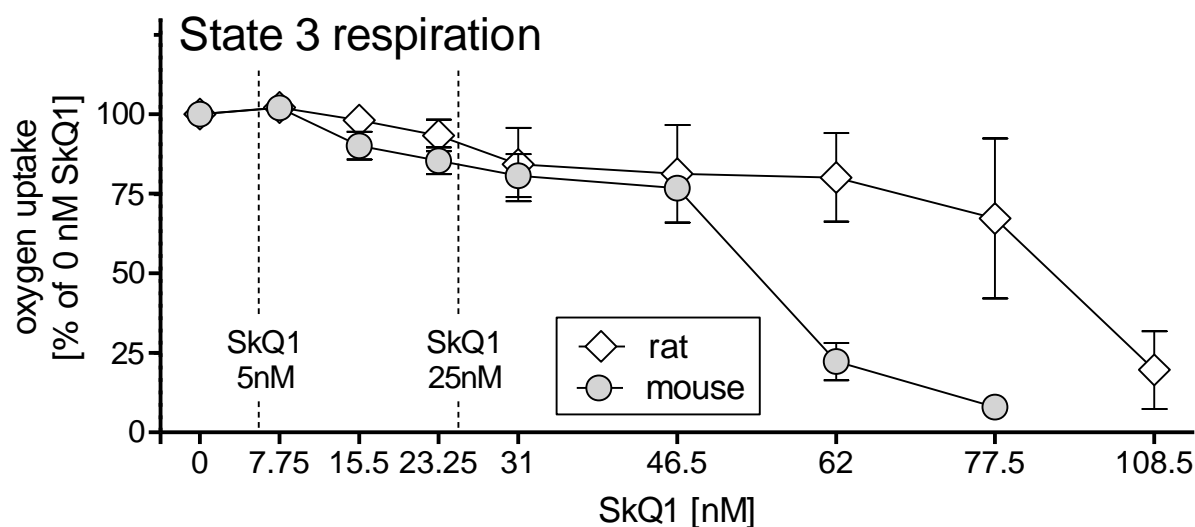


Supplementary Fig. 2. Comparison of body temperature (BT) profiles in CLP experiments performed in two different laboratories. BT profile was used as a surrogate of outcome to optimally standardize the magnitude of CLP severity between the surgery performed in Ludwig Boltzmann Institute (LBI; Vienna, Austria) and Institute of Surgical Research (ISR), University of Szeged (Hungary). Mice underwent three CLP runs at ISR and their BT profiles (at 6h, 12h and 24h) were compared to the BT profile (at 6 and 24h post-CLP) of the CLP mice enrolled in the survival study performed in LBI (solid line/dot). ISR-CLP #3 was designated as the best match and expanded to n=28 for further mitophagy analysis. LBI-CLP n=90-78; ISR-CLP#1 n=28-16; ISR-CLP#2 n=25-13; ISR-CLP#3 n=27-26. Data points shown as mean±SEM.

