

Human *CRMP4* mutation and disrupted *Crmp4* expression in mice are associated with ASD characteristics and sexual dimorphism

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Supplementary Information

History of the ASD patient

The patient weighed 3153 g at birth after a 39-week gestation and was born to a 28-year-old woman, gravida 2, para 1–2, with one previous miscarriage. This pregnancy was achieved by administration of Clomid and was complicated by maternal tobacco use, as well as weight loss of 12 lbs. Delivery was vaginal with vacuum assistance, and the baby was discharged home with his mother.

The family history is unremarkable. The patient has an older and a younger brother, both of whom are healthy with normal development, as are both parents. There is no known consanguinity. The patient had normal development until 3 years of age, during which there was a loss of language and social skills. He also developed unusual behaviours, including unusual obsessions and repetitive tapping of objects and making clicking noises. He attended a special needs preschool and repeated first grade. He has an individual education programme, receives speech therapy, and has been treated with Ritalin for attention-deficit/hyperactivity disorder.

At 8 years of age, the patient was assessed by an experienced psychologist using the Autism Diagnostic Observation Scale (ADOS) and received a score of 13 (Module 3), consistent with the diagnosis of ASD. The Autism Diagnostic Interview-Revised (ADI-R,) assessment, performed at 10 years of age, was also consistent with the ASD diagnosis. At 12 years of age, the patient received a full scale IQ standard score of 68 on the Stanford-Binet Intelligence Scale as part of CORA, falling into the ‘mildly impaired’ range (non-verbal IQ score, 82 and verbal IQ score, 57).

Previous genetic testing revealed a normal 46, XY male karyotype and normal results in DNA testing for Fragile X syndrome. As part of CORA, an oligonucleotide microarray was

performed (NimbleGen CGX-3) in 2011 that also showed no abnormalities. He underwent exome sequencing in the NCH Biomedical Genomics Core (BGC).

Whole-exome sequencing

Exome sequencing was performed on 72 simplex trios (affected and both parents). A total of 69 *de novo* damaging variants were confirmed, of which seven were considered pathogenic or likely pathogenic based on the predicted change in the protein and reports in the literature demonstrating that the gene is involved in human ASD and/or intellectual disability (G. Herman, personal communication).

DNA samples for exome sequencing were prepared from lymphoblastoid cell lines and were processed using the Agilent SureSelectXT Target Enrichment System for the Illumina Paired End Sequencing Protocol (Agilent Technologies, Santa Clara, CA, USA). DNA libraries prepared from the patient and both parents were captured using SureSelect Human All Exon v4 probes. Paired-end 100 base-pair reads were generated for exome-enriched libraries sequenced on the Illumina HiSeq 2000 (Illumina, San Diego, CA, USA) to an average depth of 74×. Secondary analysis was performed using Churchill, a pipeline at NCH, for the discovery of human genetic variation¹. A *de novo* variant of *CRMP4* found in the sequenced sample from the proband but not in the parents or siblings was confirmed by Sanger sequencing using DNA prepared directly from a sample of whole blood to avoid any possible cell-line artefacts. PCR was performed for exon 11 using the primers 5'-CAGGTCATGTTCCCCTGTCT-3' and 5'-ACGCTGCCAAGATCTTCAAC-3' to amplify a 402-bp fragment.

Crmp4 (pEGFP-Crmp4 vector) and Crmp4^{S540Y} (pEGFP-Crmp4^{S540Y}) expression vectors

