

1 Supplemental Materials

2 Title

3 The Irish DNA Atlas: Revealing Fine Scale Population Structure and History within Ireland

4 Authors

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54 **Supplemental Data 1 – Study Populations**

55 *Irish DNA Atlas*

56 The Irish DNA Atlas (henceforth the Atlas) is a DNA cohort of individuals with Irish ancestry.
57 To be included into the study, a participant must have all eight great-grandparents born within
58 50km, in Ireland. Therefore each Atlas individual is a sample of the genetics of specific regions in
59 Ireland three generations ago. The Atlas is a collaborative project between the Royal College of
60 Surgeons in Ireland, and the Genealogical Society of Ireland, and recruitment was primarily based
61 through genealogical interest groups. Saliva samples were collected using Oragene OG-250
62 (DNAGenotek, Canada) collection devices, and DNA extracted according to standard protocol.
63 Additionally each participant provided date of birth, and birth place information for all eight
64 grandparents. Samples were then genotyped on the Illumina OmniExpress chip at Edinburgh
65 Genomics, according to manufacturer’s instructions. Informed consent was obtained from all
66 individuals, and the data collection and analysis of these individuals was carried out in accordance
67 with the relevant guidelines and regulations was approved by the Royal College of Surgeons in
68 Ireland Research Committee, reference number REC0020563.

69 *Trinity Student Study Cohort*

70 This cohort consisted of 2232 students with Irish genetic ancestry recruited from Trinity
71 College Dublin [1]. The sample was included as additional samples of the Irish population. Genotype
72 information was generated using the Illumina 1M HumanOmni1-Quad chip.

73 *Peoples of the British Isles Cohort*

74 This cohort consisted of British individuals from the People of the British Isles (POBI)
75 Study[2]. Individuals are phenotyped as having ancestry from 35 geographic regions across the
76 United Kingdom. Genotype information from Illumina 1.2M platform was accessed via EBI, accession
77 number EGAD00010000632.

78 *WTCCC2 Project Multiple Sclerosis Cohort*

79 In order to provide European haplotypes, we included European individuals from the
80 WTCCC2 Multiple Sclerosis (MS) Study[3]. Genotype information from Illumina Human660-Quad chip
81 platform was accessed via the European Genome and Phenome Archive, accession number
82 EGAD00000000120.

83 **Supplemental Data 2 – Supplemental Methods**

84 **Supplemental Data 2.1 – The fineStructure Population Structure analysis.**

85 Using the final combined dataset of 2,103 individuals and 256,379 common markers we
86 phased the dataset using SHAPEIT v2.r790[4] using an effective population size of 11,418 as
87 suggested for European populations by the authors. We converted the resultant haplotype files to
88 ChromoPainter format using the “impute2chromopainter2.pl” script (downloaded at
89 <http://www.paintmychromosomes.com/>). For the phasing and conversion we used genetic map
90 build 37 downloaded with SHAPEIT.

91 Population structure analysis was performed by the combined software fineStructure (a pre-
92 release version, 2.1.0.pre)[5], which includes the software ChromoPainter, Chromocombine, and
93 fineStructure. Chromopainter was applied using default settings, with the exception of specifying the
94 number of ‘chunks’ per region to 50 as other analyses[10] have found that British and Irish
95 individuals share relatively longer haplotypes than average. We ‘painted’ each individual using every
96 other individual in the analysis as a donor using the -a 0 0 switch. Principal component analysis (PCA)
97 was performed on the resultant co-ancestry matrix. We then performed fineStructure clustering
98 MCMC analysis on the resultant co-ancestry matrix; with 1,000,000 burnin iterations, 1,000,000
99 sampling iterations, and retaining 500 MCMC samples. With the MCMC sample with the highest
100 posterior probability we performed 1,000,000 additional hill climbing moves to reach the final
101 inferred clustering and tree. When tree building we utilised the -T 1 parameter within fs-2.1.0.pre,
102 which uses the Maximum Concordance State method first reported by Leslie et al[6].

103 **Supplemental Data 2.2 – EEMS Analysis Pipeline**

104 Atlas individual latitude and longitude coordinates were generated from the average of
105 their eight great-grandparents’ birth places. The coordinates for the habitat boundaries were
106 generated with an online Google Maps API tool (<http://www.birdtheme.org/useful/v3tool.html>),
107 and the matrix of average pair-wise genetic dissimilarities was generated from plink format data
108 using the bed2diffs software included in the EEMS download package.

109 At the beginning of the analytical pipeline ten independent MCMC chains were started, each
110 with a random random-number-seed, for an initial 100,000 burnin and 100,000 sampling iterations
111 (thinning every 999 iterations), placing samples to the nearest of 600 demes. We chose the chain
112 with the highest final log-likelihood, and started 10 new EEMS chains, using this chain as a starting
113 point. This second round of chains were each started with a random random-number-seed, with
114 1,000,000 burnin iterations and 1,000,000 sampling iterations, sampling every 9,999 iteration –
115 placing samples to the nearest of 800 demes. We removed two chains (8 and 10) as these has

116 consistently lower log-likelihoods. We then checked whether their exclusion significantly changed
117 the predictive power of the EEMS model by noting the change in the r^2 value of the expected versus
118 fitted dissimilarities between demes (0.326 versus 0.325 without). We plotted the results of our
119 EEMS analysis using the R[7] package, “rEEMSplots”, which is included in the EEMS software
120 download. We used all final eight EEMS runs as input, and plotted the average estimated migration
121 and diversity surfaces, the posterior probability trace log for all eight chains, and a scatter plot of
122 observed vs fitted genetic dissimilarities within demes. The dissimilarity between observed versus
123 fitted deme pairs show a general trend with some deviation ($r^2 = 0.325$) (Supplementary Figure 5a)
124 and the log-posterior trace of the eight replicate MCMC chains (Supplementary Figure 5b) show
125 convergence of the independent EEMS runs.

126 **Supplemental Data 2.3 – The Ancestry Regression Method**

127 To investigate the genetic ancestry of any Irish clusters we observe we utilised a regression
128 based “ancestry profile” method first described by Leslie et al[6]. Briefly we estimated the
129 proportion of ancestry in each Irish and British individual that most closely resembles that
130 represented by different European, reference, individuals. These proportions can then be summed
131 across groups. We considered Y_p which is a vector of G length (where G is the number of European
132 reference populations), recording the average length of DNA genome-wide that each G -population
133 donates to Irish or British cluster P – which is then summed to unity across the vector. Additionally,
134 X_G is a vector of G -length that records the average length of DNA genome-wide that each European
135 individual copies from each European G population (with individuals unable to copy their own
136 haplotype to themselves). With,

$$137 \quad Y_p = \beta_1 X_1 + \beta_2 X_2 + \dots + \beta_G X_G,$$

138 we solve β_g assuming $\beta_g \geq 0$ and $\sum_{g=1}^G \beta_g = 1$, using an adaptation of the non-negative-least-squares
139 (nnls) function in R[7, 8]. Each inferred value of β_g is interpreted as the average proportion of
140 ancestry of genome-wide DNA each Irish or British individual from cluster P that is most closely
141 related, ancestrally, to each European cluster g .

