

SUPPORTING INFORMATION

Stereochemistry of a Second Riolozone and Other Diterpenoids from *Jatropha dioica*

Elda M. Melchor-Martínez,^{†‡} David A. Silva-Mares,[†] Ernesto Torres-López,[†] Noemí Waksman-Minsky,[†] Guido F. Pauli,[§] Shao-Nong Chen,[§] Matthias Niemitz,[‡] Mariano Sánchez-Castellanos,[‡] Alfredo Toscano,[‡] Gabriel Cuevas,^{*‡} and Verónica M. Rivas-Galindo^{*†}

[†]Universidad Autónoma de Nuevo León, Facultad de Medicina, Av. Madero y Aguirre Pequeño, Col. Mitras Centro s/n, Monterrey, N.L., C.P. 64460, México.

[‡]Instituto de Química, Universidad Nacional Autónoma de México, Circuito Exterior, Ciudad Universitaria, Delegación Coyoacán, Ciudad de México, C.P. 04510, México.

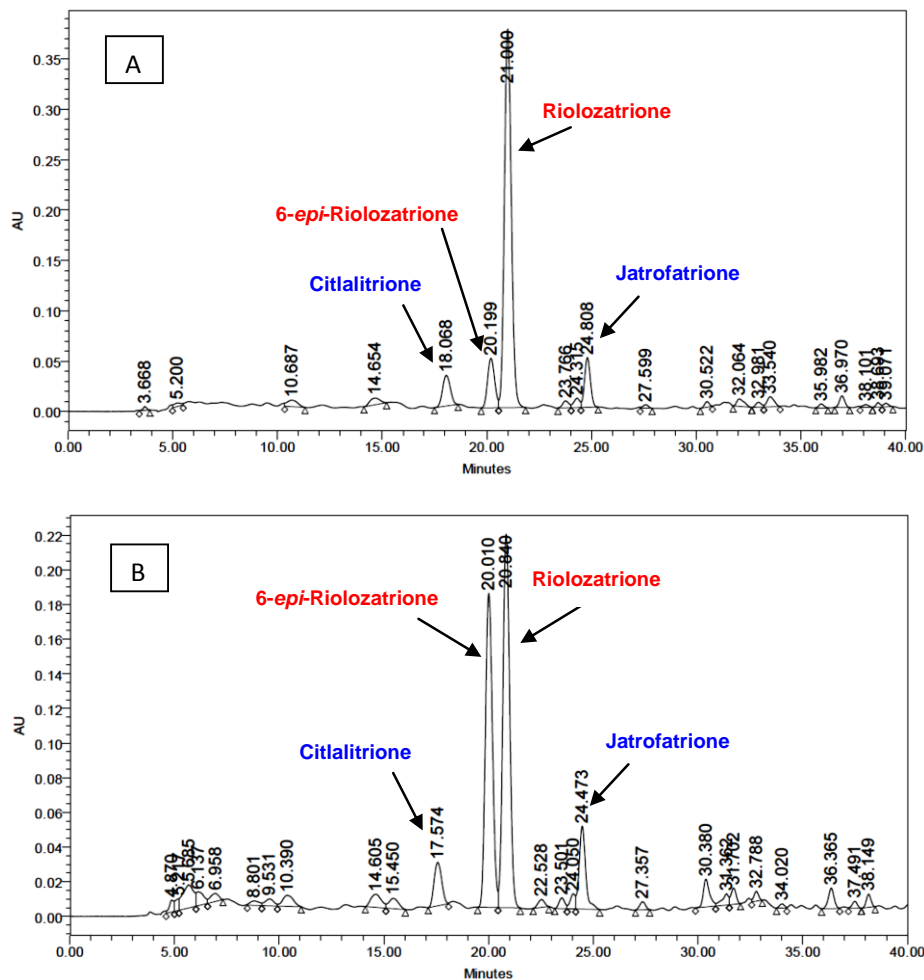
[§]Center for Natural Product Technologies (CENAPT), Department of Medicinal Chemistry and Pharmacognosy, and Institute for Tuberculosis Research; College of Pharmacy, University of Illinois at Chicago, Chicago, Illinois 60612, United States.

[‡]NMR Solutions Ltd., 70110 Kuopio, Finland.

Table of Contents	Page
S1 Chromatograms obtained from HPLC analysis for hexane fraction (A) and methylene chloride extract (B) from <i>J. dioica</i> roots	4
S2 Elemental Analysis of 6- <i>epi</i> -Riolozatrione (2)	5
S3 HRESIMS of 6- <i>epi</i> -Riolozatrione (2)	6
S4 Infrared Spectra of 6- <i>epi</i> -Riolozatrione (2)	7
S5 Experimental ¹ H NMR (400 MHz) and ¹³ C NMR (100 MHz) spectra of 1 and 2 in CDCl ₃ .	8
S6 Comparison of the 1D ¹ H NMR spectra of compounds 1 and 2 in CD ₃ OD at 700 MHz, FIDs processed with Lorentzian-Gaussian apodization	9
S7 Comparison of the ¹ H NMR signals of H-12 in compounds 1 and 2 at 400 and 750 MHz	13
S8 Comparison of the 1D ¹³ C NMR spectra of compounds 1 and 2 in CD ₃ OD at 100 MHz.	14
S9 HSQC spectrum (400 MHz, CD ₃ OD) of 6- <i>epi</i> -Riolozatrione (2)	15
S10 HMBC spectrum (400 MHz, CD ₃ OD) of 6- <i>epi</i> -Riolozatrione (2)	16
S11 NOESY spectrum (400 MHz, CD ₃ OD) of 6- <i>epi</i> -Riolozatrione (2)	17
S12 Calculated vs. experimental 1D ¹ H-NMR spectra of compounds 1 and 2 in CD ₃ OD at 700 MHz	18
S13 ¹ H NMR HiFSA data for Riolozatrione (1) in CD ₃ OD (PERCH .pms file format)	19
S14 ¹ H NMR HiFSA data for 6- <i>epi</i> -Riolozatrione (2) in CD ₃ OD (PERCH .pms file format)	20
S15 ¹ H NMR HiFSA data for Riolozatrione (1) in CDCl ₃ (PERCH .pms file format)	21
S16 ¹ H NMR HiFSA data for 6- <i>epi</i> -Riolozatrione (2) in CDCl ₃ (PERCH .pms file format)	22

- S17** Full ^1H NMR δ and J -correlation maps, termed Quantum Interaction and Linkage Tables (QuILTs), of riolozatrione (**1**) and 6-*epi*-riolozatrione (**2**) were achieved by HiFSA processing of the 700 MHz spectra in CDCl_3 . 23
- S18** Calculated (red) vs. experimental (blue) and difference (green) 1D ^1H -NMR signals in compounds **1** and **2** in CDCl_3 at 700 MHz. 24
- S19** Experimental and calculated IR and VCD spectra at the B3PW91/DGDZVP level of theory for **3** and **4**. 25

Figure S1. Chromatograms obtained from HPLC analysis for hexane fraction (A) and methylene chloride extract (B) from *J. dioica* roots.



HPLC Analysis Method of obtained extracts from *J. dioica* roots.

HPLC Method. The HPLC analysis of extracts A and B was performed on a Waters liquid chromatograph 1525 linked to a Waters diode array detector 2996, using a Waters AccQ-Tag column and isocratic elution with acetonitrile-water (50:50).

n-Hexane fraction (A): Hydro-methanolic extract of *J. dioica* roots (250 g) at RT, was evaporated at reduced pressure at 40 °C, and subsequent partitioned to obtain fractions of n-hexane, ethyl acetate, and butyl alcohol. **DCM extract (B):** Dried and powdered roots of *J. dioica* (250 g) were extracted with DCM (3 x 1L) at room temperature to give 3.5 g of concentrated crude extract at reduced pressure.

Figure S2. Elemental analysis of 6-*epi*-Riolozatrione (2)

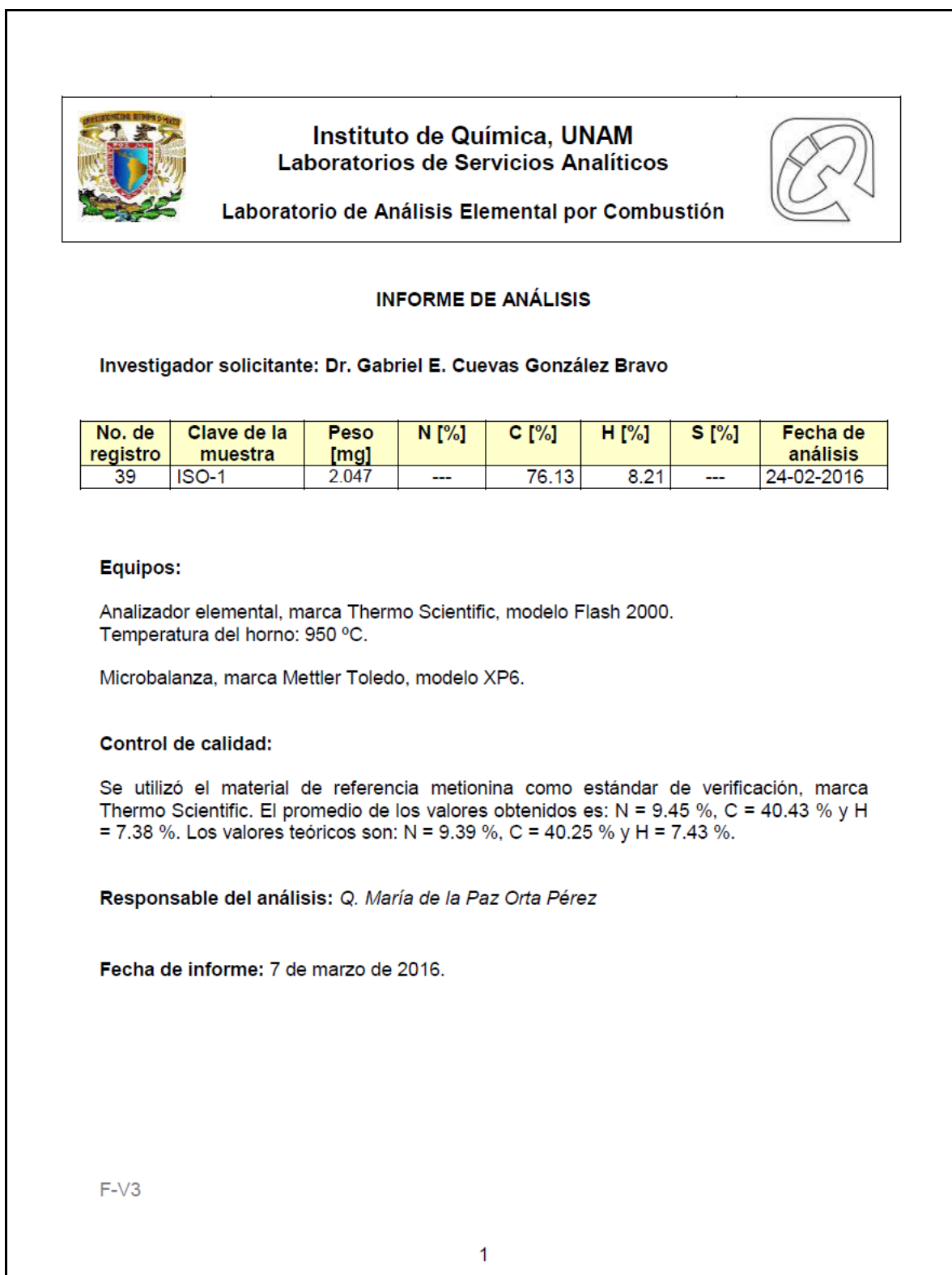


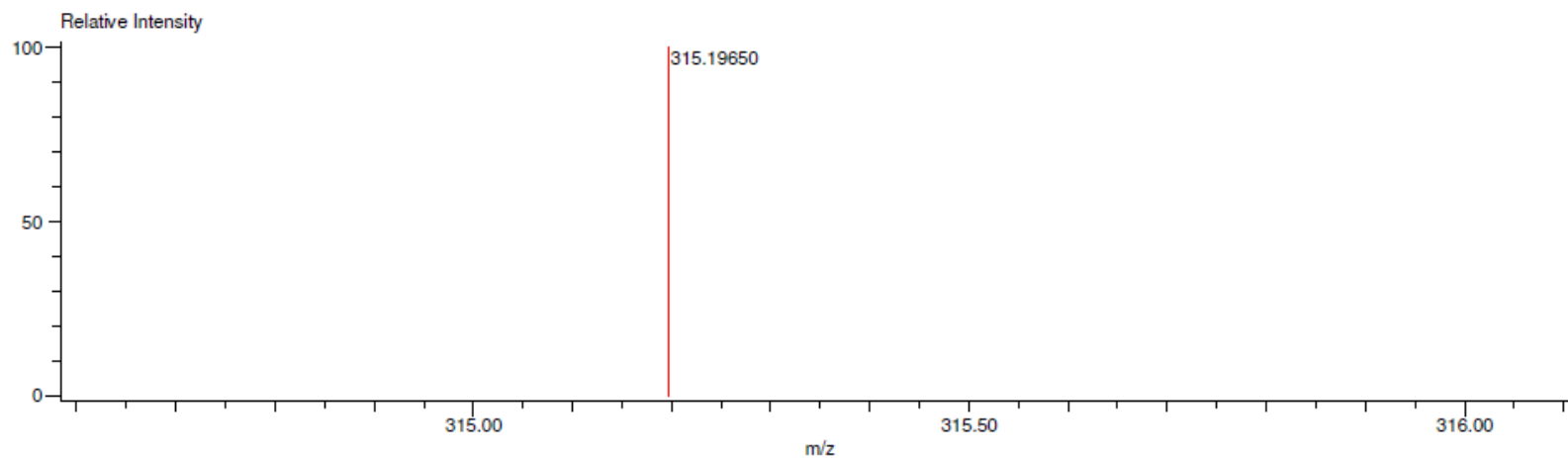
Figure S3. HRESIMS spectrum of 6-*epi*-Riolozatrione (**2**)