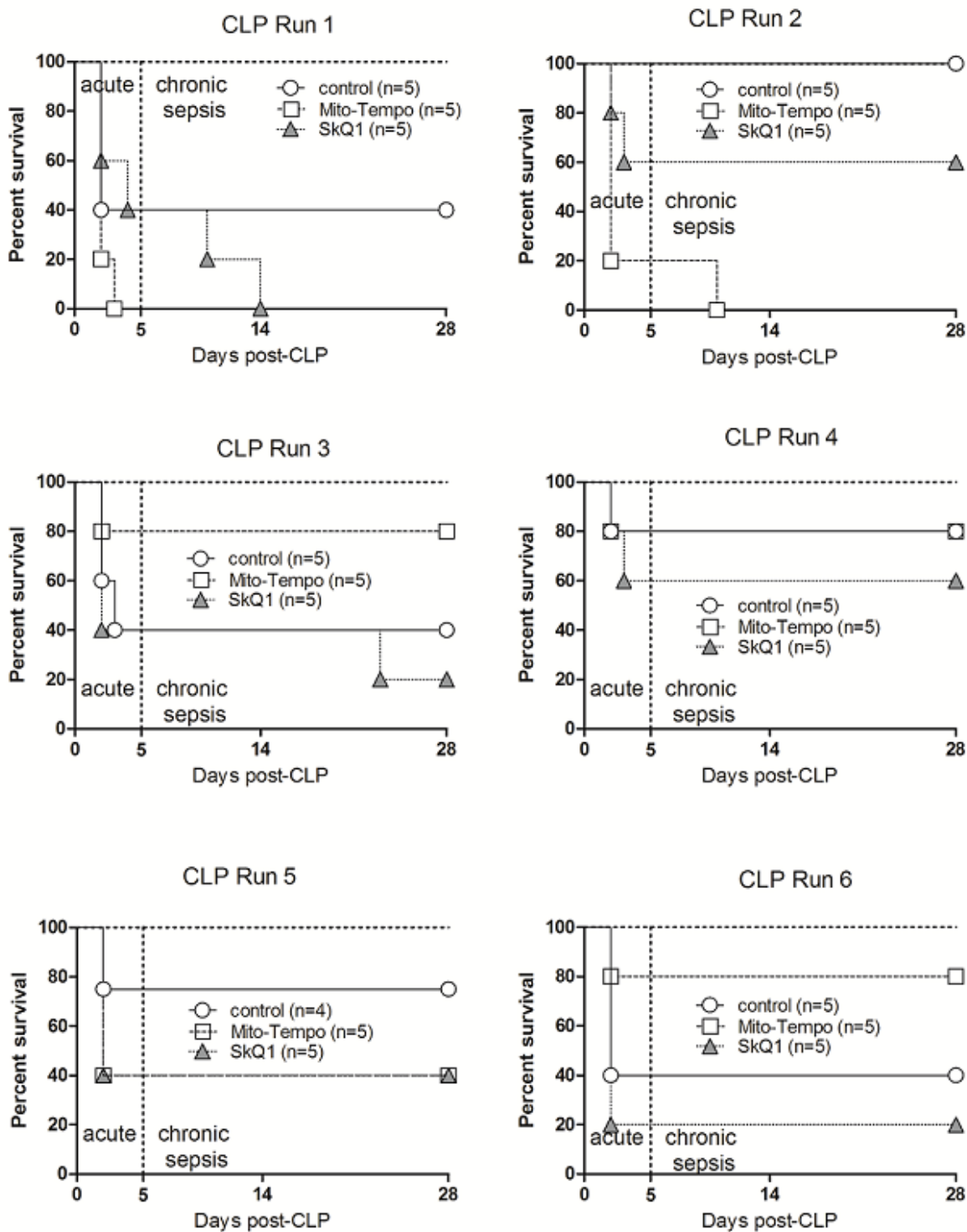


Supplementary Fig. 3. Effect of SkQ1 treatment on the state 3 mitochondrial respiration in the rat and mouse liver homogenates. State 3 respiration was measured in mouse and rat liver homogenates subjected to SkQ1 in the range of concentrations from 0 to 108.5 nM. n=4/each species. Data points shown as mean±SD. Dotted lines indicate either the single (5 nM) or cumulative (25 nM) SkQ1 dose administered to CLP mice in the main survival study (Experiment 1). The rat data serve as species comparison.