142 To calculate confidence intervals we performed bootstrapping, resampling the
143 chromosomes of the Irish and British individuals and creating pseudo-individuals from the sampled
144 chromosomes. We recalculated our estimates of β_g from these pseudo-individuals to compute the
145 95% confidence intervals over 1000 bootstrap intervals.

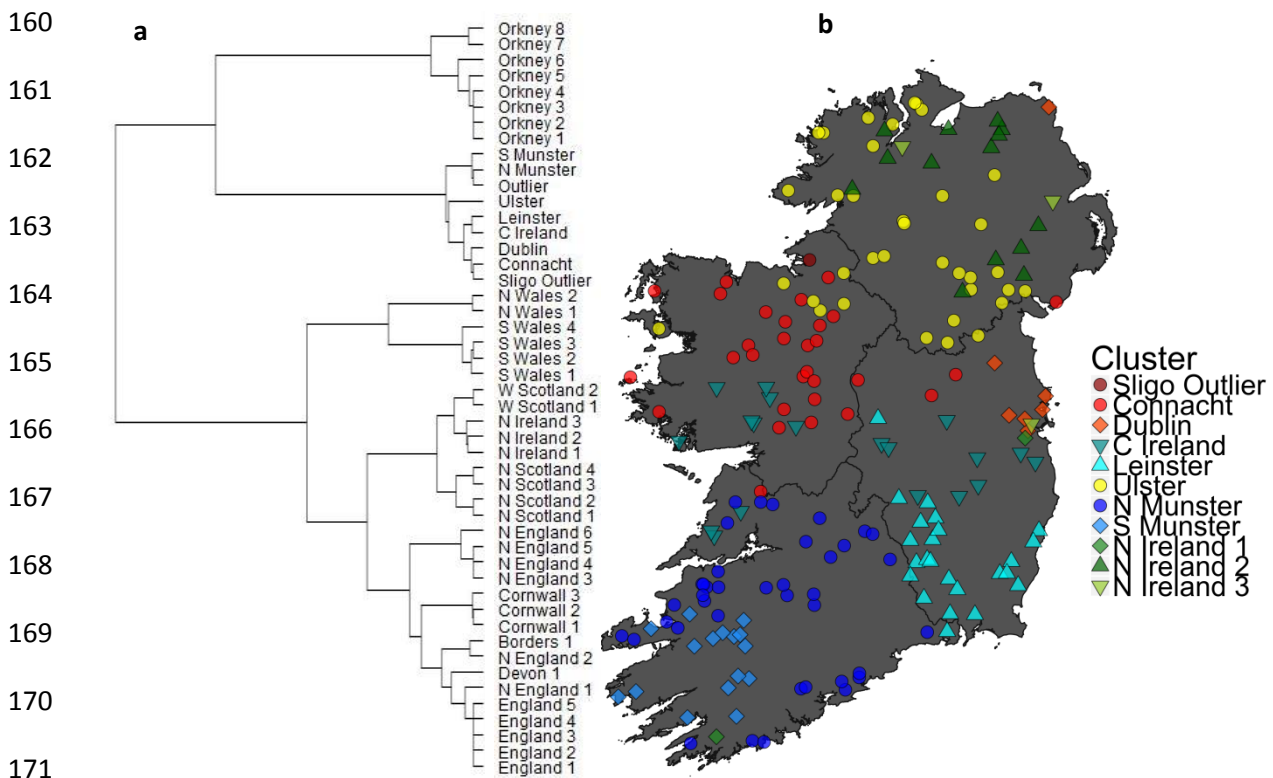
146 **Supplemental Data 3 - The Final Inferred Clustering of the Irish and British**
 147 **Datasets**

148 The final inferred state of fineStructure clusters in our analysis of Irish and British individuals
 149 (see Methods and Supplemental Data 2.1) yielded a total of 48 clusters, with many of these clusters
 150 numbers <10 individuals (n = 16). We investigated the clustering at each hierarchal value of *k*-
 151 clusters, paying attention to the cluster sizes and the degree of Irish substructure shown. We found
 152 that at *k* = 30, the 7 major clusters of predominantly Irish membership are separated (with two
 153 clusters of two Irish individuals each merged with the *Connacht* and the *N Munster* clusters
 154 respectively).

155 We show the geographic distribution of Atlas samples according to cluster membership
 156 (Figure S1A), the final inferred fineStructure tree at *k* = 48 (Figure S1B), and the membership of each
 157 *k* = 48 cluster in each *k* = 30 cluster with cluster size shown (Table S1).

158 **Supplementary Figure 1 - The Final Inferred fineStructure Clustering within Ireland**

159



172 **Supplementary Figure 1 - The final inferred fineStructure clustering state of 2,103 Irish and British**