Data:238 Isoriolo
 Sample Name:Dr Cuevas Gabriel/Operador:Carmen Garcia-Javier Perez/DART+
 Description:
 Ionization Mode:ESI+
 History:Determine m/z[Peak Detect[Centroid,30,Area];Correct Base[5.0%]];Correct Base[5.0%];Average(MS[1] 0..0)

Acquired:1/27/2016 3:11:32 PM
 Operator:AccuTOF
 Mass Calibration data:Cal_Peg_600
 Created:2/4/2016 3:51:55 PM
 Created by:AccuTOF

Charge number:1
 Element:¹²C:0 .. 100, ¹H:0 .. 200, ¹⁶O:0 .. 6

Tolerance:3.00(mmu)
 Unsaturation Number:0.0 .. 30.0 (Fraction:Both)



Mass	Intensity	Calc. Mass	Mass Difference (mmu)	Mass Difference (ppm)	Possible Formula	Unsaturation Number
315.19650	1168548.88	315.19602	0.48	1.54	¹² C ₂₀ ¹ H ₂₇ ¹⁶ O ₃	7.5

Figure S4. Infrared spectrum of 6-*epi*-Riolozatrione (2)

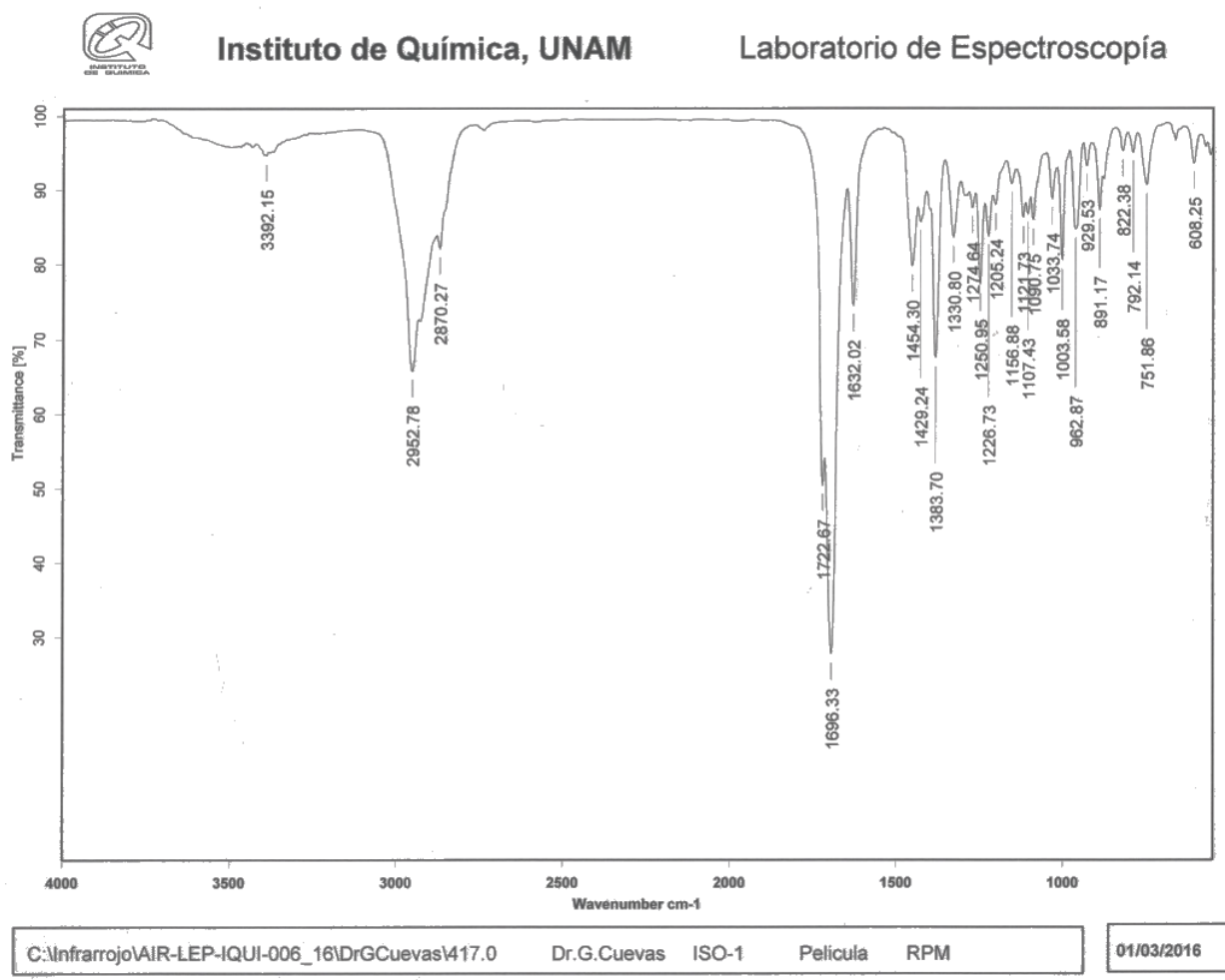


Figure S5. Experimental ^1H NMR (400 MHz) and ^{13}C NMR (100 MHz) spectra of **1** and **2** in CDCl_3 .

