## **Supporting Information**

## Direct characterization of the native structure and mechanics of cyanobacterial carboxysomes

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**Table S1** Proteomic results of isolated  $\beta$ -carboxysomes from Syn7942. The column of Normalized amount displays the amount of each of the carboxysomal proteins detected in isolated  $\beta$ -carboxysomes using mass spectroscopy, normalized against the amount of the least abundant protein CcmL.

Protein	Normalized amount (fmol)		
RbcL	4530.8 ± 432.8		
RbcS	1744.5 ± 765.1		
CcmM	1567.1 ± 412.8		
CcmK2	116.6 ± 22.0		
CcaA	81.8 ± 12.6		
CcmK4	18.7 ± 0.4		
CcmL	1		

Table S2 Comparison of the dimensions of isolated carboxysomes from different species using TEM.

Туре	Species	Diameter (nm)	Range (nm)	Reference
α-carboxysome	Halothiobacillus neapolitanus	117.3 ± 6.9	97 – 132	1
α-carboxysome	Halothiobacillus neapolitanus	100	88 - 108	2
α-carboxysome	Halothiobacillus neapolitanus	134 ± 8	116 – 169	3
α-carboxysome	Synechococcus WH8102	123 ± 5	114 – 137	4
α-carboxysome	Prochlorococcus marinus MED4	90	70 - 100	5
β-carboxysome	Synechococcus elongatus PCC7942	149.90 ± 13.78	100 - 200	This study



Fig. S1 Immunoblotting analysis of different  $\beta$ -carboxysome fractions using anti-RbcL antibody. Immunoblotting assays were carried out on the SDS-PAGE gel shown in Fig. 1C. RbcL (~50 kDa) was detected in all sucrose fractions and was most abundant in the 40 % sucrose fraction.



Fig. S2 TEM images of partial  $\beta\text{-}carboxysome$  fragments in the 20 and 30% sucrose fractions.



**Fig. S3** TEM images of intact  $\beta$ -carboxysomes in the 40% sucrose fraction. (A) TEM images of individual intact  $\beta$ -carboxysomes. The dashed arrows represent the vertex-to-vertex measurements for determining the  $\beta$ -carboxysome diameter as described in Fig. 4B. (B) TEM images of  $\beta$ -carboxysome aggregations (Fig. 4C). Scale bar: 100 nm.



**Fig. S4** Combined confocal and AFM imaging of  $\beta$ -carboxysomes fused with GFP. (A) A merged image of the transmitted and GFP channels captured using a hybrid JPK AFM-Zeiss 880 confocal microscope. The white dashed square represents a 10 × 10 µm field of view of AFM after the engage. (B) Fluorescence image of a single  $\beta$ -carboxysome in the view highlighted by the white square in panel A. (C) AFM topograph of the same  $\beta$ -carboxysome captured simultaneously with the fluorescence image (B). The combination of AFM-confocal fluorescence imaging ensures the identification of  $\beta$ -carboxysomes on AFM substrate.



**Fig. S5** AFM images of intact  $\beta$ -carboxysomes. (A) AFM topograph of single  $\beta$ -carboxysome with a vertex and three facet boundaries resolved, indicated by the green dashed lines. (B) AFM topographs of aggregated  $\beta$ -carboxysomes, reminiscent of EM results (Fig. 4C, Fig. S3B).



**Fig. S6** Statistical analysis of the nanomechanical properties of  $\beta$ -carboxysomes. (A) Histogram of the  $\beta$ -carboxysome stiffness ( $k_{CB}$ , n = 25, Equation 1). (B) Histogram of Young's moduli of  $\beta$ -carboxysomes ( $E_H$ , n = 20) using the Hertzian model (Equation 3). (C) There is no correlation between Young's moduli and the carboxysome diameter (y = -0.0039x + 1.1,  $R^2 = 0.2278$ ). (D) Histogram of Young's moduli of  $\beta$ -carboxysomes ( $E_S$ , n = 25) using the thin shell model (Equation 2).



**Fig. S7** Characterization of P22 particles. (A) TEM images of isolated P22 bacteriophage. (B) AFM topograph of a single P22 bacteriophage. The average height is  $65.1 \pm 5.9$  nm (n = 20), in good agreement with previous AFM data.<sup>6</sup> (C) Force-indentation curves of individual P22 particles. (D) The force-indentation curves of a single  $\beta$ -carboxysome (circle), a single P22 particle (square) and simulated force-indentation curves (colored dash lines) using a Hertz contact model in a sample with Young's moduli of 0.5, 1 and 10 MPa. The height of P22 particles is  $65.1 \pm 5.9$  nm (n = 20) and the spring constant of P22 is approximately 192.38  $\pm 63.77$  pN/nm (n = 8). Young's moduli of P22 fitted to the linear model and the Hertzian model are  $101.04 \pm 32.29$  MPa and  $11.06 \pm 8.77$  respectively (n = 8). Young's modulus of  $\beta$ -carboxysomes obtained using the Hertzian model ( $E_{\rm H} = 0.59 \pm 0.34$  MPa, n = 20) is significantly lower than those of P22, demonstrating the mechanical softness of  $\beta$ -carboxysome structures compared with P22.

## **Supporting References**

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