Appendix

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Figure S1.

Akt inhibition with AZD5363 can be demonstrated in a wide range of cancer cells in vitro.

A-D. Cells of differing origins were treated with 1 μ M or 10 μ M AZD5363 for 2 hours or 24 hours (A), or 2 hours only (B-D), before lysis on ice; (n = 3 experiments). (A-B), Western blots were performed with antibodies detecting pAkt, pGSK3 β , pS6 and the housekeeping protein GAPDH or vinculin. A representative is shown for each cell line, where (A) shows cells of a head and neck cancer origin, and (B), cells of a cervical cancer origin. (C-D), ELISA was used to measure levels of pPRAS40.

Data information: in (C) and (D) data is shown as mean \pm SEM; n/s P > 0.05, *P < 0.05, **P < 0.01, ***P < 0.001, **** P < 0.001). Statistical test is one-way ANOVA with Dunnett's *post hoc* test (for (C); P = 0.001-0.834, and for (D); P = 0.0001-0.0007).





Figure S2. AZD5363 has a cell line dependent effect on viability *in vitro*.

A-C. Cell lines of head and neck cancer (A), cervical cancer (B) and breast and lung cancer (C) origin, were treated with 1-10 μ M AZD5363 for 96hrs before an MTT assay was performed; (*n* = 3 experiments). Data information: Results are displayed as mean ± SÉM, normalised to a DMSO treated control and fitted to a dose-response curve.



Figure S3.

Akt inhibition with AZD5363 does not enhance the radiosensitivity of tumour cells *in vitro* and has no effect on the cell cycle of FaDu cells with or without radiation.

A-H. Clonogenic assay of (A) A549, (B) Calu-6, (C) Cal-27, (D) RPMI-2650, (E) Detroit 562, (F) MCF 7, (G) PE/CA PJ15 and (H) SiHa cells treated with 1 μ M or 10 μ M AZD5363 for 2 hours before, and 24 hours after a single dose of RT (2, 4 or 6 Gy); (*n* = 3 experiments).

I. Surviving fraction of cells of a cervical cancer origin after a single 4 Gy dose of RT combined with 10 μ M AZD5363; (*n* = 3 experiments).

J. Flow cytometric analysis of FaDu cell cycle with propidium iodide which shows no effect of 24hrs 1 μ M AZD5363 either alone or after 4 Gy irradiation; (*n* = 3 experiments).

Data information: In (A-H) values are shown of surviving fraction (mean \pm SEM) and are fitted to the linear quadratic model. In (I) values are of surviving fraction (mean \pm SEM). Statistical test is unpaired t-test, one per cell line, (P = 0.163-0.844). In (J) stacked bars represent mean percentage total cells in each treatment group; n/s P > 0.05. Statistical test is Kruskal-Wallis with Dunn's *post hoc* test (P = > 0.9999).



Figure S4.

The combination of RT with AZD5363 does not cause loss of animal weight.

A. Mean weights of FaDu tumour bearing mice treated with AZD5363 (50 mg/kg BD) or 6 Gy RT alone, or in combination, with the drug given as an adjuvant, continuously, or as a neo-adjuvant.

Data information: Plots show mean weights and are the combined data of 2 independent experiments; (n = 6-13 mice/experiment).



Figure S5.

AZD5363 targets the Akt pathway when used *in vivo*. A-B. FaDu tumour bearing mice were treated with AZD5363 (50 mg/kg) for 7 days (14 doses) after which mice were culled and lysates prepared; (n = 6-7 per group). (A), Western blot with antibodies detecting pAkt, pS6 and the housekeeping protein vinculin. (B), pPRAS40 levels as measured by ELISA; * P < 0.05. Data information: In (B) data shows individual results with line to represent median value. Statistical test is Mann-Whitney test (* P = 0.032).

А

Cell Line	Origin	Source	Media
FaDu	H&N	ATCC	Advanced MEM
			+10%FCS
			+ 10ml 100x L-Glutamine
CAL-27	H&N	ATCC	DMEM
			+10%FCS
			+ 10ml 100x L-Glutamine
Detroit-563	H&N	ATCC	EMEM
			+10%FCS
			+ 10ml 100x L-Glutamine
RPMI 2650	H&N	ATCC	EMEM
			+10%FCS
			+ 10ml 100x L-Glutamine
PE/CA PJ15	H&N	ECACC	IMDM
			+10%FCS
			+ 10ml 100x L-Glutamine
PE/CA PJ34 (Clone	H&N	ECACC	IMDM
C12)			+10%FCS
			+ 10ml 100x L-Glutamine
ME-180	Cervical	ATCC	DMEM
			+10%FCS
			+ 10ml 100x L-Glutamine
Ca Ski	Cervical	ATCC	RPMI-1640
			+10%FCS
			+ 10ml 100x L-Glutamine
SiHa	Cervical	ATCC	EMEM
			+10%FCS
			+ 10ml 100x L-Glutamine
C-33 A	Cervical	ATCC	EMEM
			+10%FCS
			+ 10ml 100x L-Glutamine
HT3	Cervical	ATCC	DMEM
			+10%FCS
			+ 10ml 100x L-Glutamine
MCF7	Breast	ATCC	EMEM
			+10%FCS
			+ 10ml 100x L-Glutamine
T-47D	Breast	ATCC	RPMI-1640
			+10%FCS
			+ 10ml 100x L-Glutamine
A549	Lung	ATCC	F12
			+10%FCS
			+ 10ml 100x L-Glutamine
Calu-6	Lung	ATCC	EMEM
			+10%FCS
			+ 10ml 100x L-Glutamine
HUVEC	Human Umbilical	Millipore	EndoGRO-LS Complete Culture
	Vein Endothelial		Media Kit
	Cells		