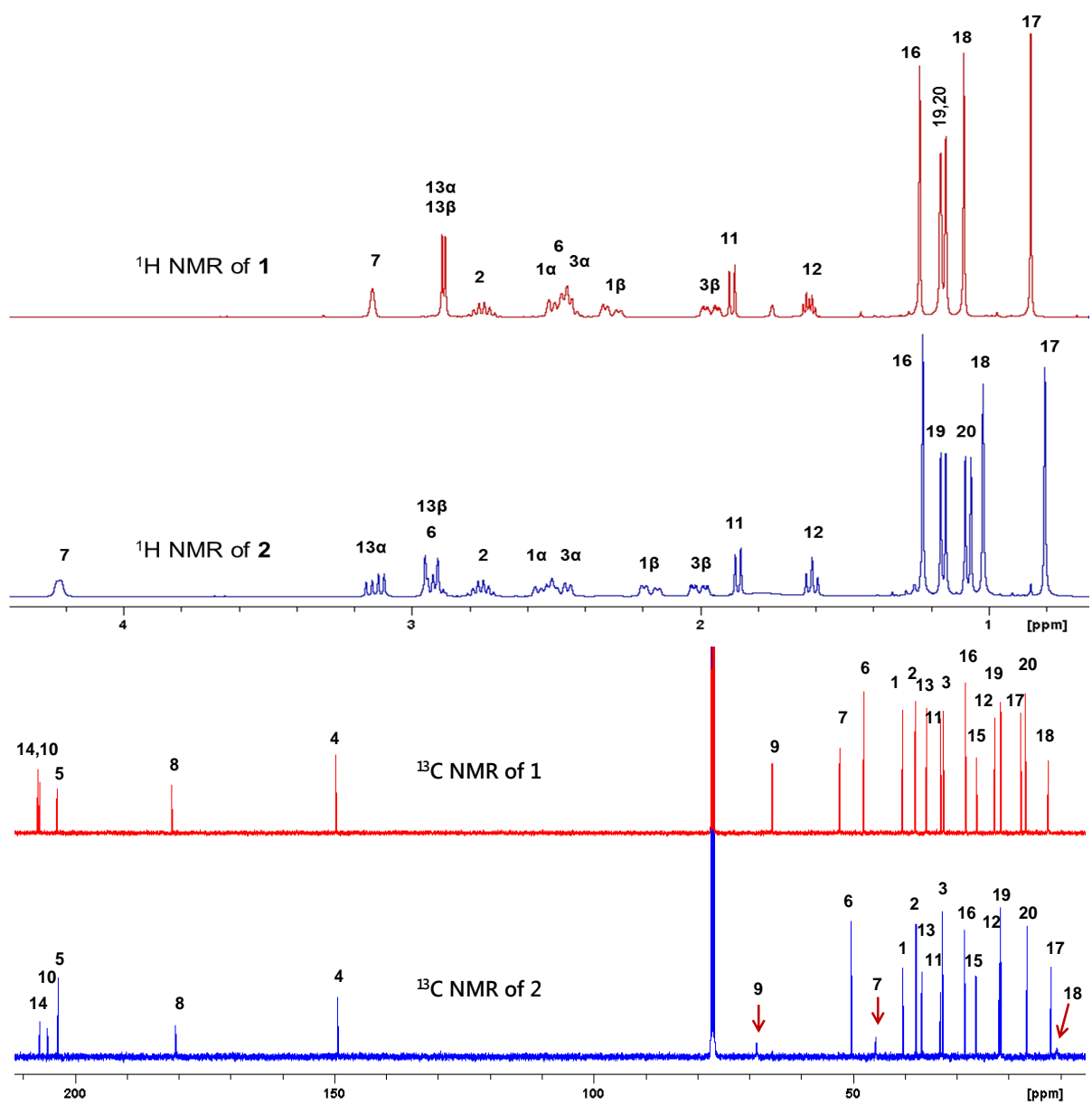
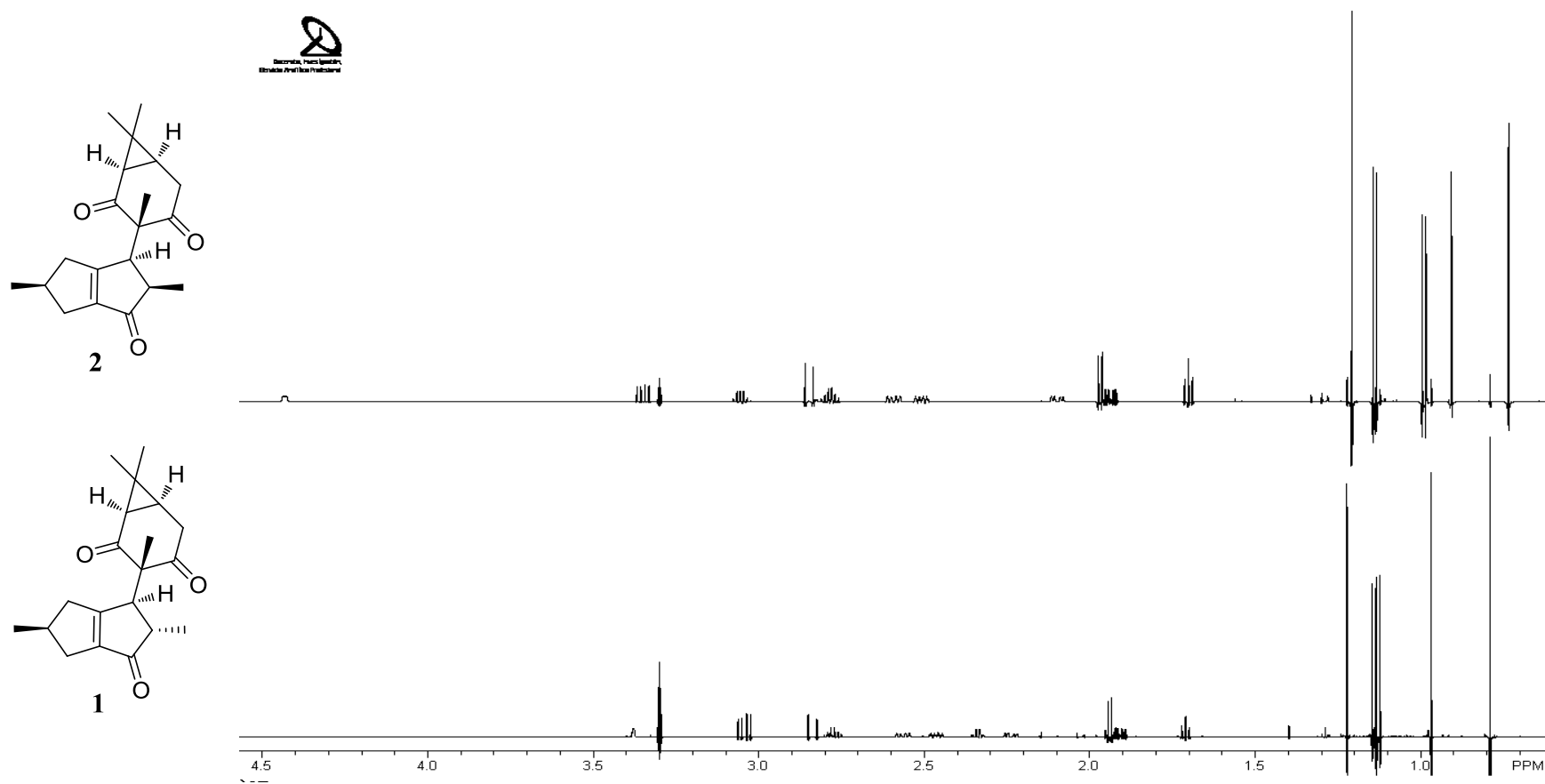
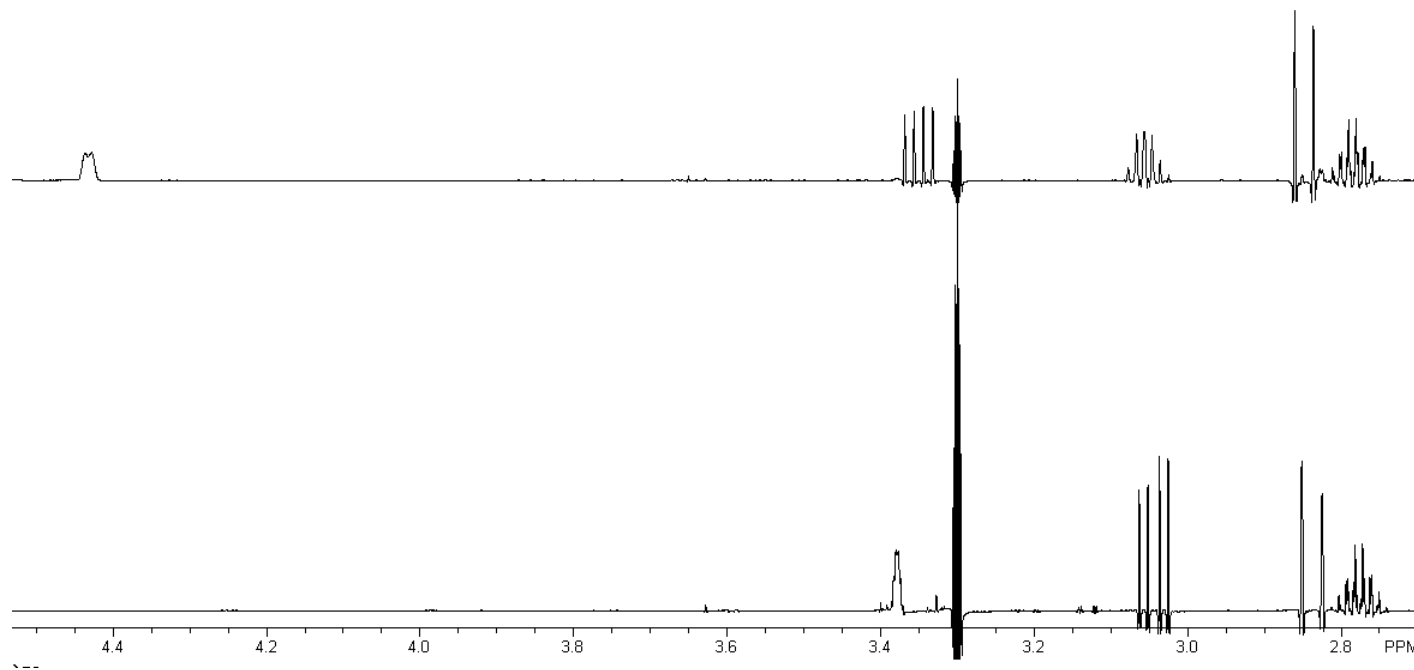
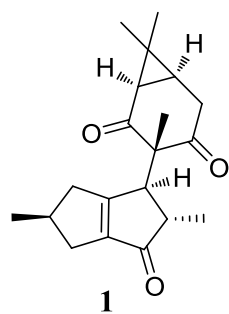
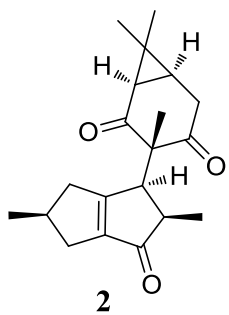
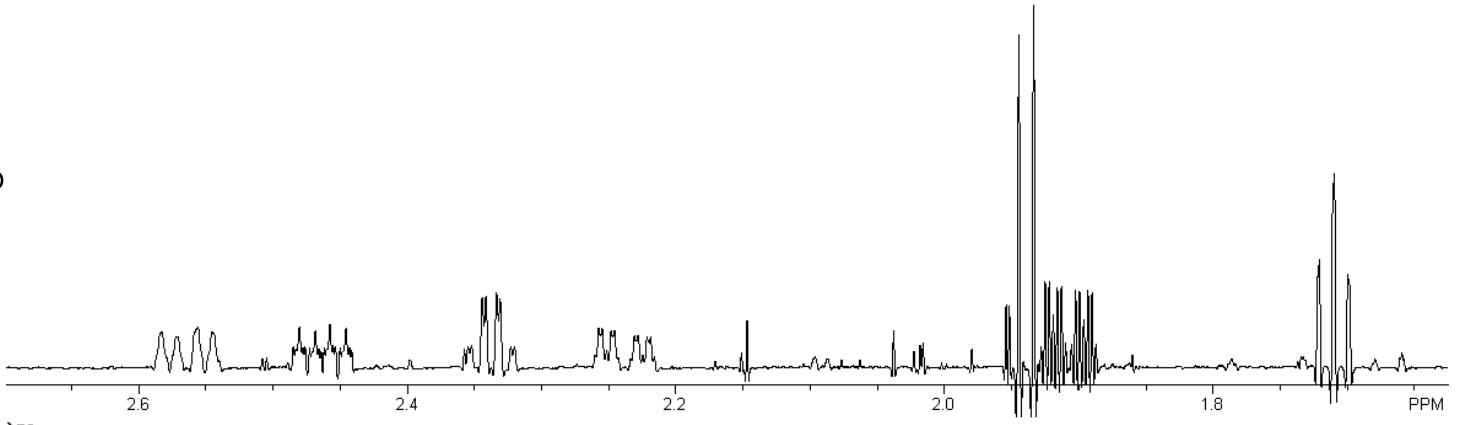
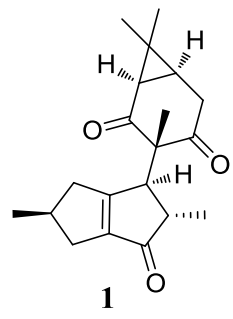
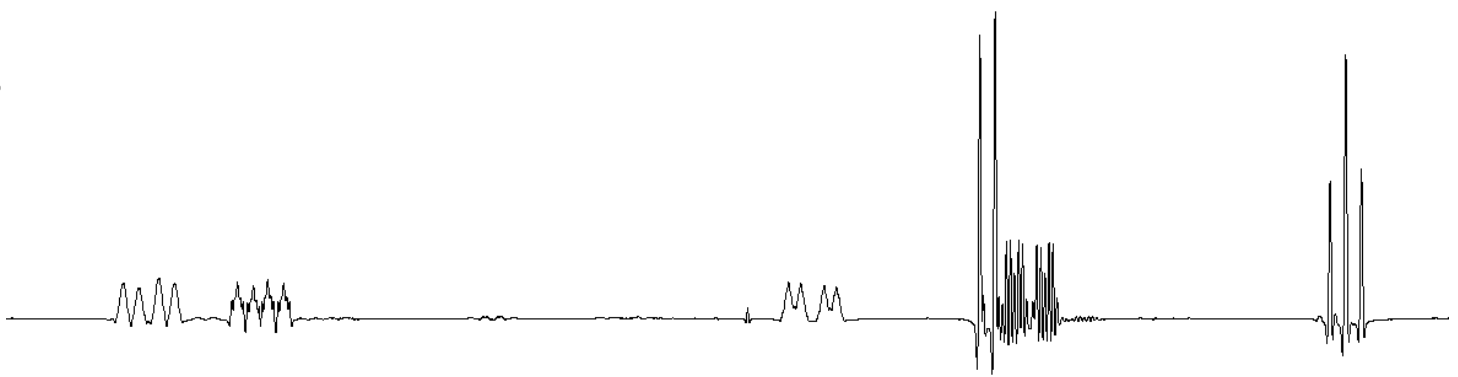
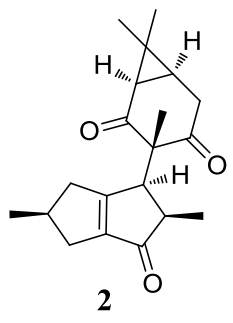


Figure S6. Comparison of the 1D ^1H -NMR spectra of compounds **1** and **2** in CD_3OD at 700 MHz, FIDs processed with Lorentzian-Gaussian apodization.