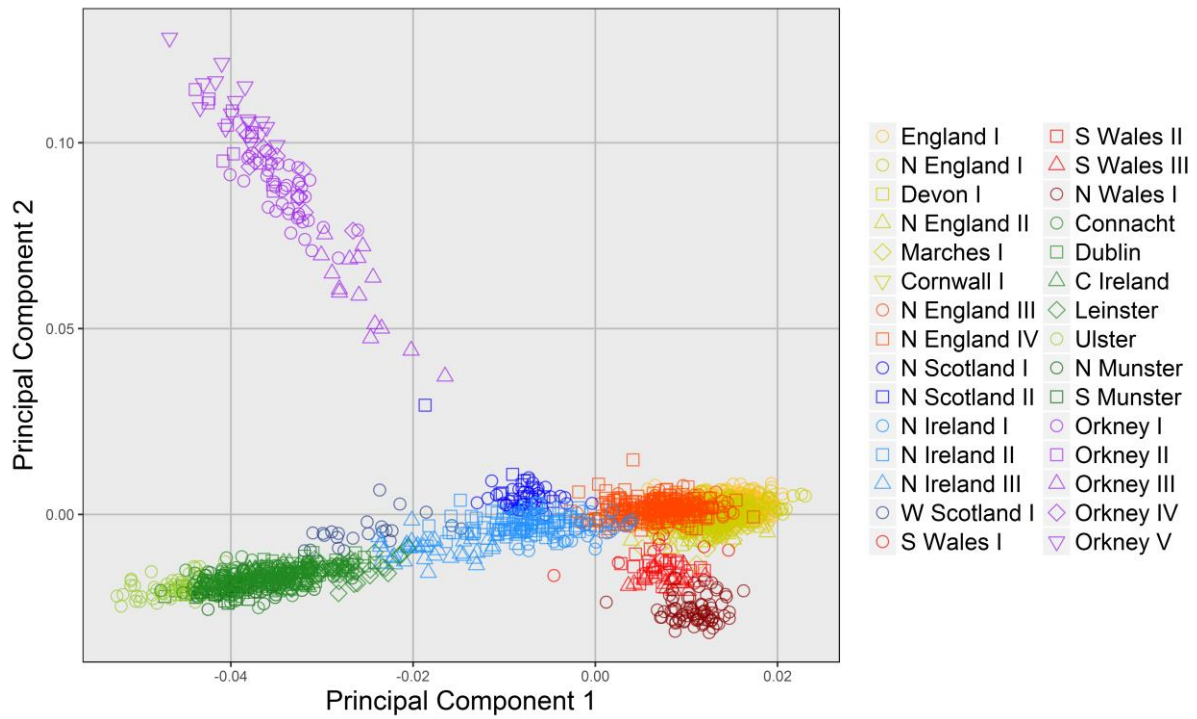
173 **individuals.** (a) The fineStructure dendrogram of the 48 final inferred clusters. (b) The geographic
 174 spread of the clusters containing Atlas Irish individuals, colour and shaped coded according to
 175 fineStructure cluster membership. Geographic location is the average of the Atlas individuals' great-
 176 grandparental birth places. Open Street Map Ireland, Copyright OpenStreetMap Contributors,
 177 (<https://www.openstreetmap.ie/>) - data available under the Open Database Licence. The figure was
 178 plotted in the statistical software language R, version 3.4.1[7], with various packages.

179
180

Supplementary Table 1 – Ireland Britain fineStructure Clustering Details			
The individual final inferred 48 clusters and which K30 cluster that cluster is a member of. Also shown are the individual sizes for each K48 and K30 cluster.			
K48 Cluster	K48 Cluster Size	K30 Cluster	K30 Cluster Size
England 1	1	England I	536
England 2	1	England I	
England 3	528	England I	
England 4	3	England I	
England 5	3	England I	
N England 1	82	N England I	82
Devon 1	73	Devon I	73
N England 2	32	N England II	32
Marches 1	78	Marches I	78
Cornwall 1	5	Cornwall I	82
Cornwall 2	13	Cornwall I	
Cornwall 3	64	Cornwall I	
N England 3	2	N England III	153
N England 4	149	N England III	
N England 5	2	N England III	
N England 6	149	N England IV	149
N Scotland 1	6	N Scotland I	39
N Scotland 2	33	N Scotland I	
N Scotland 3	7	N Scotland II	14
N Scotland 4	7	N Scotland II	
N Ireland 1	33	N Ireland I	33
N Ireland 2	94	N Ireland II	94
N Ireland 3	38	N Ireland III	38
W Scotland 1	7	W Scotland I	23
W Scotland 2	16	W Scotland I	23
S Wales 1	11	S Wales I	11
S Wales 2	3	S Wales II	26
S Wales 3	23	S Wales II	
S Wales 4	22	S Wales III	22
N Wales 1	63	N Wales I	75
N Wales 2	12	N Wales I	75
Sligo Outlier	2	Connacht	96
Connacht	94	Connacht	
Dublin	48	Dublin	48
C Ireland	77	C Ireland	77
Leinster	62	Leinster	62
Ulster	61	Ulster	61
Outlier	2	N Munster	76
N Munster	74	N Munster	
S Munster	28	S Munster	28
Orkney 1	2	Orkney I	38
Orkney 2	12	Orkney I	
Orkney 3	4	Orkney I	
Orkney 4	20	Orkney I	
Orkney 5	13	Orkney II	13
Orkney 6	15	Orkney III	15
Orkney 7	13	Orkney IV	13
Orkney 8	16	Orkney V	16

181 **Supplementary Figure 2 – Principal Component Analysis of the**
 182 **fineStructure Co-Ancestry Matrix**

183



184

185 **Supplementary Figure 2 - The Principal component analysis of the fineStructure co-ancestry**
 186 **matrix.** Shown are the first and second principal components, with individuals labelled according to
 187 $k = 30$ fineStructure cluster membership.

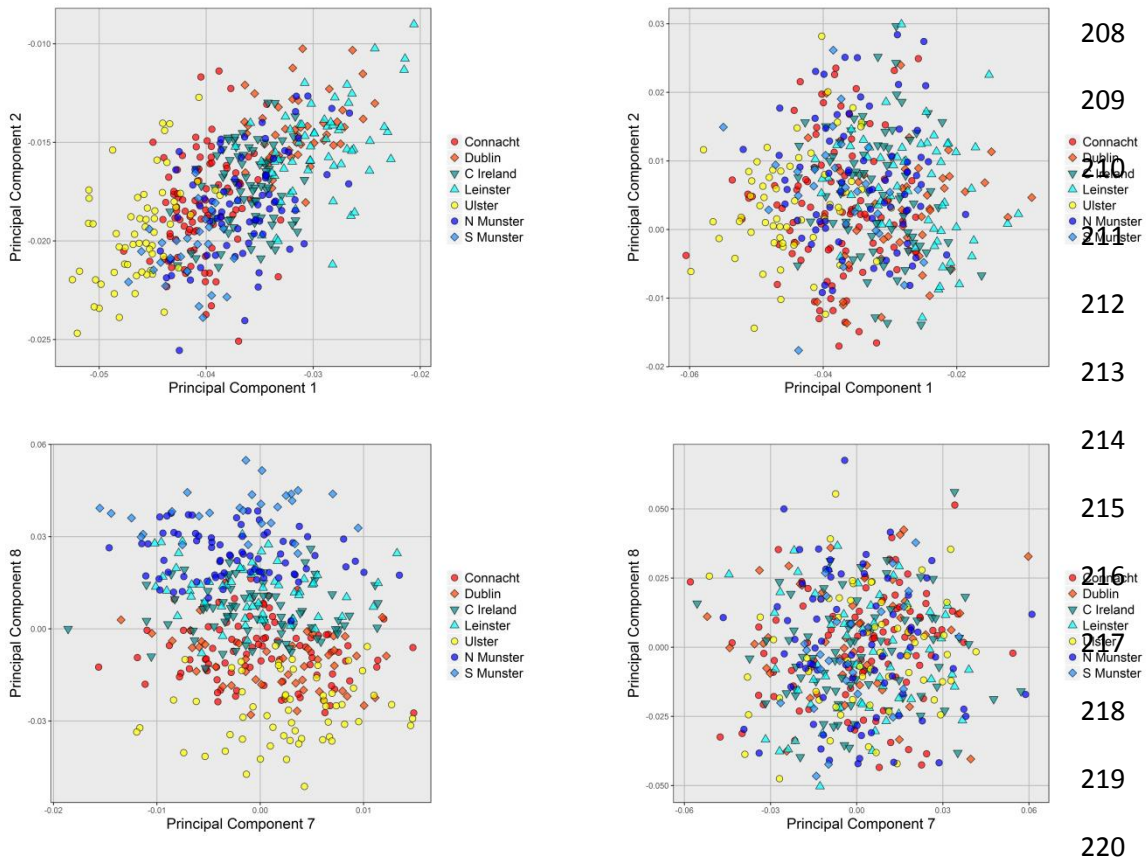
188 **Supplemental Data 4 – Comparison of fineStructure PCA to comparable**
189 **methods**

