for the singlet excited state), or T_1 state energy of photosensitizers. ε_S = static dielectric constant of the solvent, RCC = (xx Å) center-to-center separation distance determined by DFT optimization of the geometry, R_D is the radius of the BODIPY-based donor, R_A is the radius of the electron acceptor, ε_{REF} is the static dielectric constant of the solvent used for the electrochemical studies, ε_0 permittivity of free space(ε_0 = 8.854187817×10⁻¹²F/m). The solvents used in the calculation of free energy of the electron transfer is DCM (ε = 9.1, 20 °C) and Toluene (ε = 2.4, 20 °C).

For **B-1** and **C-1** in the solution,

Due to **B-1** and **C-1** can develop Hydrogen bonding in the aprotic solvent, we proposed that the center-to-center separation distance between **B-1** and **C-1** is close to the distance of covalent linkage of **B-1** and **C-1**. Here RCC = (40 Å), RA = 4.1 Å, RD = 4.1 Å, was abtained from the ground state optimized by TDDFT calculation.

In DCM,

Based on these parameters, for **B-1** and **C-1** in DCM, ΔG_S is calculated as -0.04 eV.

With reference **C-1** as electron acceptor, the $\Delta G(CS) = 1.11 - (-0.70) - 1.56 - 0.04 = 0.21 \text{ eV}$; With reference **B-1** as electron donor, the $\Delta G(CS) = 1.11 - (-0.70) - 2.45 - 0.04 = -0.68 \text{ eV}$; In Toluene, ΔG_S is calculated as 0.92 eV.

With reference C-1 as electron acceptor, the $\Delta G(CS) = 1.11 - (-0.70) - 1.56 + 0.92 = 1.17 \text{ eV}$;

With reference **B-1** as electron donor, the $\Delta G(CS) = 1.11 - (-0.70) - 2.45 + 0.92 = 0.28 \text{ eV}$;

For **B-1-Me** and **C-1** in the solution,

Due to **B-1-Me** and **C-1** cannot develop Hydrogen bonding in the aprotic solvent, we proposed that the center-to-center separation distance between **B-1** and **C-1** is infinity. Here RCC = ∞ . (Refer to JPCA)

In DCM, ΔG_S is calculated as 0.00 eV.

Based on these parameters, for **B-1** and **C-1** in DCM, ΔG_S is calculated as 0 eV.

With reference C-1 as electron acceptor, the $\Delta G(CS) = 1.11 - (-0.70) - 1.56 - 0.00 = 0.25 \text{ eV}$;

With reference **B-1-Me** as electron donor, the $\Delta G(CS) = 1.11 - (-0.70) - 2.45 - 0.00 = -0.64 \text{ eV}$;

In Toluene, ΔG_S is calculated as 1.08 eV.

With reference C-1 as electron acceptor, the $\Delta G(CS) = 1.11 - (-0.70) - 1.56 + 1.08 = 1.33 \text{ eV}$;

With reference **B-1-Me** as electron donor, the $\Delta G(CS) = 1.11 - (-0.70) - 2.45 + 1.08 = 0.44 \text{ eV}$;

For **B-2** and **C-1** in the solution,

Due to **B-2** and **C-1**can develop Hydrogen bonding in the aprotic solvent, we proposed that the center-to-center separation distance between **B-2** and **C-1** is close to the distance of covalent linkage of **B-2** and **C-1**. Here RCC = (43.9 Å), RA = 4.1 Å, RD = 11.1 Å, was obtained from the ground state optimized by TDDFT calculation.

In DCM,

Based on these parameters, for **B-2** and **C-1** in DCM, ΔG_S is calculated as -0.036 eV.

With reference **C-1** as electron acceptor, the $\Delta G(CS) = 0.84 - (-1.04) - 1.56 - 0.036 = 0.28 \text{ eV}$;

With reference **B-2** as electron donor, the $\Delta G(CS) = 0.84 - (-1.04) - 1.93 - 0.036 = -0.09 \text{ eV}$; In Toluene, ΔG_S is calculated as 0.60 eV.

With reference C-1 as electron acceptor, the $\Delta G(CS) = 0.84 - (-1.04) - 1.56 + 060 = 0.92 \text{ eV}$;

With reference **B-2** as electron donor, the $\Delta G(CS) = 0.84 - (-1.04) - 1.93 + 0.60 = 0.55 \text{ eV}$;

For B-2-Me and C-1 in the solution,

Due to **B-2-Me** and **C-1** cannot develop Hydrogen bonding in the aprotic solvent, we proposed that the center-to-center separation distance between **B-2-Me** and **C-1** is infinity. Here RCC = ∞ . (Refer to JPCA)

In DCM, ΔG_S is calculated as 0.00 eV.

