

Supplementary Figure 1: C9RAN translation reporter system allows for quantitative assessment of RAN translation in all three sense reading frames.

(a) Anti-FLAG western blot analysis of control and GA C9RAN translation reporters in HEK293 cells. #One-tenth AUG reporter was transfected into cells to prevent over-exposure. (b) Expression of control and C9RAN reporters in mRNA transfected HEK293 cells, n=6. (c) Representative expression of control and C9RAN reporter mRNAs in HeLa cell lysate, n=3. (d) The stability of control and C9RAN reporter proteins was assessed in transfected HEK293 cells by treating cells with 10 µg/mL puromycin and measuring reporter activity at 0, 6, and 24 hours later, n=9 (0 and 6 hours), n=12 (24 hours). (e)The hindrance of the DPR fusion on NLuc activity was assessed in RRL by incubating completed reactions with PSP enzyme to cleave DPRs from NLuc, n = 6-9. (f) Anti-FLAG Western blot to confirm PSP cleavage of C9RAN fusion proteins in RRL. (g) Expression from AUG-driven reporters for each sense reading frame, relative to AUG-GA70, in RRL, n=6 and (h) HEK293 cells, n=6. RRL, rabbit reticulocyte lysate. Graphs in (b) and (c) represent mean ± SD. Remaining graphs show mean ± SEM. Two-tailed Student's t test with Welch's correction, **p < 0.01; ***p < 0.001; *****p<0.0001.



Supplementary Figure 2: C9RAN translation in HEK293 cells is cap-dependent and can initiate at a near-cognate start codon.

(a) Expression of m⁷G-capped and A-capped control and C9RAN mRNA reporters in HEK293 cells, n=9. (b) Expression of all three sense C9RAN mRNAs in HEK293 cells following mutation of near-cognate codons in GA frame, n=6. (c) Expression of sense C9RAN mRNAs in HeLa cell lysate with or without CUG codon mutated to CCC, n=11. (d) Expression of GP and GR-NLuc reporters in RRL from constructs with CUG codon mutated to AUG, relative to WT sequence in RRL, n=6. (e) Insertion of an AUG codon upstream of the repeat in the GA frame enhances GA-NLuc expression in RRL, n=6. (f) Mutating CUG codon to AUG decreases expression of GP and GR-NLuc reporters in HEK293 cells, n=6. (g) Insertion of an AUG codon upstream of the repeat of the repeat in the GA frame enhances GA-NLuc reporters in HEK293 cells, n=24. Graphs represent mean \pm SEM. Two-tailed Student's t test with Welch's correction, *p < 0.05; ***p < 0.001; ****p<0.0001.



Supplementary Figure 3: CGG RAN translation in multiple reading frames is refractory to translation attenuation during ER and oxidative stress.

(a) Anti-puromycin western blot of cells treated with various cell stress inducers and 10 µg/mL puromycin to monitor global translation activity. Tubulin, GAPDH, and Coomassie stain were used as loading controls. Unt, untreated. Veh, vehicle. TG, 2µM thapsigargin. SA, 10µM sodium arsenite. Sal003, 20µM Salubrinol. CHX, 100µg/mL cycloheximide. (b) Destabilization of AUG-NLuc reporter with PEST tag results in greater decrease in AUG-NLuc expression with TG treatment. (c-d) Expression of control and CGG RAN NLuc reporters in transfected HEK293T cells when treated with (c) 2.5 µg/mL Tunicamycin, n=9 and (d) 20 µM sodium arsenite, n=9, for 5 hours. FLuc was coexpressed as an internal control with each NLuc reporter. AUG and ATF4 reporters serve as reporters that are attenuated and stimulated, respectively, during activation of the ISR. Graphs represent mean \pm SEM. Two-tailed Student's t test with Welch's correction, ***p < 0.001; ****p<0.0001.



