Structural analysis of the complex between influenza B nucleoprotein and human

importin- α

Alice LABARONNE^{1\$}, Sigrid MILLES^{1\$}, Amélie DONCHET¹, Malene Ringkjøbing JENSEN¹, Martin

BLACKLEDGE¹, Jean-Marie BOURHIS¹, Rob WH RUIGROK¹ & Thibaut CREPIN^{1*}

¹ Univ. Grenoble Alpes, CNRS, CEA, IBS, F-38000 Grenoble

Supplementary information

Supplementary Figure 1

Conformational propensities of B/NP_{TAIL}. (a) Local conformational propensities of a set of 200 conformers describing the structural propensities of B/NP_{TAIL} as derived from a combination of Flexible-Meccano and ASTEROIDS using chemical shifts (grey bars) of B/NP_{TAIL} and propensities as expected from a statistical coil (black lines). Categories as described in Fig. 2: β -sheet (β S), polyproline (β P), right (α R) and left handed (α L). (b) Secondary chemical shifts as obtained from B/NP_{TAIL} assignment (grey bars) and back calculated from the ASTEROIDS ensemble (red lines).

Supplementary Figure 2

Interaction of B/NP with importin- α 7. SDS-PAGE gels (Tris-Glycine, 4-20 % polyacrylamide) stained with coomassie blue of the control experiments done on (a) a SuperdexTM 200 increase 10/300GL column for B/NP alone, importin- α 7 alone and B/NP_{CORE} alone and on (b) a SuperdexTM 75 10/300GL column for B/NP_{TAIL} and importin- α 7.

Supplementary Figure 3

Homogeneous sample analysis of recombinant B/NP_{CORE}.

The MALLS run was performed using a SuperdexTM 200 increase 10/300GL (GE Healthcare). Sample injection and buffer flow was controlled by a Hitachi L2130 pump, following the SEC column was a L-2400 UV detector (Hitachi), Optilab T-rEX refractometer (Wyatt technologies) and a DAWN HELEOS-II multi angle light scattering detector (Wyatt technologies). Prior to injection, columns and systems were equilibrated in 5 to 10 column volumes of running buffer 20 mM Tris-HCl pH 7.5, 150 mM NaCl and 5 mM β -ME. A 50 μ L injection was performed using a recombinant B/NP_{CORE} sample concentrated at 3.2 mg.mL⁻¹ with a constant flow rate of 0.5 mL.min⁻¹. Accurate MALLS mass prediction was performed with the Astra software (Wyatt Technologies). Curves were represented with Graphpad

(Prism). The SEC-MALLS chromatograms with the UV signal is shown as a backtrace and molecular weight (red) estimate below the peak.

Supplementary Figure 4

Rg determination by Guinier extrapolation. Guinier plots of the scattering curves are shown with a line of best fit and residuals for, (a) importin- α 7, (b) B/NP, (c) the B/NP_{TAIL}:importin- α 7 complex and (d) the B/NP:importin- α 7 complex. The slopes of fits are equal to $-\text{Rg}^2/3$, and the vertical intercept is equal to the log of the zero-angle scattering intensity I(0). The maximum q in the Guinier fits was determined for qRg values less than 1.3. All fits residuals (displayed in the bottom panel) are randomly distributed around zero.







b

а







С

B/NPTAIL:importin-α7





d

7