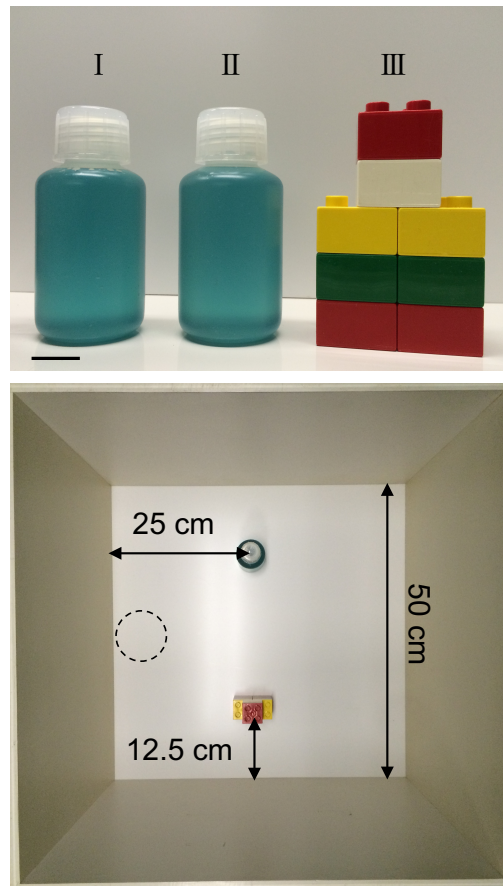
For the pEGFP-*Crmp4* vector, a cDNA fragment containing the full-length coding region of the mouse *Crmp4* gene was inserted into the *EcoRI* and *XhoI* sites of the pEGFP-N vector (Clontech Laboratories, Inc., Mountain View, CA, USA). *Crmp4*^{S540Y} was made using the PrimSTAR mutagenesis kit (Takara Bio, Japan) according to the manufacturer's protocol using the WT *Crmp4* sequence as a template. The S540 in the mouse *Crmp4* is the homologous site of S541 in the human *CRMP4* sequence (NP_001278384.1). The following primers were used to amplify the mutant mouse sequence: forward, 5'- CACCAGTATGTTGCCGAATACAACATC -3'; reverse, 5'- GGCAACATACTGGTGGTTCTTGGCAGA -3', where the underlined base results in the missense variant. Both constructs were sequenced on an ABI PRISM 310 Genetic Analyzer (Perkin Elmer Cetus), and both confirmed the WT and mutant sequences with no additional base changes.

Real-time qRT-PCR

WT and *Crmp4*-KO mice of both sexes at 8 weeks of age (n = 4 each) were deeply anesthetised with pentobarbital (50 mg/kg, i.p.), and their brains were removed from the skull. The OB, cortex, hippocampus and raphe were dissected out. RNA extraction, reverse transcription, and real-time qRT-PCR were performed as previously reported². The primer pairs used are listed in Supplementary Table S3.

Supplementary References

1. Kelly, B. J. *et al.* Churchill: an ultra-fast, deterministic, highly scalable and balanced parallelization strategy for the discovery of human genetic variation in clinical and population-scale genomics. *Genome Biol* **16**, 6 (2015).
2. Tsutiya A & Ohtani-Kaneko R. Postnatal alteration of collapsin response mediator protein 4 mRNA expression in the mouse brain. *J Anat.* **221**, 341–351 (2012).



Supplementary Fig. S1. Novel object recognition test. During the familiarisation phase, two identical objects ('I' and 'II' in top panel) were located 12.5 cm away from the wall at the central width of the box (bottom panel). The mouse was then placed into the box, always facing the wall at the same position (dotted circle area in bottom panel) and allowed to freely explore for 10 min. Exploration of an object was defined as directing the nose to the object at a distance of less than 2 cm and/or touching it with the nose. After the test, the mouse was housed again. Object b was then replaced with a novel object ('III' in top panel), and the test phase was performed 60 min after the habituation phase. Each animal was placed at the same location (dotted circle area in bottom panel) and allowed to explore freely for 5 min. The total time spent exploring the two objects was measured during both the familiarisation and test phases. The preference index is the ratio of the amount of time spent exploring any one of the two objects (familiarisation phase) or the novel object (test phase) divided by the total time spent exploring both objects: $[II/(I + II)] \times 100$ in the familiarisation phase and $[III/(I + III)] \times 100$ in the test phase.

Supplementary Table S1.

Statistical values from ANOVA for behavioural analysis. Significant results are indicated in bold.