Based on these parameters, for **B-2-Me** and **C-1** in DCM, ΔG_S is calculated as 0.00 eV.

With reference **C-1** as electron acceptor, the $\Delta G(CS) = 0.84 - (-1.04) - 1.56 - 0.00 = 0.32 \text{ eV}$;

With reference **B-2-Me** as electron donor, the $\Delta G(CS) = 0.84 - (-1.04) - 1.93 - 0.00 = -0.05$ eV;

In Toluene, ΔG_S is calculated as 0.74 eV.

With reference C-1 as electron acceptor, the $\Delta G(CS) = 0.84 - (-1.04) - 1.56 + 0.74 = 1.06 \text{ eV}$;

With reference **B-2-Me** as electron donor, the $\Delta G(CS) = 0.84 - (-1.04) - 1.93 + 0.74 = 0.69 \text{ eV}$;

For **B-3** and **C-1** in the solution,

Due to **B-3** and **C-1**can develop Hydrogen bonding in the aprotic solvent, we proposed that the center-to-center separation distance between **B-3** and **C-1** is close to the distance of covalent linkage of **B-3** and **C-1**. Here RCC = (49.5 Å), RA = 4.1 Å, RD = 13.5 Å, was obtained from the ground state optimized by TDDFT calculation.

In DCM,

Based on these parameters, for **B-3** and **C-1** in DCM, ΔG_S is calculated as -0.032 eV.

With reference **C-1** as electron acceptor, the $\Delta G(CS) = 0.74 - (-1.21) - 1.56 - 0.032 = 0.36 \text{ eV}$;

With reference **B-3** as electron donor, the $\Delta G(CS) = 0.74 - (-1.21) - 2.09 - 0.032 = -0.17 \text{ eV}$; In Toluene, ΔG_S is calculated as 0.58 eV.

With reference **C-1** as electron acceptor, the $\Delta G(CS) = 0.74 - (-1.21) - 1.56 + 0.58 = 0.97 \text{ eV}$; With reference **B-3** as electron donor, the $\Delta G(CS) = 0.74 - (-1.21) - 2.09 + 0.58 = 0.44 \text{ eV}$;

For **B-3-Me** and **C-1** in the solution,

Due to **B-3-Me** and **C-1** cannot develop Hydrogen bonding in the aprotic solvent, we proposed that the center-to-center separation distance between **B-3-Me** and **C-1** is infinity. Here RCC = ∞ . (Refer to JPCA)

In DCM, ΔG_S is calculated as 0.00 eV.

Based on these parameters, for **B-3-Me** and **C-1** in DCM, ΔG_S is calculated as 0.00 eV.

With reference **C-1** as electron acceptor, the $\Delta G(CS) = 0.74 - (-1.22) - 1.56 - 0.00 = 0.40 \text{ eV}$; With reference **B-3-Me** as electron donor, the $\Delta G(CS) = 0.74 - (-1.22) - 2.09 - 0.00 = -0.13 \text{ eV}$; In Toluene, ΔG_S is calculated as 0.70 eV.

With reference C-1 as electron acceptor, the $\Delta G(CS) = 0.74 - (-1.22) - 1.56 + 0.74 = 1.14 \text{ eV}$;

With reference **B-3-Me** as electron donor, the $\Delta G(CS) = 0.74 - (-1.22) - 2.09 + 0.74 = 0.61$ eV.



Fig. S67 Cyclic voltammogram of the compound **B-4**. Ferrocene (Fc) was used as internal reference $(E_{1/2} = +0.38 \text{ V} (\text{Fc}^+/\text{Fc}) \text{ vs. SCE})$. In deaerated DCM solutions containing 0.5 mM photosensitizers alone, or with the ferrocene, 0.10 M Bu₄NPF₆ as supporting electrolyte, Ag/AgNO₃ reference electrode, Scan rates: 0.05 V/s.