Supplementary Figure 4: Thapsigargin-induced enhancement of C9 and CGG RAN translation requires phosphorylated $elF2\alpha$

(a) Western blot showing increased phosphorylation of eIF2 α in HEK293 cells following treatment with Sal003 (20 or 40 μ M), TM (1 or 2.5 μ g/mL), and SA (20 μ M). (b) Expression of control, CGG, and C9RAN NLuc reporters normalized to co-transfected FLuc in WT and eIF2 α -S51A/A homozygous mutant MEFs following treatment with 1 μ M thapsigargin (TG), n=6-9. Graph represent mean ± SEM. Two-tailed Student's t test with Bonferroni and Welch's correction, **p < 0.01.





NLuc RLU (relative to GGG) 10000-

1000

100

10

1-





elF2α-S51D





**

**





Supplementary Figure 5: Initiation at near cognate codons is refractory to multiple stress

stimuli. (a) Expression of the control AUG-NLuc and AUG-initiated reporters harboring 100 CGG or 70 G₄C₂ repeats in multiple reading frames in HEK293 cells when treated with 20uM Sal003, for 5 hours, n=6-12. (b) Expression of NLuc reporters with varying start codon mutations in HEK293T cells relative to the negative control GGG-NLuc, n=9. (c-d) Response of the AUG-NLuc and near cognate-NLuc reporters, co-transfected with the internal FLuc control, in HEK293T cells (c) co-transfected with either WT or S51D (phosphomimetic) eIF2 α , n=12-15, or treated with (d) 2.5 µg/ml tunicamycin, n=9, or (e) 20 µM sodium arsenite, n=9, for 5 hours. Graphs represent mean ± SEM. Two-tailed Student's t test with Welch's correction, **p < 0.01; ***p < 0.001, ****p<0.0001.



b

MEFs - Stress Granules



С

MEFs - G3BP+ Stress Granules



а

Supplementary Figure 6: CGG and G₄C₂ repeats induce G3BP-positve stress granules in a phosphorylated-elF2α dependent manner.

(a) *Left*: Immunofluorescent images of HEK293 cells expressing control, $(G_4C_2)x70$, or CGGx100 reporters, scale bar=100 µm. *Right:* Quantification of the proportion of FLAG-positive cells with G3BP-positive stress granules (SGs) for each genotype, n>45. (b) Immunofluorescent images of WT and eIF2 α - S51A/A MEFs treated with vehicle or 10 µM TG for 3 hours, scale bar=100 µm. (c) *Left:* Immunofluorescent images of WT and eIF2 α -S51A/A MEFs expressing control, +1CGGx100, or +2CGGx100 RAN reporters, scale bar=100 µm. *Right:* Quantification of the proportion of FLAG-positive cells with G3BP-positive SGs for each genotype, n>40. FLAG marks reporter expressing cells, G3BP mark SGs. Fisher's exact test, ***p < 0.001.; ****p<0.0001.



Supplementary Figure 7: Full-length western blots of key experimental findings From (a) Fig. 1b (b) Fig. 3b and (c) Fig. 3f.