190 To compare fineStructure’s ability to differentiate population structure to more
191 conventional methods we performed principal component analysis (PCA) of the Irish and British
192 dataset used in our analysis of Population Structure within Ireland using gcta64[9], and compared
193 this to the PCA of the fineStructure co-ancestry matrix. We generated principal components from
194 the co-ancestry matrix using methods previously described in Supplemental Data 2.1. In order to
195 generate the principal components using gcta64, we used plink 1.9[10, 11] formatted data of the
196 2,103 Irish and British individuals and the 256,379 common markers. We first pruned the dataset of
197 SNPs with the plink command --indep-pairwise 1000 50 0.2. Then, using a pruned dataset of 79,417
198 common markers, we generated a genetic relationship matrix (grm) with gcta64, and finally
199 generated the top 10 principal components from this matrix using gcta64’s “--pca” function.

200 As previously reported[5, 6], the haplotype-based fineStructure shows a greater ability to
201 differentiate population structure than more conventional allele frequency based methods such as
202 gcta64. This is demonstrated at the higher components as well as lower principal components
203 (shown are principal components 7 and 8 in Figure S3). fineStructure’s ability to differentiate
204 population structure at lower components (where gcta64 is not able to detect structure) presumably
205 reflective of its ability to detect the fine scale structure that we observe within Ireland.

206
207

Supplementary Figure 3 – Comparison of fineStructure PCA and GCTA64 PCA



221 **Supplementary Figure 3 – Comparison of fineStructure-based principal component analysis (PCA)**
222 **and conventional PCA methods.** Shown are all individuals included in the Irish and British
223 fineStructure analysis that were placed in clusters on the Irish branch at $k = 30$. We show, with four
224 panels, their PCA coordinates along PC principal components 1 and 2 (upper two) and principal
225 components 7 and 8 (lower two) using fineStructure (left two) and gcta64 (right two).

226 **Supplemental Data 5 – Comparison to Ancient Irish Genomes**

227 We decided to compare the Atlas Irish individuals in our sample to two previously
228 published[12] high coverage ancient Irish genomes; a Neolithic farmer (Ballynahatty) and a Bronze
229 Age individual (Rathlin1). The authors of the aforementioned authors found the greatest affinity to
230 the modern Irish was found in the Bronze Age individual studied. We set out to investigate whether
231 any particular region in Ireland as represented in our Atlas Irish individuals and Irish fineStructure
232 clusters shared an affinity to either of the ancient Irish individuals.

233 We found the intersect of common shared SNPs between the individuals included in the
234 fineStructure analysis of Population Structure within Ireland (see methods for more detail)
235 individually for each ancient Irish individual (see Table S3 for SNP overlaps).

236

Supplementary Table 3 – The overlap of common SNPs between two published ancient Irish individuals and a dataset of Irish and British individuals		
Ancient Irish	Ballynahatty	Rathlin1
SNP Overlap	162,069	175,749

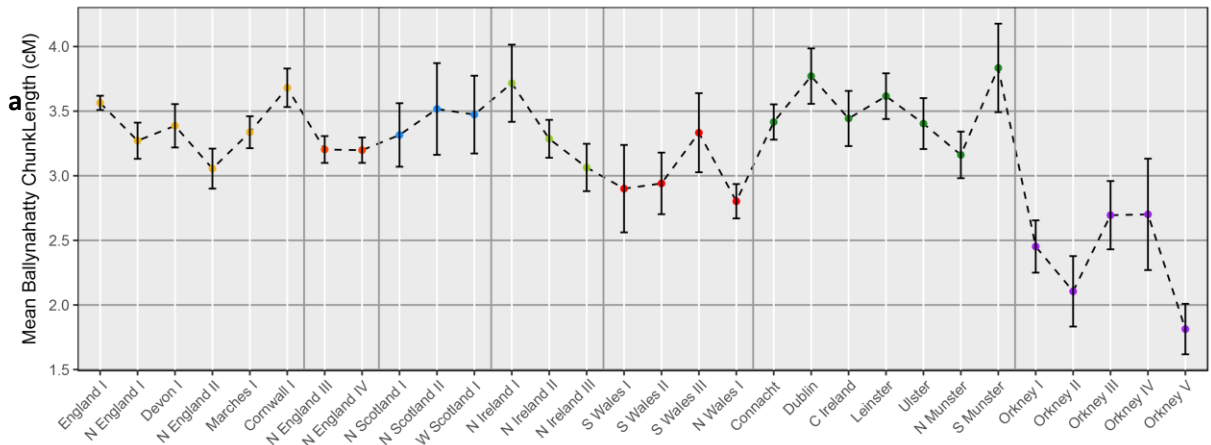
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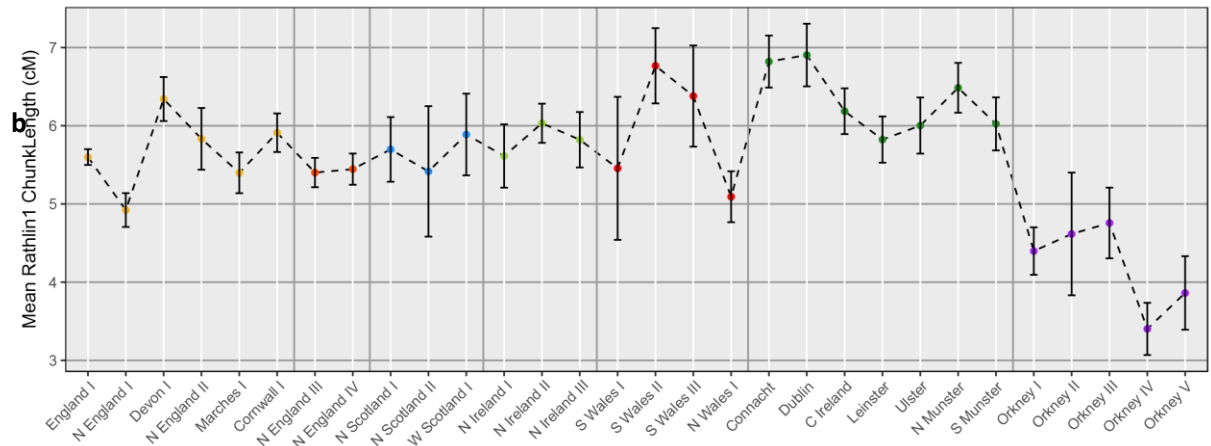
239