6.0 The detailed data of fluorenscence lifetime.



Fig. S68. Dependence of τ on the concentration of **C-1**: (a) **B-1** solution $(1.0 \times 10^{-5} \text{ M})$ in toulene, measurement wavelength 505 nm,(b) **B-2** solution $(1.0 \times 10^{-5} \text{ M})$ in toulene measurement wavelength 610 nm, (c) **B-3** solution $(1.0 \times 10^{-5} \text{ M})$ in toulene measurement wavelength 645nm. (d) **B-1-Me** solution $(1.0 \times 10^{-5} \text{ M})$ in toulene, measurement wavelength 505 nm,(e) **B-2-Me** solution $(1.0 \times 10^{-5} \text{ M})$ in toulene measurement wavelength $(1.0 \times 10^{-5} \text{ M})$ in toulene measurement wavelength $(1.0 \times 10^{-5} \text{ M})$ in toulene measurement wavelength $(1.0 \times 10^{-5} \text{ M})$ in toulene, measurement wavelength $(1.0 \times 10^{-5} \text{ M})$ in toulene measurement wavelength $(1.0 \times 10^{-5} \text{ M})$ in toulene measurement wavelength $(1.0 \times 10^{-5} \text{ M})$ in toulene measurement wavelength $(1.0 \times 10^{-5} \text{ M})$ in toulene measurement wavelength $(1.0 \times 10^{-5} \text{ M})$ in toulene measurement wavelength $(1.0 \times 10^{-5} \text{ M})$ in toulene measurement wavelength $(1.0 \times 10^{-5} \text{ M})$ in toulene measurement wavelength $(1.0 \times 10^{-5} \text{ M})$ in toulene measurement wavelength $(1.0 \times 10^{-5} \text{ M})$ in toulene measurement wavelength $(1.0 \times 10^{-5} \text{ M})$ in toulene measurement wavelength $(1.0 \times 10^{-5} \text{ M})$ in toulene measurement wavelength $(1.0 \times 10^{-5} \text{ M})$ in toulene measurement wavelength $(1.0 \times 10^{-5} \text{ M})$ in toulene measurement wavelength $(1.0 \times 10^{-5} \text{ M})$ in toulene measurement wavelength $(1.0 \times 10^{-5} \text{ M})$ in toulene measurement wavelength $(1.0 \times 10^{-5} \text{ M})$ in toulene measurement wavelength $(1.0 \times 10^{-5} \text{ M})$ in toulene measurement wavelength $(1.0 \times 10^{-5} \text{ M})$ in toulene measurement wavelength $(1.0 \times 10^{-5} \text{ M})$ in toulene measurement wavelength $(1.0 \times 10^{-5} \text{ M})$ in toulene measurement wavelength $(1.0 \times 10^{-5} \text{ M})$ in toulene measurement wavelength $(1.0 \times 10^{-5} \text{ M})$ in toulene measurement wavelength $(1.0 \times 10^{-5} \text{ M})$ in toulene measurement wavelength $(1.0 \times 10^{-5} \text{ M})$ in toulene measurement wavele

7.0 Molecular dynamics (MD) simulations.



Fig. S69 Conformations of **B-1-C-1** complex in aqueous solution. Left: the starting extended conformation used in MD simulations. Right: Conformation taken from MD simulation at 9.5 ns, showing the hydrogen bonds (<3 Å as indicated using black dotted lines) between **B-1** and **C-1** motif, and π - π interactions between B-1 and fullerene motif of **C-1**. The white surfaces represent the solvent (water molecules) accessible surface areas for **B-1** and fullerene motif of **C-1** structures. Water molecules are not shown in the conformation at 9.5 ns for clarity.

The possible conformation of complex including hydrogen acceptor and donor was explored by molecular dynamics (MD) simulations performed in aqueous solution. It was found that a relative stable complex can be formed in less than 10 ns, indicating that the recognition of hydrogen donor and acceptor is rather feasible in aqueous solution. Note that three pairs of hydrogen bonds in the initial conformation are largely preserved in the compact conformation, and the π - π interactions between **B-1** and fullerene motif of **C-1** molecules further contribute the stability of such complex.

Fig. S70 Atomic partial charges calculated for (a) B-1, (b) C-1, and (c) B-3 molecules by DFT method and used in the MD simulations.

8.0 The detailed data of Triplet-Triplet annihilation upconversion.

Fig. S71 TTA upconversions of **C-1** with H-bonding modules of **B-3**. **1-CBPEA** was used as triplet acceptor. The emission of **B-3** and **1-CBPEA** with increasing **C-1** added. Excited with 589 nm laser (4.8 mW, power density is 65 mW cm $^{-2}$) and a band-pass filter (470 nm – 570 nm) was used to suppress the scattered laser. c[Photosensitizers] = 1.0×10^{-5} M. c [1-CBPEA] = 1.2×10^{-4} M in deaerated toluene. 20 °C.

Fig. S72 The spectra of Transmittance as Y-axis and Wavelength as X-axis on the band-pass filter (470 nm-570 nm). The data was supplied by Giai photnics Co., Ltd (ShenZhen, China).