Test	Statistical method	<i>F</i> -statistics, degree of freedom, and <i>p</i> -values for ANOVA	<i>F</i> -statistics, degree of freedom, and <i>p</i> -values for ANOVA (for each effect and interaction)								
			Genotype		Sex		Genotype × Sex				
Open field	Distance		F(3,54) = 0.805, <i>p</i> = 0.497	F(1,54) = 0.472, <i>p</i> = 0.495		F(1,54) = 0.434, <i>p</i> = 0.513		F(1,54) = 1.269, <i>p</i> = 0.265			
	In zone center	Two-way ANOVA	F(3,54) = 1.066, <i>p</i> = 0.371	F(1,54) = 1.110, <i>p</i> = 0.297		F(1,54) = 1.082, <i>p</i> = 0.303		F(1,54) = 1.027, <i>p</i> = 0.315			
	Velocity		F(3,54) = 0.788, <i>p</i> = 0.506	F(1,54) = 0.479, <i>p</i> = 0.492		F(1,54) = 0.393, <i>p</i> = 0.533		F(1,54) = 1.255, <i>p</i> = 0.268			
Elevated plus maze	Time in open arms		F(3,39) = 0.625, <i>p</i> = 0.603	F(1,39) = 1.122, <i>p</i> = 0.296		F(1,39) = 0.308, <i>p</i> = 0.582		F(1,39) = 0.529, <i>p</i> = 0.471			
	Time in closed arms		F(3,39) = 0.204, <i>p</i> = 0.893	F(1,39) = 0.159, <i>p</i> = 0.692		F(1,39) = 0.444, <i>p</i> = 0.509		F(1,39) = 0.043, <i>p</i> = 0.837			
	No. of entries to open arms	Two-way ANOVA	F(3,39) = 0.208, <i>p</i> = 0.891	F(1,39) = 0.013, <i>p</i> = 0.909		F(1,39) = 0.001, <i>p</i> = 0.979		F(1,39) = 0.606, <i>p</i> = 0.441			
	No. of entries to closed arms		F(3,39) = 1.736, <i>p</i> = 0.175	F(1,39) = 1.787, <i>p</i> = 0.190		F(1,39) = 3.967, <i>p</i> = 0.053		F(1,39) = 0.012, <i>p</i> = 0.912			
Novel object recognition	Familiarisation	Preference	F(3,52) = 0.697, <i>p</i> = 0.558	F(1,52) = 0.801, <i>p</i> = 0.372		F(1,52) = 0.616, <i>p</i> = 0.436		F(1,52) = 0.762, <i>p</i> = 0.387			
		Total time	F(3,52) = 0.065, <i>p</i> = 0.978	F(1,52) = 0.028, <i>p</i> = 0.868		F(1,52) = 0.038, <i>p</i> = 0.847		F(1,52) = 0.140, <i>p</i> = 0.709			
	Test	Preference	F(3,52) = 0.075, <i>p</i> = 0.973	F(1,52) = 0.014, <i>p</i> = 0.906		F(1,52) = 0.077, <i>p</i> = 0.782		F(1,52) = 0.143, <i>p</i> = 0.707			
		Total time	F(3,52) = 0.877, <i>p</i> = 0.459	F(1,52) = 1.172, <i>p</i> = 0.284		F(1,52) = 0.943, <i>p</i> = 0.336		F(1,52) = 0.224, <i>p</i> = 0.638			
Three-chamber test	6W	Three-way ANOVA	Duration (s)	F(11,120) = 15.829, <i>p</i> < 0.001	F(1,120) = 0.000, <i>p</i> = 1.000	F(1,120) = 0.000, <i>p</i> = 1.000	F(2,120) = 81.648, <i>p</i> < 0.001	F(1,120) = 0.000, <i>p</i> = 1.000	F(2,120) = 1.484, <i>p</i> = 0.231	F(2,120) = 0.902, <i>p</i> = 0.409	F(2,120) = 2.078, <i>p</i> = 0.130
			No. of entries	F(7,80) = 0.334, <i>p</i> = 0.936	F(1,80) = 1.083, <i>p</i> = 0.301	F(1,80) = 0.523, <i>p</i> = 0.472	F(1,80) = 0.002, <i>p</i> = 0.961	F(1,80) = 0.452, <i>p</i> = 0.503	F(1,80) = 0.260, <i>p</i> = 0.611	F(1,80) = 0.102, <i>p</i> = 0.750	F(1,80) = 0.011, <i>p</i> = 0.917