240 We merged the Irish and British dataset individually with each ancient individual, and as
241 each ancient individuals was of high coverage (>10x), we phased and performed fineStructure
242 analysis separately. We phased each of the datasets using SHAPEIT[4], using the same method
243 outlined by Cassidy et al[12]. We painted each individual donating haplotypes to every other
244 individual in the analysis (the “-a” switch). We report the average haplotypic donation from each
245 ancient Irish genome to each modern Irish or British cluster in cM. We calculated the standard error
246 for each Irish or British cluster from the standard error of the individual sample lengths within each
247 cluster.

248 **Supplementary Figure 4 – Haplotypic affinity of Irish and British clusters to two ancient**
 249 **Irish genomes**



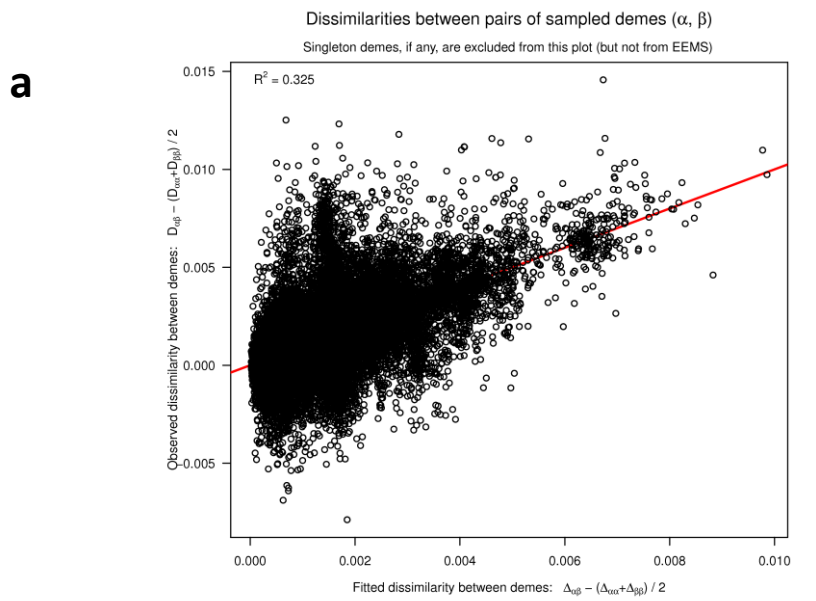
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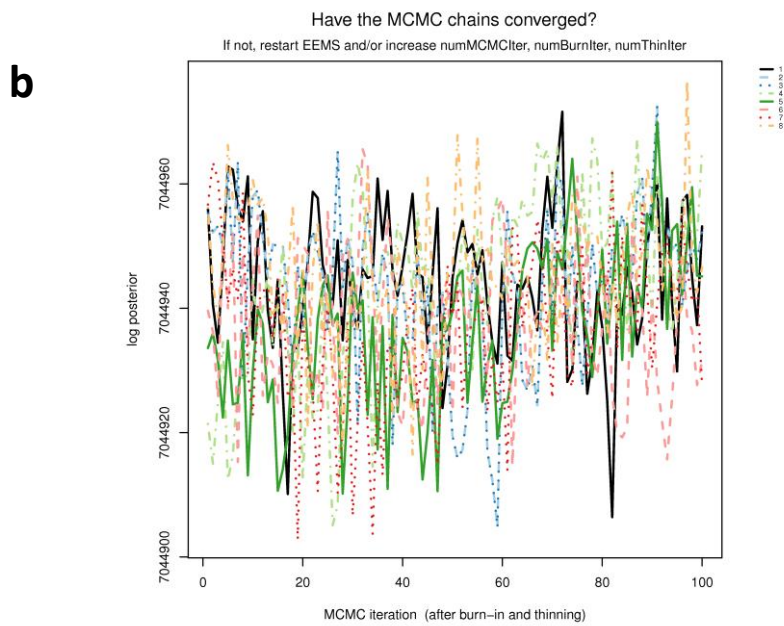
251

252 **Supplementary Figure 4 – Haplotypic Affinity of each Ancient Irish genome to each modern Irish**
 253 **and British cluster.** Shown is the average length of haplotype donation from each ancient individual,
 254 Ballynahatty (a) and Rathlin (b), to each $k = 30$ Irish and British cluster in cM. Also shown are the
 255 standard error bars calculated from the standard error of the sample lengths of the individuals
 256 within each of the individual clusters.

257 **Supplementary Figure 5 – Estimated Effective Migration Surface Diagnostic**
258 **Plots**



259



260

261 **Supplementary Figure 5 – Estimated Effective Migration Surface Diagnostic plots.** (a) The observed
262 versus expect dissimilarity between pairs of demes (with demes with only one individual not
263 considered). Strong deviations from the fitted line (red) indicate pairs of demes much more
264 genetically distant than expected. (b) The posterior probability log of the six replicates of the EEMS
265 run, indicating whether the MCMC chains have converged.