9
Date: _____
Page No: _____



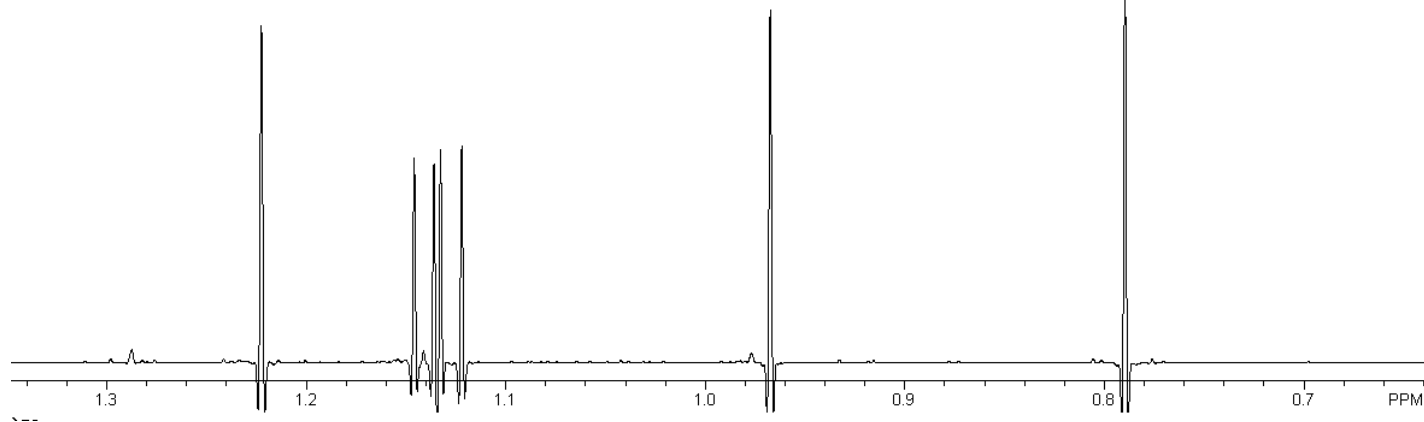
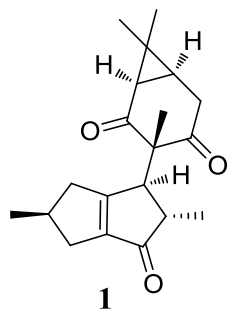
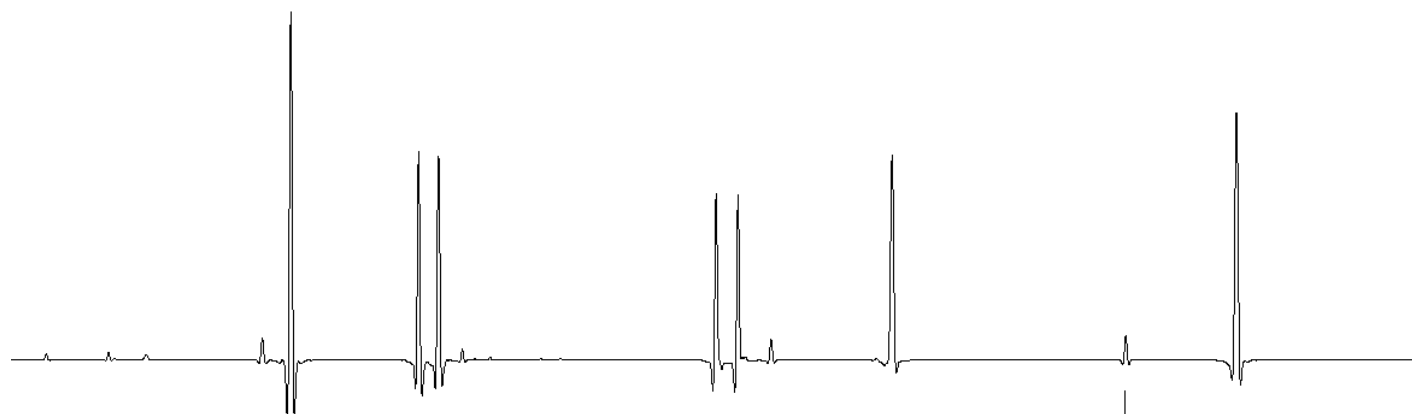
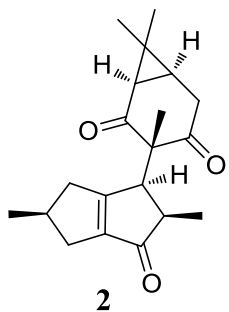


Figure S7. Comparison of the ^1H -NMR signals of the H-12 proton in compound **1** at 400 and 750 MHz and compound **2** at 750 MHz. Only in **1**, the highly coupled vicinal methylene protons, 2H-13, form an AB pair with very close chemical shifts ($\Delta\delta=0.0014\text{ppm}$). Therefore, H-12 in **1** gives rise to a higher order resonance pattern with a ddd-like multiplicity that can be readily misinterpreted when working under first-order assumptions. In contrast, as the methylene protons, 2H-13, resonate as relatively distant AM/AX nuclei in **2**, H-12 gives rise to an apparent triplet (pseudo triplet; with two additional long-range couplings). In fact, as the *6-epi* relationship between **1** and **2** does essentially not affect the geometry of the six-membered ring, its coupling patterns have to be identical in both molecules – only the resonances are not, due to the higher order effects caused by the differences in *relative* chemical shifts.

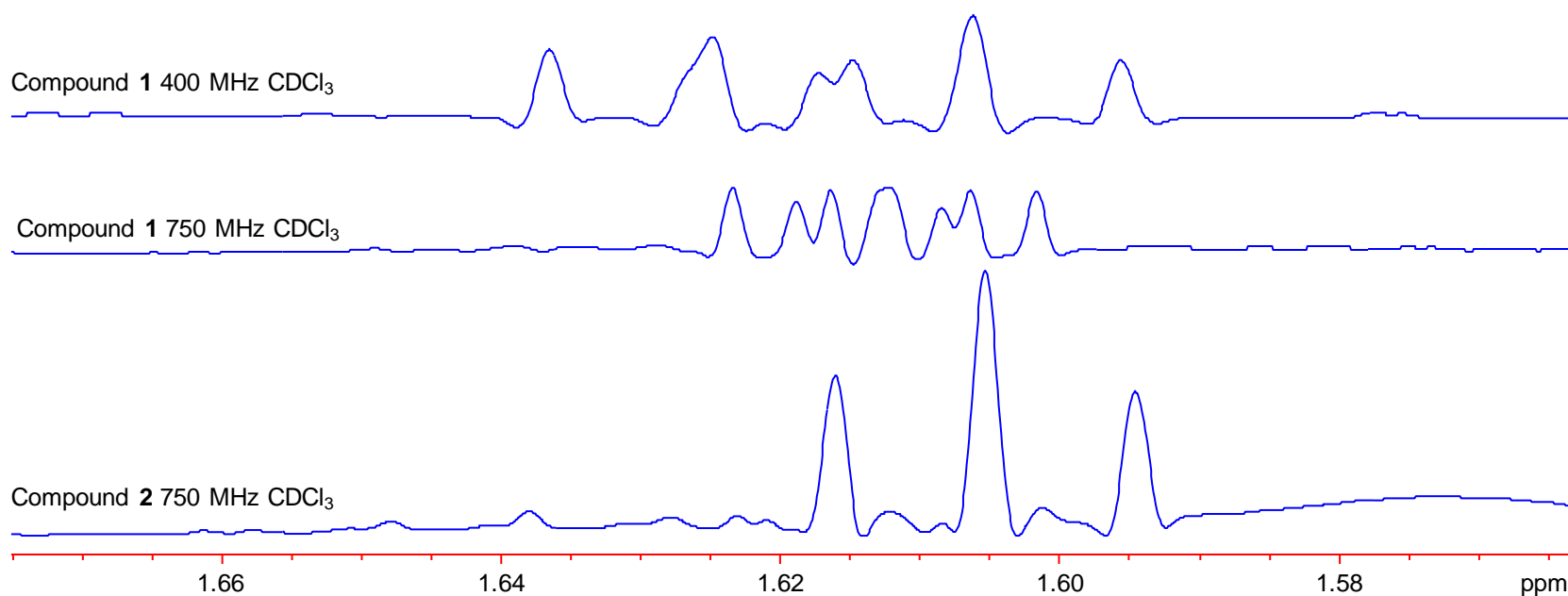


Figure S8. Comparison of the 100 MHz 1D ^{13}C -NMR spectra of compounds **1** and **2** in CD_3OD .

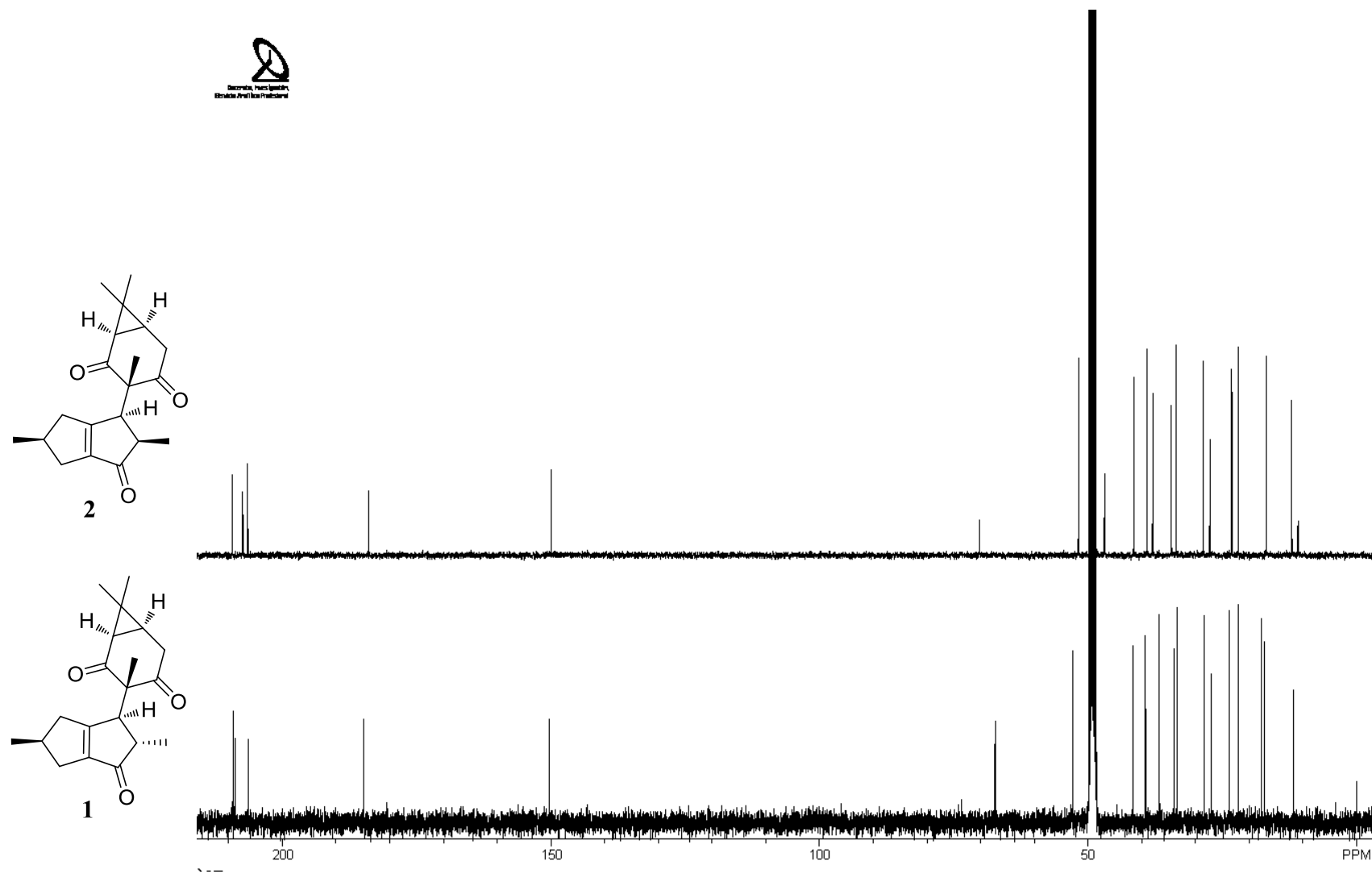


Figure S9. HSQC spectrum (400 MHz, CD₃OD) of 6-*epi*-Riolozatrione (**2**).