	10W	Three-way ANOVA	Sniffing (s)	F(7,80) = 4.410, <i>p</i> < 0.001	F(1,80) = 0.166, <i>p</i> = 0.685	F(1,80) = 0.043, <i>p</i> = 0.835	F(1,80) = 19.626, <i>p</i> < 0.001	F(1,80) = 0.195, <i>p</i> = 0.660	F(1,80) = 4.988, <i>p</i> = 0.028	F(1,80) = 5.299, <i>p</i> = 0.024	F(1,80) = 1.127, <i>p</i> = 0.292
			Duration (s)	F(11,102) = 21.618, <i>p</i> < 0.001	F(1,102) = 0.000, <i>p</i> = 1.000	F(1,102) = 0.000, <i>p</i> = 1.000	F(2,102) = 101.6, <i>p</i> < 0.001	F(1,102) = 0.000, <i>p</i> = 1.000	F(2,102) = 5.904, <i>p</i> = 0.004	F(2,102) = 9.791, <i>p</i> < 0.001	F(2,102) = 0.532, <i>p</i> = 0.589
		Three-way ANOVA	No. of entries	F(7,68) = 0.609, <i>p</i> = 0.746	F(1,68) = 1.049, <i>p</i> = 0.309	F(1,68) = 1.152, <i>p</i> = 0.287	F(1,68) = 0.370, <i>p</i> = 0.545	F(1,68) = 0.039, <i>p</i> = 0.844	F(1,68) = 0.130, <i>p</i> = 0.719	F(1,68) = 0.064, <i>p</i> = 0.801	F(1,68) = 1.058, <i>p</i> = 0.307
			Sniffing (s)	F(7,68) = 9.675, <i>p</i> < 0.001	F(1,68) = 4.128, <i>p</i> = 0.046	F(1,68) = 3.453, <i>p</i> = 0.067	F(1,68) = 26.493, <i>p</i> < 0.001	F(1,68) = 0.130, <i>p</i> = 0.720	F(1,68) = 13.125, <i>p</i> = 0.001	F(1,68) = 14.596, <i>p</i> < 0.001	F(1,68) = 4.869, <i>p</i> = 0.031
Social interaction	Two-way ANOVA	F(3,57) = 6.207, <i>p</i> = 0.01	F(1,57) = 14.966, <i>p</i> < 0.001		F(1,57) = 1.120, <i>p</i> = 0.294		F(1,57) = 1.891, <i>p</i> = 0.174				
Tube test	Binomial test		Male WT vs. KO <i>p</i> = 1.000 (Win, WT : KO = 6 : 6)				Female WT vs. KO <i>p</i> = 0.815 (Win, WT : KO = 8 : 10)				
Food exploring	Two-way ANOVA	7W	F(3,54) = 1.785, <i>p</i> = 0.161	F(1,54) = 1.874, <i>p</i> = 0.177		F(1,54) = 3.416, <i>p</i> = 0.070		F(1,54) = 0.207, <i>p</i> = 0.551			
		11W	F(3,47) = 0.961, <i>p</i> = 0.419	F(1,47) = 0.154, <i>p</i> = 0.697		F(1,47) = 0.505, <i>p</i> = 0.481		F(1,47) = 2.493, <i>p</i> = 0.121			
Hot plate	Two-way ANOVA	F(3,83) = 0.869, <i>p</i> = 0.461	F(1,83) = 2.119, <i>p</i> = 0.149		F(1,83) = 0.240, <i>p</i> = 0.625		F(1,83) = 0.186, <i>p</i> = 0.668				
UV experiment	Three-way ANOVA	Bedding	Genotype	F(7,98) = 3.336, <i>p</i> = 0.003	F(1,98) = 5.357, <i>p</i> = 0.023		F(1,98) = 10.577, <i>p</i> = 0.002	F(1,98) = 4.129, <i>p</i> = 0.045	F(1,98) = 5.313, <i>p</i> = 0.023	F(1,98) = 0.000, <i>p</i> = 0.991	F(1,98) = 0.037, <i>p</i> = 0.848
			Sex	F(1,98) = 0.001, <i>p</i> = 0.976	F(1,98) = 0.001, <i>p</i> = 0.976		F(1,98) = 0.001, <i>p</i> = 0.976		F(1,98) = 0.001, <i>p</i> = 0.976		F(1,98) = 0.001, <i>p</i> = 0.976
Temperature	Three-way ANOVA	Temperature	Genotype	F(11,128) = 3.931, <i>p</i> < 0.001	F(1,128) = 0.279, <i>p</i> = 0.599		F(2,128) = 8.288, <i>p</i> < 0.001	F(1,128) = 0.426, <i>p</i> = 0.515	F(2,128) = 7.243, <i>p</i> = 0.001	F(2,128) = 1.689, <i>p</i> = 0.189	F(2,128) = 4.827, <i>p</i> = 0.010
			Sex	F(1,128) = 0.367, <i>p</i> = 0.546	F(1,128) = 0.367, <i>p</i> = 0.546		F(1,128) = 0.367, <i>p</i> = 0.546		F(1,128) = 0.367, <i>p</i> = 0.546		F(1,128) = 0.367, <i>p</i> = 0.546