Supplementary Table 1: Primers for reporter generation

Reporter	Forward Primer Sequence	Reverse Primer Sequence	
PSP-GGG- NL-3xFLAG	CCGGTCTCGAGGTCCTCTTCCAGGGACCCA	CCGGTGGGTCCCTGGAAGAGGACCTCGAGA	
GA frame PSP site	CCAGGGACCCGATGGGGTCTTCAC	AAGAGGACCTCGAGACCG	
GP frame PSP site	CCTCGAGGTCCTCTTCCAGGGACCCGATGG	ACCOCTOCCCCCCCCC	
GR frame PSP site	CCCTCGAGGTCCTCTTCCAGGACCCGATGG	ACCGGTGGGCGCGCCCGG	
Intron1 CTG- CCC	GTAGCAAGCTCCCGAACTCAGGAGTCGC	AGGCTGCGGTTGTTTCCC	
Intron1 AGG- AAA	TCTGGAACCAAAAGTCGCGCGC	GCTTGCTACAGGCTGCGG	
Intron1 CTG- CCC and AGG-AAA	CTAAAAGTCGCGCGCTAGCGGCC	TTCGGGAGCTTGCTACAGGCTGCGTTG	
Intron CTG- ATG	GTAGCAAGCTATGGAACTCAGG	AGGCTGCGGTTGTTTCCC	
Intron1 AUG	ATGAGTCGCGCGCTATCTA	CCTGAGTTCCAGAGCTTG	
AUG-GA	CTAGCTAACTAACACCATGGC	GGCCGCCATGGTGTTAGTTAG	
AUG-GP	CTAGCTAACTAACACCATGGGGC	GGCCGCCCCATGGTGTTAGTTAG	
AUG-GR	CTAGCTAACTAACACCATGGGC	GGCCGCCCATGGTGTTAGTTAG	
CUG-NLuc	ACCCTGGTCTTCACACTCGAAGATTTC		
GUG-NLuc	ACCGTGGTCTTCACACTCGAAGATTTC	GGUTTATTTACCAACAGTACCGGATTG	
ACG-NLuc	ACCACGGTCTTCACACTCGAAGATTTC		

Supplementary Table 2: C9RAN construct sequences

Construct	Sequence
Nhe1-Intron1-GA70-PSP-	GCTAGCGTGTGTGTTTTTGTTTTTCCCACCCTCTCTCCCCACTACTTGCTCTCACAGTACTCG
GGG-NLuc-3xFLAG-PspOMI	CTGAGGGTGAACAAGAAAAGACCTGATAAAGATTAACCAGAAGAAAACAAGGAGGGAAACAA
	CCGCAGCCTGTAGCAAGCTCTGGAACTCAGGAGTCGCGCGCTAGCGGCCGGGGCCGGGGC
	CGGGGCCGGGGCCGGGGCCGGGGCCGGGGCCGGGGCCGGGG
	GGCCGGGGCAGGGGCCGGGGCCGGGGCCGGGGCCGGGGCCGGGGCCGGGG
	CGGGGCCGGGGCCGGGGCCGGGGCCGGGGCCGGGGCCGGGG
	GGCCGGGGCCGGGGCCGGGGCCGGGGCCGGGGCCGGGGCCGGGG
	CGGGGCCGGGGCCGGGGCCGGGGCCGGGGCCGGGGCCGGGG
	GGCCGGGGCCGGGGCCGGGGCCGGGGCCGGGGCCGGGGCCGGGG
	CGGGGCCGGGGCCGGGGCCGGGGCCGGGGCCGGGGCCGGGG
	GGCCGGGGCCGGTCGTGGAAGGGTGGGCGCGCCCCACCGGTCTCGAGGTCCTCTTCCAGGG
	ACCCGATGGGGTCTTCACACTCGAAGATTTCGTTGGGGACTGGCGACAGACA
	AACCTGGACCAAGTCCTTGAACAGGGAGGTGTGTCCAGTTTGTTT
	CGTAACTCCGATCCAAAGGATTGTCCTGAGCGGTGAAAATGGGCTGAAGATCGACATCCATG
	TCATCATCCCGTATGAAGGTCTGAGCGGCGACCAAATGGGCCAGATCGAAAAAATTTTTAAG
	GTGGTGTACCCTGTGGATGATCATCACTTTAAGGTGATCCTGCACTATGGCACACTGGTAATC
	GACGGGGTTACGCCGAACATGATCGACTATTTCGGACGGCCcTATGAAGGCATCGCCGTGTT
	CGACGGCAAAAAGATCACTGTAACAGGGACCCTGTGGAACGGCAACAAAATTATCGACGAGC
	GCCTGATCAACCCCGACGGCTCCCTGCTGTTCCGAGTAACCATCAACGGAGTGACCGGCTG
	GCGGCTGTGCGAACGCATTCTGGCGGACTACAAAGACCATGACGGTGATTATAAAGATCATG
	ACATCGATTACAAGGATGACGATGACAAGTAAGGCCGCGACTCGAGAGGGCCC
Nhe1-Intron1-GP70-PSP-	GCTAGCGTGTGTGTTTTTGTTTTTCCCACCCTCTCTCCCCACTACTTGCTCTCACAGTACTCG
GGG-NLuc-3xFLAG-PspOMI	CTGAGGGTGAACAAGAAAAGACCTGATAAAGATTAACCAGAAGAAAACAAGGAGGGAAACAA
	CCGCAGCCTGTAGCAAGCTCTGGAACTCAGGAGTCGCGCGCG
	CGGGGCCGGGGCCGGGGCCGGGGCCGGGGCCGGGGCCGGGG
	GGCCGGGGCAGGGGCCGGGGCCGGGGCCGGGGCCGGGGCCGGGGCCGGGG
	CGGGGCCGGGGCCGGGGCCGGGGCCGGGGCCGGGGCCGGGG
	GGCCGGGGCCGGGGCCGGGGCCGGGGCCGGGGCCGGGGCCGGGG
	CGGGGCCGGGGCCGGGGCCGGGGCCGGGGCCGGGGCCGGGG
	GGCCGGGGCCGGTCGTGGAAGGGTGGGCGCGCCcACCGGTcCTCGAGGTCCTCTTCCAGG
	GACCCGATGGGGTCTTCACACTCGAAGATTTCGTTGGGGACTGGCGACAGACA