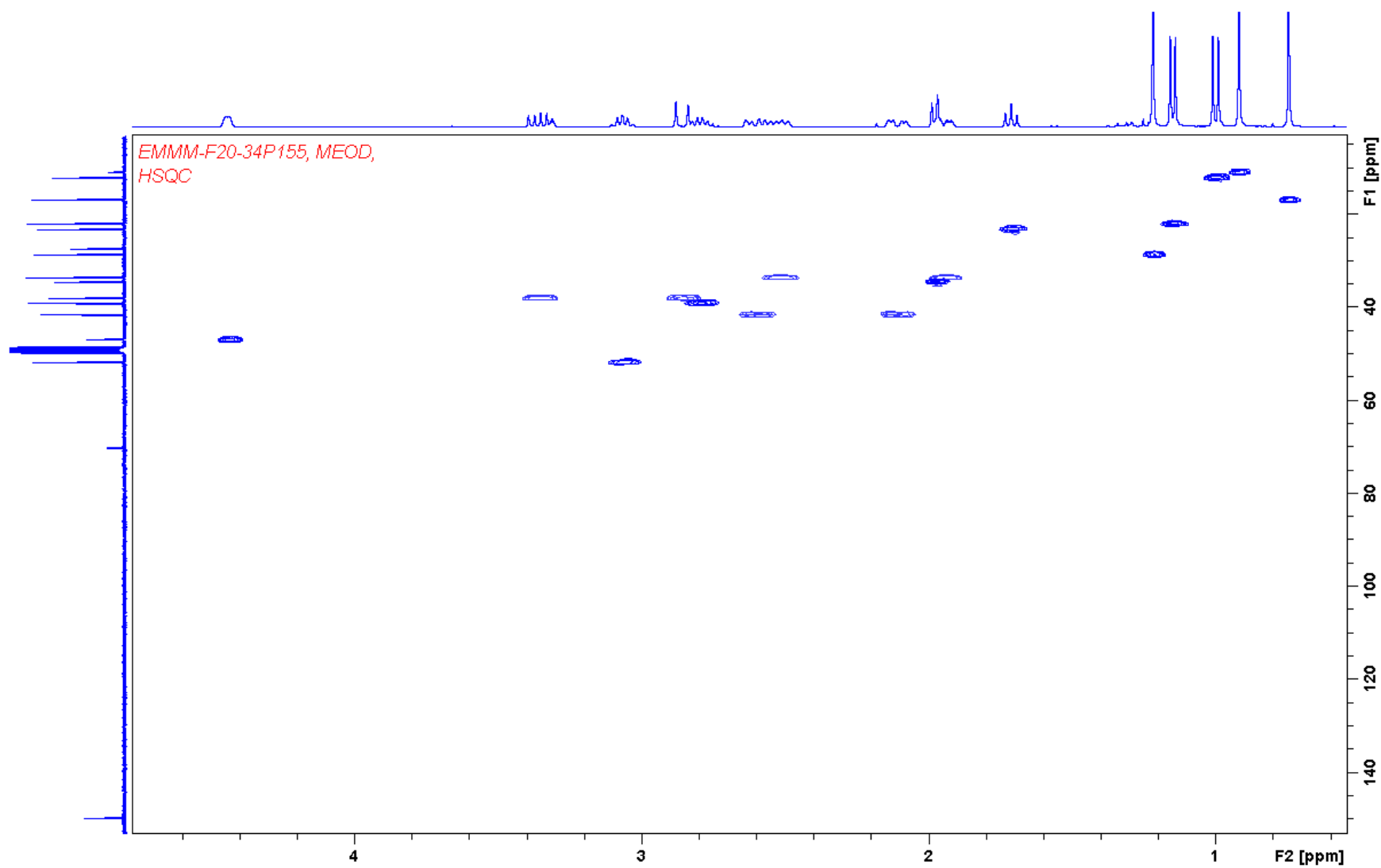


Figure S10. HMBC spectrum (400 MHz, CD₃OD) of 6-*epi*-Riolozatrione (**2**).

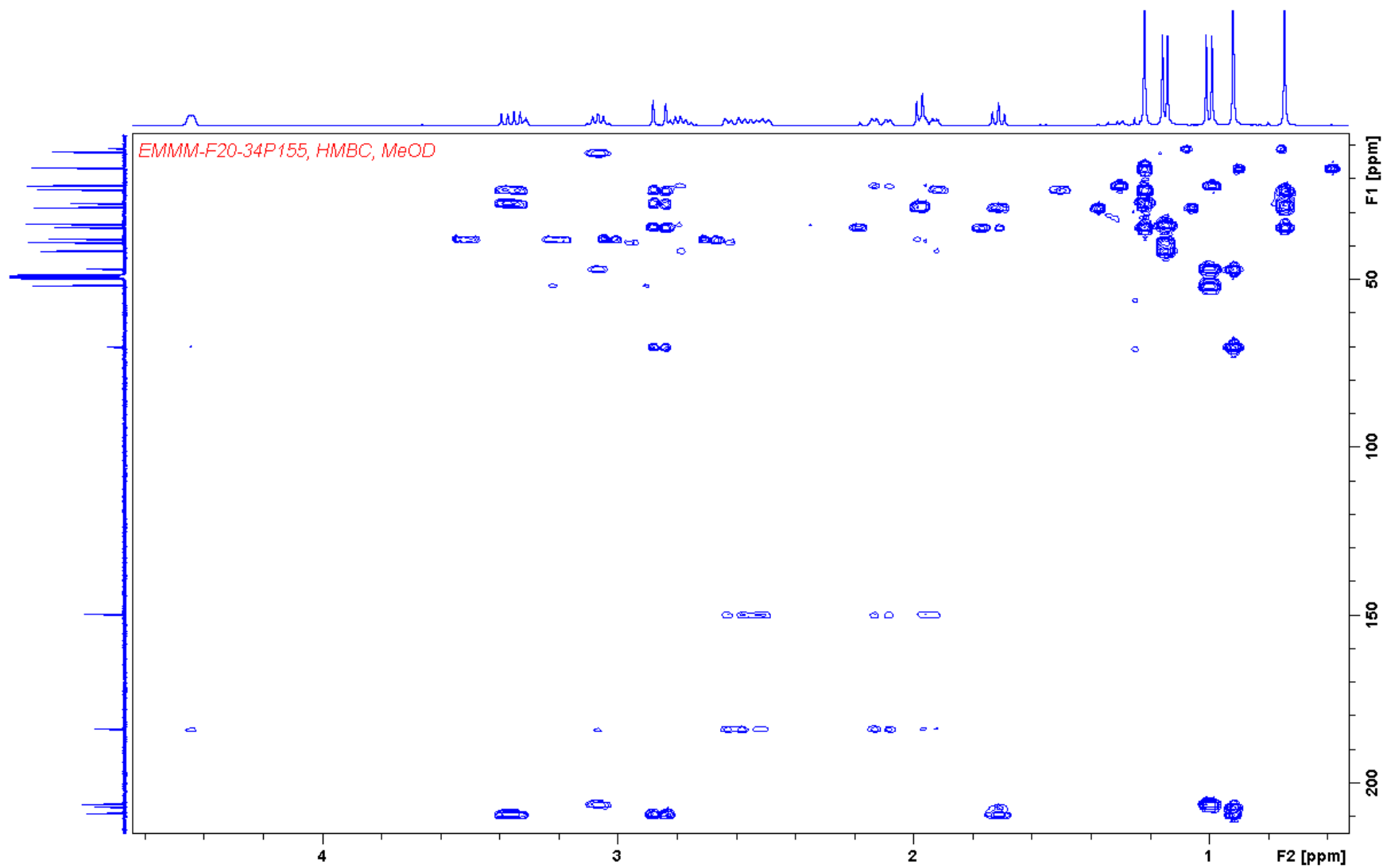


Figure S11. NOESY spectrum (400 MHz, CD₃OD) of 6-*epi*-Riolozatrione (**2**).

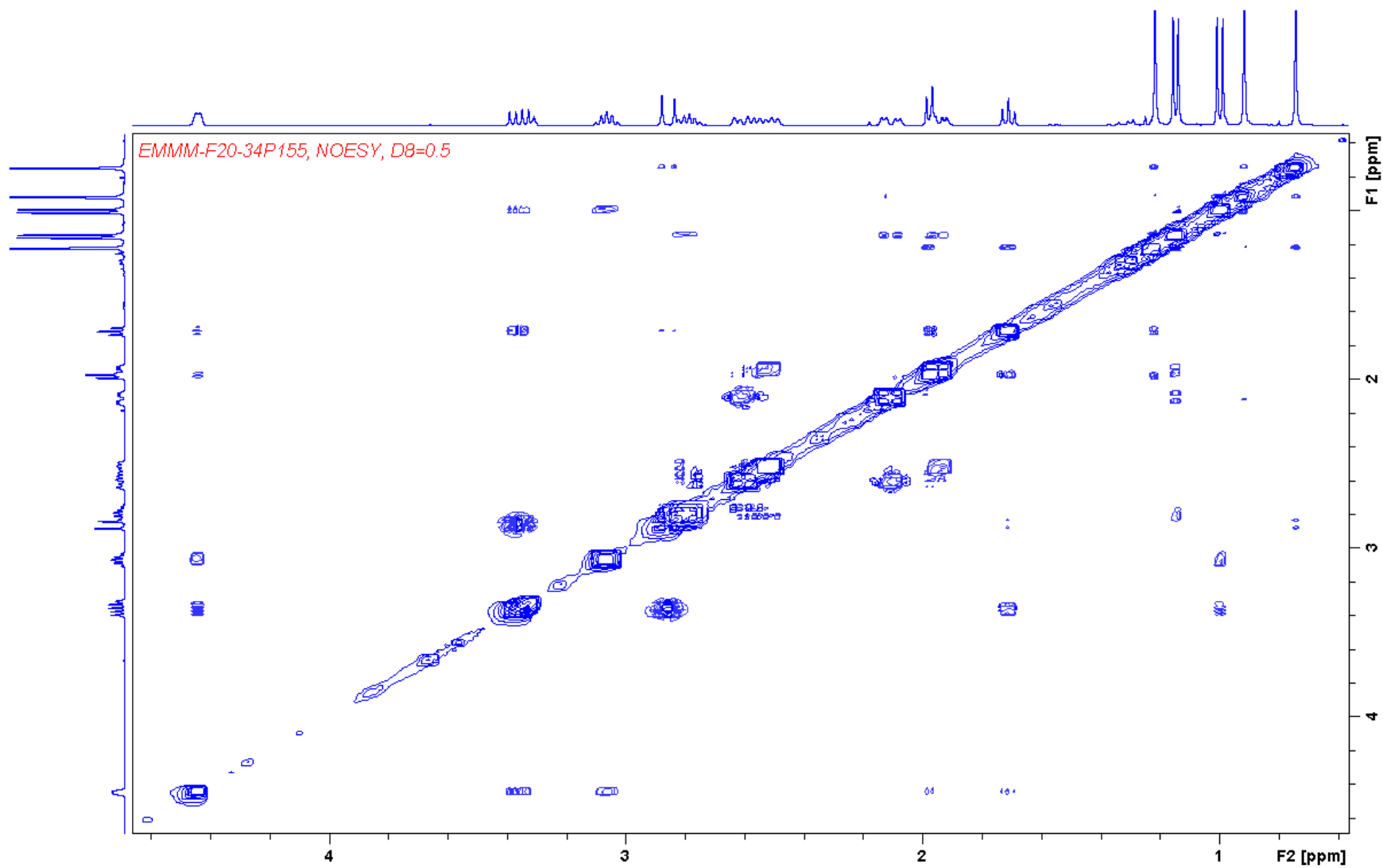


Figure S12. Calculated (red) vs. experimental (blue) and difference (green) 1D ^1H -NMR signal of H-1b in compound 1 in CD_3OD at 700 MHz.