Supplementary Table S2.

Statistical *p*-values from multiple comparisons (PLSD, Bonferroni, Sidak, and Tukey HSD) for behavioural studies. Significant results are indicated in bold.

Test	Subject	Comparison	<i>p</i> -values from multiple comparison				
			PLSD	Bonferroni	Sidak	Tukey HSD	
Three chamber test (6W)	Duration	Male KO	Stranger vs. Object	0.001	0.034	0.033	0.025
		Male WT	Stranger vs. Object	0.008	0.226	0.203	0.133
	Sniffing	Male KO	Stranger vs. Object	0.560	1.000	1.000	0.999
		Female WT	Stranger vs. Object	<0.001	0.009	0.009	0.007
		Female KO	Stranger vs. Object	0.005	0.131	0.123	0.084
Three chamber test (10W)	Duration	Male KO	Stranger vs. Object	0.009	0.577	0.440	0.255
		Female WT	Stranger vs. Object	<0.001	0.009	0.009	0.007
	Sniffing	Male WT	Stranger vs. Object	<0.001	0.012	0.012	0.010
		Male KO	Stranger vs. Object	0.041	1.000	0.692	0.438
		Female WT	Stranger vs. Object	<0.001	<0.001	<0.001	<0.001
Female KO	Stranger vs. Object	<0.001	0.003	0.003	0.003		
Social interaction test	Male	WT vs. KO	<0.001	0.002	0.002	0.002	
	Female	WT vs. KO	0.093	0.556	0.442	0.328	
UV experiment	Bedding	Male WT	Familiar vs. Unfamiliar	0.015	0.433	0.354	0.223
		Male KO	Familiar vs. Unfamiliar	0.560	1.000	1.000	0.999
		Female WT	Familiar vs. Unfamiliar	0.007	0.188	0.172	0.115
		Female KO	Familiar vs. Unfamiliar	0.671	1.000	1.000	1.000
		Male Unfamiliar	WT vs. KO	0.002	0.061	0.059	0.044
	Temperature	Male WT	19°C vs. 9°C	<0.001	<0.001	<0.001	<0.001
Male KO		19°C vs. 9°C	0.037	1.000	0.916	0.617	
Female WT		19°C vs. 9°C	0.004	0.240	0.213	0.132	
Female KO		19°C vs. 9°C	0.026	1.000	0.822	0.513	
Male 9°C		WT vs. KO	0.001	0.078	0.075	0.051	
KO 9°C		Male vs. Female	0.011	0.745	0.527	0.308	

Supplementary Table S3.

Statistical *p*-values from multiple comparison (PLSD, Bonferroni, Sidak, and Tukey HSD) for gene expression analysis. Significant results are indicated in bold.

Brain region	Gene	Comparison	<i>p</i> -values from multiple comparison				
			PSLD	Bonferroni	Sidak	Tukey HSD	
OB	<i>GluR2</i>	Female WT vs. KO	0.019	0.115	0.110	0.079	
		Male WT vs. KO	0.010	0.057	0.056	0.041	
	<i>VGluT1</i>	Female WT vs. KO	<0.001	0.001	0.001	0.001	
		WT male vs. female	0.003	0.021	0.020	0.016	
	<i>VGluT2</i>	KO male vs. female	0.014	0.087	0.083	0.061	
	<i>GABAAα1</i>	Female WT vs. KO	<0.001	<0.001	<0.001	<0.001	
		KO male vs. female	<0.001	<0.001	<0.001	<0.001	
	<i>GABAAγ2</i>	Female WT vs. KO	<0.001	0.001	0.001	0.001	
		KO male vs. female	0.006	0.039	0.038	0.029	
	<i>GABABR1</i>	Female WT vs. KO	0.002	0.012	0.012	0.009	
		KO male vs. female	0.029	0.174	0.162	0.114	
	<i>VGAT</i>	Female WT vs. KO	<0.001	0.003	0.003	0.002	
		KO male vs. female	0.012	0.073	0.071	0.052	
	<i>Ncam1</i>	Male WT vs. KO	0.009	0.053	0.051	0.038	
		Female WT vs. KO	0.007	0.039	0.039	0.029	
	Hippocampus	<i>GluR1</i>	Male WT vs. KO	0.014	0.082	0.079	0.057
			KO male vs. female	0.015	0.088	0.085	0.062
		<i>GluR2</i>	Male WT vs. KO	0.015	0.092	0.089	0.063
<i>GABAAα1</i>		KO male vs. female	0.031	0.186	0.172	0.121	
<i>GABABR1</i>		KO male vs. female	0.034	0.204	0.187	0.132	
Cortex	<i>GABAAγ2</i>	Female WT vs. KO	0.003	0.016	0.016	0.012	
		WT male vs. Female	0.012	0.072	0.070	0.051	
	<i>GABABR1</i>	Male WT vs. KO	0.014	0.083	0.080	0.058	
		Male WT vs. KO	0.003	0.020	0.019	0.015	
	<i>Ncam1</i>	Female WT vs. KO	0.002	0.010	0.010	0.008	
		Male WT vs. KO	0.002	0.012	0.012	0.009	
<i>N-cadherin</i>	Male WT vs. KO	0.002	0.012	0.012	0.009		
	Female WT vs. KO	<0.001	<0.001	<0.001	<0.001		

Supplementary Table S4. Gene expressions without significant differences among groups in adults