Table S1- Cell lines and culture requirements

Abreviations: H&N, Head and Neck; FCS, Foetal Calf Serum; MEM, Minimum Essential Medium; DMEM, Dulbecco's Modified Eagle Medium; EMEM, Eagle's Minimum Essential Medium; IMDM, Iscove's Modified Dulbecco's Medium; RPMI-1640, Roswell Park Memorial Institute 1640 Medium; Suppliers: ATCC, American Type Culture Collection (Manassas, USA); ECACC, European Collection of Authenticated Cell Cultures (Salisbury, UK); Millipore (Watford, UK).

Antibody	Supplier	Species	Size (kDa)	Dilution
Phospho-Akt (Ser473)	Cell Signaling	Rabbit	66	1:500
	Technology,			
	Danver, USA			
Phospho-GSK3β (Ser9)	Cell Signaling	Rabbit	46	1:500
	Technology,			
	Danver, USA			
Phospho-S6 Ribosomal	Cell Signaling	Rabbit	32	1:1000
Protein (Ser235/236)	Technology,			
	Danver, USA			
Vinculin	Sigma-Aldrich,	Mouse	116	1:5000
	Dorset, UK			
GAPDH	Sigma-Aldrich,	Rabbit	37	1:10000
	Dorset, UK			
Anti-rabbit IgG, HRP-	Cell Signaling	Goat	N/A	1:2000
linked Antibody	Technology,			
	Danver, USA			
Anti-mouse IgG, HRP-	Cell Signaling	Horse	N/A	1:2000
linked Antibody Technology,				
	Danver, USA			

Table S2- Antibodies used in Western blot analysis

Table S3- ELISA kits used

ELISA Kit	Supplier	
PRAS40 (pT246)	Invitrogen	
Phospho-ELISA Kit	(Paisley, UK).	
(Human)		
Human VEGF	R&D	
Quantikine [®] ELISA Kit	(Abingdon, UK)	
Mouse VEGF	R&D	
Quantikine [®] ELISA Kit	(Abingdon, UK)	

Stain	Antigen Retrieval	Blocking Steps	Primary Antibody (supplier*, dilution and incubation)	Detection (supplier*)
CD31	HIER (100°C, pH6, 20mins)	EPB, 10mins PPB, 30mins	Anti-CD31 antibody, rabbit polyclonal (Abcam, 1:225, 15mins, RT)	NovoLink Polymer Refine Kit (Leica)
HIF-1α	HIER (100°C, pH9, 40mins)	EPB, 10mins PPB, 30mins BS, 30 mins	Anti-HIF-1-alpha antibody [EP1215Y], rabbit mAb (Abcam, 1:1000, 60mins, RT)	NovoLink Polymer Refine Kit (Leica)
Human Ki-67	HIER (125°C 1min & 90°C 10sec, pH6)	EPB, 10mins BR, 20mins	Monoclonal Mouse Anti-Human Ki-67 Antigen Clone MIB-1 (Dako, 1:100, 20mins, RT)	ARK™ (Animal Research Kit) Peroxidase, (Dako)
Pimonidazole	HIER (125°C 1min & 90°C 10sec, pH6)	EPB, 10mins CB, 20mins	Affinity purified rabbit anti- pimonidazole antibody (Hydroxyprobe, 1:300, 120mins, RT)	EnVision-HRP Kit (Dako)
CD11b	HIER (pH6, 95°C 32mins)	DI, 8mins	Anti-CD11b antibody [EPR1344], rabbit mAb (Abcam, 1:4000, 40mins, 37°C)	DISCOVERY® OmniMap anti- Rabbit HRP (Ventana)
Mouse Ki-67	HIER (pH8.5, 95°C, 64 mins	DI, 8mins	Ki-67, rabbit anti-mouse (Bethyl, 1:2000, 60mins, 37°C)	DISCOVERY® OmniMap anti- Rabbit HRP (Ventana)

Table S4- Summary of Immunohistochemical staining techniques

Abreviations: HIER, Heat-induced Epitope Retrieval; EPB, Endogenous Peroxidase Block (3% H₂O₂, Sigma); PPB, Post Primary Block (Leica Refine Kit); BS, Blocking Solution (Candor); BR, Blocking Reagent (normal mouse serum, Dako); CB, 10% Casein Block (Vector); DI, Discovery Inhibitor (Ventana); RT, Room Temperature; mAb, monoclonal Antibody. *Suppliers: Abcam (Cambridge, UK); Dako (Ely, UK); Vector Laboritories (Peterborough, UK); Leica, Leica Microsystems (Milton Keynes, UK); Ventana (Tucson, USA); Hydroxyprobe (Burlington, USA); Candor, (Candor Bioscience, Wangen, Germany); Sigma, Sigma-Aldrich (Dorset, UK); Dako (Ely, UK); Bethyl (Bethyl Laboratories, Montgomery, USA).