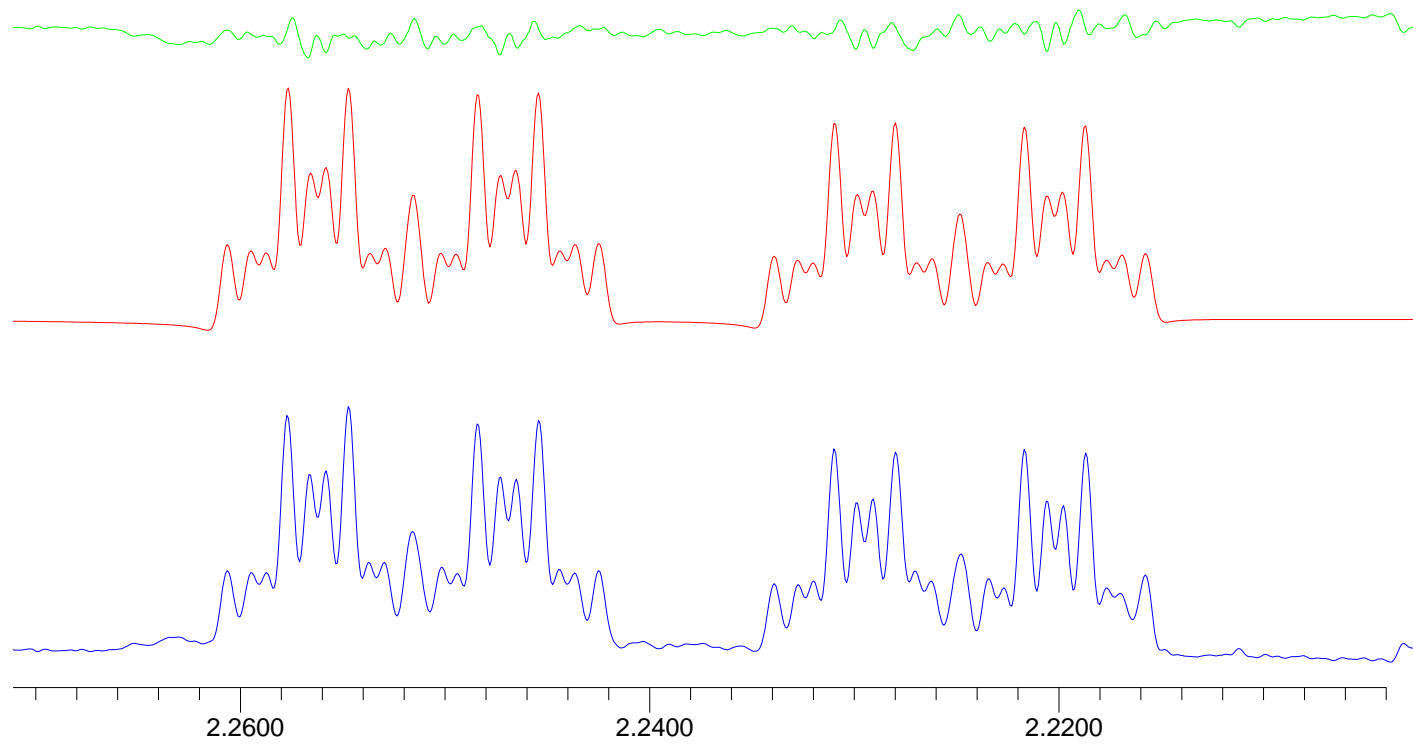


Figure S13. ¹H NMR HiFSA data for Riolozatrione (**1**) in CD₃OD (PERCH .pms file format)

```

ACTIVE SPECIES:1H
CHEMICAL SHIFTS (PPM): FIT-INDEX= 28.957
PROTON 2*SPIN=1 SPECIES=1H POPULATION= 1.0000
H7 /1 3.378440 1*1*1 STAT=Y PRED= 3.159 RANGE= 0.474 WIDTH= 1.033 RESP= 1.0000 HSQC=C7
H12 /1 1.710480 1*1*1 STAT=Y PRED= 1.558 RANGE= 0.411 WIDTH= 0.609 RESP= 1.0000 HSQC=C12
H15 /1 1.939020 1*1*1 STAT=Y PRED= 1.832 RANGE= 0.462 WIDTH= 0.876 RESP= 1.0000 HSQC=C15
H6 /1 2.337510 1*1*1 STAT=Y PRED= 2.872 RANGE= 0.468 WIDTH= 0.578 RESP= 1.0000 HSQC=C6
H1A /1 2.238770 1*1*1 STAT=Y PRED= 1.678 RANGE= 0.451 WIDTH= 0.521 RESP= 1.0000 HSQC=C1
H1B /1 2.563580 1*1*1 STAT=Y PRED= 2.328 RANGE= 0.444 WIDTH= 0.585 RESP= 1.0000 HSQC=C1
H2 /1 2.776340 1*1*1 STAT=Y PRED= 2.460 RANGE= 0.441 WIDTH= 0.483 RESP= 1.0000 HSQC=C2
H3A /1 1.907940 1*1*1 STAT=Y PRED= 2.069 RANGE= 0.391 WIDTH= 0.442 RESP= 1.0000 HSQC=C3
H3B /1 2.462910 1*1*1 STAT=Y PRED= 2.460 RANGE= 0.370 WIDTH= 0.506 RESP= 1.0000 HSQC=C3
H19 /1 1.140890 1*1*3 STAT=Y PRED= 1.058 RANGE= 0.264 WIDTH= 0.500 RESP= 1.0000 HSQC=C19
H13A/1 3.043140 1*1*1 STAT=Y PRED= 3.063 RANGE= 0.407 WIDTH= 0.653 RESP= 1.0000 HSQC=C13
H13B/1 2.838660 1*1*1 STAT=Y PRED= 2.343 RANGE= 0.416 WIDTH= 0.956 RESP= 1.0000 HSQC=C13
H16 /1 1.222410 1*1*3 STAT=Y PRED= 1.221 RANGE= 0.216 WIDTH= 0.750 RESP= 1.0000 HSQC=C16
H17 /1 0.789750 1*1*3 STAT=Y PRED= 0.965 RANGE= 0.332 WIDTH= 0.562 RESP= 1.0000 HSQC=C17
H18 /1 0.967460 1*1*3 STAT=Y PRED= 1.183 RANGE= 0.392 WIDTH= 0.676 RESP= 1.0000 HSQC=C18
H20 /1 1.127430 1*1*3 STAT=Y PRED= 1.173 RANGE= 0.269 WIDTH= 0.410 RESP= 1.0000 HSQC=C20

COUPLING CONSTANTS (HZ): FIT-INDEX= 0.000
J10_27 1.8960 J H7 H6 STAT=Y PRED= 5.629 RANGE= 3.222
J10_28 1.3566 J H7 H1A STAT=Y PRED= -0.413 RANGE= 5.000
J10_29 1.6867 J H7 H1B STAT=Y PRED= -0.424 RANGE= 5.000
J10_31 2.2395 J H7 H3A STAT=Y PRED= 0.000 RANGE= 0.001
J10_32 2.8447 J H7 H3B STAT=Y PRED= 0.000 RANGE= 0.001
J23_24 7.7901 J H12 H15 STAT=Y PRED= 10.333 RANGE= 2.000
J23_36 7.9276 J H12 H13A STAT=Y PRED= 9.837 RANGE= 5.000
J23_37 0.9628 J H12 H13B STAT=Y PRED= 1.118 RANGE= 5.000
J24_36 -0.2019 J H15 H13A STAT=Y PRED= 0.000 RANGE= 0.001
J24_37 -0.5000 J H15 H13B STAT=Y PRED= 0.000 RANGE= 0.001
J27_47 7.4637 J H6 H20 STAT=Y PRED= 7.032 RANGE= 2.237
J27_28 0.7750 J H6 H1A STAT=Y PRED= 0.000 RANGE= 0.001
J27_29 0.7459 J H6 H1B STAT=Y PRED= 0.000 RANGE= 0.001
J27_31 0.2028 J H6 H3A STAT=Y PRED= 0.000 RANGE= 0.001
J27_32 0.0884 J H6 H3B STAT=Y PRED= 0.000 RANGE= 0.001
J28_29 -18.7214 J H1A H1B STAT=Y PRED= -15.891 RANGE= 2.632
J28_30 6.4954 J H1A H2 STAT=Y PRED= 1.976 RANGE= 3.218
J28_31 2.0396 J H1A H3A STAT=Y PRED= 1.737 RANGE= 1.000
J28_32 2.0349 J H1A H3B STAT=Y PRED= 2.066 RANGE= 1.000
J29_30 8.2098 J H1B H2 STAT=Y PRED= 7.227 RANGE= 4.500
J29_31 2.0738 J H1B H3A STAT=Y PRED= 2.072 RANGE= 1.000
J29_32 1.0952 J H1B H3B STAT=Y PRED= 1.709 RANGE= 1.763
J30_33 6.9495 J H2 H19 STAT=Y PRED= 7.051 RANGE= 1.000
J30_31 6.4032 J H2 H3A STAT=Y PRED= 3.938 RANGE= 2.611
J30_32 8.4003 J H2 H3B STAT=Y PRED= 10.316 RANGE= 2.853
J31_32 -15.9028 J H3A H3B STAT=Y PRED= -16.751 RANGE= 3.272
J36_37 -18.5145 J H13A H13B STAT=Y PRED= -16.154 RANGE= 1.303

CONTROL PARAMETERS:
SOLVENT = None
REFERENCE = TMS
0.00000000 = CONCENTRATION (vol%)
298.000 = TEMPERATURE (vol%, def=1.0%)
699.95430000 = FIELD(1H,MHZ), used to transform shifts to ppm
0.00100000 = Minimum line-intensity
0.00100000 = Diagonalization criterium (not used)
0.00000000 = Left frequency limit (ppm)
0.00000000 = Right frequency limit (ppm)
0.410 = Default line-width (Hz)
0.000000000 = Data-point resolution (Hz)
0.000 = Gaussian contribution%
0.000 = Dispersion contribution%
0.00000000 = Decoupling frequency (for DORES only)

CONSTRAINTS (in equations X0 = 1.0)...use no empty lines

END of FILE

```

Figure S14. ¹H NMR HiFSA data for 6-*epi*-Riolozatrione (**2**) in CD₃OD (PERCH .pms file format)

```

ACTIVE SPECIES:1H
CHEMICAL SHIFTS (PPM): FIT-INDEX= 14.469
PROTON 2*SPIN=1 SPECIES=1H POPULATION= 1.0000
H7A /1 4.432000 1*1*1 STAT=Y PRED= 3.575 RANGE= 0.470 WIDTH= 1.653 RESP= 1.0000 HSQC=C7
H12A/1 1.701460 1*1*1 STAT=Y PRED= 1.508 RANGE= 0.411 WIDTH= 1.014 RESP= 1.0000 HSQC=C12
H15A/1 1.967950 1*1*1 STAT=Y PRED= 1.809 RANGE= 0.459 WIDTH= 0.946 RESP= 1.0000 HSQC=C15
H6A /1 3.056290 1*1*1 STAT=Y PRED= 3.203 RANGE= 0.435 WIDTH= 0.650 RESP= 1.0000 HSQC=C6
H1A /1 2.098740 1*1*1 STAT=Y PRED= 2.012 RANGE= 0.455 WIDTH= 0.811 RESP= 1.0000 HSQC=C1
H1B /1 2.592310 1*1*1 STAT=Y PRED= 2.186 RANGE= 0.432 WIDTH= 0.829 RESP= 1.0000 HSQC=C1
H2A /1 2.784900 1*1*1 STAT=Y PRED= 2.317 RANGE= 0.415 WIDTH= 0.376 RESP= 1.0000 HSQC=C2
H3A /1 1.936890 1*1*1 STAT=Y PRED= 1.990 RANGE= 0.387 WIDTH= 0.495 RESP= 1.0000 HSQC=C3
H3B /1 2.509160 1*1*1 STAT=Y PRED= 2.566 RANGE= 0.380 WIDTH= 0.449 RESP= 1.0000 HSQC=C3
H19 /1 1.138690 1*1*3 STAT=Y PRED= 1.063 RANGE= 0.230 WIDTH= 0.522 RESP= 1.0000 HSQC=C19
H13A/1 3.349960 1*1*1 STAT=Y PRED= 3.077 RANGE= 0.424 WIDTH= 1.410 RESP= 1.0000 HSQC=C13
H13B/1 2.848410 1*1*1 STAT=Y PRED= 2.336 RANGE= 0.438 WIDTH= 1.286 RESP= 1.0000 HSQC=C13
H17 /1 0.906450 1*1*3 STAT=Y PRED= 1.222 RANGE= 0.216 WIDTH= 1.158 RESP= 1.0000 HSQC=C17
H16 /1 0.733900 1*1*3 STAT=Y PRED= 0.881 RANGE= 0.351 WIDTH= 0.971 RESP= 1.0000 HSQC=C16
H18 /1 1.207750 1*1*3 STAT=Y PRED= 1.255 RANGE= 0.414 WIDTH= 0.486 RESP= 1.0000 HSQC=C18
H20 /1 0.989150 1*1*3 STAT=Y PRED= 1.169 RANGE= 0.359 WIDTH= 0.567 RESP= 1.0000 HSQC=C20

COUPLING CONSTANTS (HZ): FIT-INDEX= 0.000
J10_27 6.1386 J H7A H6A STAT=Y PRED= 7.606 RANGE= 3.326
J10_28 1.3117 J H7A H1A STAT=Y PRED= -0.351 RANGE= 5.000
J10_29 1.6114 J H7A H1B STAT=Y PRED= -0.432 RANGE= 5.000
J10_31 2.2248 J H7A H3A STAT=Y PRED= 0.000 RANGE= 0.001
J10_32 2.7617 J H7A H3B STAT=Y PRED= 0.000 RANGE= 0.001
J10_30 0.5000 J H7A H2A STAT=Y PRED= 0.000 RANGE= 0.001
J10_44 -0.4145 J H7A H18 STAT=Y PRED= 0.000 RANGE= 0.001
J23_24 7.9358 J H12A H15A STAT=Y PRED= 10.333 RANGE= 2.000
J23_37 0.6424 J H12A H13B STAT=Y PRED= 0.269 RANGE= 5.000
J23_36 8.4397 J H12A H13A STAT=Y PRED= 8.850 RANGE= 5.000
J27_47 7.7129 J H6A H20 STAT=Y PRED= 7.032 RANGE= 2.237
J27_28 0.9992 J H6A H1A STAT=Y PRED= 0.000 RANGE= 0.001
J27_29 0.9352 J H6A H1B STAT=Y PRED= 0.000 RANGE= 0.001
J27_32 0.2720 J H6A H3B STAT=Y PRED= 0.000 RANGE= 0.001
J27_30 0.0029 J H6A H2A STAT=Y PRED= 0.000 RANGE= 0.001
J28_29 -18.5844 J H1A H1B STAT=Y PRED= -15.884 RANGE= 2.632
J28_30 6.3365 J H1A H2A STAT=Y PRED= 7.062 RANGE= 4.500
J28_31 1.9488 J H1A H3A STAT=Y PRED= 1.716 RANGE= 1.758
J28_32 2.0035 J H1A H3B STAT=Y PRED= 2.072 RANGE= 1.000
J29_30 8.2174 J H1B H2A STAT=Y PRED= 7.210 RANGE= 4.500
J29_31 2.0716 J H1B H3A STAT=Y PRED= 2.069 RANGE= 1.000
J29_32 1.1656 J H1B H3B STAT=Y PRED= 1.750 RANGE= 1.000
J30_33 6.9555 J H2A H19 STAT=Y PRED= 7.051 RANGE= 1.000
J30_31 6.3205 J H2A H3A STAT=Y PRED= 7.100 RANGE= 2.288
J30_32 8.3159 J H2A H3B STAT=Y PRED= 10.313 RANGE= 2.855
J31_32 -15.8538 J H3A H3B STAT=Y PRED= -16.750 RANGE= 3.273
J36_37 -16.9872 J H13A H13B STAT=Y PRED= -16.158 RANGE= 1.302

CONTROL PARAMETERS:
SOLVENT = None
REFERENCE = TMS
0.00000000 = CONCENTRATION (vol%)
298.000 = TEMPERATURE (vol%, def=1.0%)
699.95430000 = FIELD(1H,MHZ), used to transform shifts to ppm
0.00100000 = Minimum line-intensity
0.00100000 = Diagonalization criterium (not used)
0.00000000 = Left frequency limit (ppm)
0.00000000 = Right frequency limit (ppm)
0.376 = Default line-width (Hz)
0.0000000000 = Data-point resolution (Hz)
0.000 = Gaussian contribution%
0.000 = Dispersion contribution%
0.00000000 = Decoupling frequency (for DORES only)

CONSTRAINTS (in equations X0 = 1.0)...use no empty lines:

END of FILE