Brain region	Genes	F-statistics, degree of freedom, and p-values for two-way ANOVA				Expression levels					
		For each factor and interaction				Fold differences		Male		Female	
		Genotype	Sex	Genotype × Sex		WT vs. <i>Crmp4</i> -KO (<i>Crmp4</i> -KO/WT)	Male vs. Female (Female/Male)	WT	<i>Crmp4</i> -KO	WT	<i>Crmp4</i> -KO
Olfactory bulb	<i>Crmp4</i>	Not performed				Not performed	1.102	2.578 ± 0.145	ND	2.958 ± 0.099	ND
	<i>GluR1</i>	F(3,12) = 0.131, p = 0.940	F(1,12) = 0.218, p = 0.649	F(1,12) = 0.173, p = 0.685	F(1,12) = 0.001, p = 0.976	1.102	1.090	0.861 ± 0.255	0.959 ± 0.040	0.949 ± 0.211	1.034 ± 0.205
	<i>DIR</i>	F(3,8) = 1.166, p = 0.381	F(1,8) = 3.097, p = 0.116	F(1,8) = 0.003, p = 0.961	F(1,8) = 0.398, p = 0.546	1.895	1.018	0.263 ± 0.118	0.395 ± 0.178	0.195 ± 0.068	0.474 ± 0.064
	<i>D2R</i>	F(3,8) = 0.871, p = 0.495	F(1,8) = 0.236, p = 0.640	F(1,8) = 2.369, p = 0.162	F(1,8) = 0.007, p = 0.938	1.144	1.542	0.300 ± 0.084	0.346 ± 0.018	0.466 ± 0.206	0.531 ± 0.049
	<i>5HT_{1A}</i>	F(3,12) = 0.625, p = 0.612	F(1,12) = 0.847, p = 0.375	F(1,12) = 0.684, p = 0.424	F(1,12) = 0.344, p = 0.569	1.277	0.804	0.550 ± 0.143	0.789 ± 0.236	0.511 ± 0.100	0.564 ± 0.123
	<i>5HT_{2A}</i>	F(3,12) = 1.418, p = 0.286	F(1,12) = 0.017, p = 0.898	F(1,12) = 2.742, p = 0.124	F(1,12) = 1.494, p = 0.245	0.938	2.374	0.110 ± 0.067	0.295 ± 0.115	0.596 ± 0.302	0.368 ± 0.070
	<i>5HT₇</i>	F(3,8) = 0.143, p = 0.474	F(1,8) = 1.617, p = 0.239	F(1,8) = 4.821, p = 0.059	F(1,8) = 0.772, p = 0.405	1.425	1.869	0.612 ± 0.111	0.716 ± 0.017	0.959 ± 0.102	1.524 ± 0.503
	<i>GABAAβ2</i>	F(3,12) = 1.629, p = 0.235	F(1,12) = 2.730, p = 0.124	F(1,12) = 0.215, p = 0.651	F(1,12) = 1.942, p = 0.189	1.216	1.056	2.784 ± 0.302	2.872 ± 0.317	2.464 ± 0.481	3.509 ± 0.218
Hippocampus	<i>Crmp4</i>	Not performed				Not performed	0.879	0.700 ± 0.060	ND	0.615 ± 0.042	ND
	<i>VGluT2</i>	F(3,12) = 0.666, p = 0.589	F(1,12) = 0.936, p = 0.352	F(1,12) = 0.901, p = 0.361	F(1,12) = 0.161, p = 0.695	0.847	1.175	0.779 ± 0.056	0.700 ± 0.077	0.963 ± 0.196	0.775 ± 0.164
	<i>D1R</i>	F(3,9) = 3.546, p = 0.061	F(1,9) = 3.611, p = 0.090	F(1,9) = 3.156, p = 0.109	F(1,9) = 4.288, p = 0.068	1.290	0.789	0.674 ± 0.118	0.660 ± 0.044	0.368 ± 0.016	0.683 ± 0.014
	<i>D2R</i>	F(3,8) = 0.215, p = 0.883	F(1,8) = 0.216, p = 0.655	F(1,8) = 0.176, p = 0.686	F(1,8) = 0.253, p = 0.629	1.169	1.153	0.132 ± 0.053	0.130 ± 0.009	0.128 ± 0.016	0.174 ± 0.079
	<i>5HT_{1A}</i>	F(3,12) = 1.365, p = 0.301	F(1,12) = 2.665, p = 0.129	F(1,12) = 1.424, p = 0.256	F(1,12) = 0.005, p = 0.943	1.303	1.213	0.836 ± 0.303	1.326 ± 0.228	1.241 ± 0.170	1.602 ± 0.170
	<i>5HT_{2A}</i>	F(3,11) = 1.657, p = 0.233	F(1,11) = 0.059, p = 0.813	F(1,11) = 4.094, p = 0.068	F(1,11) = 0.512, p = 0.489	1.117	0.377	0.136 ± 0.062	0.183 ± 0.067	0.071 ± 0.038	0.048 ± 0.019
	<i>5HT₇</i>	F(3,8) = 1.211, p = 0.366	F(1,8) = 0.781, p = 0.403	F(1,8) = 1.004, p = 0.346	F(1,8) = 1.849, p = 0.211	1.373	1.435	0.297 ± 0.057	0.241 ± 0.085	0.255 ± 0.061	0.517 ± 0.200