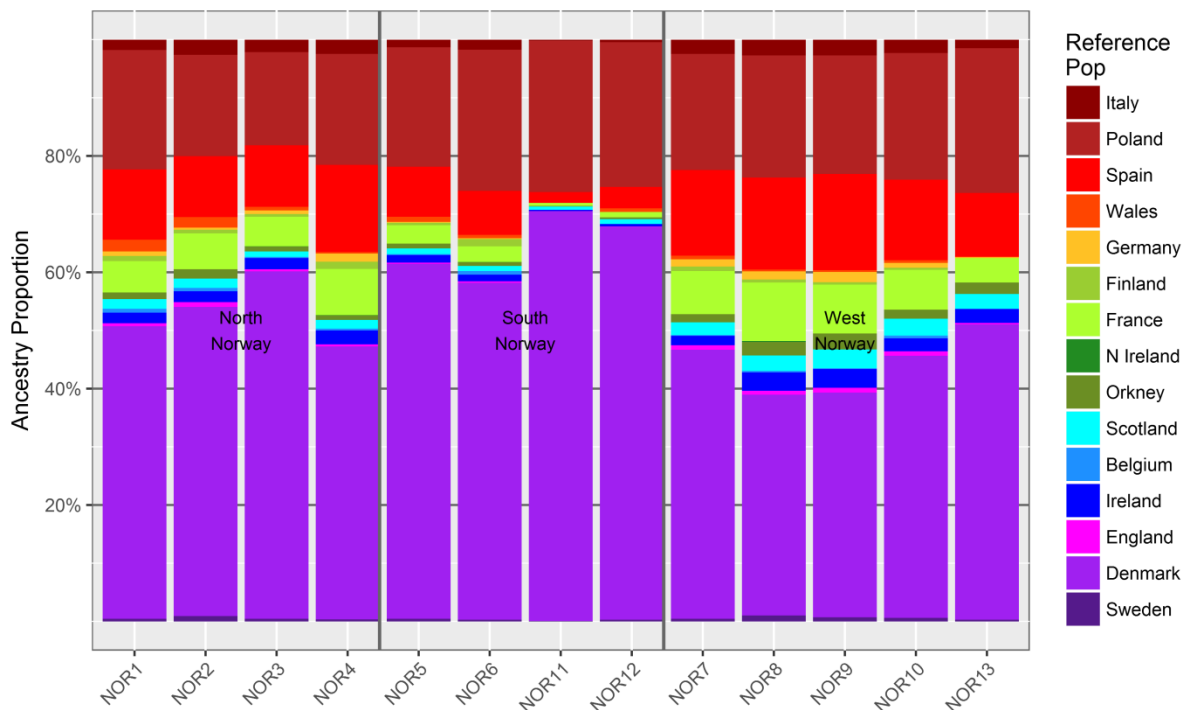
266

267 **Supplemental Data 6 – Checking Ancestry Proportions**

268 Our regression based ancestry profile analysis of Irish and British haplotypes revealed a
269 surprising amount of Norwegian-like ancestry in our Irish samples. To investigate whether this was
270 due wholly or in part to Irish haplotypes existing in our modern Norwegian sample we performed an
271 additional regression based ancestry analysis. We modelled the Norwegian $k = 51$ clusters as a
272 mixture of the other $k = 51$ European clusters, as well as the $k = 30$ Irish and British clusters using the
273 same methodology as described in the Materials and Methods, as well as Supplementary Data 2.1.
274 We present the data as the average reference country contribution to each Norwegian cluster,
275 where Norwegian clusters are organised by region of origin within Norway.

276 **Supplementary Figure 6 – Ancestry profiles of 12 Norwegian populations**

277



278 **Supplementary Figure 6 – Ancestry profiles of 13 Norwegian populations modelled as a mixture of**
279 **Irish and British, and mainland European populations.** Shown are the average total ancestry
280 contributions of all reference fineStructure clusters where the majority of individuals originate from
281 each country, in each Norwegian cluster. Norwegian clusters are organised in three groups based on
282 where the majority of individuals are from in Norway in each cluster.

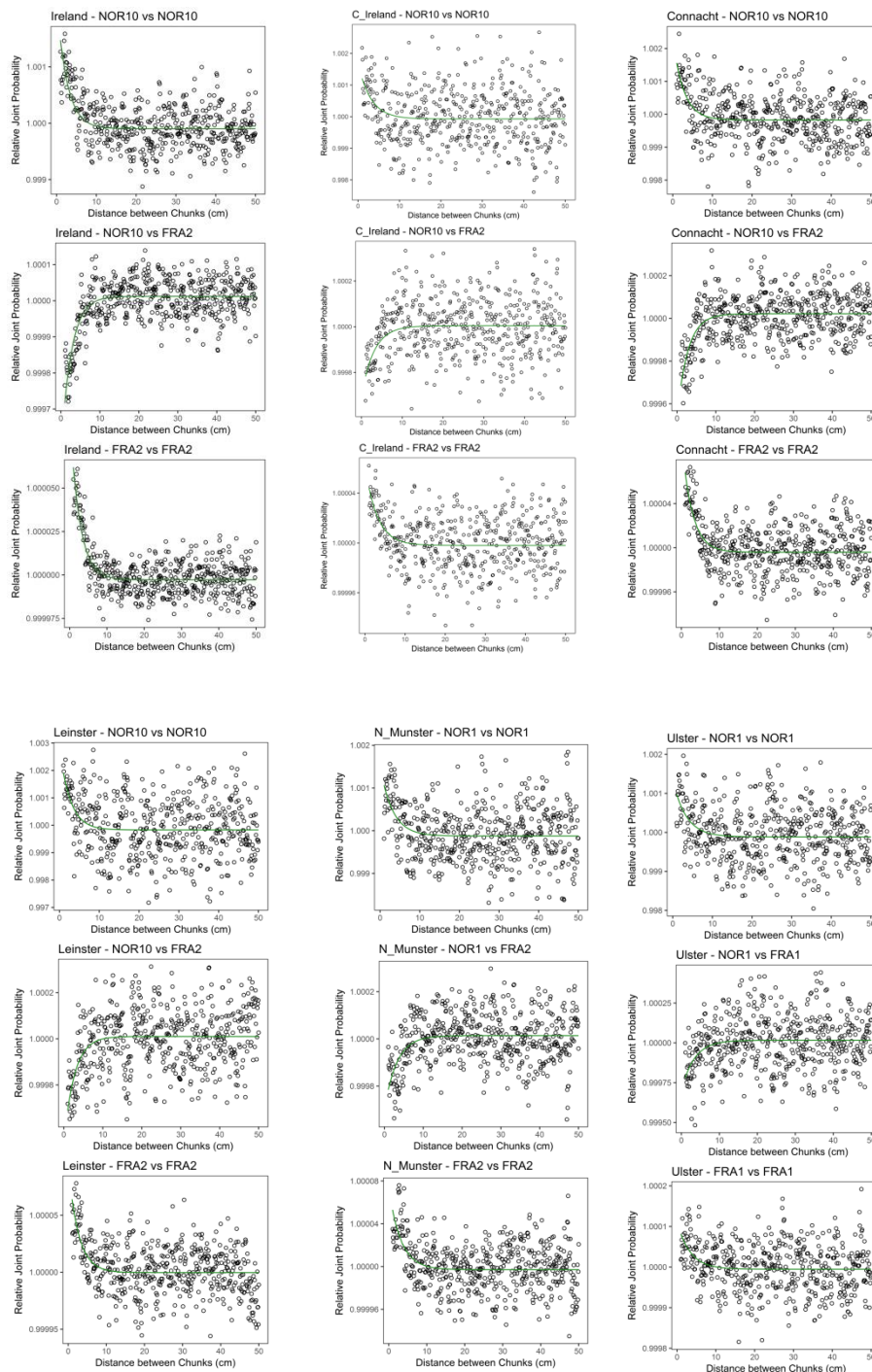
283 **Supplementary Data 7 – GLOBETROTTER Joint Probability Curves**

284 To investigate the evidence of admixture events within Ireland we performed
285 Globetrotter[13] analysis on a number of different fineStructure identified clusters of Irish
286 membership. Firstly, we investigated the evidence of European admixture into Ireland. We present
287 the joint probability plot for each Irish cluster, showing the primary Norwegian cluster versus the
288 main component on the putative Gaelic source Figure S7.1. Secondly we investigated evidence that
289 the three clusters of joint Irish and British membership were the result of admixture event(s)
290 between Irish and British clusters, the results of which are shown in Table S8. We also present the
291 joint probability curves for the major surrogate components for each source population, or each *N*
292 *Ireland* cluster analysis (see below, Figures S7.2-4).