```

Figure S15. ¹H NMR HiFSA data for Riolozatrione (**1**) in CDCl₃ (PERCH .pms file format)

```

ACTIVE SPECIES:1H
CHEMICAL SHIFTS (PPM) :
PROTON      2*SPIN= 1 SPECIES=1H      POPULATION (Y)= 0.80740
H7 / 1      3.126458 1*1*1 STAT=Y PRED= 3.125 RANGE= 0.073 WIDTH(Y)= 1.324 RESP(Y)= 1.0000
H12 / 1     1.614077 1*1*1 STAT=Y PRED= 1.614 RANGE= 0.073 WIDTH(Y)= 0.892 RESP(Y)= 1.0000
H15 / 1     1.883196 1*1*1 STAT=Y PRED= 1.883 RANGE= 0.073 WIDTH(Y)= 0.646 RESP(Y)= 1.0000
H6 / 1      2.456355 1*1*1 STAT=Y PRED= 2.457 RANGE= 0.073 WIDTH(Y)= 1.023 RESP(Y)= 1.0000
H1A / 1     2.310340 1*1*1 STAT=Y PRED= 2.310 RANGE= 0.073 WIDTH(Y)= 0.878 RESP(Y)= 1.0000
H1B / 1     2.485253 1*1*1 STAT=Y PRED= 2.483 RANGE= 0.073 WIDTH(Y)= 0.726 RESP(Y)= 1.0000
H2 / 1      2.753404 1*1*1 STAT=Y PRED= 2.755 RANGE= 0.073 WIDTH(Y)= 0.940 RESP(Y)= 1.0000
H3A / 1     1.961866 1*1*1 STAT=Y PRED= 1.961 RANGE= 0.073 WIDTH(Y)= 0.907 RESP(Y)= 1.0000
H3B / 1     2.494908 1*1*1 STAT=Y PRED= 2.497 RANGE= 0.073 WIDTH(Y)= 0.916 RESP(Y)= 1.0000
H19 / 1     1.153539 1*1*3 STAT=Y PRED= 1.155 RANGE= 0.073 WIDTH(Y)= 0.886 RESP(Y)= 1.0000
H13A / 1    2.882158 1*1*1 STAT=Y PRED= 2.888 RANGE= 0.073 WIDTH(Y)= 0.928 RESP(Y)= 1.0000
H13B / 1    2.884841 1*1*1 STAT=Y PRED= 2.878 RANGE= 0.073 WIDTH(Y)= 1.124 RESP(Y)= 1.0000
H16 / 1     1.235937 1*1*3 STAT=Y PRED= 1.235 RANGE= 0.073 WIDTH(Y)= 1.143 RESP(Y)= 1.0000
H17 / 1     0.852886 1*1*3 STAT=Y PRED= 0.853 RANGE= 0.073 WIDTH(Y)= 0.933 RESP(Y)= 1.0000
H18 / 1     1.088124 1*1*3 STAT=Y PRED= 1.089 RANGE= 0.073 WIDTH(Y)= 1.031 RESP(Y)= 1.0000
H20 / 1     1.157132 1*1*3 STAT=Y PRED= 1.157 RANGE= 0.073 WIDTH(Y)= 0.846 RESP(Y)= 1.0000

COUPLING CONSTANTS (HZ) :
J10_27      2.0096 J H7 H6 STAT=Y PRED= 1.896 RANGE= 0.100
J10_28      1.3779 J H7 H1A STAT=Y PRED= 1.357 RANGE= 0.100
J10_29      1.5871 J H7 H1B STAT=Y PRED= 1.687 RANGE= 0.100
J10_31      2.2919 J H7 H3A STAT=Y PRED= 2.240 RANGE= 0.100
J10_32      2.8463 J H7 H3B STAT=Y PRED= 2.845 RANGE= 0.100
J23_24      7.6821 J H12 H15 STAT=Y PRED= 7.790 RANGE= 0.100
J23_36      8.0127 J H12 H13A STAT=Y PRED= 7.928 RANGE= 0.100
J23_37      0.6452 J H12 H13B STAT=Y PRED= 0.963 RANGE= 0.100
J24_36     -0.4748 J H15 H13A STAT=Y PRED= -0.202 RANGE= 0.100
J24_37     -0.1206 J H15 H13B STAT=Y PRED= -0.500 RANGE= 0.100
J27_28      0.7631 J H6 H1A STAT=Y PRED= 0.775 RANGE= 0.100
J27_29      0.6335 J H6 H1B STAT=Y PRED= 0.746 RANGE= 0.100
J27_31      0.0080 J H6 H3A STAT=Y PRED= 0.203 RANGE= 0.100
J27_32      0.0673 J H6 H3B STAT=Y PRED= 0.088 RANGE= 0.100
J27_47      7.4599 J H6 H20 STAT=Y PRED= 7.464 RANGE= 0.100
J28_29     -18.5611 J H1A H1B STAT=Y PRED= -18.721 RANGE= 0.100
J28_30      6.6499 J H1A H2 STAT=Y PRED= 6.495 RANGE= 0.100
J28_31      2.0453 J H1A H3A STAT=Y PRED= 2.040 RANGE= 0.100
J28_32      2.0472 J H1A H3B STAT=Y PRED= 2.035 RANGE= 0.100
J29_30      8.2769 J H1B H2 STAT=Y PRED= 8.210 RANGE= 0.100
J29_31      2.1368 J H1B H3A STAT=Y PRED= 2.072 RANGE= 1.000
J29_32      1.0197 J H1B H3B STAT=Y PRED= 1.095 RANGE= 0.100
J30_31      6.6044 J H2 H3A STAT=Y PRED= 6.403 RANGE= 0.100
J30_32      8.4073 J H2 H3B STAT=Y PRED= 8.400 RANGE= 0.100
J30_33      6.9716 J H2 H19 STAT=Y PRED= 6.949 RANGE= 0.100
J31_32     -16.0398 J H3A H3B STAT=Y PRED= -15.903 RANGE= 0.100
J36_37     -18.1640 J H13A H13B STAT=Y PRED= -18.515 RANGE= 0.100

CONTROL PARAMETERS:
Solvent = none (def. 99% enriched)
REFERENCE = TMS
1.000 = Concentration (vol%, def=1.0%)
0.001000000 = Minimum line-intensity
0.001000000 = Diagonalization criterium (not in use)
699.95432200 = FIELD(1H,MHz), used to transform shifts to ppms
16.23558651 = Left frequency (ppm)
-3.90528483 = Right frequency (ppm)
0.000 = Acquisition time (s, for QMtls)
0.397 = Line-width (for modes D, P & T, 0=use defaults)
0.053778627 = Data-point resolution (Hz)
52.609 = GAUSSIAN (% , 0=use default from INF)
0.000 = Dispersion contribution (% , 0=use default from INF)
0.000000000 = Decoupling frequency (for DORES)

CONSTRAINTS (in equations X0 = 1.0)...use no empty lines

END of FILE