	<i>GABAAβ2</i>	F(3,12) = 0.416, p = 0.745	F(1,12) = 0.216, p = 0.650	F(1,12) = 0.992, p = 0.339	F(1,12) = 0.040, p = 0.845	0.916	1.223	0.281 ± 0.012	0.244 ± 0.044	0.328 ± 0.095	0.314 ± 0.045
	<i>GABAAγ2</i>	F(3,12) = 0.396, p = 0.758	F(1,12) = 0.475, p = 0.504	F(1,12) = 0.357, p = 0.561	F(1,12) = 0.357, p = 0.561	1.096	0.917	0.193 ± 0.012	0.229 ± 0.042	0.193 ± 0.026	0.194 ± 0.020
	<i>VGAT</i>	F(3,12) = 0.503, p = 0.688	F(1,12) = 0.197, p = 0.665	F(1,12) = 0.911, p = 0.359	F(1,12) = 0.401, p = 0.539	1.047	0.906	0.302 ± 0.017	0.296 ± 0.053	0.255 ± 0.016	0.287 ± 0.014
	<i>Ncam1</i>	F(3,12) = 1.535, p = 0.256	F(1,12) = 2.512, p = 0.139	F(1,12) = 2.077, p = 0.175	F(1,12) = 0.014, p = 0.906	1.084	1.076	0.082 ± 0.041	0.141 ± 0.071	1.035 ± 0.159	1.318 ± 0.223
Cortex	<i>Crmp4</i>	Not performed				Not performed	1.182	0.615 ± 0.059	ND	0.727 ± 0.058	ND
	<i>GluR1</i>	F(3,8) = 2.287, p = 0.155	F(1,8) = 0.547, p = 0.481	F(1,8) = 4.394, p = 0.069	F(1,8) = 1.921, p = 0.203	0.854	1.574	0.294 ± 0.031	0.162 ± 0.023	0.339 ± 0.056	0.379 ± 0.104
	<i>GluR2</i>	F(3,10) = 1.874, p = 0.198	F(1,10) = 0.004, p = 0.948	F(1,10) = 1.455, p = 0.255	F(1,10) = 3.410, p = 0.095	1.006	0.860	0.392 ± 0.059	0.319 ± 0.019	0.267 ± 0.015	0.345 ± 0.035
	<i>VGluT1</i>	F(3,12) = 1.719, p = 0.216	F(1,12) = 1.631, p = 0.226	F(1,12) = 2.892, p = 0.115	F(1,12) = 0.663, p = 0.442	1.004	1.011	1.085 ± 0.008	1.084 ± 0.004	1.091 ± 0.009	1.101 ± 0.004
	<i>D1R</i>	F(3,8) = 0.403, p = 0.755	F(1,8) = 0.000, p = 0.985	F(1,8) = 0.048, p = 0.833	F(1,8) = 1.163, p = 0.312	1.005	0.943	1.932 ± 0.451	1.467 ± 0.476	1.362 ± 0.151	1.845 ± 0.567
	<i>D2R</i>	F(3,8) = 1.369, p = 0.320	F(1,8) = 0.396, p = 0.547	F(1,8) = 1.701, p = 0.228	F(1,8) = 2.009, p = 0.194	1.288	1.709	0.746 ± 0.034	0.483 ± 0.207	0.709 ± 0.242	1.393 ± 0.587
	<i>5HT_{1A}</i>	F(3,8) = 1.789, p = 0.227	F(1,8) = 0.143, p = 0.715	F(1,8) = 1.520, p = 0.253	F(1,8) = 3.703, p = 0.091	0.936	1.241	1.010 ± 0.071	1.369 ± 0.158	1.742 ± 0.407	1.208 ± 0.138
	<i>5HT_{2A}</i>	F(3,8) = 0.815, p = 0.521	F(1,8) = 1.502, p = 0.255	F(1,8) = 0.944, p = 0.360	F(1,8) = 0.000, p = 0.984	1.367	0.779	0.168 ± 0.029	0.222 ± 0.014	0.126 ± 0.023	0.179 ± 0.079
	<i>5HT₇</i>	F(3,8) = 0.028, p = 0.993	F(1,8) = 0.021, p = 0.888	F(1,8) = 0.043, p = 0.841	F(1,8) = 0.021, p = 0.889	0.977	0.969	0.953 ± 0.105	0.953 ± 0.111	0.944 ± 0.231	0.902 ± 0.096
	<i>GABAAα1</i>	F(3,12) = 1.880, p = 0.187	F(1,12) = 3.673, p = 0.079	F(1,12) = 1.198, p = 0.295	F(1,12) = 0.770, p = 0.398	0.784	0.874	0.169 ± 0.029	0.149 ± 0.016	0.165 ± 0.014	0.113 ± 0.008
	<i>GABAAβ2</i>	F(3,12) = 0.792, p = 0.521	F(1,12) = 0.749, p = 0.404	F(1,12) = 1.489, p = 0.246	F(1,12) = 0.138, p = 0.717	0.830	1.297	0.077 ± 0.017	0.068 ± 0.013	0.105 ± 0.026	0.083 ± 0.010
	<i>VGAT</i>	F(3,12) = 2.826, p = 0.084	F(1,12) = 3.913, p = 0.071	F(1,12) = 1.120, p = 0.311	F(1,12) = 3.445, p = 0.088	0.808	1.121	0.597 ± 0.039	0.383 ± 0.026	0.553 ± 0.062	0.546 ± 0.080
Raphe	<i>SERT</i>	F(3,10) = 1.175, p = 0.911	F(1,10) = 0.437, p = 0.523	F(1,10) = 0.062, p = 0.808	F(1,10) = 0.063, p = 0.807	1.251	0.918	0.438 ± 0.070	0.590 ± 0.109	0.438 ± 0.093	0.506 ± 0.183

