Supplemental Data

Isomer activation controls stereospecificity of class I fructose-1,6-bisphosphate aldolases

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FIGURE S1. Structural comparison of the Schiff-base FBP complexes in TgALD and rabbit muscle aldolase

(*A*) Structural superposition of Schiff-base FBP TgALD (*green*) corresponding to the *30min* structure with Schiff-base FBP from rabbit muscle aldolase (*pink*) (PDB ID: 1ZAI) based on PyMol alignment of equivalent Ca atoms (r.m.s.d. = 0.40 for 273 atoms). (*B*) Active site view of key residues implicated in the reaction mechanism. Lys-231 is responsible for Schiff-base formation with ketone substrates (FBP or DHAP). TgALD Thr-39 corresponds to Ser-38 in rabbit muscle aldolase; all other key active residues are identical. Alignment of active site residues shown (9 residues) based on equivalent Ca atoms yields an r.m.s.d. = 0.14. Numbering shown is based on TgALD sequence. Equivalent residues in rabbit muscle aldolase correspond to: Asp-33, Ser-35, Ser-38, Lys-107, Lys-146, Glu-187, Glu-189, Lys-229, and Arg-303.



FIGURE S2. Multiple sequence alignment showing active site conservation.

Highlights the extensive conservation of active site residues in TgALD with respect to other class I aldolases (including rabbit muscle aldolase). The blue gradient illustrates the level of similarity, and the different colours correspond to key active site residues (shown in Fig. 1A), colored according to their biochemical properties: *red* - acidic; *green* - polar; *blue* - basic. The numbering above the alignment corresponds to residue positions for TgALD. Alignment was performed in Jalview v.2.10.1 using the Clustal algorithm. The blue arrows are to denote the stretches of contiguous sequences lining the active site that contribute residues implicated in catalysis.



FIGURE S3. Configurational rearrangements during aldol addition.

(A) Panel showing nucleophilic attack on the electrophilic C1 carbonyl atom by the DHAP C3 carbanion. The modeled trans-configuration (*cyan*) of D-G3P is shown with the appropriate torsion angle at the C2-C3 bond enabling aldol addition with the DHAP-enamine to form FBP (*pink*). (B) The *Imin* and *30min* structures were superposed (alignment of Ca atoms) to illustrate the conformational change that occurs during C3-C4 bond formation, and is consistent with least atomic motion. Binding by the phosphate groups of D-G3P and FBP at the P6-site is isostructural and the trajectory requires a slight hinge rotation perpendicular to the viewing plane (~ 10°) about the D-G3P phosphate to superpose D-G3P atoms C1, C2, and C3 with FBP atoms C4, C5, and C6. Atoms in the DHAP enamine are essentially identically positioned as equivalent FBP atoms (r.m.s.d. = 0.20 Å). The distance between DHAP C3 and D-G3P C1 prior to addition is 2.5 Å (*black dashes*). The nascent C4-hydroxyl group is stabilized by a short hydrogen bend with Glu-189 and by charged hydrogen-bonding with Lys-146 (*pink dashes*).





Geometry of enzymatic intermediates was used to identify the chemical identity of the trapped intermediatealdolase complex. (A) DHAP-enamine intermediate (green) was identified in the 0.5min, 1min and 2min structures of native TgALD soaked with substrate. The observed hydrogen bond formation (blue dash – 3.1 Å) between the Ser-301 hydroxyl and Lys-231 N ζ is consistent with sp3 hybridization of Lys-231 N ζ in the enamine intermediate. Planar stereochemistry is observed about the DHAP C2 atom, consistent with enamine formation. Structure shown illustrates the 1min structure. (B) The iminium FBP intermediate was identified in the 10min and 30min structures corresponding to the Schiff base complex with FBP (pink). sp2 hybridization of Lys-231 N ζ would preclude hydrogen bond formation with Ser-301 (blue dash – 3.7 Å) and is indicated by the planarity of the electron density about the N ζ atom. Structure shown illustrates the 10min structure. Both maps depict difference electron density maps calculated from simulated annealing $F_o - F_c$ omit maps encompassing Lys-231 and bound substrate and contoured at 3 σ .