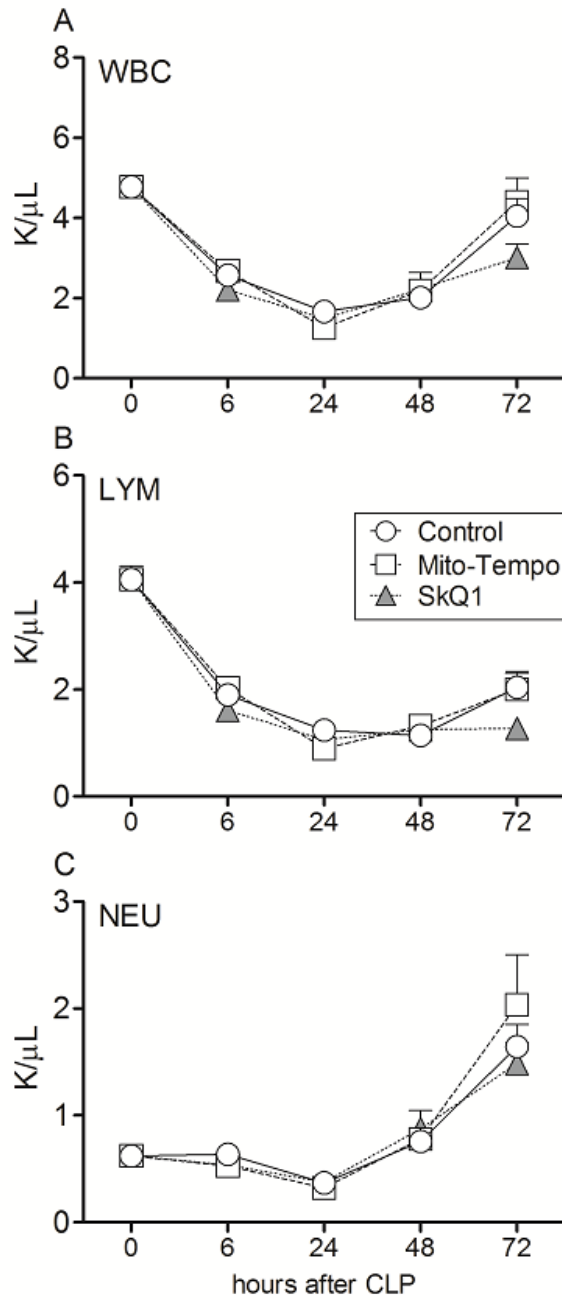
Materials and Methods

Measurement of mitochondrial respiration in liver homogenate

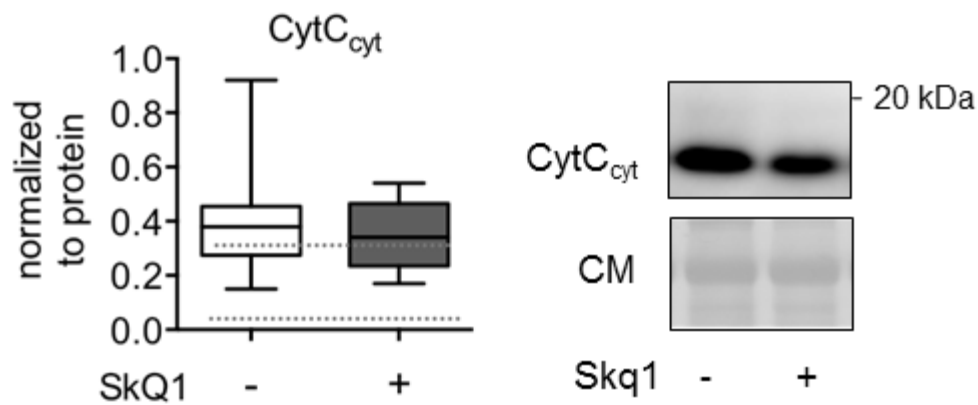
Fifty milligrams of rat/mouse liver were homogenized (RW 1 basic homogenizer, IKA, Wilmington, NC, USA) with ten volumes (1:11 wt/vol) of buffer (0.25M saccharose, 10mM Tris, 0.5 mM EDTA, 5 mg/ml fatty acid-free bovine serum albumin, pH 7.2). For measurement of mitochondrial respiration, 25µL of liver homogenate was incubated in buffer containing 105mM KCl, 5mM KH₂PO₄, 20mM Tris-HCl, 0.5mM EDTA and 5 mg/ml fatty acid-free bovine serum albumin (pH 7.2, 37°C). State 3 respiration was monitored using a high-resolution respirometer (Oxygraph-2k, Oroboros Instruments, Austria) by addition of 10mM succinate, 1 ng/mL rotenone and 1mM ADP. All reagents were obtained from Sigma-Aldrich (Vienna, Austria).



Supplementary Fig. 4. Visualization of outcome for each individual CLP run. CLP was performed in six independent reiterations with 14-15 mice at each repetition (typically 5 mice/each group; the precise n indicated on each panel). Statistical assessment of outcome was performed on the combined data set (Fig. 2).



Supplementary Fig. 5. Trajectory of white blood cells (WBC; A), lymphocyte (LYM; B) and neutrophil (NEU; C) counts for SkQ1, MitoTEMPO and placebo mice (control). For A-C: at BL n=50; at 6h control n=20, SkQ1 n=25, MitoTEMPO n=23; at 24h control n=23, SkQ1 n=23, MitoTEMPO n=22; at 48h control n=18, SkQ1 n=15, MitoTEMPO n=15; at 72h control n=18, SkQ1 n=14, MitoTEMPO n=15. Data points shown as mean +/- SEM.



Supplementary Fig. 6. Assessment of Cytochrome C release in the liver of placebo-treated control vs. SkQ1-treated group at 24h post-CLP. CLP mice received total of three SkQ1/placebo injections before sacrifice at 24h. Data is shown as densitometric analysis of the Western blot from cytosolic. Total number of CLP mice loaded on three different gels: SkQ1 n=16; control (placebo) n=12. Data as (min-to-max) box-and-whiskers plots. Dotted lines indicate upper/lower standard deviation calculated based on eight healthy control mice (no CLP, no treatment) that were analyzed together with the CLP mice. CM: coumassie stained gel.