293 Each joint probability curve shows the probability that two positions, separated by genetic
294 distance x , corresponds to ancestry donated by population A and population B, where populations A
295 and B can be the same population. These can be used to assess the strength of the admixture signal
296 detected by Globetrotter, the cleaner the signal around the fitted curve and the steeper the curve,
297 the stronger the admixture signal. When population A=A or B=B and the fitted curve is negative, and
298 the fitted curve when A as compared to B is positive it is indicative of admixture.

299
300

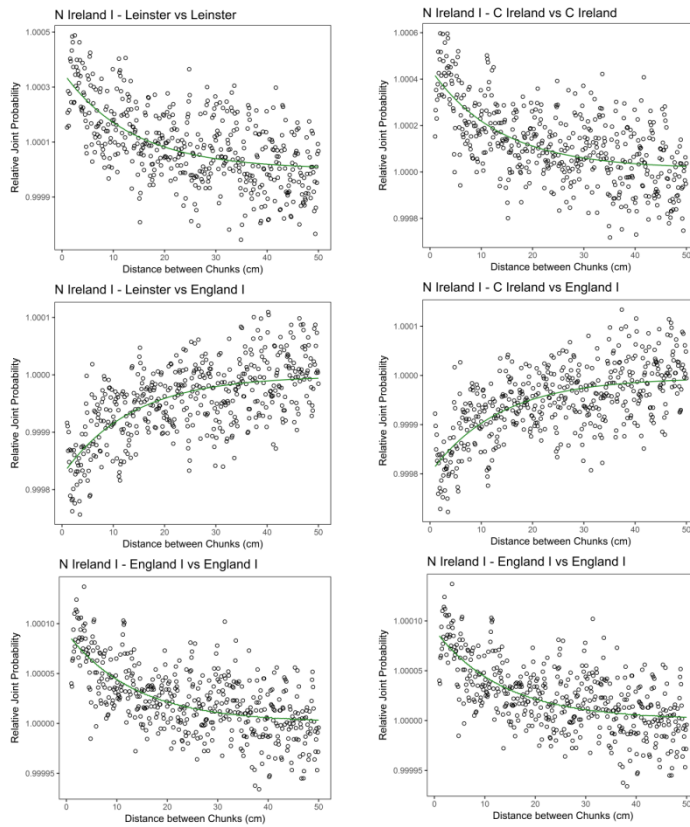
Supplementary Figure 7.1 – Joint Probability Curves of Irish clusters comparing Irish signal to primary Norwegian signal



301

302 **Supplementary Figure 7.1 – The admixture probability curves of the major components of the**
303 **Gaelic Irish admixture events.** The joint probability curves of the 6 Gaelic Irish clusters that show
304 evidence of a significant admixture signal; (top left to bottom right), *Ireland*, *C Ireland*, *Connacht*,
305 *Leinster*, *N Munster*, and *Ulster*. The scaled probability data is shown in black, with the fitted curve
306 shown in green.

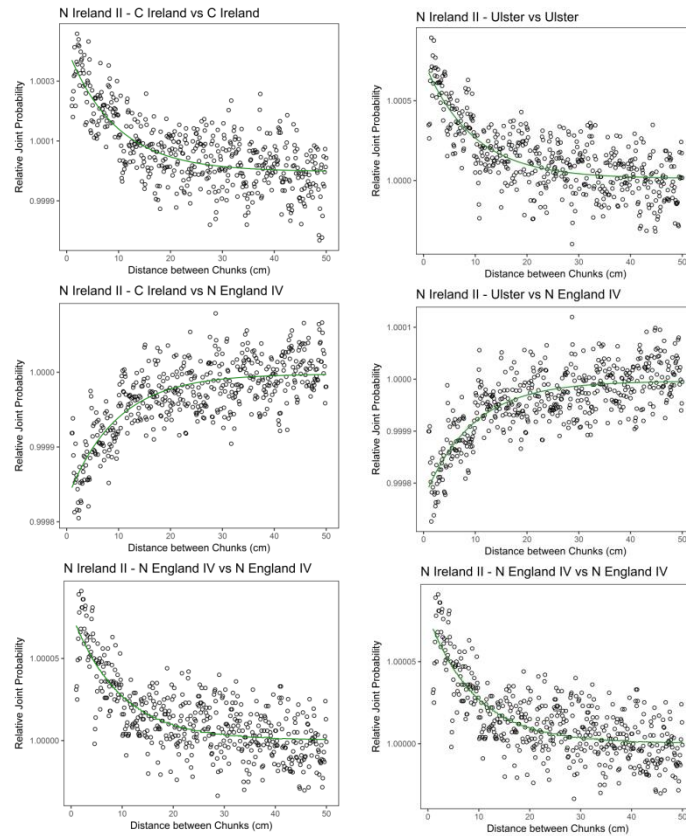
307 **Supplementary Figure 7.2 – Joint Probability Curves of *N Ireland I***



308

309 **Supplementary Figure 7.2 – The admixture probability curves of the major components of the *N***
 310 ***Ireland I* admixture event.** (left) The three sets of probability curves showing the primary British
 311 (*England I*) and Irish (*Leinster*) components of the admixture event, and the probability that two sites
 312 separated by genetic distance x are donated from population A and B. (right) The same as the left
 313 panels, with the secondary Irish component, *C Ireland*. For both the scaled probability data is shown
 314 in black, with the fitted curve shown in green.