Supplementary Table S5. Real-time qRT-PCR primer sequences

Target genes	Primer sequence (5'-3')		Product size (bp)
	Forward	Reverse	
<i>Crmp4</i>	GGTACAGAGCCTCAGCAAGG	TTATCCCCATTTCCAGCATC	200
<i>VGluT1</i>	TGGGTTTCTGTATCAGCTTTG	TGTGCTGTTGTTGACCATGGACACG	74
<i>VGluT2</i>	CGTGAAGAATGGCAGTATGTCTTC	TGAGGCAAATAGTGCATAAAATATGACT	81
<i>GluR1</i>	CCCTTTACAACGTGGAGGAA	GAACAAGGGCGTCTCTTCTG	152
<i>GluR2</i>	ACGAGTGGCACACTGAGGAA	GCACCCAAGGAAAACCAGAG	103
<i>GAD67</i>	CTCAGGCTGTATGTCAGATGTTC	AAGCGAGTCACAGAGATTGGTC	111
<i>VGAT</i>	CCATTGGCATCATCGTGTT	CCAGTTCATCATGCAGTGGAA	101
<i>GABAAα1</i>	AAGGACCCATGACAGTGCTC	CAGAGTGCCATCCTCTGTGA	149
<i>GABAAβ2</i>	AACGCCTTCCATCATTGTTC	ATCACCCTCCACGACATCA	148
<i>GABAAγ2</i>	AGTGTGGGATGGCAAGGAC	AGGAGTCCATTTTGGCAATG	111
<i>GABABR1</i>	TCTGGTTGTGCTCTTTGTGC	TCCTCATTGTTGTTGGTGGA	111
<i>DAT</i>	CCAGCAATTCAGTGATGACATC	CAGCATAGCCGCCAGTACAG	69
<i>DIR</i>	CTCCTGATGGAACACCATTG	GCTTAGCCCTCACGTTCTTG	119
<i>D2R</i>	TGCCATTGTTCTTGGTGTTG	AGAGGACTGGTGGGATGTTG	98
<i>5HT_{1A}</i>	CAAGACGGTCAAGAAGGTGG	CACTACCTGGCTGACCATTC	101
<i>5HT_{2A}</i>	TTCGGGCTACAGGATGATTC	TGATGGTTAGGGGGATGAAA	112
<i>5HT₇</i>	TGCGGTAAGCACACTACAGC	CACAAAGCCTAGACGGGAAG	135
<i>SERT</i>	TGCCTTTTATATCGCCTCCTAC	CAGTTGCCAGTGTTC CAAGA	123
<i>Ncam1</i>	TGTCAAGTGGCAGGAGATGC	GGCGTTGTAGATGGTGAGGGT	138
<i>N-cadherin</i>	AGCGCAGTCTTACCGAAGG	TCGCTGCTTTCATACTGAAC TTT	101
<i>β-actin</i>	GCTACAGCTTACCACCACA	TCTCCAGGGAGGAAGAGGAT	123