```

Figure S16. ¹H NMR HiFSA data for 6-*epi*-Riolozatrione (**2**) in CDCl₃ (PERCH .pms file format)

```
ACTIVE SPECIES:1H
CHEMICAL SHIFTS (PPM) :
PROTON      2*SPIN= 1 SPECIES=1H      POPULATION (Y)= 0.91072
H7A / 1      4.219272 1*1*1  STAT=Y  PRED= 4.222 RANGE= 0.073 WIDTH (Y)= 3.500 RESP (Y)= 1.0000
H12A / 1     1.607877 1*1*1  STAT=Y  PRED= 1.607 RANGE= 0.073 WIDTH (Y)= 0.629 RESP (Y)= 1.0000
H15A / 1     1.864763 1*1*1  STAT=Y  PRED= 1.862 RANGE= 0.073 WIDTH (Y)= 0.946 RESP (Y)= 1.0000
H6A / 1      2.921293 1*1*1  STAT=Y  PRED= 2.920 RANGE= 0.073 WIDTH (Y)= 0.718 RESP (Y)= 1.0000
H1A / 1      2.174499 1*1*1  STAT=Y  PRED= 2.172 RANGE= 0.073 WIDTH (Y)= 3.107 RESP (Y)= 1.0000
H1B / 1      2.479418 1*1*1  STAT=Y  PRED= 2.481 RANGE= 0.073 WIDTH (Y)= 1.775 RESP (Y)= 1.0000
H2A / 1      2.757998 1*1*1  STAT=Y  PRED= 2.757 RANGE= 0.073 WIDTH (Y)= 0.554 RESP (Y)= 1.0000
H3A / 1      2.002126 1*1*1  STAT=Y  PRED= 2.002 RANGE= 0.073 WIDTH (Y)= 0.576 RESP (Y)= 1.0000
H3B / 1      2.540613 1*1*1  STAT=Y  PRED= 2.541 RANGE= 0.073 WIDTH (Y)= 0.605 RESP (Y)= 1.0000
H19B / 1     1.155959 1*1*3  STAT=Y  PRED= 1.156 RANGE= 0.073 WIDTH (Y)= 0.643 RESP (Y)= 1.0000
H13A / 1     3.121516 1*1*1  STAT=Y  PRED= 3.123 RANGE= 0.073 WIDTH (Y)= 1.498 RESP (Y)= 1.0000
H13B / 1     2.931317 1*1*1  STAT=Y  PRED= 2.929 RANGE= 0.073 WIDTH (Y)= 1.868 RESP (Y)= 1.0000
H17B / 1     1.019603 1*1*3  STAT=Y  PRED= 1.019 RANGE= 0.073 WIDTH (Y)= 2.914 RESP (Y)= 1.0000
H16B / 1     0.804109 1*1*3  STAT=Y  PRED= 0.804 RANGE= 0.073 WIDTH (Y)= 2.037 RESP (Y)= 1.0000
H18B / 1     1.226542 1*1*3  STAT=Y  PRED= 1.225 RANGE= 0.073 WIDTH (Y)= 0.780 RESP (Y)= 1.0000
H20B / 1     1.070224 1*1*3  STAT=Y  PRED= 1.069 RANGE= 0.073 WIDTH (Y)= 0.619 RESP (Y)= 1.0000
```

```
COUPLING CONSTANTS (HZ) :
J10_27      6.3191  J H7A  H6A  STAT=Y  PRED= 7.604 RANGE= 3.320
J48         1.4989  J H7A  H1A  STAT=Y  PRED= -0.500 RANGE= 1.000
J49         1.5066  J H7A  H1B  STAT=Y  PRED= -0.500 RANGE= 1.000
J45         2.1803  J H7A  H3A  STAT=Y  PRED= 2.000 RANGE= 1.000
J46         2.8238  J H7A  H3B  STAT=Y  PRED= 2.000 RANGE= 1.000
20          -0.3510  J H7A  H18B STAT=Y
J23_24      7.8141  J H12A H15A STAT=Y  PRED= 9.000 RANGE= 2.000
J23_36      8.3722  J H12A H13A STAT=Y  PRED= 8.821 RANGE= 5.000
J23_37      0.6381  J H12A H13B STAT=Y  PRED= 0.280 RANGE= 5.000
18          0.8346  J H6A  H1A  STAT=Y  PRED= 0.280 RANGE= 5.000
19          0.8728  J H6A  H1B  STAT=Y  PRED= 0.280 RANGE= 5.000
J27_47      7.7524  J H6A  H20B STAT=Y  PRED= 7.032 RANGE= 2.230
J28_29     -18.4677  J H1A  H1B  STAT=Y  PRED= -15.926 RANGE= 2.560
J28_30      6.5099  J H1A  H2A  STAT=Y  PRED= 7.071 RANGE= 4.500
J28_31      2.1374  J H1A  H3A  STAT=Y  PRED= 1.716 RANGE= 1.750
J28_32      2.0492  J H1A  H3B  STAT=Y  PRED= 2.072 RANGE= 1.000
J29_30      8.2393  J H1B  H2A  STAT=Y  PRED= 7.215 RANGE= 4.500
J29_31      2.2020  J H1B  H3A  STAT=Y  PRED= 2.069 RANGE= 1.000
J29_32      1.1195  J H1B  H3B  STAT=Y  PRED= 1.749 RANGE= 1.000
J30_31      6.4856  J H2A  H3A  STAT=Y  PRED= 7.096 RANGE= 2.280
J30_32      8.3419  J H2A  H3B  STAT=Y  PRED= 10.031 RANGE= 3.200
J30_33      6.9456  J H2A  H19B STAT=Y  PRED= 7.056 RANGE= 1.000
J31_32     -16.0054  J H3A  H3B  STAT=Y  PRED= -16.802 RANGE= 3.250
J36_37     -16.9517  J H13A H13B STAT=Y  PRED= -15.509 RANGE= 1.330
```

```
CONTROL PARAMETERS:
Solvent = none (def. 99% enriched)
REFERENCE = TMS
1.000 = Concentration (vol%, def=1.0%)
0.00100000 = Minimum line-intensity
0.00100000 = Diagonalization criterium (not in use)
699.95432200 = FIELD(1H,MHz), used to transform shifts to ppm
16.23496007 = Left frequency (ppm)
-3.90591196 = Right frequency (ppm)
0.000 = Acquisition time (s, for QMFLS)
0.495 = Line-width (for modes D, P & T, 0=use defaults)
0.053778627 = Data-point resolution (Hz)
142.439 = GAUSSIAN (% , 0=use default from INF)
0.000 = Dispersion contribution (% , 0=use default from INF)
0.00000000 = Decoupling frequency (for DORES)
```

CONSTRAINTS (in equations X0 = 1.0)...use no empty lines

END of FILE

Figure S17. Full ^1H NMR δ and J -correlation maps, termed Quantum Interaction and Linkage Tables (QuILTs), of riolozatrione (**1**) and 6-*epi*-riolozatrione (**2**) were achieved by HiFSA processing of the 700 MHz spectra in CDCl_3 . Multiplicities within parentheses are due to couplings of ≤ 1 Hz. Couplings with absolute value of ≤ 0.10 Hz are given as “ \emptyset ”.

δ 1	M	H	1b(β)	1a(α)	2	3b(β)	3a(α)	6	7	11	12	13a(α)	13b(β)	16	17	18	19	20
2.3103	dddd(d)	1b(β)	d	2	3	4	4	5	4	7	8	7	7	9	9	6	4	6
2.4853	dddd(d)	1a(α)	-18.56	d	3	4	4	5	4	7	8	7	7	9	9	6	4	6
2.7534	ddqdd	2	6.65	8.28	d	3	3	6	5	8	9	8	8	10	10	7	3	7
1.9619	dddd	3b(β)	2.05	2.14	6.60	d	2	5	5	8	9	8	8	10	10	7	4	6
2.4949	dddd	3a(α)	2.05	1.02	8.41	-16.04	d	5	5	8	9	8	8	10	10	7	4	6
2.4564	qd(dd)	6	0.76	0.63	\emptyset	\emptyset	\emptyset	d	3	6	7	6	6	8	8	5	7	3
3.1265	dddd	7	1.38	1.59	\emptyset	2.29	2.85	2.01	d	5	6	5	5	7	7	4	6	4
1.8832	d(dd)	11	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	d	3	4	4	4	4	5	9	7
1.6141	dd(d)	12	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	7.68	d	3	3	4	4	6	10	8
2.8822	dd(d)	13a(α)	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	-0.47	8.01	d	2	5	5	5	9	7	
2.8848	d(dd)	13b(β)	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	-0.12	0.65	-18.16	d	5	5	5	9	7	
1.2359	s	16	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	d	4	7	11	9
0.8529	s	17	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	d	7	11	9
1.0881	s	18	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	d	8	6
1.1535	d	19	\emptyset	\emptyset	6.972	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	d	8
1.1571	d	20	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	7.46	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	d

δ 2	M	H	1b(β)	1a(α)	2	3b(β)	3a(α)	6	7	11	12	13a(α)	13b(β)	16	17	18	19	20
2.1745	dddd(d)	1b(β)	d	2	3	4	4	5	4	7	8	7	7	9	9	6	4	6
2.4794	dddd(d)	1a(α)	-18.47	d	3	4	4	5	4	7	8	7	7	9	9	6	4	6
2.7580	ddqdd	2	6.51	8.24	d	3	3	6	5	8	9	8	8	10	10	7	3	7
2.0021	dddd	3b(β)	2.14	2.20	6.486	d	2	5	5	8	9	8	8	10	10	7	4	6
2.5406	dddd	3a(α)	2.05	1.12	8.34	-16.01	d	5	5	8	9	8	8	10	10	7	4	6
2.9213	qd(dd)	6	0.83	0.87	\emptyset	\emptyset	\emptyset	d	3	6	7	6	6	8	8	5	7	3
4.2193	dddd(d)	7	1.50	1.51	\emptyset	2.18	2.82	6.32	d	5	6	5	5	7	7	4	6	4
1.8648	d	11	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	d	3	4	4	4	4	5	9	7
1.6079	dd(d)	12	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	7.81	d	3	3	4	4	6	10	8
3.1215	dd	13a(α)	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	8.37	d	2	5	5	5	9	7
2.9313	d(d)	13b(β)	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	0.64	-16.95	d	5	5	5	9	7
1.2265	s	16	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	d	4	7	11	9
0.8041	s	17	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	d	7	11	9
1.0196	(d)	18	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	-0.35	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	d	8	6
1.1560	d	19	\emptyset	\emptyset	6.95	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	d	8
1.0702	d	20	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	7.75	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	\emptyset	d

Figure S18. Calculated (red) vs. experimental (blue) and difference (green) 1D ^1H -NMR signals in compounds **1** and **2** in CDCl_3 at 700 MHz.

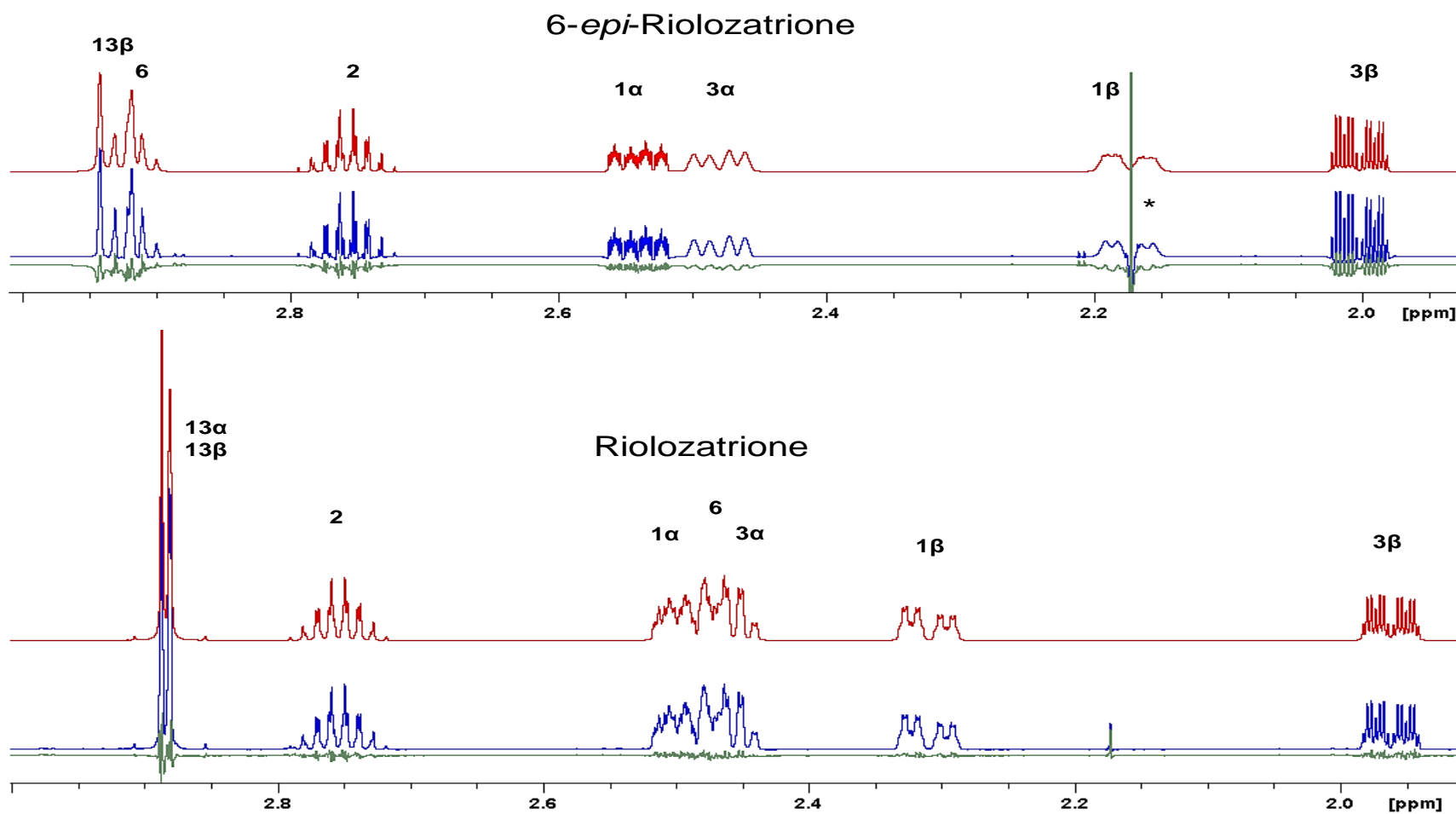


Figure S19. Experimental and calculated IR and VCD spectra at the B3PW91/DGDZVP level of theory for **3** (left) and **4** (right).