	TCGACGGCAAAAAGATCACTGTAACAGGGACCCTGTGGAACGGCAACAAAATTATCGACGAG
	CGCCTGATCAACCCCGACGGCTCCCTGCTGTTCCGAGTAACCATCAACGGAGTGACCGGCT
	GGCGGCTGTGCGAACGCATTCTGGCGGACTACAAAGACCATGACGGTGATTATAAAGATCAT
	GACATCGATTACAAGGATGACGATGACAAGTAAGGCCGCGACTCGAGAGGGCCC
Nhe1-Intron1-GR70-PSP-	GCTAGCGTGTGTGTTTTTGTTTTTCCCACCCTCTCTCCCCACTACTTGCTCTCACAGTACTCG
GGG-NLuc-3xFLAG-PspOMI	CTGAGGGTGAACAAGAAAAGACCTGATAAAGATTAACCAGAAGAAAACAAGGAGGGAAACAA
	CCGCAGCCTGTAGCAAGCTCTGGAACTCAGGAGTCGCGCGCTAGCGGCCGGGGCCGGGGC
	CGGGGCCGGGGCCGGGGCCGGGGCCGGGGCCGGGGCCGGGG
	GGCCGGGGCAGGGGCCGGGGCCGGGGCCGGGGCCGGGGCCGGGGCCGGGG
	GGCCGGGGCCGGGGCCGGGGCCGGGGCCGGGGCCGGGGCCGGGG
	CGGGGCCGGGGCCGGGGCCGGGGCCGGGGCCGGGGCCGGGG
	GGCCGGGGCCGGGGCCGGGGCCGGGGCCGGGGCCGGGGCCGGGG
	000000000000000000000000000000000000000
	GGCCGGGGCCGGTCGTGGAAGGGTGGGCGCGCCCACCGGTccCTCGAGGTCCTCTCCAGG
	GACCCGATGGGGTCTTCACACTCGAAGATTTCGTTGGGGGACTGGCGACAGACA
	GGCGGCTGTGCGAACGCATTCTGGCGGACTACAAAGACCATGACGGTGATTATAAAGATCAT
	GACATCGATTACAAGGATGACGATGACAAGTAAGGCCGCGACTCGAGAGGGCCC