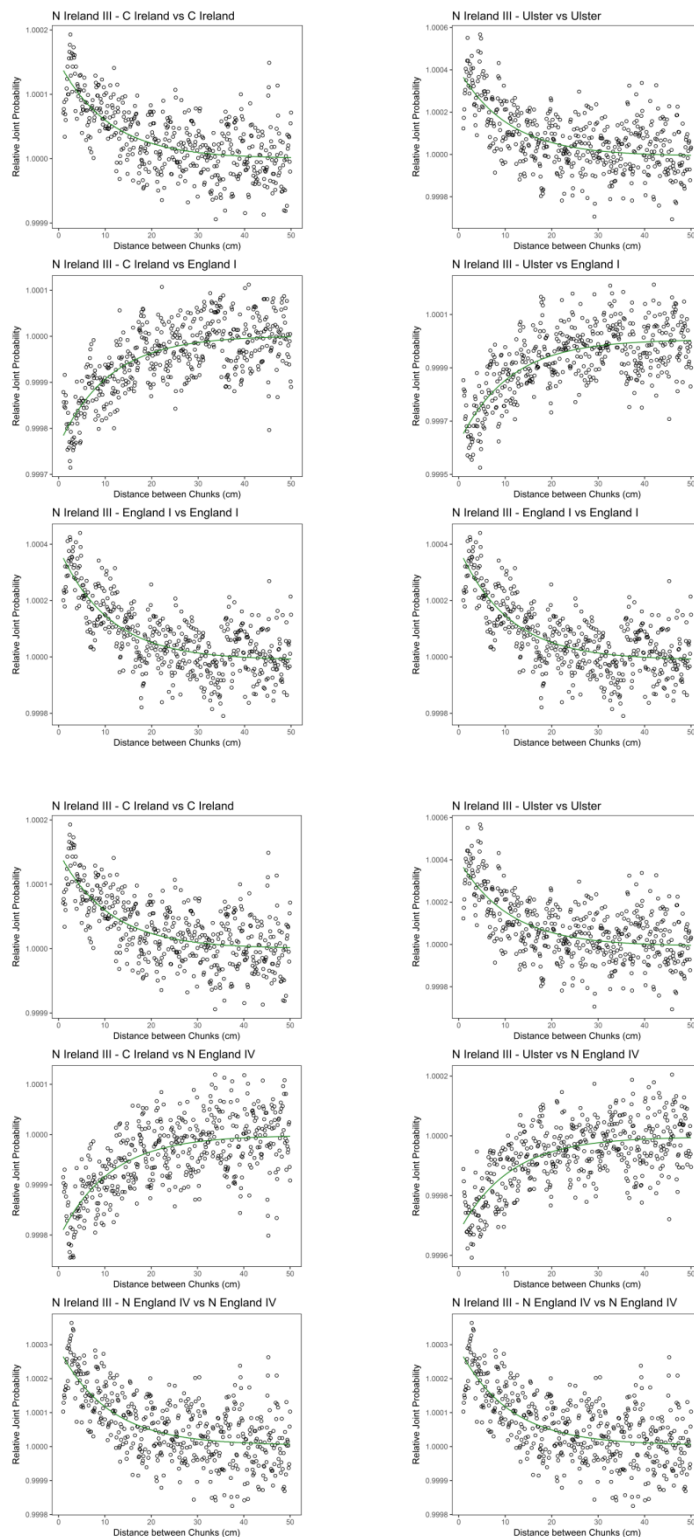
315 **Supplementary Figure 7.3 – Joint Probability Curves of *N Ireland II***



316

317 **Supplementary Figure 7.3 – The admixture probability curves of the major components of the *N***
 318 ***Ireland II* admixture event.** (left) The three sets of probability curves showing the primary British (*N*
 319 *England IV*) and Irish (*Leinster*) components of the admixture event, and the probability that two
 320 sites separated by genetic distance x are donated from population A and B. (right) The same as the
 321 left panels, with the secondary Irish component, *C Ireland*. For both the scaled probability data is
 322 shown in black, with the fitted curve shown in green.

323 **Supplementary Figure 7.4 – Joint Probability Curves of *N Ireland III***



324

325 **Supplementary Figure 7.4 – The admixture probability curves of the major components of the *N***
 326 ***Ireland III* admixture event. (top-left) The three sets of probability curves showing the primary**
 327 **British (*England I*) and Irish (*C Ireland*) components of the admixture event, and the probability that**
 328 **two sites separated by genetic distance x are donated from population A and B. (top-right) The same**

329 as the left panels, with the secondary Irish component, *Ulster*. (bottom-left) As top-left, with the
330 secondary British component, *N England IV*, and the primary Irish component, *C Ireland*. (bottom-
331 right) As top-left, but with the secondary British component, *N England IV*, and the secondary Irish
332 component, *Ulster*. For both the scaled probability data is shown in black, with the fitted curve
333 shown in green.

334 **References**

335 1. Desch KC, et al., *Linkage analysis identifies a locus for plasma von Willebrand factor*
336 *undetected by genome-wide association*. Proc Natl Acad Sci USA, 2013. **110**(2): p. 588-93.

337 2. Winney B, et al., *People of the British Isles: preliminary analysis of genotypes and surnames*
338 *in a UK-control population*. Eur J Hum Genet, 2012. **20**(2): p. 203-10.

339 3. IMSSGC WTCCC2, *Genetic risk and a primary role for cell-mediated immune mechanisms in*
340 *multiple sclerosis*. Nature, 2011. **476**(7359): p. 214-219.

341 4. Delaneau O, Marchini J, and Zagury JF, *A linear complexity phasing method for thousands of*
342 *genomes*. Nature Methods, 2011. **9**(2): p. 179-81.

343 5. Lawson DJ, et al., *Inference of population structure using dense haplotype data*. PLoS Genet,
344 2012. **8**(1): p. e1002453.

345 6. Leslie S, et al., *The fine-scale population structure of the British population*. Nature, 2015.
346 **519**: p. 309-14.

347 7. R Core Team, *R: A Language and Environment for Statistical Computing*. 2017, R Foundation
348 for Statistical Computing: Vienna, Austria.

349 8. Lawson CL and Hanson RJ, *Solve Least Squares Problems*. 1995: Reprinted by the Society for
350 Industrial and Applied Mathematics.

351 9. Yang J, et al., *GCTA: a tool for genome-wide complex trait analysis*. Am J Hum Genet, 2011.
352 **88**(1): p. 76-82.

353 10. Purcell S, et al., *PLINK: a tool set for whole-genome association and population-based linkage*
354 *analyses*. Am J Hum Genet, 2007. **81**(3): p. 559-75.

355 11. Chang CC, et al., *Second-generation PLINK: rising to the challenge of larger and richer*
356 *datasets*. Gigascience, 2015. **4**: p. 7.

357 12. Cassidy LM, et al., *Neolithic and Bronze Age migration to Ireland and establishment of the*
358 *insular Atlantic genome*. Proc Natl Acad Sci USA., 2016. **113**(2): p. 368-73.

359 13. Hellenthal G, et al., *A genetic atlas of human admixture history*. Science, 2014. **343**(6172): p.
360 747-51.

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